nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a	Confirmed	
☐	☒	The exact sample size (*n*) for each experimental group/condition, given as a discrete number and unit of measurement
☐	☒	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
☐	☒	The statistical test(s) used AND whether they are one- or two-sided *Only common tests should be described solely by name; describe more complex techniques in the Methods section.*
☒	☐	A description of all covariates tested
☐	☒	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
☐	☒	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
☐	☒	For null hypothesis testing, the test statistic (e.g. *F*, *t*, *r*) with confidence intervals, effect sizes, degrees of freedom and *P* value noted *Give P values as exact values whenever suitable.*
☒	☐	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
☒	☐	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
☒	☐	Estimates of effect sizes (e.g. Cohen's *d*, Pearson's *r*), indicating how they were calculated

Our web collection on statistics for biologists contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection	Image Lab software v6.1.0 (Bio-rad), Quantstudio Real Time PCR software v1.7.2 (Applied Biosystems), Leica Las X v5.02 (Leica),
Data analysis	FlowJo v10 (BD Biosciences), Prism 8.0 (GraphPad Software, Inc.)

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Source data are provided with this paper. TCGA dataset was accessing using the UCSC Xena Browser (http://xenabrowser.net/)

Human research participants

Policy information about studies involving human research participants and Sex and Gender in Research.

Reporting on sex and gender	The informed consent for the study specimens were obtained from all subjects. This study was approved by Clinical Research Ethics Committee of the Chinese University of Hong Kong. All patients were treatment naive CRC patients. The age and gender of the patients have been documented.
Population characteristics	Tumor tissues were collected from patients with pathologically confirmed CRC undergoing surgery at the Prince of Wales Hospital, the Chinese University of Hong Kong. The patients were Chinese with average age at resection was 67.1 years. 116 patients were male and 89 female.
Recruitment	Patients were enrolled in by their oncologist at the Chinese University of Hong Kong. With informed consent from patients/ immediate family of patients, tumor specimens were collected after surgical removal of tumors.
Ethics oversight	This study was approved by Clinical Research Ethics Committee of the Chinese University of Hong Kong.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

☒ Life sciences ☐ Behavioural & social sciences ☐ Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	Sample size for in vitro and animal work were decided based on experience from similar experiments in our laboratory (Wong et al. Nature Communications, 2022 Jul 8;13(1):3971. Wong et al. Gastroenterology, 2020 Dec;159(6):2163-2180.e6.)
Data exclusions	No exclustion of data was performed.
Replication	All in vitro work was repeated in at least 2 independent experiments. Details of experimental replicates are given in the figure legends. All reported attempts at replication were successful.
Randomization	All samples and animals were analyzed and allocated randomly.
Blinding	Pathological evaluation was performed in a blinded manner by experienced pathologists.

Behavioural & social sciences study design

All studies must disclose on these points even when the disclosure is negative.

Study description	*Briefly describe the study type including whether data are quantitative, qualitative, or mixed-methods (e.g. qualitative cross-sectional, quantitative experimental, mixed-methods case study).*
Research sample	*State the research sample (e.g. Harvard university undergraduates, villagers in rural India) and provide relevant demographic information (e.g. age, sex) and indicate whether the sample is representative. Provide a rationale for the study sample chosen. For studies involving existing datasets, please describe the dataset and source.*
Sampling strategy	*Describe the sampling procedure (e.g. random, snowball, stratified, convenience). Describe the statistical methods that were used to predetermine sample size OR if no sample-size calculation was performed, describe how sample sizes were chosen and provide a rationale for why these sample sizes are sufficient. For qualitative data, please indicate whether data saturation was considered, and what criteria were used to decide that no further sampling was needed.*
Data collection	*Provide details about the data collection procedure, including the instruments or devices used to record the data (e.g. pen and paper, computer, eye tracker, video or audio equipment) whether anyone was present besides the participant(s) and the researcher, and whether the researcher was blind to experimental condition and/or the study hypothesis during data collection.*

Timing	*Indicate the start and stop dates of data collection. If there is a gap between collection periods, state the dates for each sample cohort.*
Data exclusions	*If no data were excluded from the analyses, state so OR if data were excluded, provide the exact number of exclusions and the rationale behind them, indicating whether exclusion criteria were pre-established.*
Non-participation	*State how many participants dropped out/declined participation and the reason(s) given OR provide response rate OR state that no participants dropped out/declined participation.*
Randomization	*If participants were not allocated into experimental groups, state so OR describe how participants were allocated to groups, and if allocation was not random, describe how covariates were controlled.*

Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

Study description	*Briefly describe the study. For quantitative data include treatment factors and interactions, design structure (e.g. factorial, nested, hierarchical), nature and number of experimental units and replicates.*
Research sample	*Describe the research sample (e.g. a group of tagged Passer domesticus, all Stenocereus thurberi within Organ Pipe Cactus National Monument), and provide a rationale for the sample choice. When relevant, describe the organism taxa, source, sex, age range and any manipulations. State what population the sample is meant to represent when applicable. For studies involving existing datasets, describe the data and its source.*
Sampling strategy	*Note the sampling procedure. Describe the statistical methods that were used to predetermine sample size OR if no sample-size calculation was performed, describe how sample sizes were chosen and provide a rationale for why these sample sizes are sufficient.*
Data collection	*Describe the data collection procedure, including who recorded the data and how.*
Timing and spatial scale	*Indicate the start and stop dates of data collection, noting the frequency and periodicity of sampling and providing a rationale for these choices. If there is a gap between collection periods, state the dates for each sample cohort. Specify the spatial scale from which the data are taken*
Data exclusions	*If no data were excluded from the analyses, state so OR if data were excluded, describe the exclusions and the rationale behind them, indicating whether exclusion criteria were pre-established.*
Reproducibility	*Describe the measures taken to verify the reproducibility of experimental findings. For each experiment, note whether any attempts to repeat the experiment failed OR state that all attempts to repeat the experiment were successful.*
Randomization	*Describe how samples/organisms/participants were allocated into groups. If allocation was not random, describe how covariates were controlled. If this is not relevant to your study, explain why.*
Blinding	*Describe the extent of blinding used during data acquisition and analysis. If blinding was not possible, describe why OR explain why blinding was not relevant to your study.*

Did the study involve field work? ☐ Yes ☐ No

Field work, collection and transport

Field conditions	*Describe the study conditions for field work, providing relevant parameters (e.g. temperature, rainfall).*
Location	*State the location of the sampling or experiment, providing relevant parameters (e.g. latitude and longitude, elevation, water depth).*
Access & import/export	*Describe the efforts you have made to access habitats and to collect and import/export your samples in a responsible manner and in compliance with local, national and international laws, noting any permits that were obtained (give the name of the issuing authority, the date of issue, and any identifying information).*
Disturbance	*Describe any disturbance caused by the study and how it was minimized.*

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involved in the study
☐	☒ Antibodies
☐	☒ Eukaryotic cell lines
☒	☐ Palaeontology and archaeology
☐	☒ Animals and other organisms
☒	☐ Clinical data
☒	☐ Dual use research of concern

Methods

n/a	Involved in the study
☒	☐ ChIP-seq
☐	☒ Flow cytometry
☒	☐ MRI-based neuroimaging

Antibodies

Antibodies used

CD4 (Abcam, ab183685, EPR19514, 1:2000), CD8 (Abcam, ab217344, EPR21769, 1:2000), S100A8 (Proteintech, 15792-1-AP, 1:800), SLC25A22 (Sigma-Aldrich, HPA014662, 1:200 for IHC and IF; 1:2000 for WB); CD11b (Abcam, ab133357, EPR1344,1:500); Gr-1 (BioLegend, #108448, RB6-8C5, 1:100) ; ETS2 (ThermoFisher, #PA5-28053, Polyclonal, 5μg per reaction for Chip, 1:1000 for WB). Phospho-ETS2 (ThermoFisher, #44-1105G, Polyclonal, 1:700); Rabbit IgG (Santa Cruz Biotechnology, sc-2027, 5μg per reaction for Chip); Src (Abcam, ab231081, GD11, 1:5000); Phospho-Src (Abcam, ab185617, EPR17734, 1:1000); ASNS (Abclonal, A1030, Polyclonal, 1:1000); Lamin A/C (Cell signaling, #4777, 4C11, 1:3000); GAPDH (Cell signaling, #5174, D16H11, 1:2000); β-actin (Cell signaling, #4970, 13E5, 1:2000); Goat anti-Rabbit IgG (H+L) Cross-Adsorbed Secondary Antibody, Alexa Fluor™ 488 (Invitrogen, #A-11008, Polyclonal, 1:1000); Goat anti-Rat IgG (H+L) Cross-Adsorbed Secondary Antibody, Alexa Fluor™ 594 (Invitrogen, # A-11007, Polyclonal, 1:1000); Anti-mouse IgG, HRP-linked Antibody (Cell Signaling Technology, #7076, 1:10000); Anti-rabbit IgG, HRP-linked Antibody (Cell Signaling Technology, #7074, 1:10000);
Human CD45 (Biolegend, #304042, HI30, 1:100); Human CD3 (Biolegend, #317308, OKT3, 1:100); Human CD4 (Biolegend, #357405, A161A1, 1:100); Human CD8 (Biolegend, #344770, SK1, 1:100); Human MHC-II (HLA-DR) (Biolegend, #327007, LN3, 1:100); Human CD11b (Biolegend, #301330, ICRF44, 1:100); Human CD33 (Biolegend, #366618, P67.6, 1:100); Mouse CD45 (Biolegend, 30-F11, 1:100); Mouse CD3 (Biolegend, #100206, 17A2, 1:100); Mouse CD4 (Biolegend, #100509, RM4-5, 1:100); Mouse CD8 (Biolegend, #100738, 53-6.7, 1:100); Mouse IFN-γ (Biolegend,#505806, XMG1.2, 1:100); Mouse TNF-α (Biolegend, #506324, MP6-XT22, 1:100); Mouse Granzyme B (Biolegend, #372212, QA16A02, 1:100); Mouse CD11b (Biolegend, #101228, M1/70, 1:100); Mouse Ly-6G/Ly-6C (Gr-1) (Biolegend, #108406, RB6-8C5, 1:100); Mouse Ly-6G (Biolegend, #127618, 1A8, 1:100); Mouse Ly-6C (Biolegend, #128008, HK1.4, 1:100); Mouse CD274 (B7-H1, PD-L1) (Biolegend, #124319, 10F.9G2, 1:100); Mouse CD11c (Biolegend, #117308, N418, 1:100); Mouse CD206 (Biolegend, #141717, C068C2, 1:100); Mouse F4/80 (Biolegend, #123108, BM8, 1:100);

Validation

All antibodies are obtained from commercial sources, and validation was available from the websites of respective vendors;
CD4 (Abcam, ab183685, EPR19514, 1:2000): https://www.abcam.cn/products/primary-antibodies/cd4-antibody-epr19514-ab183685.html;
CD8 (Abcam, ab217344, EPR21769, 1:2000): https://www.abcam.cn/products/primary-antibodies/cd8-alpha-antibody-epr21769-ab217344.html;
S100A8 (Proteintech, 15792-1-AP, 1:800): https://www.ptglab.com/products/S100A8-Antibody-15792-1-AP.htm;
SLC25A22 (Sigma-Aldrich, HPA014662, 1:200 for IHC and IF; 1:2000 for WB): https://www.sigmaaldrich.com/HK/zh/product/sigma/hpa014662;
CD11b (Abcam, ab133357, EPR1344,1:500): https://www.abcam.cn/products/primary-antibodies/cd11b-antibody-epr1344-ab133357.html;
Gr-1 (BioLegend, #108448, RB6-8C5, 1:100): https://www.biolegend.com/en-us/products/alexa-fluor-594-anti-mouse-ly-6g-ly-6c-gr-1-antibody-9672?GroupID=BLG4876;
ETS2 (ThermoFisher, #PA5-28053, Polyclonal, 5μg per reaction for Chip, 1:1000 for WB): https://www.thermofisher.cn/cn/zh/antibody/product/ETS2-Antibody-Polyclonal/PA5-28053;
Phospho-ETS2 (ThermoFisher, #44-1105G, Polyclonal, 1:700): https://www.thermofisher.cn/cn/zh/antibody/product/Phospho-ETS2-Thr72-Antibody-Polyclonal/44-1105G;
Rabbit IgG (Santa Cruz Biotechnology, sc-2027, 5μg per reaction for Chip): https://datasheets.scbt.com/sc-2027.pdf;
Src (Abcam, ab231081, GD11, 1:5000): https://www.abcam.com/products/primary-antibodies/src-antibody-gd11-ab231081.html;
Phospho-Src (Abcam, ab185617, EPR17734, 1:1000): https://www.abcam.cn/products/primary-antibodies/src-phospho-y419-antibody-epr17734-ab185617.html;
ASNS (Abclonal, A1030, Polyclonal, 1:1000): https://abclonal.com.cn/Datasheet/Antibodies/A1030.pdf;
Lamin A/C (Cell signaling, #4777, 4C11, 1:3000): https://media.cellsignal.com/pdf/4777.pdf;
GAPDH (Cell signaling, #5174, D16H11, 1:2000): https://media.cellsignal.com/pdf/5174.pdf;
β-actin (Cell signaling, #4970, 13E5, 1:2000): https://www.cellsignal.com/products/primary-antibodies/b-actin-13e5-rabbit-mab/4970;
Goat anti-Rabbit IgG (H+L) Cross-Adsorbed Secondary Antibody, Alexa Fluor™ 488 (Invitrogen, #A-11008, Polyclonal, 1:1000): https://www.thermofisher.cn/order/genome-database/dataSheetPdf?producttype=antibody&productsubtype=antibody_secondary&productId=A-11008&version=316;
Goat anti-Rat IgG (H+L) Cross-Adsorbed Secondary Antibody, Alexa Fluor™ 594 (Invitrogen, # A-11007, Polyclonal, 1:1000): https://www.thermofisher.cn/order/genome-database/dataSheetPdf?producttype=antibody&productsubtype=antibody_secondary&productId=A-11007&version=316;
Anti-mouse IgG, HRP-linked Antibody (Cell Signaling Technology, #7076, 1:10000): https://www.cellsignal.com/products/secondary-antibodies/anti-mouse-igg-hrp-linked-antibody/7076;
Anti-rabbit IgG, HRP-linked Antibody (Cell Signaling Technology, #7074, 1:10000): https://www.cellsignal.com/products/secondary-antibodies/anti-rabbit-igg-hrp-linked-antibody/7074;
Human CD45 (Biolegend, #304042, HI30, 1:100): https://www.biolegend.com/en-us/products/brilliant-violet-605-anti-human-cd45-antibody-8521?GroupID=BLG5926;

Human CD3 (Biolegend, #317308, OKT3, 1:100): https://www.biolegend.com/en-us/products/purified-anti-human-cd3-antibody-3642?GroupID=BLG4203;
Human CD4 (Biolegend, #357405, A161A1, 1:100): https://www.biolegend.com/en-us/products/fitc-anti-human-cd4-antibody-8738?GroupID=BLG11451;
Human CD8 (Biolegend, #344770, SK1, 1:100): https://www.biolegend.com/en-us/products/pe-cyanine5-anti-human-cd8-antibody-21385?GroupID=BLG10167;
Human MHC-II (HLA-DR) (Biolegend, #327007, LN3, 1:100): https://www.biolegend.com/en-us/products/pe-anti-human-hla-dr-antibody-4166?GroupID=BLG10409;
Human CD11b (Biolegend, #301330, ICRF44, 1:100): https://www.biolegend.com/en-us/products/apc-cyanine7-anti-human-cd11b-antibody-9611?GroupID=BLG9916;
Human CD33 (Biolegend, #366618, P67.6, 1:100): https://www.biolegend.com/en-us/products/apc-anti-human-cd33-antibody-877?GroupID=BLG10601;
Mouse CD45 (Biolegend, #103140, 30-F11, 1:100): https://www.biolegend.com/en-us/products/brilliant-violet-605-anti-mouse-cd45-antibody-8721?GroupID=BLG6831;
Mouse CD3 (Biolegend, #100206, 17A2, 1:100): https://www.biolegend.com/en-us/products/pe-anti-mouse-cd3-antibody-47?GroupID=BLG242;
Mouse CD4 (Biolegend, #100509, RM4-5, 1:100): https://www.biolegend.com/en-us/products/fitc-anti-mouse-cd4-antibody-480;
Mouse CD8 (Biolegend, #100738, 53-6.7, 1:100): https://www.biolegend.com/en-us/products/brilliant-violet-421-anti-mouse-cd8a-antibody-7138;
Mouse IFN-γ (Biolegend, #505806, XMG1.2, 1:100): https://www.biolegend.com/en-us/products/fitc-anti-mouse-ifn-gamma-antibody-995;
Mouse TNF-α (Biolegend, #506324, MP6-XT22, 1:100): https://www.biolegend.com/en-us/products/pe-cyanine7-anti-mouse-tnf-alpha-antibody-5866;
Mouse Granzyme B (Biolegend, #372212, QA16A02, 1:100): https://www.biolegend.com/en-us/products/percp-cyanine5-5-anti-humanmouse-granzyme-b-recombinant-antibody-15597;
Mouse CD11b (Biolegend, #101228, M1/70, 1:100): https://www.biolegend.com/en-us/products/percp-cyanine5-5-anti-mouse-human-cd11b-antibody-4257;
Mouse Ly-6G/Ly-6C (Gr-1) (Biolegend, #108406, RB6-8C5, 1:100): https://www.biolegend.com/en-us/products/fitc-anti-mouse-ly-6g-ly-6c-gr-1-antibody-458;
Mouse Ly-6G (Biolegend, #127618, 1A8, 1:100): https://www.biolegend.com/en-us/products/pe-cyanine7-anti-mouse-ly-6g-antibody-6139;
Mouse Ly-6C (Biolegend, #128008, HK1.4, 1:100): https://www.biolegend.com/en-us/products/pe-anti-mouse-ly-6c-antibody-4904;
Mouse CD274 (B7-H1, PD-L1) (Biolegend, #124319, 10F.9G2, 1:100): https://www.biolegend.com/en-us/products/brilliant-violet-711-anti-mouse-cd274-b7-h1-pd-l1-antibody-9808;
Mouse CD11c (Biolegend, #117308, N418, 1:100): https://www.biolegend.com/en-us/products/pe-anti-mouse-cd11c-antibody-1816;
Mouse CD206 (Biolegend, #141717, C068C2, 1:100): https://www.biolegend.com/en-us/products/brilliant-violet-421-anti-mouse-cd206-mmr-antibody-8638;
Mouse F4/80 (Biolegend, #123108, BM8, 1:100): https://www.biolegend.com/en-us/products/fitc-anti-mouse-f4-80-antibody-4067;

Eukaryotic cell lines

Policy information about cell lines and Sex and Gender in Research

Cell line source(s)	293T, CRC cells line (DLD1, CT26 and MC38) were obtained from Tthe American Type Culture Collection (Rockville, MD). Colo26 cells were provided by Celia Dong from Chinese University of Hong Kong.
Authentication	All commercial cell lines (293T, DLD1, CT26 and MC38) were authenticated by STR Profiling. Non-commercial cell lines (Colo 26) were not authenticated.
Mycoplasma contamination	All commercial cell lines (293T, DLD1, CT26 and MC38) were negative for mycoplasma. Non-commercial cell lines (Colo 26) were not tested.
Commonly misidentified lines (See ICLAC register)	No commonly misidentified cell lines were used in the study.

Palaeontology and Archaeology

Specimen provenance	*Provide provenance information for specimens and describe permits that were obtained for the work (including the name of the issuing authority, the date of issue, and any identifying information). Permits should encompass collection and, where applicable, export.*
Specimen deposition	*Indicate where the specimens have been deposited to permit free access by other researchers.*
Dating methods	*If new dates are provided, describe how they were obtained (e.g. collection, storage, sample pretreatment and measurement), where they were obtained (i.e. lab name), the calibration program and the protocol for quality assurance OR state that no new dates are provided.*

☐ Tick this box to confirm that the raw and calibrated dates are available in the paper or in Supplementary Information.

Ethics oversight	*Identify the organization(s) that approved or provided guidance on the study protocol, OR state that no ethical approval or guidance was required and explain why not.*

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Animals and other research organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research, and Sex and Gender in Research

Laboratory animals	Male C57BL/6J mice (6-8 weeks old), BALB/c mice (6-8 weeks old) and NOD-SCID -γ (NSG) (4 weeks old) mice were obtained from Chinese University of Hong Kong mouse repository. SLC25A22fl/+ mice, Apcmin/+KrasG12D and Apcmin/+KrasG12DSlc25a22-/- mice were used between 3 to 7 weeks, which were purchased commercially from Nanjing Biomedical Research Institute, Nanjing University. All mice were maintained under specific pathogen-free conditions at the animal facility of CUHK. These mice were maintained in 12 hour light/dark cycle, and the housing temperature and humidity were at 23 degrees and 45%, respectively.
Wild animals	Not involved.
Reporting on sex	All mice used in the experiments were male.
Field-collected samples	Not involved.
Ethics oversight	All animal experiments were approved by the Animal Experimentation Ethics Committees at CUHK.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Clinical data

Policy information about clinical studies

All manuscripts should comply with the ICMJEguidelines for publication of clinical research and a completedCONSORT checklist must be included with all submissions.

Clinical trial registration	*Provide the trial registration number from ClinicalTrials.gov or an equivalent agency.*
Study protocol	*Note where the full trial protocol can be accessed OR if not available, explain why.*
Data collection	*Describe the settings and locales of data collection, noting the time periods of recruitment and data collection.*
Outcomes	*Describe how you pre-defined primary and secondary outcome measures and how you assessed these measures.*

Dual use research of concern

Policy information about dual use research of concern

Hazards

Could the accidental, deliberate or reckless misuse of agents or technologies generated in the work, or the application of information presented in the manuscript, pose a threat to:

No	Yes	
☐	☐	Public health
☐	☐	National security
☐	☐	Crops and/or livestock
☐	☐	Ecosystems
☐	☐	Any other significant area

Experiments of concern

Does the work involve any of these experiments of concern:

No	Yes	
☐	☐	Demonstrate how to render a vaccine ineffective
☐	☐	Confer resistance to therapeutically useful antibiotics or antiviral agents
☐	☐	Enhance the virulence of a pathogen or render a nonpathogen virulent
☐	☐	Increase transmissibility of a pathogen
☐	☐	Alter the host range of a pathogen
☐	☐	Enable evasion of diagnostic/detection modalities
☐	☐	Enable the weaponization of a biological agent or toxin
☐	☐	Any other potentially harmful combination of experiments and agents

ChIP-seq

Data deposition

☐ Confirm that both raw and final processed data have been deposited in a public database such as GEO.

☐ Confirm that you have deposited or provided access to graph files (e.g. BED files) for the called peaks.

Data access links *May remain private before publication.*	*For "Initial submission" or "Revised version" documents, provide reviewer access links. For your "Final submission" document, provide a link to the deposited data.*
Files in database submission	*Provide a list of all files available in the database submission.*
Genome browser session (e.g. UCSC)	*Provide a link to an anonymized genome browser session for "Initial submission" and "Revised version" documents only, to enable peer review. Write "no longer applicable" for "Final submission" documents.*

Methodology

Replicates	*Describe the experimental replicates, specifying number, type and replicate agreement.*
Sequencing depth	*Describe the sequencing depth for each experiment, providing the total number of reads, uniquely mapped reads, length of reads and whether they were paired- or single-end.*
Antibodies	*Describe the antibodies used for the ChIP-seq experiments; as applicable, provide supplier name, catalog number, clone name, and lot number.*
Peak calling parameters	*Specify the command line program and parameters used for read mapping and peak calling, including the ChIP, control and index files used.*
Data quality	*Describe the methods used to ensure data quality in full detail, including how many peaks are at FDR 5% and above 5-fold enrichment.*
Software	*Describe the software used to collect and analyze the ChIP-seq data. For custom code that has been deposited into a community repository, provide accession details.*

Flow Cytometry

Plots

Confirm that:

☒ The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).

☒ The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).

☒ All plots are contour plots with outliers or pseudocolor plots.

☒ A numerical value for number of cells or percentage (with statistics) is provided.

Methodology

Sample preparation	Tumor samples were dissected and digested in Hank's balanced salt solution with 0.5 mg/mL collagenase D (Roche) and 0.25mg/L DNase I (Roche) for 30 min at 37 °C. Sample was filtered by 40 µm cell strainer to obtain single cell.

Instrument	BD FACS Celesta
Software	FlowJo v10
Cell population abundance	Abundance and purity of cell population determined by FlowJo V10 software with identical gating strategy across each set of samples.
Gating strategy	FSC-A/SSC-A was used for gating immune cells. FSC-W/FSC-H was used for gating singlets. The specific gating strategies are presented in Supplementary Figure.

☒ Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.

Magnetic resonance imaging

Experimental design

Design type	*Indicate task or resting state; event-related or block design.*
Design specifications	*Specify the number of blocks, trials or experimental units per session and/or subject, and specify the length of each trial or block (if trials are blocked) and interval between trials.*
Behavioral performance measures	*State number and/or type of variables recorded (e.g. correct button press, response time) and what statistics were used to establish that the subjects were performing the task as expected (e.g. mean, range, and/or standard deviation across subjects).*

Acquisition

Imaging type(s)	*Specify: functional, structural, diffusion, perfusion.*
Field strength	*Specify in Tesla*
Sequence & imaging parameters	*Specify the pulse sequence type (gradient echo, spin echo, etc.), imaging type (EPI, spiral, etc.), field of view, matrix size, slice thickness, orientation and TE/TR/flip angle.*
Area of acquisition	*State whether a whole brain scan was used OR define the area of acquisition, describing how the region was determined.*

Diffusion MRI ☐ Used ☐ Not used

Preprocessing

Preprocessing software	*Provide detail on software version and revision number and on specific parameters (model/functions, brain extraction, segmentation, smoothing kernel size, etc.).*
Normalization	*If data were normalized/standardized, describe the approach(es): specify linear or non-linear and define image types used for transformation OR indicate that data were not normalized and explain rationale for lack of normalization.*
Normalization template	*Describe the template used for normalization/transformation, specifying subject space or group standardized space (e.g. original Talairach, MNI305, ICBM152) OR indicate that the data were not normalized.*
Noise and artifact removal	*Describe your procedure(s) for artifact and structured noise removal, specifying motion parameters, tissue signals and physiological signals (heart rate, respiration).*
Volume censoring	*Define your software and/or method and criteria for volume censoring, and state the extent of such censoring.*

Statistical modeling & inference

Model type and settings	*Specify type (mass univariate, multivariate, RSA, predictive, etc.) and describe essential details of the model at the first and second levels (e.g. fixed, random or mixed effects; drift or auto-correlation).*
Effect(s) tested	*Define precise effect in terms of the task or stimulus conditions instead of psychological concepts and indicate whether ANOVA or factorial designs were used.*

Specify type of analysis: ☐ Whole brain ☐ ROI-based ☐ Both

Statistic type for inference (See Eklund et al. 2016)	*Specify voxel-wise or cluster-wise and report all relevant parameters for cluster-wise methods.*
Correction	*Describe the type of correction and how it is obtained for multiple comparisons (e.g. FWE, FDR, permutation or Monte Carlo).*

Models & analysis

n/a	Involved in the study
☐	☐ Functional and/or effective connectivity
☐	☐ Graph analysis
☐	☐ Multivariate modeling or predictive analysis

Functional and/or effective connectivity	*Report the measures of dependence used and the model details (e.g. Pearson correlation, partial correlation, mutual information).*
Graph analysis	*Report the dependent variable and connectivity measure, specifying weighted graph or binarized graph, subject- or group-level, and the global and/or node summaries used (e.g. clustering coefficient, efficiency, etc.).*
Multivariate modeling and predictive analysis	*Specify independent variables, features extraction and dimension reduction, model, training and evaluation metrics.*