

Supplementary information

Caveolin-mediated cytosolic delivery of spike nanoparticle enhances antitumor immunity of neoantigen vaccine for hepatocellular carcinoma

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Table S1. Encapsulation efficiency and loading capacity of NeoAgs and ODN-1826 in V-scVLPs and SPS, respectively.

scVLPs	NeoAgs	ODN-1826
Entrapment efficiency	91.8%	76.6%
Loading efficiency	31.46%	4.57%

SPS	NeoAgs	ODN-1826
Entrapment efficiency	63.1%	38.8%
Loading efficiency	23.98%	2.37%

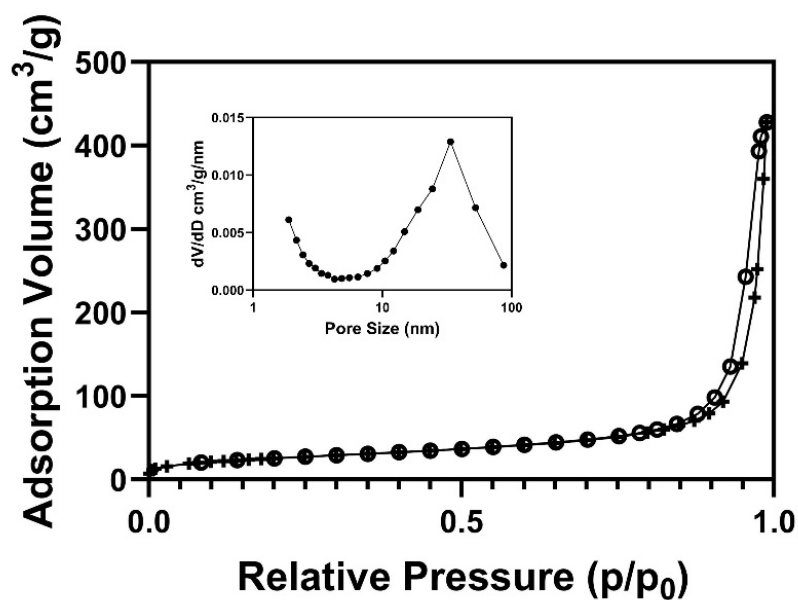


Figure. S1. N₂ sorption isotherm for scVLPs and DFT pore size distribution for scVLPs.

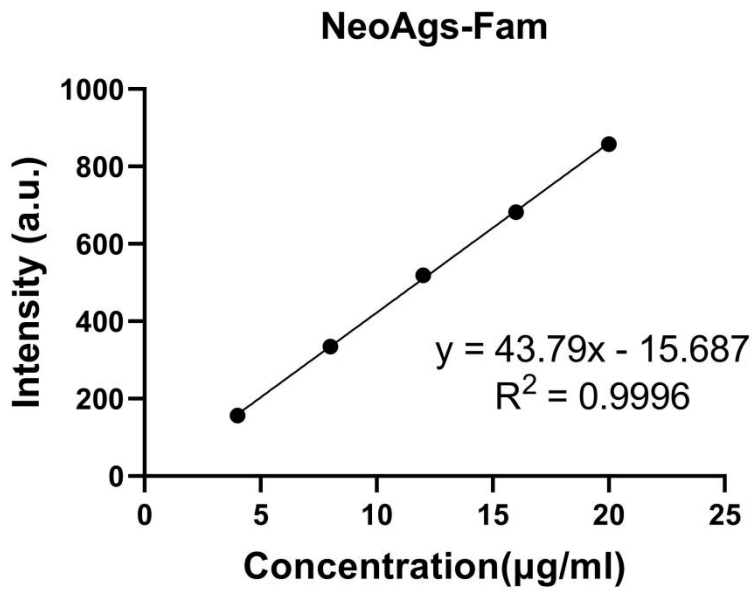


Figure. S2. The concentration of NeoAgs from scVLPs (^{FAM}-NeoAgs) was quantified according to the linear fit ($Y = 43.79x - 15.687$, $R^2 = 0.9996$) of fluorescence intensity at 520 nm, the correlation curve was prepared from 4.0~20.0 µg/mL.

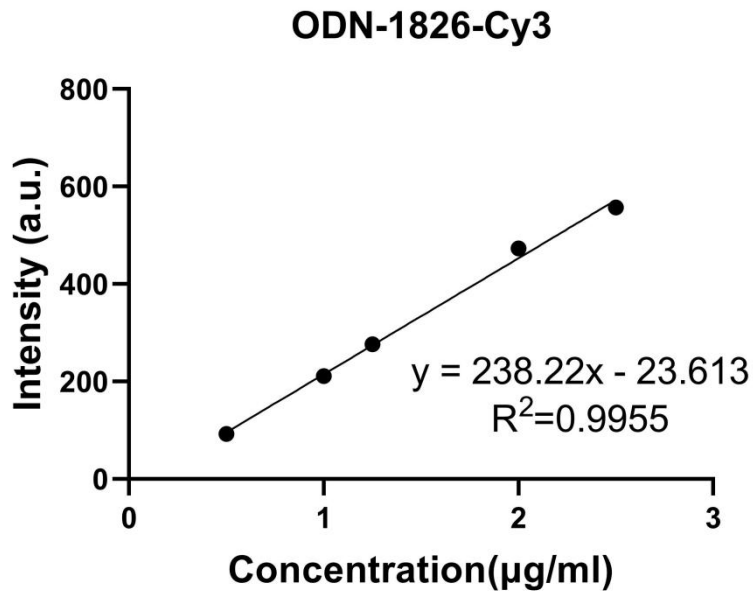


Figure. S3. The concentration of ODN-1826 from scVLPs was quantified according to the linear fit ($Y = 238.22x - 23.613$, $R^2 = 0.9955$) of fluorescence intensity at 570 nm, the correlation curve was prepared from 0.5~2.5 µg/mL.

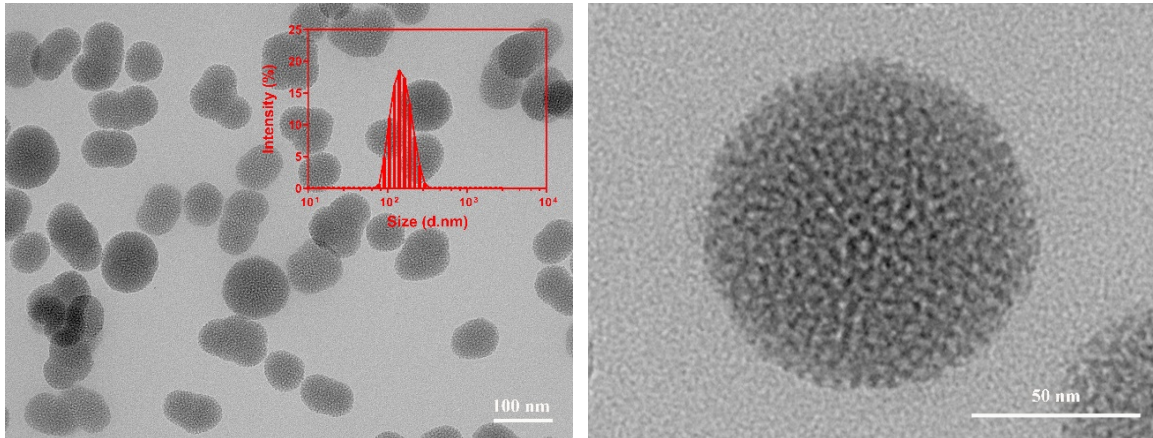


Figure. S4. TEM image of SPS.

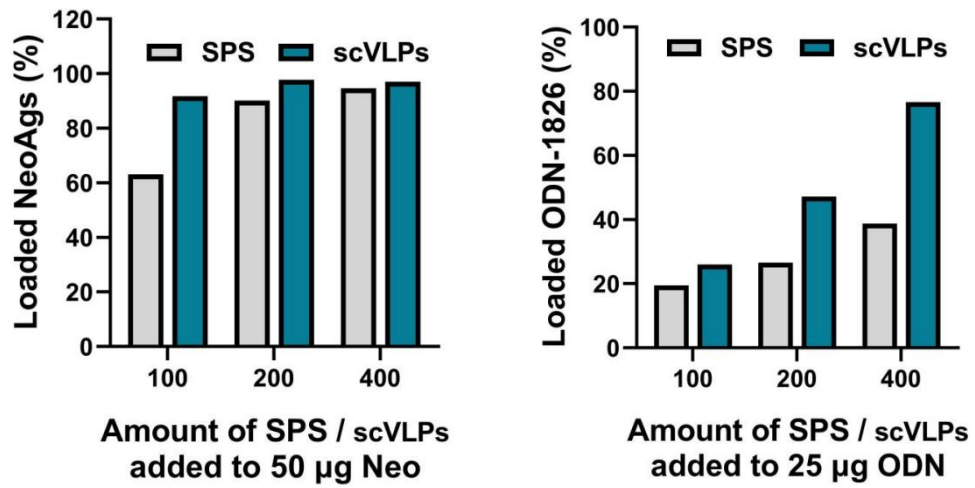


Figure. S5. The loading percentage of NeoAgs and ODN-1826 in SPS or scVLPs, respectively.

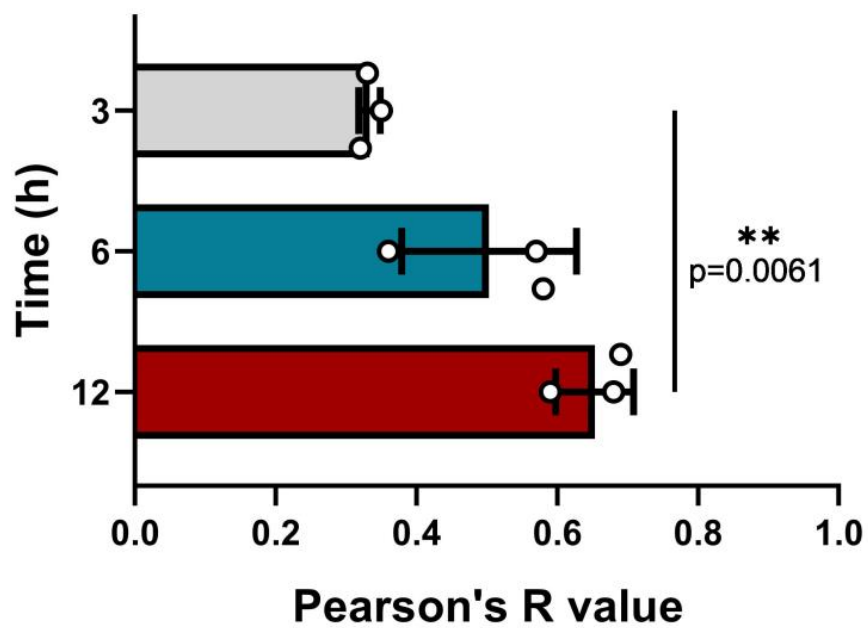
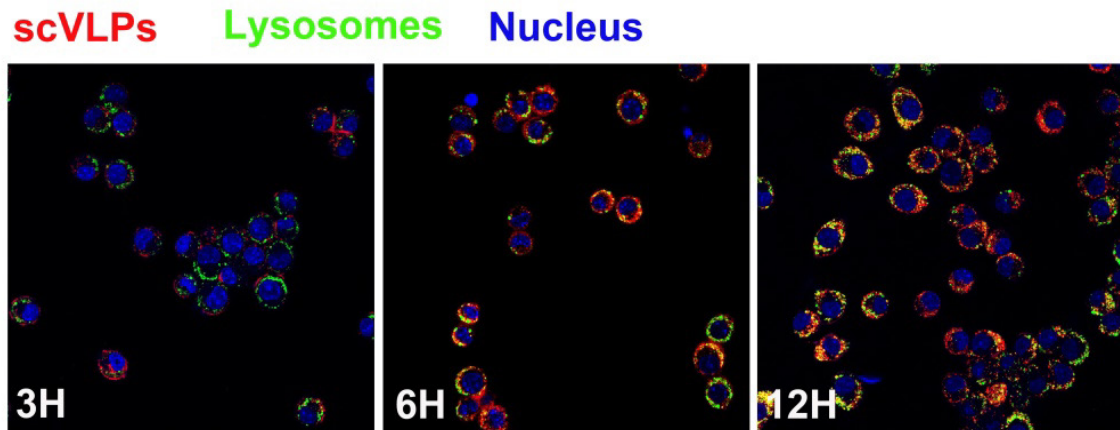


Figure. S6. PCCs of Cy5 labeled scVLPs with lysosomes at different incubation times, (n=3). The statistical analysis was performed with ANOVA analysis, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$, (n = 3). Data are presented as mean \pm SD.

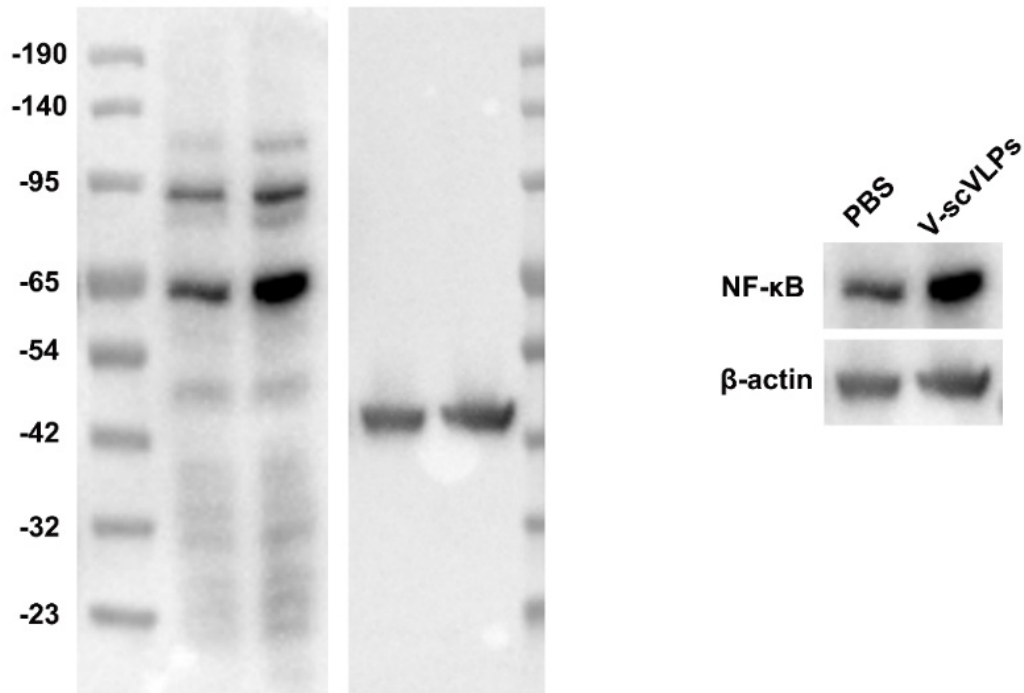


Figure. S7. Western blot analysis of NF-κB in BMDCs after co-incubation with V-scVLPs for 48 h, and PBS treated cell were used as control.

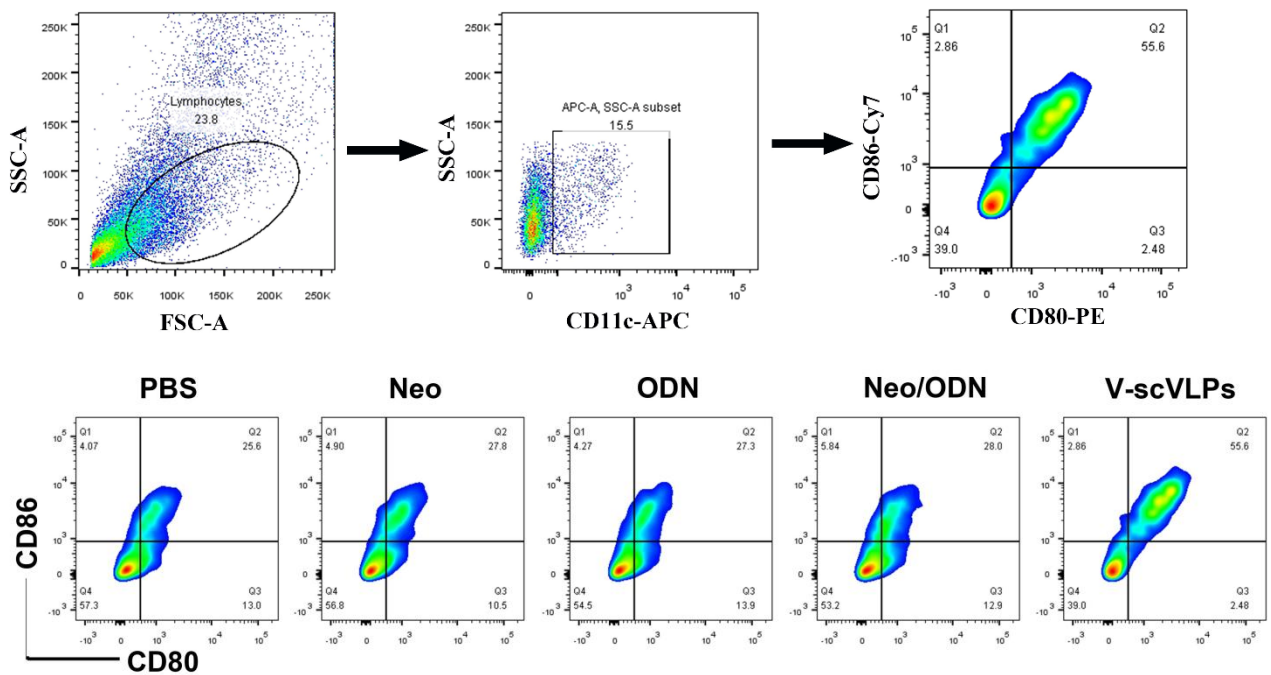


Figure. S8. Identify the maturation of BMDCs after receiving different treatment through FACS with staining anti-CD11c-APC, anti-CD80-PE, and anti-CD86-PE-Cy7 antibodies.

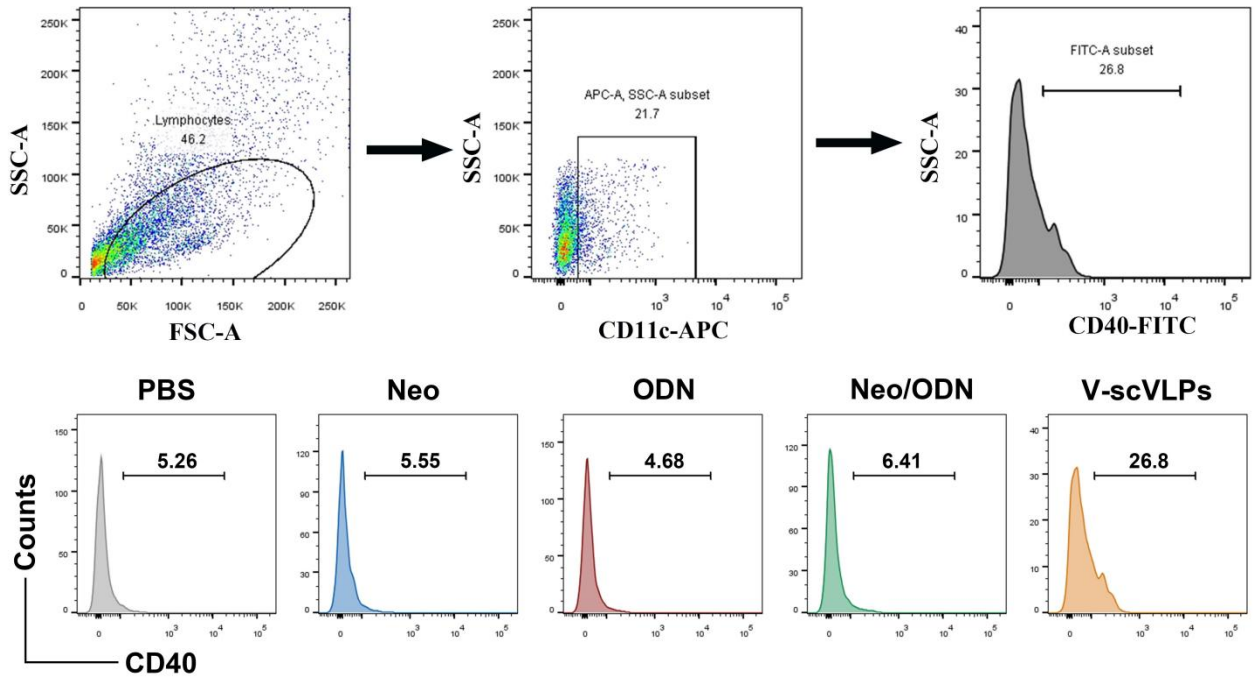


Figure. S9. Identify the maturation of BMDCs after receiving different treatment through FACS with staining anti-CD11c-APC, anti-CD40-FITC antibodies.

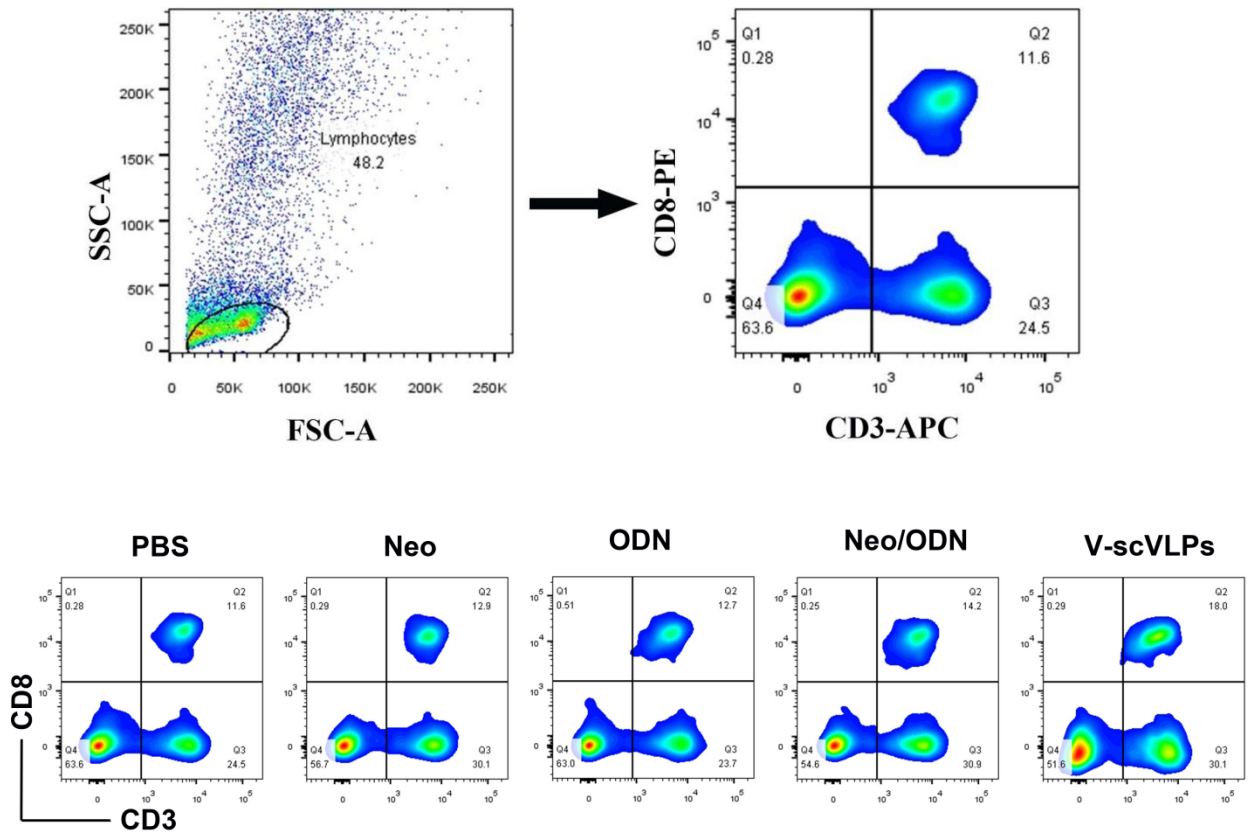


Figure. S10. Identify the population of T cells after receiving different treatment through FACS with staining anti-CD3-APC, anti-CD8-PE antibodies.

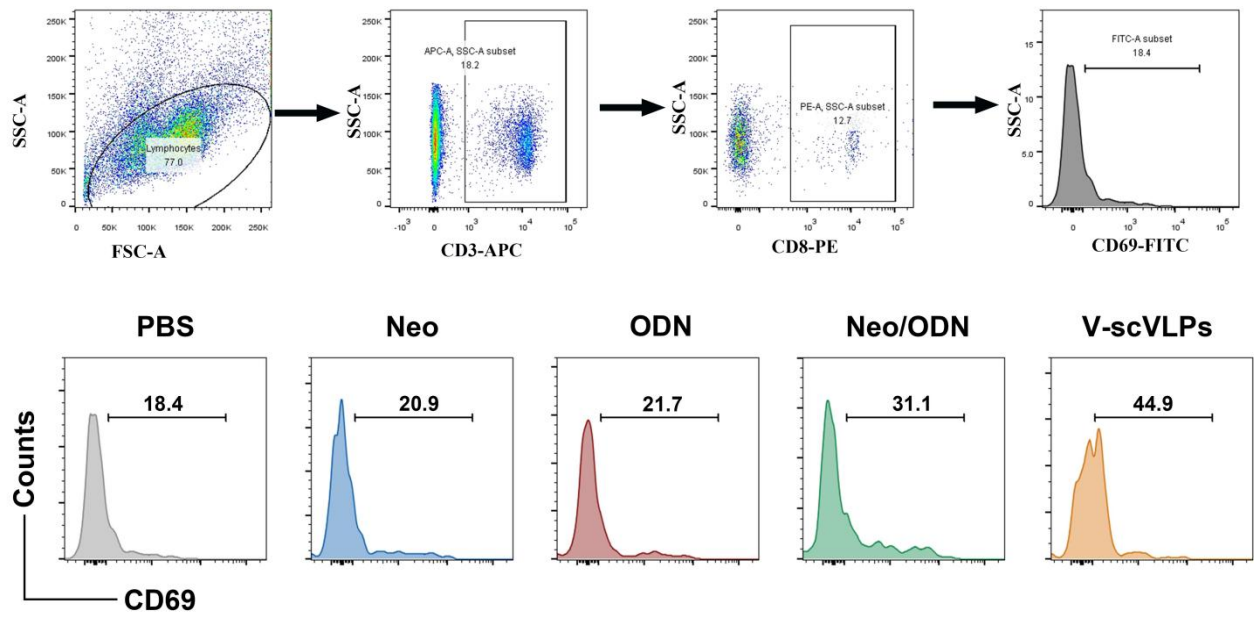


Figure. S11. Identify the activation of T cells after receiving different treatment through FACS with staining anti-CD3-APC, anti-CD8-PE and anti-CD69-FITC antibodies.

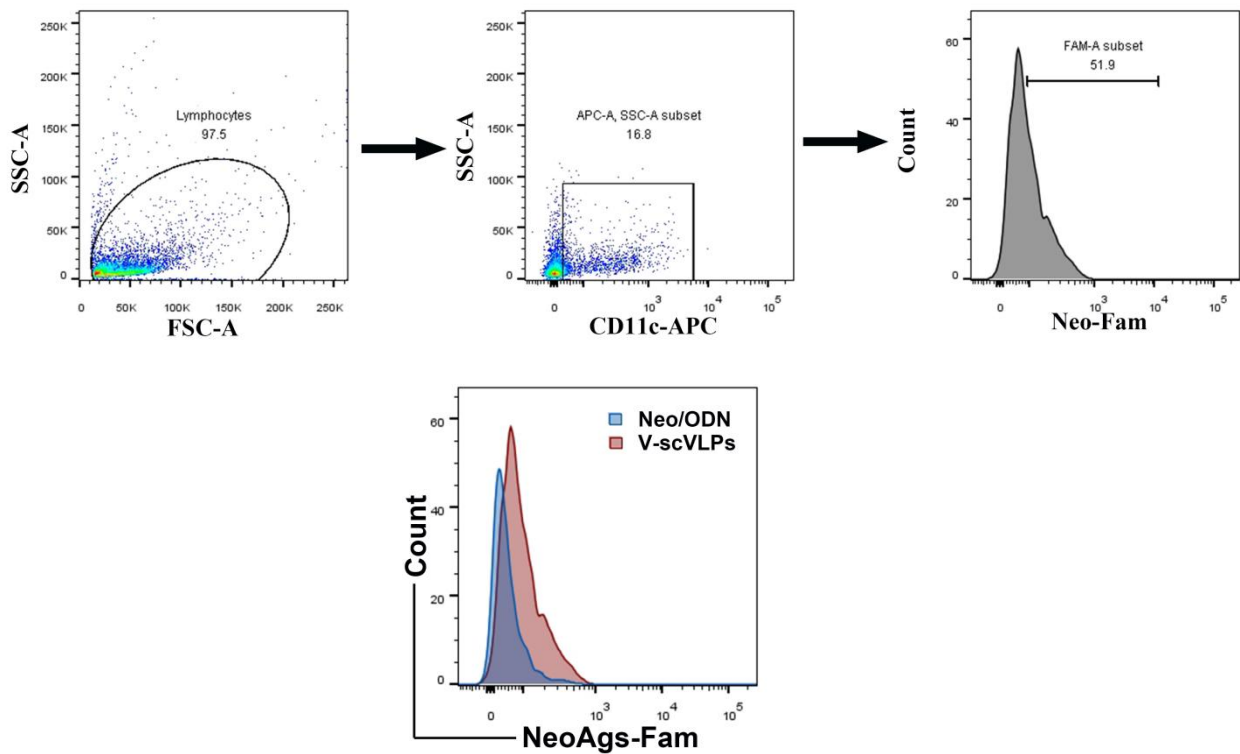


Figure. S12. Representative fluorescence intensity of ^{FAM}NeoAg_s positive cells gated on DCs in lymph node after 12h of Neo/ODN and scVLPs injection.

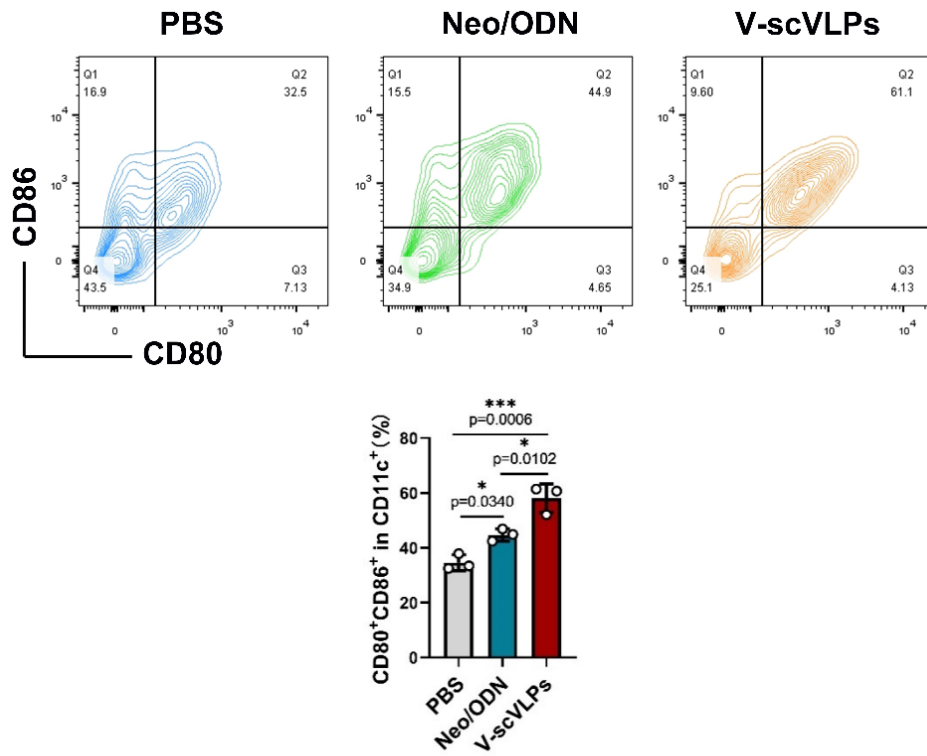
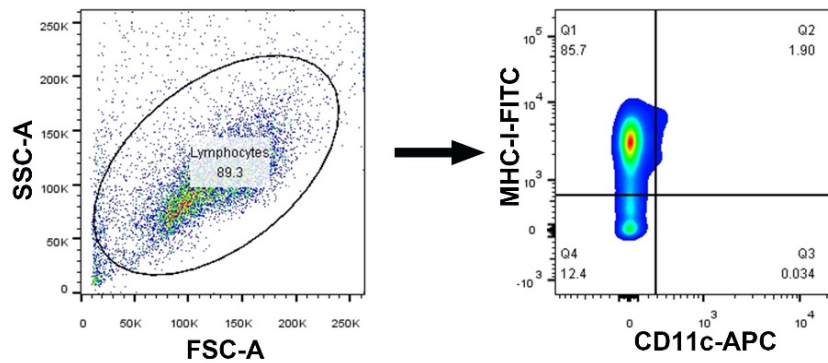


Figure. S13. Identify the maturation of DCs in LNs after receiving different treatment through FACS with staining anti-CD11c-APC, and anti-80-PE and anti-86-PECy7 antibodies.



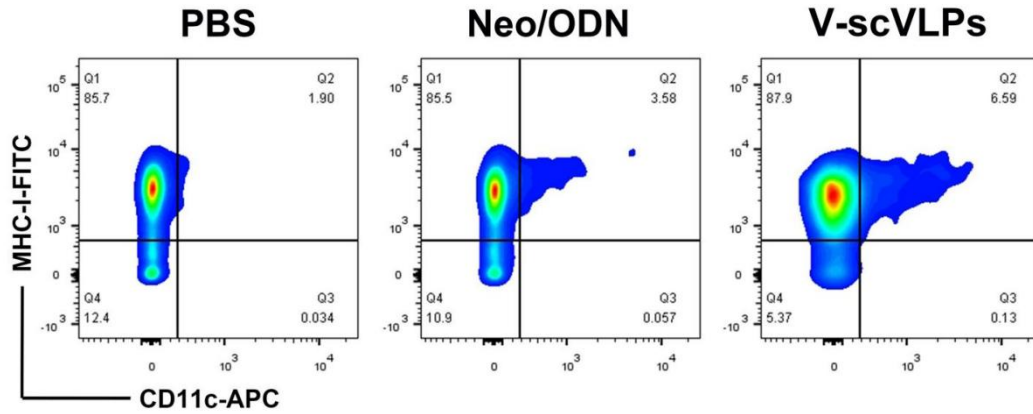


Figure. S14. Identify the maturation of DCs in LNs after receiving different treatment through FACS with staining anti-CD11c-APC, and anti-MHC-I-PE antibodies.

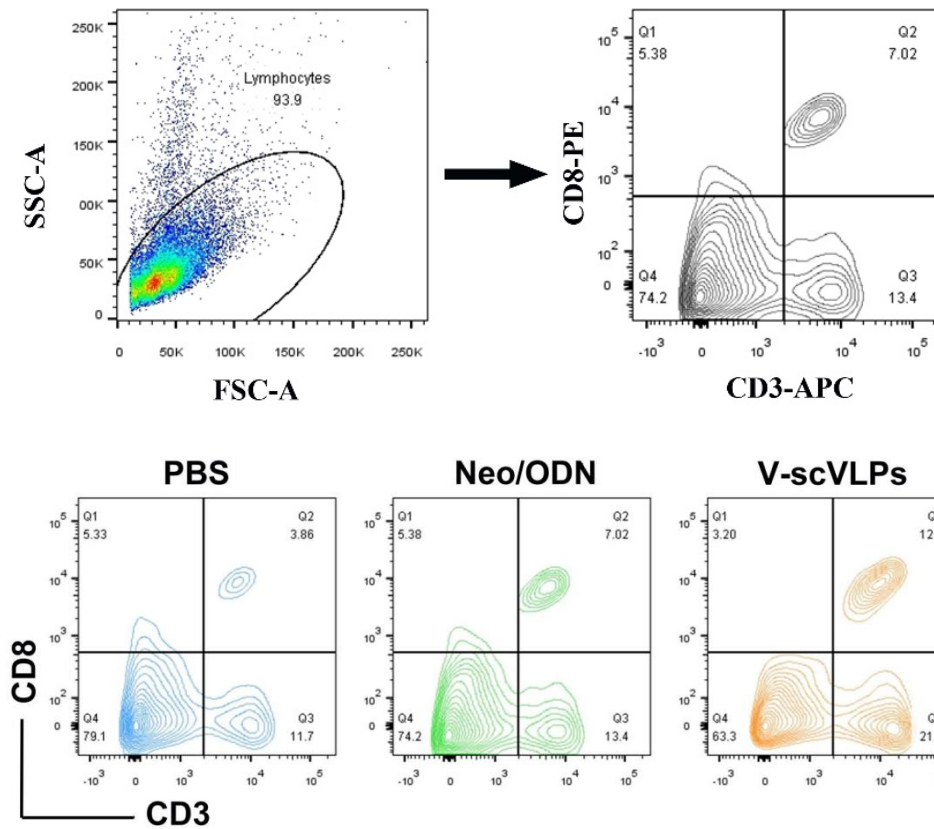


Figure. S15. Identify the CD8⁺T cells in spleen after receiving different treatment through FACS with staining anti-CD3-APC and anti-CD8-PE antibodies.

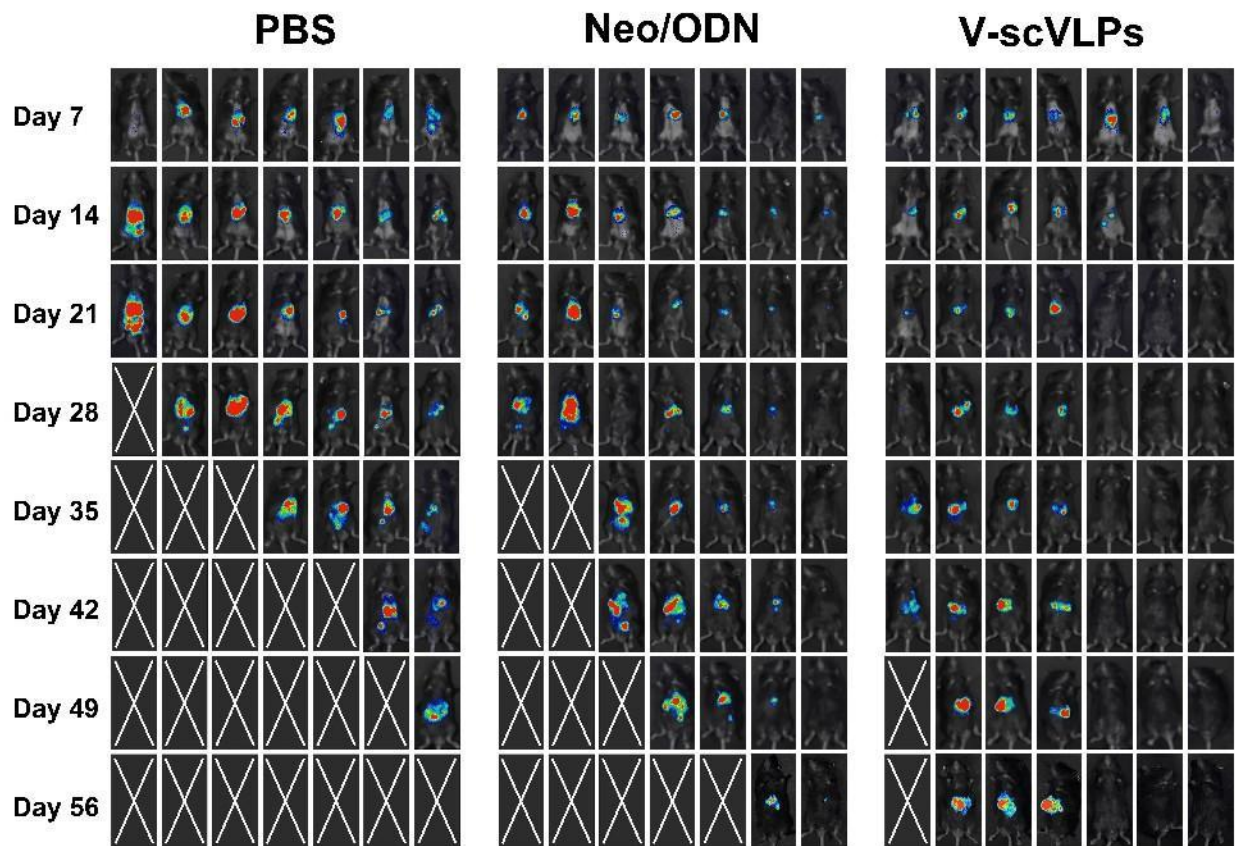


Figure. S16. Bioluminescence imaging of mice at the 7, 14, 21, 28, 35, 42, 49 and 56 days before and after receiving different treatments as indicated, n = 7.

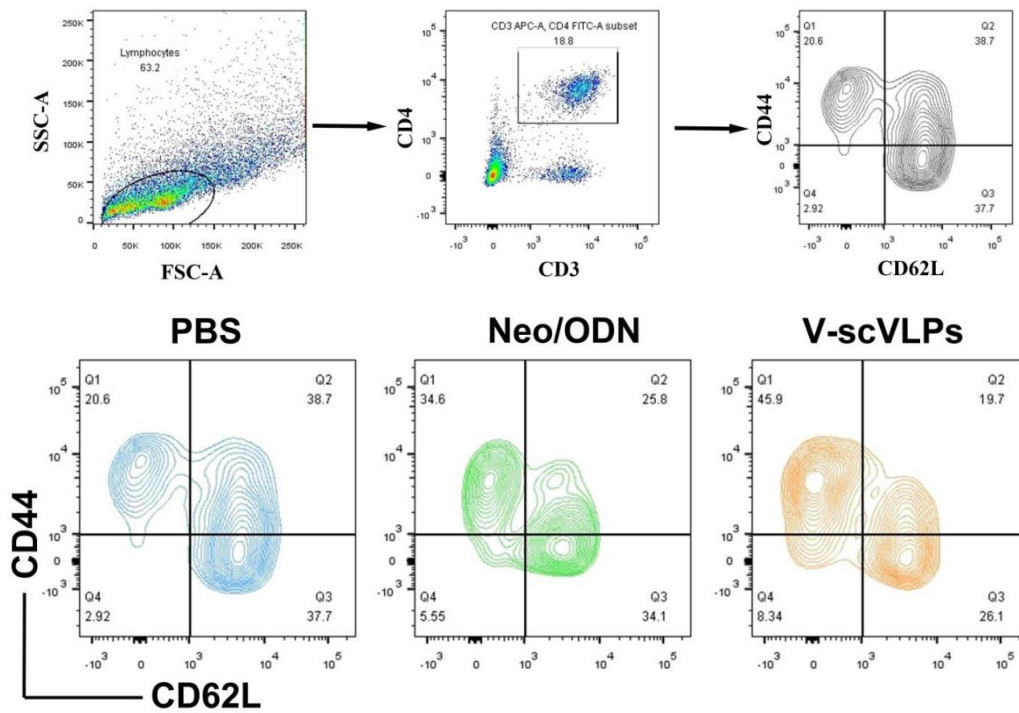


Figure. S17. Identify the CD3⁺CD4⁺CD44⁺CD62L⁺T cells in the spleen after receiving different treatments through FACS with staining anti-CD3-APC, anti-CD4-FITC antibodies, anti-CD44-PE-Cy7 and anti-CD62-PerCP-Cy5.5 antibodies.

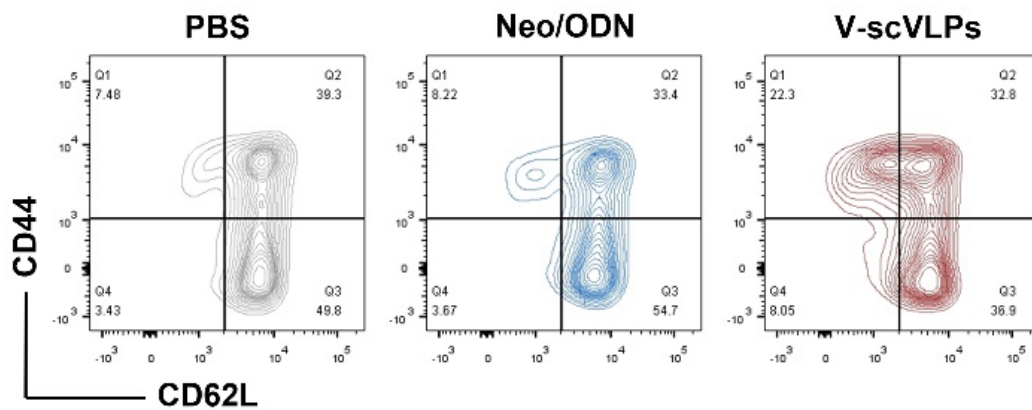


Figure. S18. Identify the CD3⁺CD8⁺CD44⁺CD62L⁺T cells in the spleen after receiving different treatments through FACS with staining anti-CD3-APC, anti-CD8-PE antibodies, anti-CD44-PE-Cy7 and anti-CD62-PerCP-Cy5.5 antibodies.

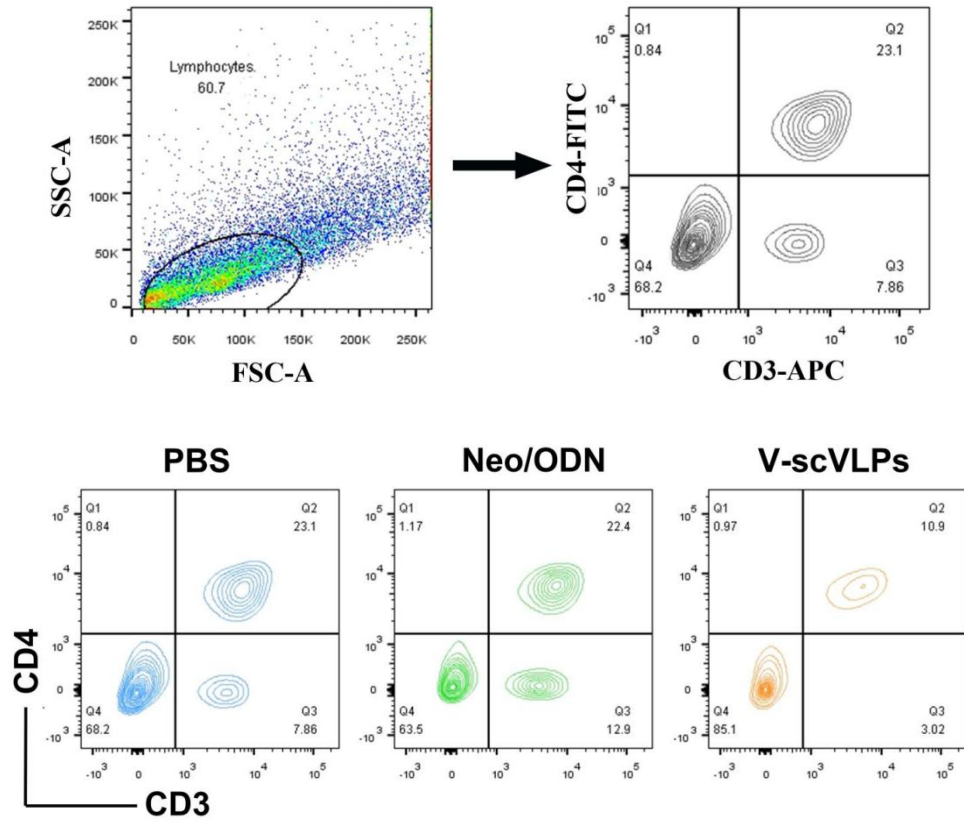


Figure. S19. Identify the CD3⁺CD4⁺T cells in the spleen after receiving different treatments through FACS with staining anti-CD3-APC and anti-CD4-FITC antibodies.

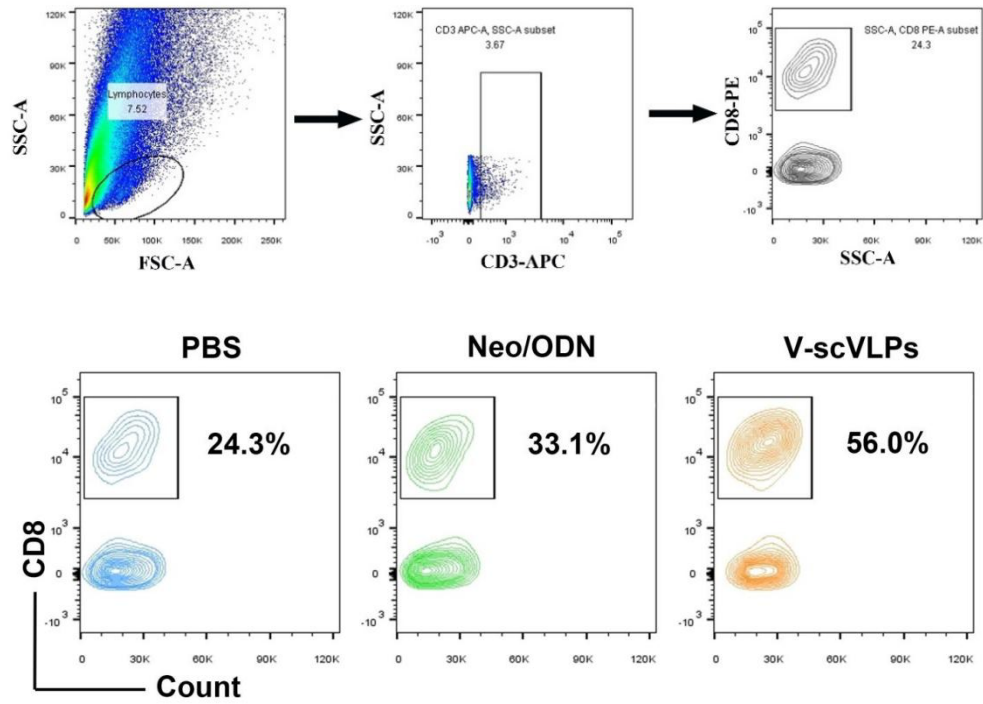


Figure. S20. Identify the CD8⁺T cells in tumor after receiving different treatment through FACS with staining anti-CD3-APC and anti-CD8-PE antibodies.

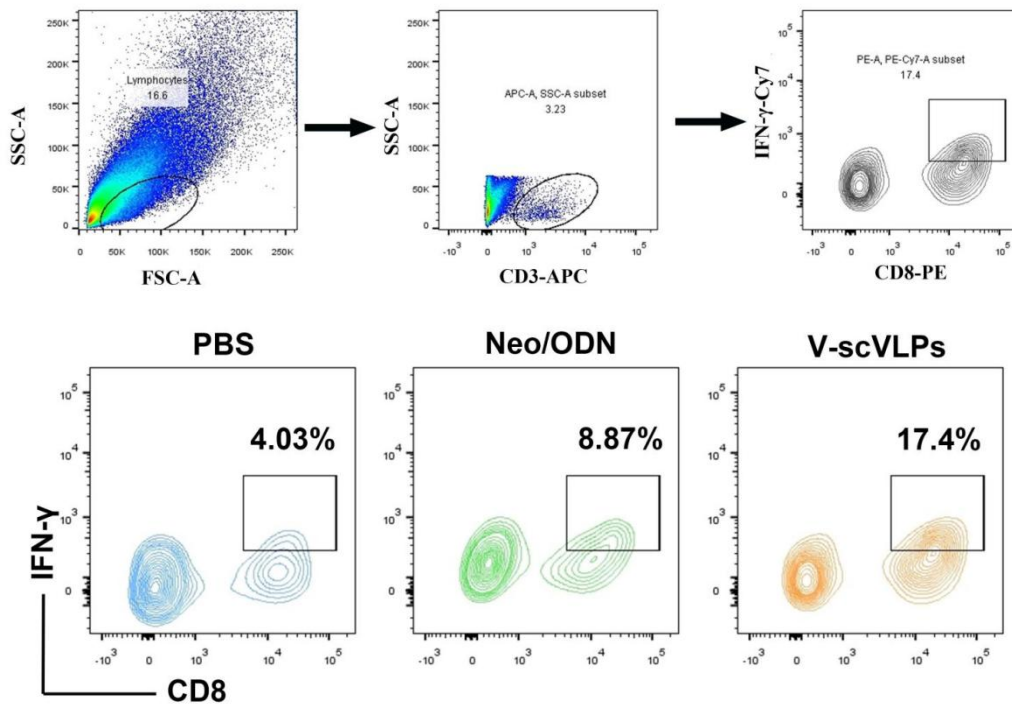


Figure. S21. Identify different types of IFN- γ ⁺CD8⁺CD3⁺T cells in tumors after receiving different treatments through FACS with staining anti-CD3-APC, anti-CD8-PE, IFN- γ - PE-Cy7 antibodies.

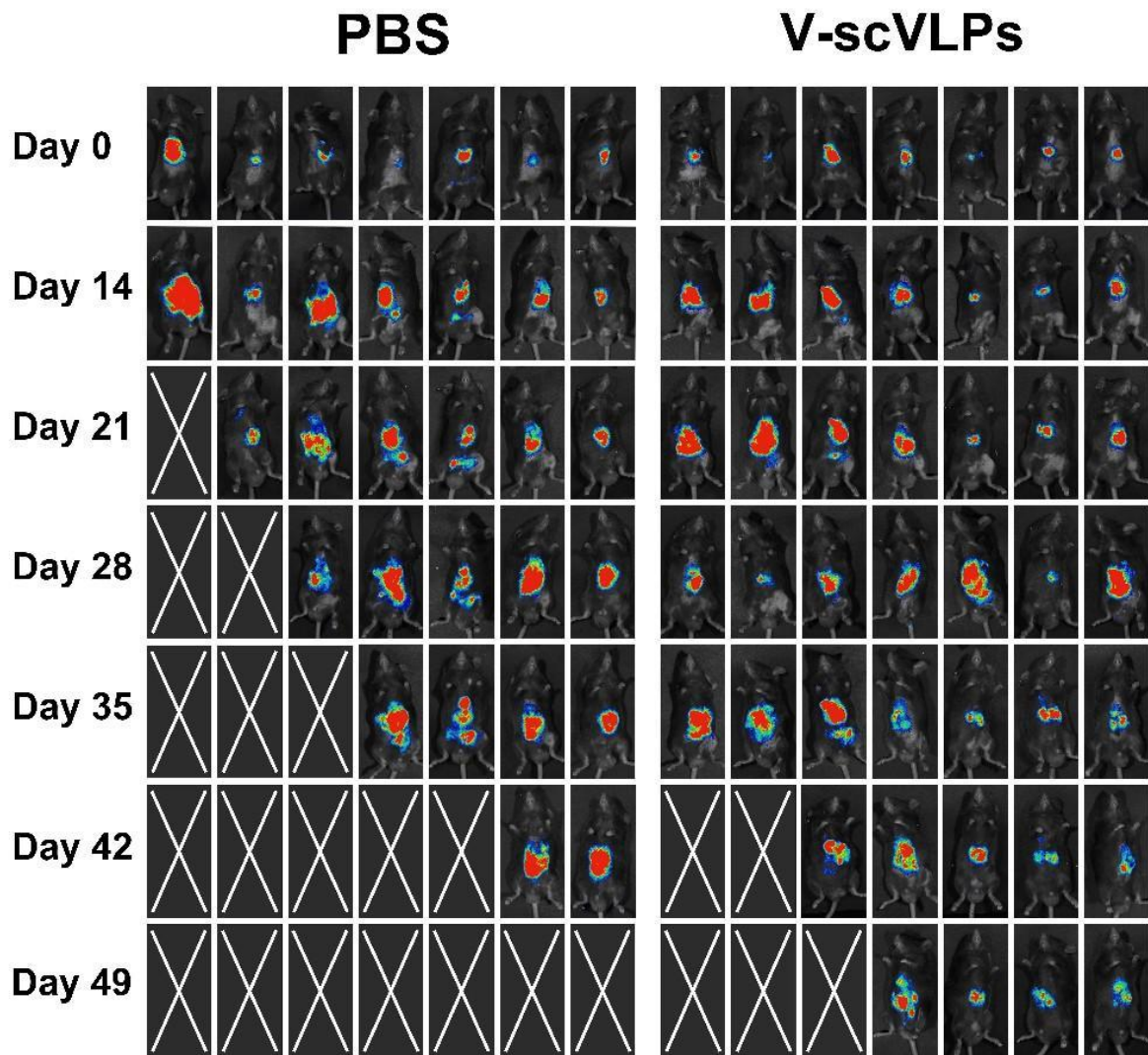


Figure. S22. Bioluminescence imaging of mice at the day 0, 14, 21, 28, 35,42 and 49 after receiving different treatments as indicated, n = 7.

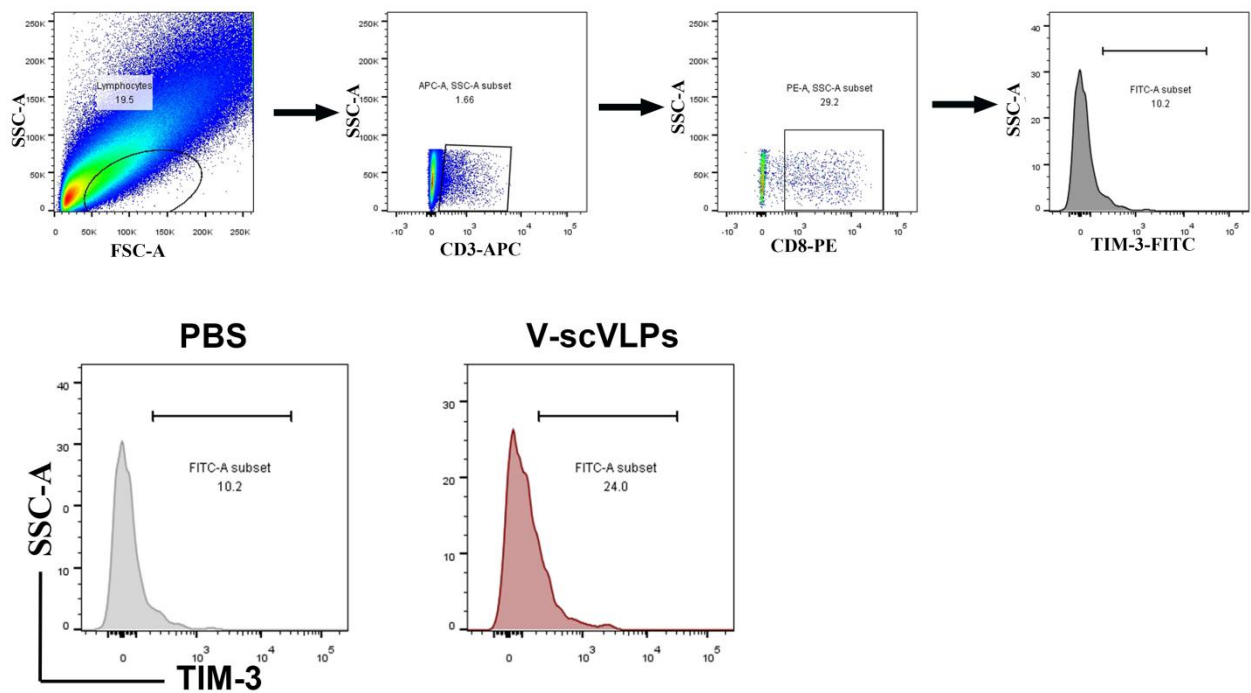


Figure. S23. Identify the different types of TIM-3⁺CD8⁺CD3⁺T cells in tumors after receiving different treatments by FCM with staining anti-CD3-APC, anti-CD8-PE, TIM-3-FITC antibodies.

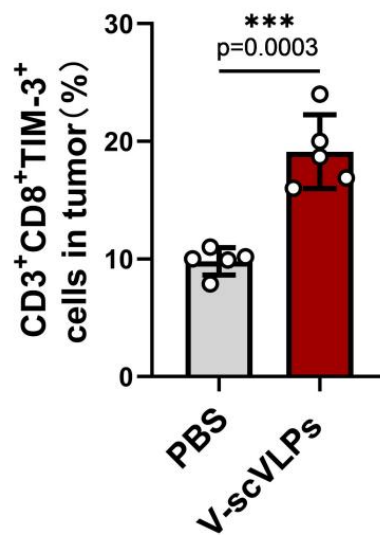


Figure. S24. The percentage of CD3⁺CD8⁺TIM-3⁺T cells in orthotopic tumors after receiving different treatment as indicated, n = 5. Statistical analysis was performed with T test, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$. Data are expressed as mean \pm SD.

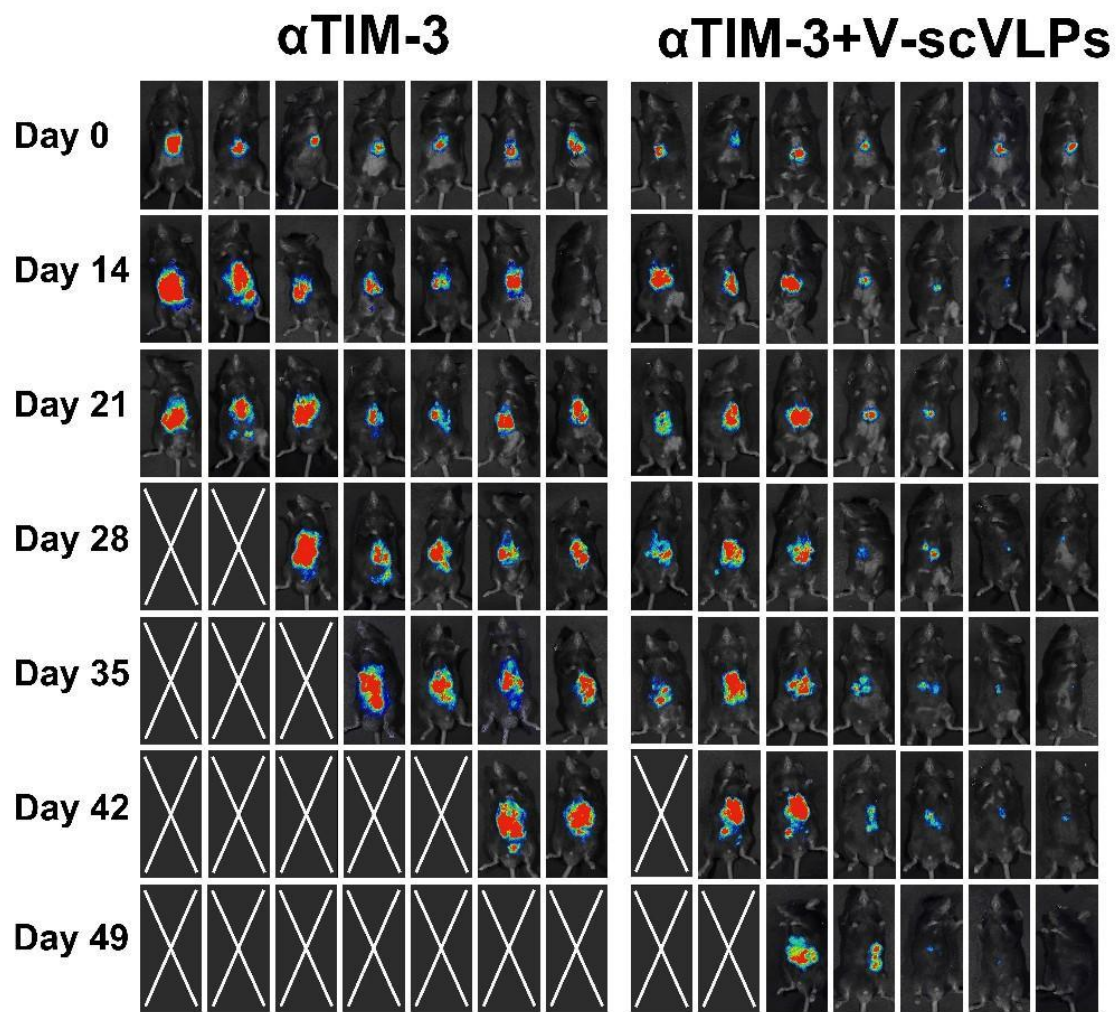


Figure. S25. Bioluminescence imaging of mice at the day 0, 14, 21, 28, 35,42 and 49 before and after receiving different treatments as indicated, n = 7.

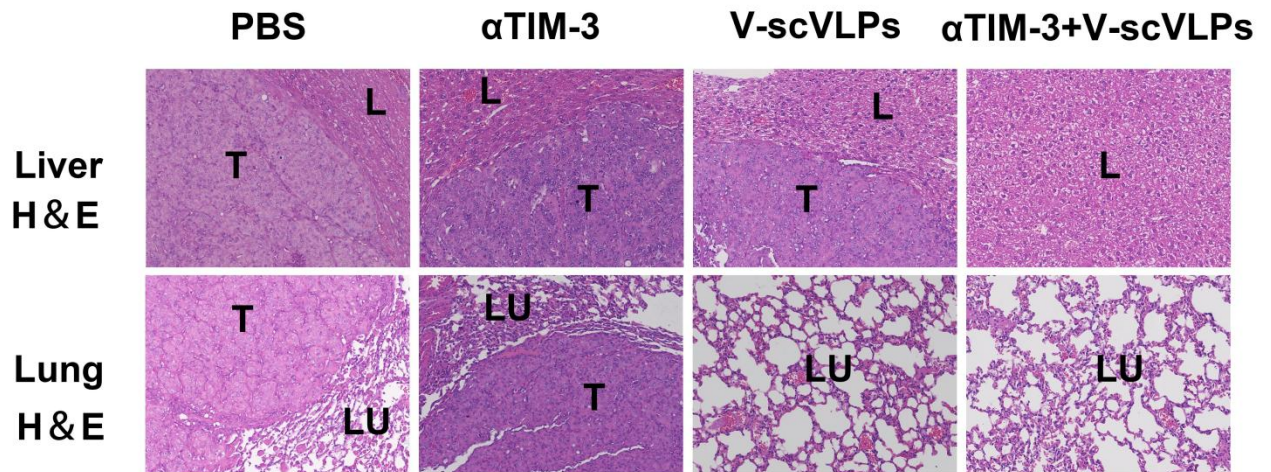


Figure. S26. The H&E staining of liver and lung tissues after receiving different treatments at the 49th day.

T represents tumor tissues, L was liver tissues, and Lu was lung tissues.

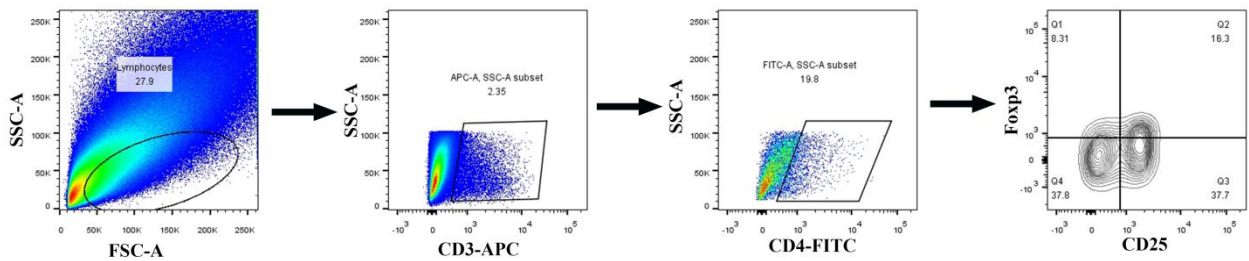


Figure. S27. Identify the different types of $CD3^+CD4^+CD25^+Foxp3^+$ T cells in tumors after receiving different treatments by FCM with staining anti-CD3-APC, anti-CD8-PE, TIM-3-FITC antibodies.

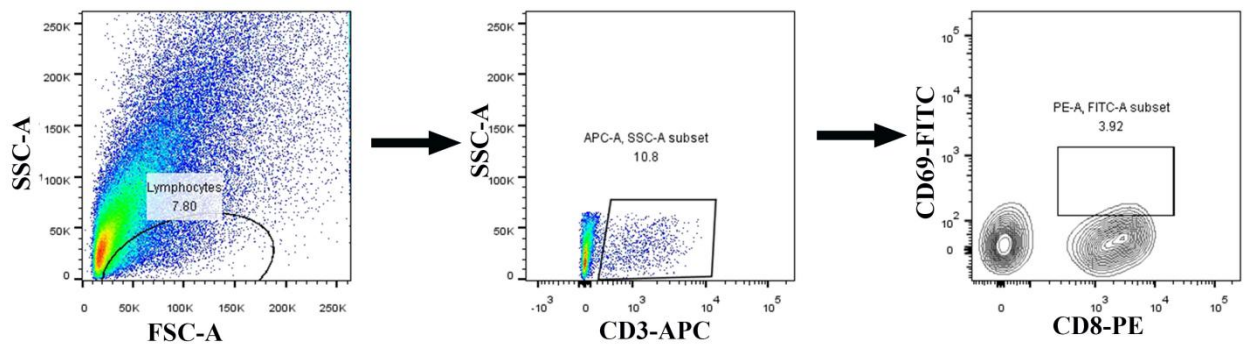


Figure. S28. Identify the different types of $CD3^+CD8^+CD69^+$ T cells in tumors after receiving different treatments by FCM with staining anti-CD3-APC, anti-CD8-PE, CD69-FITC antibodies.

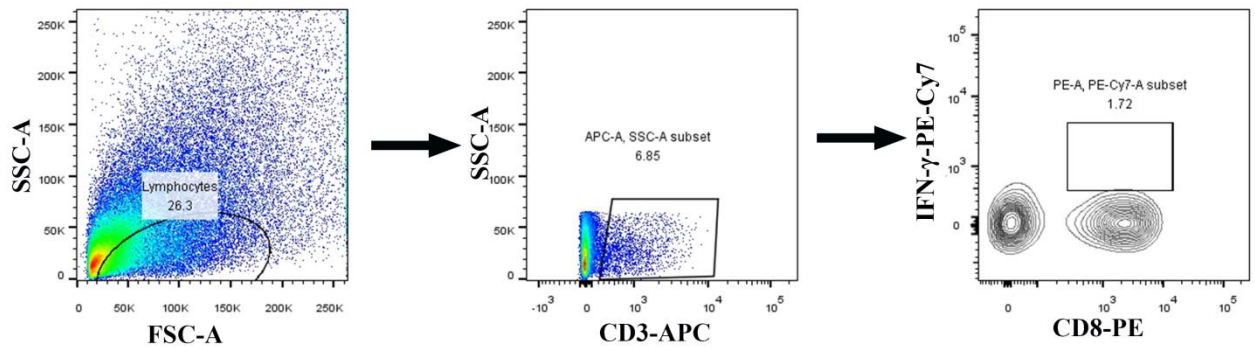


Figure. S29. Identify the different types of $CD3^+CD8^+IFN-\gamma^+$ T cells in tumors after receiving different treatments by FCM with staining anti-CD3-APC, anti-CD8-PE, IFN- γ -PE-Cy7 antibodies.

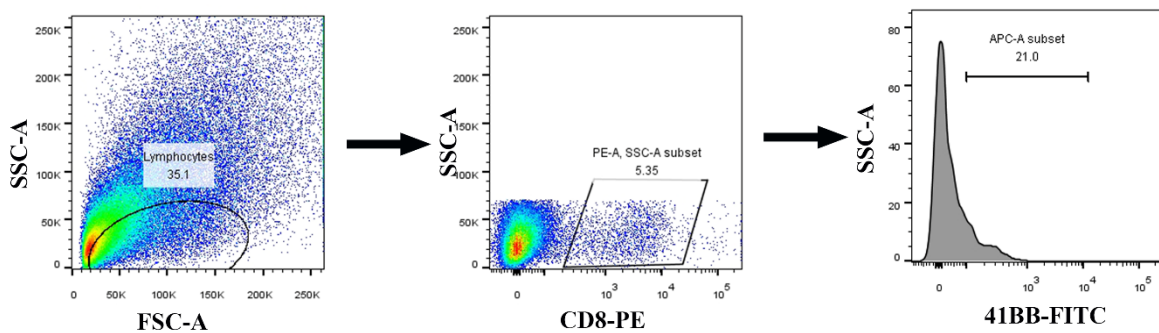


Figure. S30. Identify the different types of $CD8^+41BB^+$ T cells in tumors after receiving different treatments by FCM with staining anti-CD8-PE, 41BB-FITC antibodies.

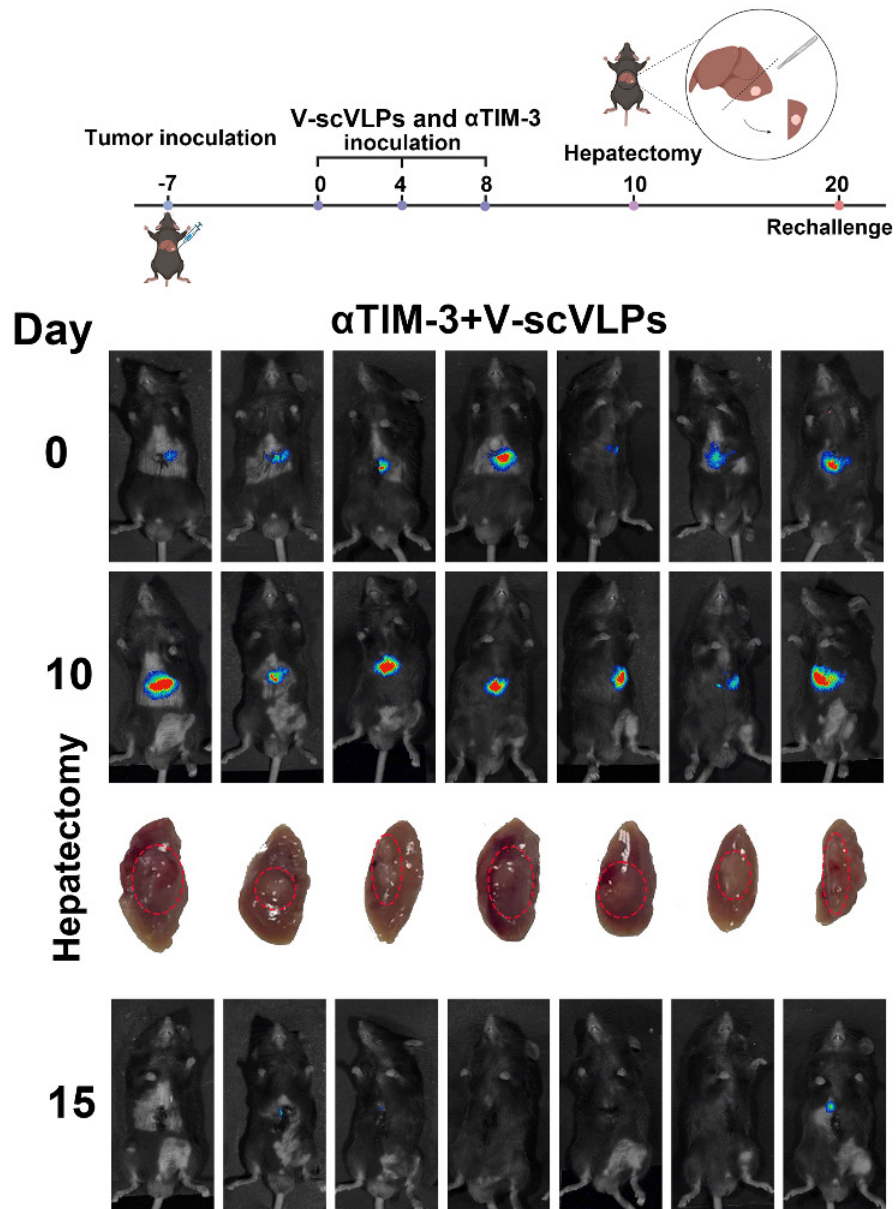


Figure. S31. Bioluminescence imaging of mice at the day 0, 10, 15 and photographs of orthotopic liver tumors and their H&E staining that excised from mice after inoculation Hepa1-6 tumor cells for 10 days, n = 7.

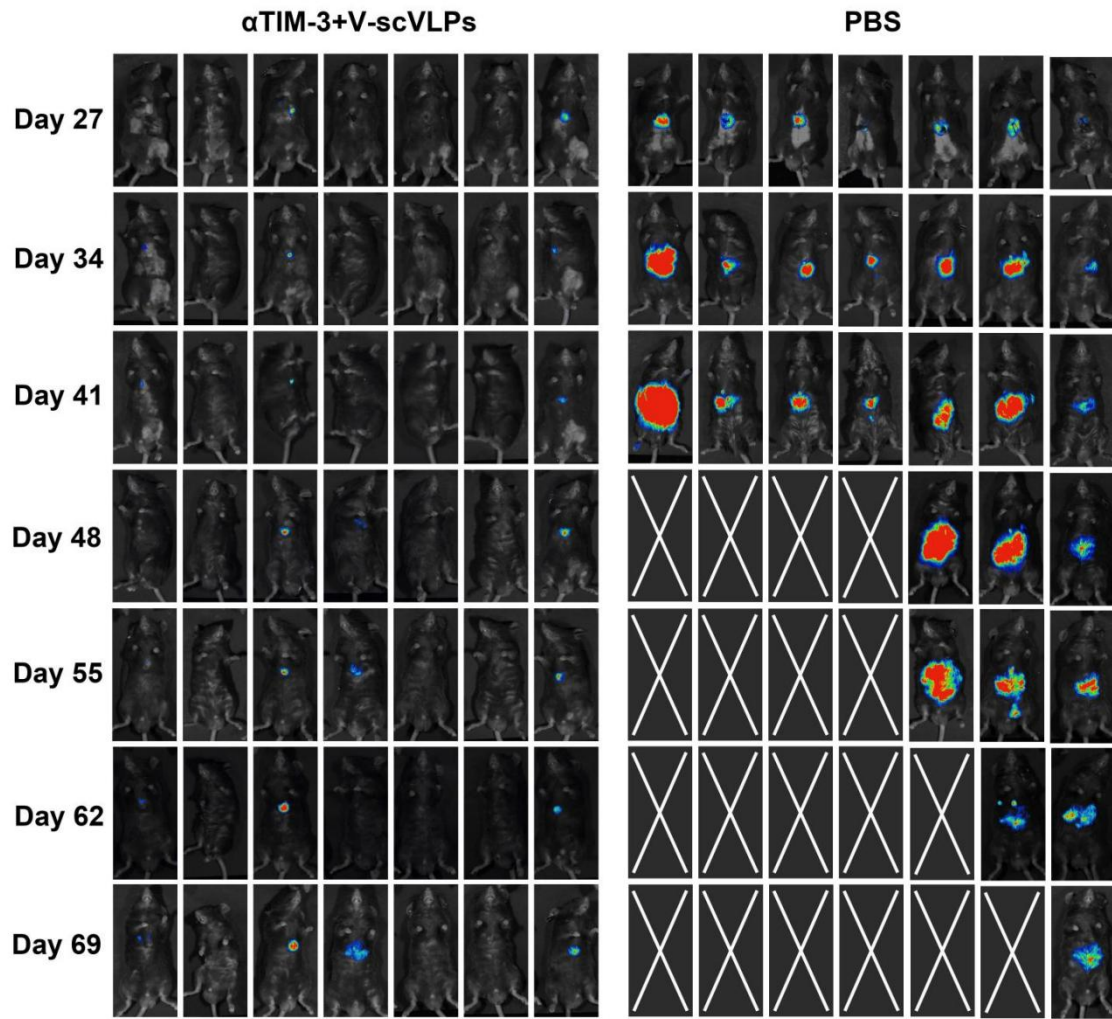
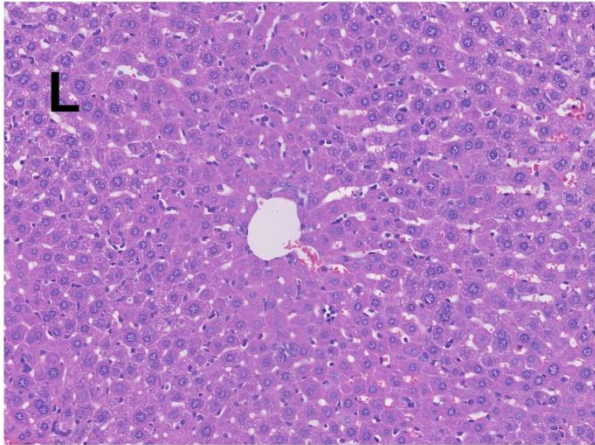


Figure. S32. Bioluminescence imaging of mice at the day 27, 34, 41, 48, 55, 62 and 69 after rechallenge as indicated, n = 7.

α TIM-3+V-scVLPs



PBS

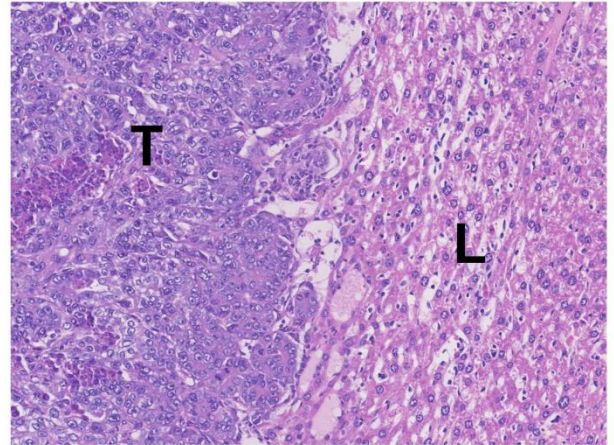


Figure. S33. The H&E staining of liver tissues and tumors after receiving different treatments at the day 69 day. T represents tumor tissues, L was liver tissues.

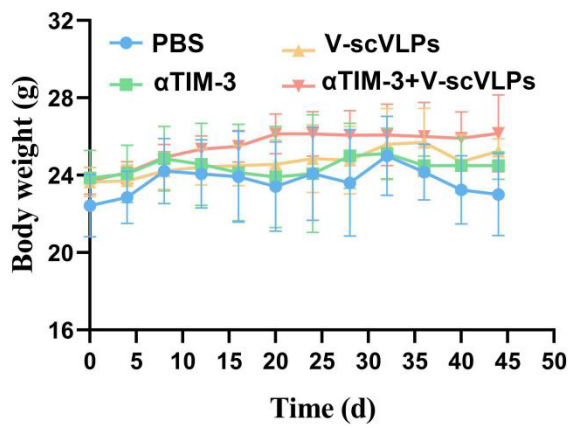
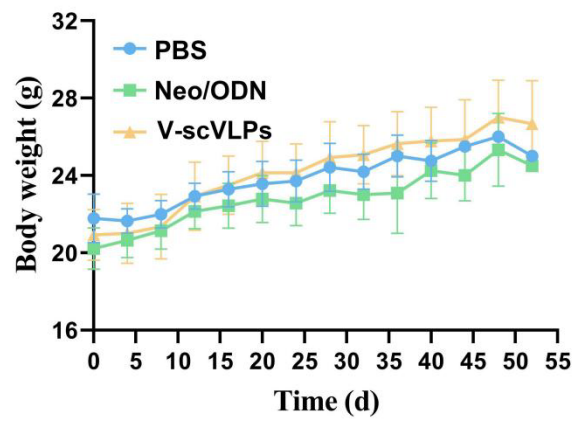
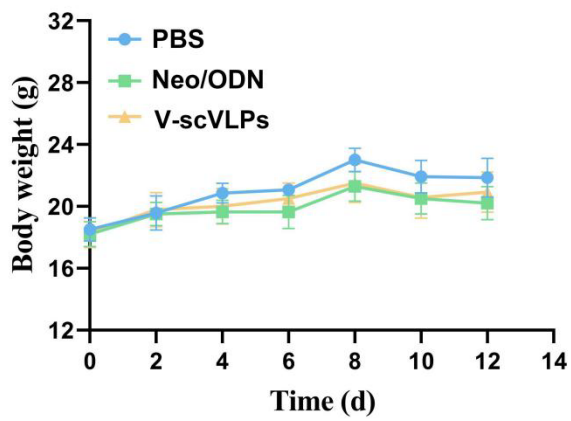


Figure. S34. Weight loss of Hepa1-6 tumor-bearing mice during different treatments as indicated. Data are presented as mean \pm SD (n = 7).

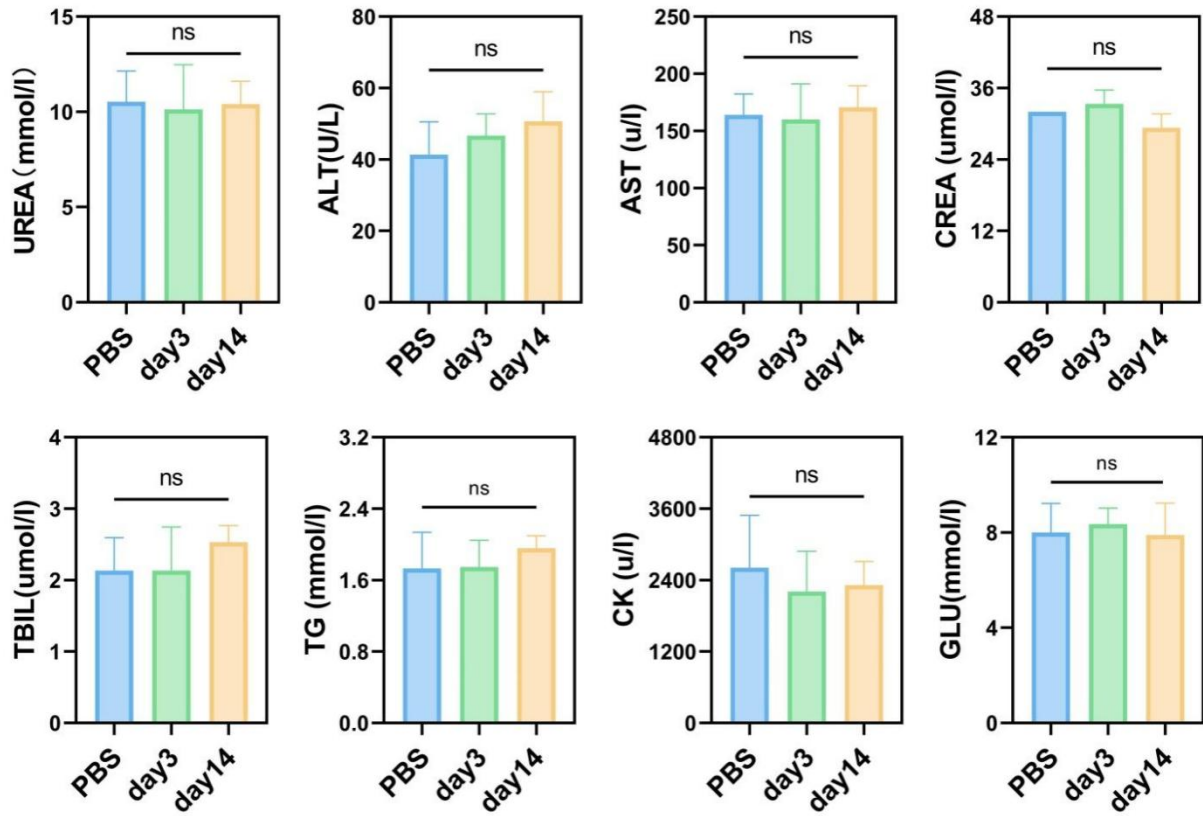


Figure. S35. Biochemical analysis (aspartate aminotransferase, AST; alanine aminotransferase, ALT; creatinine, UREA, Serum urea; CREA; total bilirubin, TBIL; thyroglobulin, TG; Alkaline phosphatase, CK; Glucose, GLU) in healthy mice at 3rd and 14th day after *s.c.* injection of V-scVLPs. Data are presented as mean \pm SD (n = 3). The statistical analysis was performed with ANOVA analysis.

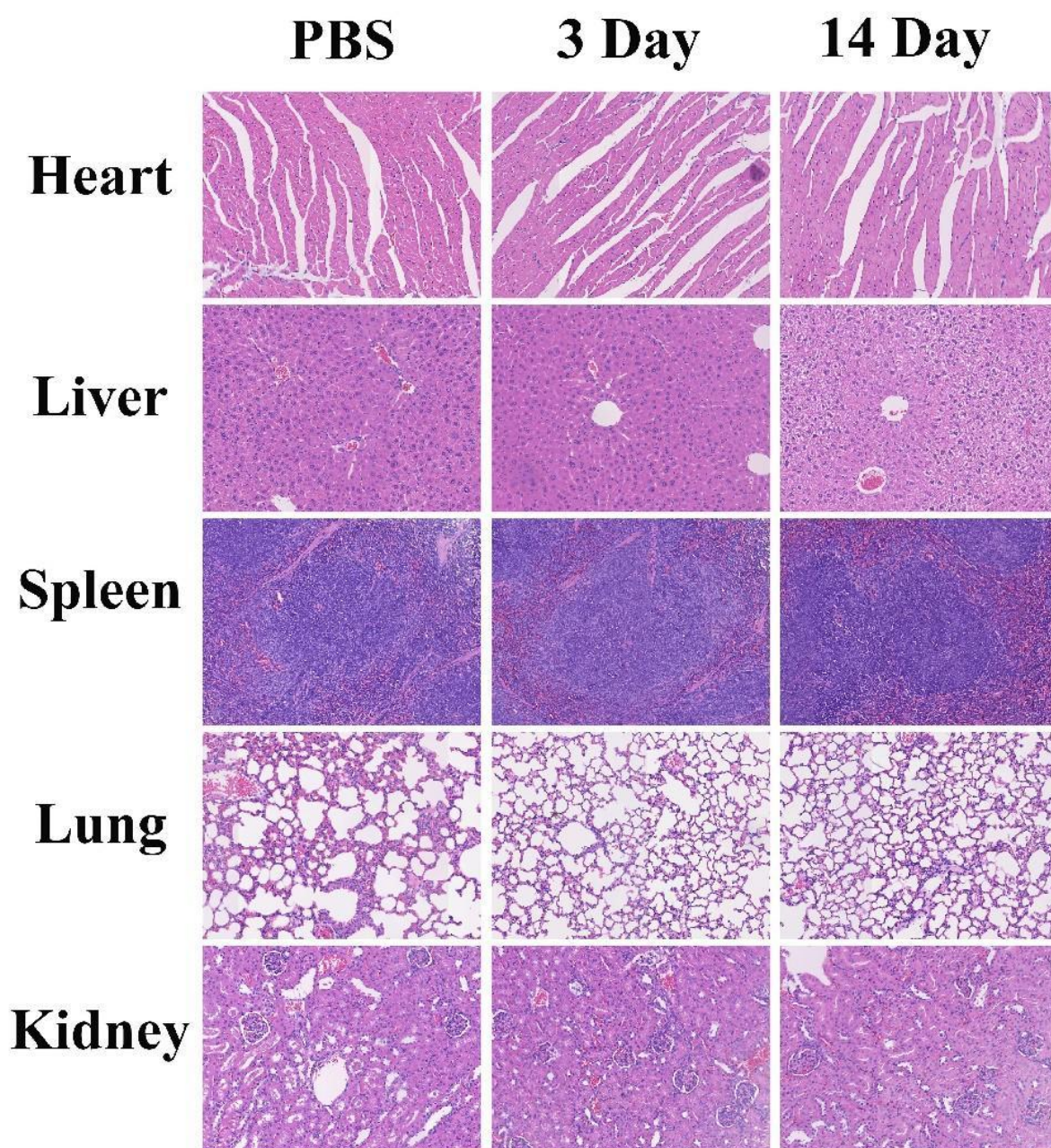


Figure. S36. H&E imaging of major organs (heart, spleen, lung and kidney) from healthy mice after *s.c.* injection with V-scVLPs for 3 or 14 days later.