Hypoxic viscosity and diabetic retinopathy

T Rimmer, J Fleming, E M Kohner

Abstract

Diabetic and sickle retinopathy have features in common – for example, venous dilatation, microaneurysms, and capillary closure preceding neovascularisation. Bearing in mind that haemoglobin in poorly controlled diabetes is abnormal and that extremely low oxygen tensions (known to cause sickling) exist in the healthy cat retina, we wished to explore the possibility that diabetic blood, like that of sickle cell disease, may become more viscous when deoxygenated. To do this we measured whole blood viscosity, under oxygenated and deoxygenated conditions, of 23 normal persons, 23 diabetic patients without retinopathy, and 34 diabetic patients with retinopathy. The shear rate used was 230 s^{-1} , which is similar to that thought to prevail in the major retinal veins. The viscosity of blood from normal persons, corrected for packed cell volume, did not change significantly on deoxygenation: mean 4.54 (SD 0.38) cps, versus, 4.57 (0.39) paired t test, p=0.66. Similarly the blood from diabetics without retinopathy showed no change: 4.42 (0.45) versus 4.42 (0.30), p=0.98; whereas the blood from patients with retinopathy changed from 4.82 (0.48) to 4.95 (0.63), p=0.027. The hypoxic viscosity ratio (deoxygenated divided by oxygenated viscosity) correlated with total serum cholesterol (r=0.44, p=0.018) but not with HbA1, serum glucose, triglycerides, or age. A disproportionate increase in venous viscosity relative to arterial viscosity would lead to increased intraluminal and transmural pressure and therefore exacerbate leakage across capillary walls.

Diabetic retinopathy is the commonest cause of blindness in the working age group in England¹ and the United States.² Since its pathophysiology is still not understood, all possible factors should be explored. We considered the well known fact that diabetic retinopathy has features in common with sickle cell retinopathy, namely, venous dilatation, microaneurysms, and capillary closure³ preceding neovascularisation,⁴ even though the new vessels are usually in different areas.

Both diseases are also characterised by abnormal levels of different haemoglobin components, which in sickle cell disease, under conditions of hypoxia, cause a marked rise in viscosity. Even in health the functional reserve of oxygen of the human retina lasts only seconds,⁵ and the oxygen tension (in cats) just proximal to the outer nuclear layer has been shown to be 12 mmHg⁶ and even zero,⁷ and the PO₂ of blood in human retinal veins has recently been estimated to be as low as 25 mmHg.⁸ If low oxygen tensions interfere with blood flow anywhere, it is likely to happen in the retina. Venous dilatation, one of the first clinical signs of diabetic retinopathy, could be the result of the blood becoming more viscous as it enters the venous side of the circulation. The aim of this study was to test the hypothesis that blood from diabetic patients becomes more viscous under conditions of extreme hypoxia such as are thought to exist even in the healthy retina.

Patients and methods

PATIENTS

Consecutive patients were entered into the study from the Diabetic Retinopathy and General Diabetic Clinics of the Hammersmith Hospital. The 57 patients in the study were later grouped, from clinical records and reference to fundus photographs, according to the retinopathy status of the worse eye: 23 had no retinopathy, 11 had background retinopathy (at least one microaneurysm but no macular oedema nor preproliferative changes), nine had more severe background retinopathy with exudative maculopathy (one also having cotton-wool spots), and 14 had previously undergone argon laser panretinal photocoagulation (at least 2000 burns 500 µm in size and strong enough to cause moderate blanching) for proliferative retinopathy. No patients in the study had active proliferative retinopathy. Twenty-three normal subjects were recruited from staff and relatives of patients. The details of patients and controls are shown in Table I.

Methods

After informed consent was obtained, blood samples were collected from the antecubital vein in lithium heparin containers (15 IU/ml). The samples were collected from five subjects at a time. From each sample 1.5 ml was transferred to each of two 10 ml syringes, which were immediately capped. There were five syringes containing blood to be equilibrated with an oxygenating gas mixture and five syringes containing blood to be equilibrated with a deoxygenating gas mixture. A frame was constructed which allowed five syringes of blood/gas to be equilibrated simultaneously.

TABLE I Details of patients and normal person	TABLE I	Details of	patients and	normal	persons
---	---------	------------	--------------	--------	---------

	Number	Age (vr)*	Duration of diabetes (yr)
Normal	23	38.5 (15.5)	-
No retinopathy	23	55.7 (15.0)	7·7 (6·0)
Background	11	53.7 (15.2)	9.7 (4.2)
Maculopathy	10	55.1 (8.2)	12.1 (5.4)
Postphotocoagulation	14	50.1 (11.5)	16.1 (10.9)

* Means (with standard deviations).

Department of Medicine, Royal Postgraduate Medical School, London W12 0HS T Rimmer E M Kohner

Department of Surgery J Fleming

Correspondence to: Timothy Rimmer, FRCS, Biomedical Engineering Department, Northwestern University, Evanston, Illinois 60208, USA.

Accepted for publication 15 February 1990

The oxygenating gas mixture was 95% air/5% CO₂ by volume and the deoxygenating mixture 1.4% $0_2/7.0\%$ CO₂/balance N₂. Both gas mixtures were passed through a humidifyer (Sintaglass Ltd, London) before being exposed to the blood samples. The first five syringes containing blood samples were deoxygenated by filling the remaining 8.5 ml of the syringes with the appropriate gas mixture and then rotating the syringes at approximately 30 RPM for 20 minutes. This duration was chosen because a preliminary test had shown that another 10 minutes only lowered the PO_2 by a further 1 or 2 mmHg. The syringes were submerged in water at 37°C. At 5-minute intervals the gas volumes were expelled and replaced with fresh gas mixture. The blood samples in the second set of five syringes were then oxygenated in the same manner as above with the oxygenating gas mixture.

After agitation a small volume from each svringe was injected into the Corning 178 pH/ blood gas analyser (Ciba Corning Diagnostics Ltd, Halstead, UK) and drawn into capillary tubes for packed cell volume PCV) measurement by the microhaematocrit procedure (Hawksley Ltd, England). The remaining volume in each syringe was reduced to 1 ml for viscometry. Apparent whole blood viscosities were measured at 37°C with a Wells-Brookfield Cone-Plate rotational viscometer (model LVT, Stanford, CA) which was calibrated immediately prior to the study. Our viscosity assays were 'apparent' because they were only at one shear rate, whereas viscosity of a non-Newtonian fluid is theoretically the slope of the tangent of the curved plot of shear stress against shear rate. The viscometer was purged with the appropriate gas mixture to minimise gas exchange during the period of the assay. This was achieved by passing the gas through a nozzle attached to the barrel of the viscometer which did not interfere with any moving parts. It took 31/2 hours to complete the above procedure for five subjects from venepuncture to the last viscosity measurement. The investigator who did all the studies was masked for the patients' retinopathy status.

Besides measuring whole blood viscosity we also report on the viscosity corrected for PCV variation by the methods described by Dormandy.⁹ At venepuncture samples were also taken for HbA1 levels. All diabetic patients underwent regular biochemical screening. Plasma glucose (Beckman Glucose analyser),

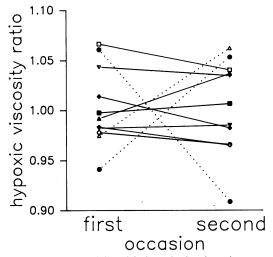


Figure 1: Reproducibility of the hypoxic viscosity ratio (deoxygenated divided by oxygenated viscosity) of 11 normal subjects, the two measurements being several weeks apart and both at the shear rate of 230 s^{-1} . The solid lines represent reproducibility scores greater than 95%, whereas the dotted lines represent those between 84 and 95%.

and HbA1 were measured at every visit. Total serum cholesterol and triglyceride levels, however, were assayed less frequently but, because of an association with retinopathy,¹⁰¹¹ were recorded for those patients who had these tests on blood taken at the same time as that for viscometry.

To study the reproducibility of the relationship between the oxygenated and deoxygenated viscosities the viscosity measurements were repeated several weeks later on 11 normal persons. The results on each occasion were expressed as a ratio of the deoxygenated divided by the oxygenated viscosity, which was called the 'hypoxic viscosity ratio'. Reproducibility scores were calculated for each individual by expressing the difference between the two ratios (the signs being ignored) as a percentage of the mean and then subtracting this from 100. Correlation tests (least squares method) were carried out between viscosity values and ratios described above and independent variables - that is, HbA1, blood glucose, serum total cholesterol, serum triglycerides, and age. Student's t tests were used to compare groups of data, and p values less than 0.05 were considered significant.

Results

For the 11 normal persons the reproducibility

TABLE 11 Results of viscometry of normal persons and diabetics. The shear rate was 230 s⁻¹, which is similar to that thought to prevail in the major retinal veins

	Normal		Diabetics without retinopathy		Diabetics with retinopathy	
Number	23	p value*	23	p value ^s	34	
Measured viscosities (cps)†						
Oxygenated viscosity	4.31 (0.53)	0.73	4.26 (0.40)	0.001	4.72 (0.56)	
Deoxygenated viscosity	4.32 (0.47)	0.53	4.22 (0.55)	<0.001	4.84 (0.71)	
Paired t test	0.89		0.53		0.041	
Corrected viscosities (cps)‡						
Oxygenated viscosity	4.57 (0.39)	0.14	4.42 (0.30)	<0.001	4.82 (0.48)	
Deoxygenated viscosity	4.54 (0.38)	0.31	4.42 (0.45)	0.001	4.95 (0.63)	
Paired t test, p value	0.66		0.98		0.027	
Hypoxic viscosity ratio	0.996 (0.065)	0.62	0.990 (0.017)	0.014	1.025 (0.064)	

* Unpaired t tests between normal persons and diabetics without retinopathy. † Expressed as means (with standard deviations). ‡ Corrected to standard PCV of 45%. § Unpaired t tests between diabetics with and without retinopathy.

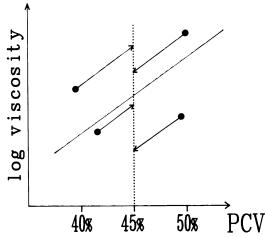


Figure 2: Correction of viscosity for variations of packed cell volume (PCV) for all subjects and patients. Data points (four sample ones are shown) were moved by computer to the 45% PCV line in a direction parallel to the linear regression line relating the log of the measured viscosity to the PCV for the group concerned.

scores ranged from 84.6 to 99.7 (mean 95.4). These scores are illustrated in Figure 1.

The viscosities of oxygenated and deoxygenated blood of normal persons and diabetics without and with retinopathy are presented in Table II. The PCV corrected viscosity is shown in Figure 2, where the slope of the linear regression line was calculated separately for each group concerned, at the same shear rate of 230 s⁻¹. The log of viscosity is used because of its greater correlation with PCV. The whole blood viscosity of normal persons and diabetics without retinopathy did not change significantly when deoxygenated (paired t tests, p=0.66 and 0.98 respectively for corrected values). In contrast the blood from patients with retinopathy became more viscous when deoxygenated (p=0.027). This pattern was seen with both the measured and corrected values.

Blood gas analyses on 56 samples (20 normals and 36 diabetics) showed that after oxygenation the mean values for PO₂ were 124·7 (SD 17·2) mmHg and for PCO₂ 40·4 (1·6) mmHg. After deoxygenation the values for PO₂ were 18·3 (SD 2·8) mmHg and for PCO₂ were 54·1 (2·3) mmHg. The desired changes in blood gases were achieved with good reproducibility, indicated by the relatively small standard deviations.

The results of corrected viscosities of separate retinopathy groups together with the results of the other blood tests performed are presented in Table III. The biggest differences of viscosities

are between the no-retinopathy group and the maculopathy and postphotocoagulation groups. In particular there were no significant differences between the HbA1 levels of the retinopathy groups. A scatterplot of the corrected deoxygenated viscosities is shown in Figure 3 to illustrate the spread of data.

Correlation studies showed a significant relationship between total serum cholesterol and the hypoxic viscosity ratio for the 29 patients who had these assays at the same time as the viscometry:

 $HVR = 0.02 \times C + 0.89$, r=0.44, p=0.018.

where HVR is the hypoxic viscosity ratio and C is the total serum cholesterol level (mmol/l). The relationship was the same for corrected and measured viscosities and is illustrated in Figure 4. Cholesterol also correlated with deoxygenated viscosity ($0.24 \times C+3.3$, r=0.45, p=0.013) but not with oxygenated values. There were no other statistically significant correlations between any of the viscosity values and any of the other independent variables tested, namely, glucose, HbA1, triglycerides, and age. There was no correlation between cholesterol and either HbA1 or blood glucose.

The first 18 samples (6 normal and 12 diabetic) had PCVs measured of both oxygenated and deoxygenated portions. Of these, seven (three normal) showed a slight increase of PCV on deoxygenation (mean 2.6%) seven (three normal) showed a decrease (mean 2.5%), and four (one normal) showed no change. Only seven of the 18 pairs of PCVs reflected the change in viscosity. Owing to this lack of pattern all the following samples only had one estimate of PCV, performed on the original venous sample.

Discussion

The study was undertaken to test the hypothesis that blood from diabetic patients becomes more viscous when deoxygenated. The hypothesis has been supported in part: blood from diabetics with retinopathy was significantly more viscous when deoxygenated in contrast to that of normals and diabetics without retinopathy. However, this tendency (expressed as a 'hypoxic viscosity ratio') did not correlate with HbA1 but did with total serum cholesterol to a modest degree (r = 0.44).

The correlation with cholesterol indicates that, of the factors influencing the hypoxic ratio, only 19% (r^2) can be attributed to the serum cholesterol level. Although cholesterol has been reported to affect membrane viscosity¹² and rigidity¹³ and to correlate with the severity of

TABLE III Results of viscometry and blood tests for 57 diabetic patients divided into retinopathy groups. The viscosities have been corrected to a PCV of 45%. The values are expressed as means (with standard deviations). The shear rate was 230 s'. The figures in parentheses, unless otherwise obvious, are p values (<0.05) for unpaired t tests with the no-retinopathy group

	No retinopathy	Background	Maculopathy	Postphotocoagulation
Number	23	11	9	14
Oxygenated viscosity cps	4.42 (0.30)	4.47 (0.47)	4.86 (0.51) (0.005)	4.86 (0.50) (0.002)
Deoxygenated viscosity cps	4.42 (0.45)	4.77 (0.59)	5.00 (0.58) (0.005)	5.05 (0.70) (0.002)
Hypoxic viscosity ratio	0.990 (0.017)	1.007 (0.069)	1.028(0.030)(<0.046)	1.037 (0.075)
PCV%*	42.7 (3.8)	43.4 (4.5)	42.7 (4.9)	43.9 (4.3)
HbA1%	7.7(2.1)	7.7 (1.5)	7.6(2.1)	8.0 (2.0)
Blood glucose mmol/1	9.2 (3.6)	14.2(6.7)	10.2(2.8)	9.6 (6.2)
Total serum cholesterol mmol/l	$5 \cdot 3(1 \cdot 4)(n=5)$	5.6(1.4)(n=8)	7.3(1.8)(n=5)	6.5(1.2)(n=11)
Serum triglycerides mmol/l	1.2(0.6)(n=5)	$2 \cdot 3 (2 \cdot 2) (n=8)$	4.0(4.8)(n=5)	1.7(1.2)(n=11)

*Normal persons in this study: PCV 42·2 (3·7)%, HbA1 4·3 (0·6)%. Normal ranges for the hospital: blood glucose 3·5-5·0 mmol/l, total serum cholesterol 4·0-6·5 mmol/l, and triglycerides 0-2·0 mmol/l.

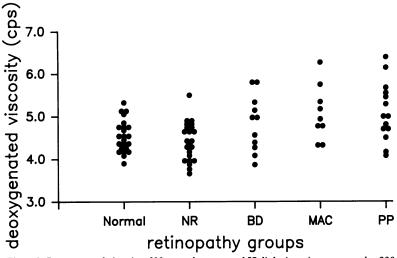
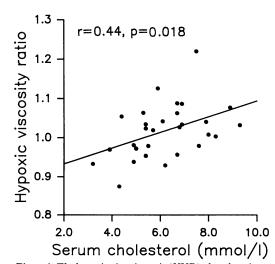
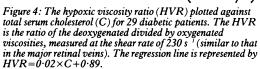


Figure 3: Deoxygenated viscosity of 23 normal persons and 57 diabetic patients measured at 230 s⁻¹ and corrected to a PCV of 45%. NR=no retinopathy. BD=background retinopathy only. MAC=maculopathy. PP=postphotocoagulation. The deoxygenated viscosity of the patients in the maculopathy and postphotocoagulation groups were 13% (p=0.013) and 14% (p=0.007) higher than that of patients without retinopathy respectively.

diabetic retinopathy,10 it is just as likely that the diabetic metabolic state itself is interfering directly with blood viscosity rather than through abnormal cholesterol levels. It was recently shown that red cell deformability was the same in diabetics with and without retinopathy (n=30,presumably under conditions of full oxygenation).¹⁴ In the same study red cell membrane cholesterol content was lower in patients with retinopathy than those without (p < 0.025) in contrast to the plasma levels, which were the other way around (p < 0.05). In our study there was no relationship between cholesterol and HbA1 (or blood glucose), which might have been expected, since bad control is associated with increased levels of cholesterol.

The deoxygenating gas mixture was designed to bring the PO_2 down from the normal mixed venous level of about 40 mmHg to just below that of retinal venous blood, which is approximately 25 mmHg.⁸ It is probable that the oxygen tension of blood in the deepest retinal capillaries will be lower still than that of mixed retinal venous blood. The PO_2 of the oxygenated samples is





about 25 mmHg higher than true arterial blood, but the difference in saturation of the erythrocytes is of no consequence. The phenomenon of hypoxic viscosity may be a result of the combination of highly glycated haemoglobin and high cholesterol or other components of the blood. Haemoglobin is glycated at the terminal valine of the β chain – the same site of attachment of 2,3 diphosphoglycerate (DPG).15 This substance, apart from facilitating the release of oxygen from haemoglobin, favourably affects the solubility of normal deoxygenated haemoglobin,¹⁶ which may therefore be reduced if DPG is displaced by glycation. Fibrinogen is one blood component, for example, which has been reported to influence blood viscosity in diabetes¹⁷ but was not measured in this study.

Some increase in viscosity on deoxygenation may be expected in normal persons because the chloride shift is supposed to cause a 3% rise in PCV in venous blood.¹⁸ This rise was not seen consistently in this study perhaps because the microhaematocrit method was not sensitive enough. The viscosity measurements in this study were carried out at the shear rate of 230 s⁻¹ because this most closely resembles the shear rate thought to prevail in the retinal veins. The data of Riva et al^{19} and Feke et al^{20} show that the centre-line velocity of blood in a retinal vein of 180 μ m diameter is 1.9 and 3.2 cm/s respectively, indicating average shear rates of 281 and 474 s⁻¹.* Red blood cells are subjected to changing shear rates during their passage through the vascular system. The plug flow in capillaries requires them to change shape rapidly in order to maximise contact between their cell membranes and the capillary endothelium. The shear rate in this situation is not well defined but is probably high.

Blood from the patients with retinopathy was 12% more viscous when deoxygenated than deoxygenated blood from diabetic patients without retinopathy (p=0.001, Figure 3, Table III). This could mean reductions in blood flow, which may not be adequately corrected by autoregulation because the PO₂, PCO₂ and pressure of blood in the retinal arteries is normal. Previous studies on retinal vascular autoregulation have demonstrated vascular responses to abnormal changes in arterial blood including PO₂, PCO₂²¹, systemic blood pressure,²² blood glucose,^{23 24} and pharmacological agents.^{25 26} Initial demonstrations of an increase in blood flow in response to dark adaptation^{27 28} were later questioned²⁹ and explained as an artifact of the laser doppler system.³⁰ There is no evidence that retinal blood flow can increase in the absence of changes in arterial blood parameters, but the vessels may be able to respond directly to tissue hypoxia.

To maintain volume blood flow in the face of increased viscosity there must be an increase in pressure gradient and/or dilatation of vessels (and hence a reduction in velocity). Venous dilatation and lower velocities are seen in practice in diabetic retinopathy.³¹ Any increase in pressure gradient would cause a rise in capillary intraluminal and transmural pressure and predispose to oedema. In the event of volume blood flow not being fully corrected the consequence

*Average shear rate= $(-4/3)\times(\max \text{ velocity/radius})$. This is derived from an integration of the radially dependent shear rate over the cross sectional area, assuming Poiseuille flow since the Reynolds number is only 0.7.

could be chronic mild hypoperfusion of the retinal tissue, which could lead to hypoxia and a rise in concentration of waste products of metabolism, such as toxic oxidative radicles, which are considered by some to be important in the development of diabetic complications.³² In advanced retinopathy increased hypoxic viscosity could set up a vicious circle by leading to reduced blood flow and lower oxygen tensions, which would cause further increase in viscosity. However, the fact that it was only the patients with the most severe retinopathy who showed an increase in hypoxic viscosity suggests that this is the result of the same diabetic microenvironment which caused the retinopathy rather than the cause of the retinopathy itself; nevertheless the retinopathy could be worsened by it.

The phenomenon of hypoxic viscosity could be important because it might present a new target for pharmacological attack. A future study will examine whether the deformability of red blood cells from diabetic patients is affected by oxygen tension.

The authors would like to express their appreciation to Victoria Rimmer, RGN, and Drs Robert Linsenmeier, Thomas Goldstick and Paul Sullivan for helpful discussions, Professor Stephen Bloom for allowing his patients to be included in this study, Rod Braun MS for shear rate calculations, and Dr Emile Chen for invaluable help in preparing the manuscript.

This work was supported by the British Council for the Prevention of Blindness

- Department of Health and Social Security. Reports on Public Health and Medical Subjects No. 129. Blindness and partial sight in England 1969-1976. London: HMSO, 1979.
- Signi in Enguna 1709-1710. London: HMSO, 1919.
 Weingeist TA. Congressional testimony in support of the Citizens' Budget Proposal for the National Eye Institute for fiscal year 1990. New York: Research to Prevent Blindness, 1989.
 Condon PI, Serjeant GR. Ocular findings in homozygous sickle cell anemia in Jamaica. Am f Ophthalmol 1972; 73: 533-43
- 33-43
- Goldberg MF. Classification and pathogenesis of proliferative sickle retinopathy. Am J Ophthalmol 1971; 71: 649–65.
 Anderson B, Saltzmann HA. Retinal oxygen utilization measured by hyperbaric blackout. Arch Ophthalmol 1964; 727-702 5 **72:** 792–5.
- 6 Alder VA, Cringle SJ, Constable IJ. The retinal oxygen profile in cats. Invest Ophthalmol Vis Sci 1983; 24: 30-6. 7 Linsenmeier RA. Effects of light and darkness
- on oxygen distribution and consumption in the cat retina. J Gen Physiol
- distribution and consumption in the cat retina. J Gen Physiol 1986; 88: 521-42.
 8 Sebag J, Delori FC, Feke GT, Weiter JJ. Effects of optic atrophy on retinal blood flow and oxygen saturation in humans. Arch Ophthalmol 1989; 107: 222-6.
 9 Dormandy JA. Measurement of whole blood viscosity. In: Lowe GD, Barbenel JC, Forbes CD, ed Clinical aspects of blood viscosity and cell deformability. New York: Springer 1981: 67-78.

- Dornan TL, Carter RD, Bron A, et al. Low density lipoprotein cholesterol: an association with the severity of diabetic retinopathy. *Diabetologia* 1982; 22: 167-70.
 Mohan R, Mohan V, Susheela L, et al. Increased LDL cholesterol in non-insulin dependent diabetics with maculopathy. *Acta Diabetol Lat* 1984; 21: 85-9.
 Feinstein MB, Fernandez SM, Sha'afi RI. Fluidity of natural membranes and phosphatidylserine and ganglioside dis-persions. Effects of local anesthetics, cholesterol and protein. *Biochim Biophys Acta* 1975; 413: 354-70.
 Duax WL, Wawrzak Z, Griffin JF, Cheer C. Sterol confor-mation and molecular properties. In: Yeagle PL. *Biology of Cholesterol*. Boca Raton: CRC Press, 1988: 1-18.
 Zamorano AF, Arnalich F, Ferro-Sanchez A, Grande C, Lahoz C, Pallardo LF. Deformabilidad eritrocitaria y lipidos de la membrana eritrocitaria en la diabetes mellitus. *Med Clin (Barc)* 1987; 89: 717-20.

- Inpucos de la memorana eritrocitaria en la diabetes mellitus. Med Clin (Barc) 1987; 89: 717-20.
 15 Bunn HF, Gabbay KH, Gallop PM. The glycosylation of hemoglobin: relevance to diabetes mellitus. Science 1978; 200: 21-7.
- 16 Friederichs E, Lakomek M, Winkler H, Tillman W. Effects of
- 16 Frederichs E, Lakomer M, Winker A, Himlan W. Effects of calcium ions and 2,3 DPG on the solubility of deoxygenated human hemoglobin. *Clin Hemorheol* 1988; 8: 707–14.
 17 Barnes AJ, Locke P, Saidde PR, et al. Is hyperviscosity a treatable component of diabetic microvascular disease? *Lancet* 1977; ii: 789–91.
- Bancer WF. Review of medical physiology. 6th ed Los Altos: Lange, 1973: 487.
 Riva CE, Grunwald JE, Sinclair SH, Petrig BL. Blood velocity
- and volumetric flow rate in human retinal vessels. Invest Ophthalmol Vis Sci 1985; 26: 1124-32.
 Feke GT, Tagawa H, Deupree DM, et al. Blood flow in the normal human retina. Invest Ophthalmol Vis Sci 1989; 30: 500 ft.
- 58-65
- 21 Hickam JB, Frayser R. Studies of the retinal circulation in man. Circulation 1966; 33: 302-16.
- man. Circulation 1906; 35: 502-10.
 Robinson F, Riva CE, Grunwald JE, Petrig BL, Sinclair SH. Retinal blood flow autoregulation in response to an acute increase in blood pressure. Invest Ophthalmol Vis Sci 1986; 27: 722-6.
- 23 Atherton A, Hill DW, Keen H, Young S, Edwards EJ. The
- effect of acute hyperlycaemia on the retinal circulation of the normal cat. *Diabetologia* 1980; 18: 223–7.
 Fallon TJ, Sleightholm MA, Merrick C, Chahal P, Kohner EM. The effect of acute hyperglycaemia on flow velocity in the macular capillaries. *Invest Ophthalmol Vis Sci* 1987; 28:
- 1027-30.
 ffytche TJ, Bulpitt CJ, Archer D, et al. Effects of papaverine on the retinal microcirculation. Br J Ophthalmol 1973; 57: 910-20.
- Yan HY, Chiou GC. Effects of L-timolol D-timolol, halo-peridol and domperidone on rabbit retinal blood flow 26 measured with laser doppler method. Ophthalmic Res 1987; 19:45-8.
- 27 Feke GT, Zuckerman R, Green GJ, Weiter JJ. Response of human retinal blood flow to light and dark. Invest Ophthalmol
- Numan retinal blood how to light and dark. Invest Ophinalmol Vis Sci 1983; 24: 136–41.
 28 Riva CE, Grunwald JE, Petrig BL. Reactivity of the human retinal circulation to darkness: a laser doppler velicometry study. Invest Ophthalmol Vis Sci 1983; 24: 737–40.
 29 Hill DW, Houseman J. Retinal blood flow in the cat following provide of light and darkness. Even By 1085; 41: 210–25.
- periods of light and darkness. Exp Eye Res 1985; 41: 219-25. 30 Riva CE, Petrig BL, Grunwald JE. Near infrared retinal laser
- Avado CE, Feling DE, Grunwald JE. Vean infrared retinat aser doppler velicometry. Laser Ophthalmol 1987; 1: 211–5. Grunwald JE, Riva CE, Sinclair SH, et al. Laser doppler velocimetry study of retinal circulation in diabetes mellitus. Arch Ophthalmol 1986; 104: 991–6.
- Wolff SP. The potential role of oxidative stress in diabetes and its complications: novel implications for theory and therapy. In: Crabbe MJC, ed. Diabetic complications, scientific and clinical aspects. Edinburgh: Churchill Livingstone, 1987: 167 167-220.