Supplementary Figure S1.

Validation and utilization of patient-derived tumor samples for in vitro HTS. A, Culture conditions for solid and hematologic tumor samples from ZERO patients. NEAA = nonessential amino acid; AAS = antibiotic antimycotic solution; bFGF = basic fibroblast growth factor; EGF = epidermal growth factor; PDGF = platelet-derived growth factor; FBS = fetal bovine serum; Pen-Strep = penicillin-streptomycin; MSC-hTERT cells = human telomerase reverse transcriptase-immortalized bone marrow mesenchymal stem cells; GM-CSF = granulocyte-macrophage colony-stimulating factor; ITS = insulin-transferrin-selenium; DMEM/F12 = Dulbecco's Modified Eagle Medium/Nutrient Mixture F-12; RPMI 1640 = Roswell Park Memorial Institute 1640; StemSpan SFEM II = StemSpan Serum-Free Expansion Medium II; IMDM = Iscove Modified Dulbecco Medium; MEM α = Minimum Essential Medium α . (Created with BioRender.com). Examples of a successful validation of an *in vivo* expanded Wilms' tumor (B and C) and unsuccessful validation of an in vitro expanded osteosarcoma (D and E). B, Circos plot showing the genome-wide profile of the original Wilms' tumor detected by WGS. Inner areas show the structural rearrangements and inner to outer circles show the minor allele ploidy, copy number changes (red deletions, green amplifications), regions of loss of heterozygosity, and mutations. C, Copy number profiles of the original Wilms' tumor (detected by WGS; top) versus the matching in vivo expanded sample generated for in vitro HTS (detected by SNP array profiling; bottom). The copy number profile of the original Wilms' tumor was retained upon in vivo expansion. D, Circos plot showing the genome-wide profile of the original osteosarcoma detected by WGS. Inner areas show the structural rearrangements and inner to outer circles show the minor allele ploidy, copy number changes (red deletions, green amplifications), regions of loss of heterozygosity, and mutations. E, Copy number profiles of the original osteosarcoma (detected by WGS; top) versus the matching in vitro expanded sample generated for in vitro HTS (detected by SNP array profiling; bottom). The copy number variations present in the original osteosarcoma disappeared upon in vitro expansion due to the outgrowth of non-neoplastic cells. HTS of this sample was therefore considered failed. F, Time in days required to expand the patient-derived tumor cells for HTS, highlighted by cancer type.



D

0.







Expansion time (days)





Osteosarcoma

Supplementary Figure S2.

Drug response profiles are consistent across freshly dissociated and expanded samples and drug-drug correlations identify effective combination therapies. A, Heatmap of the chemotherapeutic drug response profiles across the 125-sample cohort. Drug response profiles were established by calculating the median AUC Z scores for chemotherapeutic drugs grouped according to the same MOA. Median AUC Z scores are represented by a color scale from blue (resistant) to red (sensitive). MOAs and tumor samples are ordered by unsupervised hierarchal clustering. Top annotations indicate tumor type and type of sample. **B**, Correlation analysis of drug AUC (left) and log₂[IC₅₀] (right) values comparing freshly dissociated cells and *in vivo* expanded cells derived from the same rhabdomyosarcoma tumor. Each dot represents a drug and colors indicate their MOA. C, Pearson's correlation between MEK inhibitor AUC Z scores across the sample cohort: trametinib versus selumetinib (left), cobimetinib versus selumetinib (middle) and trametinib versus cobimetinib (right). **D**, *In vivo* effects of vehicle control (black), irinotecan plus temozolomide (IRN + TMZ; green), alisertib monotherapy (blue) and alisertib in combination with irinotecan plus temozolomide (red) on tumor volume in patient-derived xenograft (PDX) models for a malignant rhabdoid tumor (MRT; left), Wilms' tumor (WT; middle), and neuroectodermal tumor (NET; right). Effects on tumor volume were established during and after a 28-day treatment period. Each line represents an individual mouse. E, Percentages change in body weight for each individual mouse treated with alisertib monotherapy (= A, blue) or alisertib in combination with irinotecan plus temozolomide (= C, red). Grey areas indicate treatment schedules for A and C. Tumor type key: brain tumors (BT), hematologic malignancies (HM), neuroblastoma (NB), sarcoma tumors (SAR), solid other (SO).





Control

Days post treatment initiation

IRN + TMZ

Alisertib

40

Days post treatment initiation

60

0

0

20

Treatment

Alisertib + IRN + TMZ

80

100

Treatment

0

40

80

Days post treatment initiation

120

-10

-20

0

7

21

28

14

Days post treatment initiation

200

160

Supplementary Figure S3.

In vitro HTS on patient-derived tumor samples confirms known associations between drug sensitivity and driver aberrations. A, $Log_2[IC_{50}]$ distribution across all samples for targeted drugs. Drugs are ordered from most (left) to least (right) effective based on lowest median $\log_2[IC_{50}]$ followed by lowest quartile $\log_2[IC_{50}]$ and lowest detected $\log_2[IC_{50}]$, respectively. Colors indicate drug MOAs. B, Volcano plots of tumor type-specific sensitivity and resistance to drugs based on the $\log_2[IC_{50}]$ Z scores for each drug, using the following formula: [average $\log_2[IC_{50}]$ Z score for tumor type of interest] - [average $\log_2[IC_{50}]$ Z score for the remaining cohort]. A difference in average $\log_2[IC_{50}]$ Z score < -0.5 and adjusted P value < 0.01 is considered relatively resistant (blue) and a difference in average $\log_2[IC_{50}] Z$ score > 0.5 and adjusted P value < 0.01 is considered relatively sensitive (red). Only significant targeted drugs are named in the figure. Circle size represents the number of samples within the given tumor type on which the indicated drug has been tested. C, Log₂[IC₅₀] values for TRK inhibitor Larotrectinib sulfate, BRAF inhibitors dabrafenib and vemurafenib, and ALK inhibitors ceritinib, crizotinib, and alectinib with samples harboring an NTRK fusion, BRAF V600E mutation, or ALK aberration (i.e., ALK F1245I mutation or EML4-ALK fusion) highlighted in red, respectively. Arrows indicate the NTRK fusion-positive HGG, BRAF V600E-mutated HGG and ALK F1245I-mutated NB sample with available clinical data shown in E, F, and G, respectively. D, Cell viability curves of larotrectinib, dabrafenib, and crizotinib for two resistant wildtype samples versus samples harboring genomic alterations in NTRK, BRAF, and ALK, respectively. E, Post-contrast T1-weighted MRI scans demonstrating the NTRK fusionpositive high-grade glioma prior to treatment (left) and a complete remission following treatment (right) with larotrectinib. F, Post-contrast T1-weighted MRI scans demonstrating the BRAF V600E-mutated high-grade glioma prior to treatment (left) and a complete remission following treatment (right) with dabrafenib plus trametinib. G, I-123 MIBG scans demonstrating widespread metastatic bone disease in the ALK F1245I-mutated neuroblastoma patient prior to treatment (left) and a partial response following treatment with the ALK inhibitor lorlatinib (right). H, Log₂[IC₅₀] values of MEK inhibitors trametinib (left), cobimetinib (middle), and selumetinib (right) in samples with (colored) versus without (grey) bona fide aberrations in RAS-MAPK signaling. Shapes of the symbols indicate tumor type. Tumor type key: brain tumors (BT), hematologic malignancies (HM), neuroblastoma (NB), sarcoma tumors (SAR), solid other (SO). ***P < 0.001, ns=not significant.



Before Fo treatment tr

Following treatment

Supplementary Figure S4.

Sensitivity to mTORC1 and CDK4/6 inhibitors is not predicted by clinically applied predictive biomarkers. A, AUC and B, log₂[IC₅₀] values of mTORC1 inhibitors everolimus (left), sirolimus (middle) and temsirolimus (right) in samples without (grey) versus with (colored) genomic alterations in PI3K-AKT-mTOR signaling. Shapes of the symbols indicate tumor type. C, In vivo best responses to temsirolimus for an osteosarcoma PDX model harboring a *TSC2* mutation (left) and an Ewing sarcoma PDX model harboring a *PIK3CA* mutation (right) demonstrating average and high in vitro sensitivity to temsirolimus, respectively. Heatmaps of **D**, AUC and **E**, $\log_2[IC_{50}]$ Z scores for CDK4/6 inhibitors abemaciclib mesylate, palbociclib, and ribociclib for the 111-sample cohort with matching drug efficacy and WGS data. AUC and $\log_2[IC_{50}]$ Z scores are represented by a color scale from red (low Z score; sensitive) to blue (high Z score; resistance). Tumor samples are ordered by mean AUC and $\log_2[IC_{50}]$ Z score values, respectively, with lowest values (most sensitive) shown on the left and highest values (most resistant) shown on the right. Top annotation indicates the tumor type. Loss-of-function events in CDKN2A, CDKN2B, and RB1 and gain-of-function events in CDK4 are shown at the bottom. Tumor type key: brain tumors (BT), hematologic malignancies (HM), neuroblastoma (NB), sarcoma tumors (SAR), solid other (SO). ns=not significant.



Supplementary Figure S5.

Integrative analysis links adavosertib efficacy to aberrant cell cycle phase transition and DNA replication. **A**, GSEA enrichment plots for top enriched GO biological processes and hallmark gene sets for the positively correlating genes with adavosertib efficacy. Gene sets enriched among the genes for which log₂[TPM] values negatively correlated with adavosertib AUC *Z* scores (high expression correlates with increased efficacy) with FDR *q* value < 0.01, list > 10%, and NES \leq -2 were selected as most relevant. Genes with $|r| \geq 0.3$ are highlighted in the barcode regions. **B**, Correlation between gene log₂[TPM] values and adavosertib AUC *Z* scores for the 25 top correlating genes to adavosertib response. Blue lines are the regression lines of best fit and the grey shadings represent the 95% confidence intervals.



Supplementary Figure S6.

Neuroblastoma sensitivity to adavosertib is linked to MYCN status. A, Reactome functional interaction (FI) network of the adavosertib target gene WEE1 with the 25 top correlating genes to adavosertib response. WEE1 is highlighted in green, positively correlated genes in blue and negatively correlated genes in red. Genes linking at least 4 out of the 26 genes are indicated in grey, other linker genes in white. CCDC138, MIR17HG, LRRC30, and DSCC1 are not included in the FI network as they are not covered in the reactome database. B, Heatmap of the expression profiles (top), copy number (CN) (middle), and SNV (bottom) for the 25-gene set associated with adavosertib efficacy plus WEE1 and the linker genes identified by network analysis in the 63 pediatric solid tumor cohort with matching RNA-seq and adavosertib efficacy data. Adavosertib AUC Z scores are represented by a color scale from red (low Z score; sensitive) to blue (high Z score; resistance); Gene TPM Z scores are represented by a color scale from blue (low Z score; low expression) to red (high Z score; high expression); Gene CN are represented by a color scale from green (low normalized CN) to orange (high normalized CN). Middle annotations indicate the tumor purity and ploidy as estimated by WGS. For four samples, no matching WGS data was available (tumor purity, tumor ploidy, CN and SNV/SNVg indicated in gray). Right annotation represents the Pearson's correlation coefficient between advosertib AUC Z scores and gene log₂[TPM], corrected copy number for ploidy or SNV/SNVg presence (red; negatively correlated, blue; positively correlated). Tumor samples are ordered by adavosertib AUC Z score values, with lowest values (most sensitive) on the left and highest values (most resistant) on the right. Top annotation indicates the tumor type, and the black arrow indicates the adavosertib-sensitive neuroblastoma sample with a deviated expression of the 25-gene set. C, Adavosertib AUC (left) and log₂[IC₅₀] (right) values in patient-derived neuroblastoma samples with normal MYCN (nMYCN) versus amplified or mutated MYCN (MYCNA/mutated). The MYCN status of two neuroblastoma samples was confirmed by WGS of an earlier received biopsy (hexagon symbols) and the neuroblastoma sample harboring a MYCN activating mutation (P44L) is indicated by the open circle. Horizontal lines indicate median values. D, Higher potency of adavosertib in neuroblastoma cells with normal versus amplified MYCN was confirmed using the publicly available datasets Genomics of Drug Sensitivity in Cancer 2 (GDSC2) and DepMap Public 22Q4 to retrieve matching adavosertib AUC values and MYCN statuses for classical neuroblastoma cell lines. Horizontal lines indicate median values. E, Effects of 24 hours treatment with 1.9 µmol/L adavosertib on G2-M transition in SHEP-21N neuroblastoma cells with MYCN on (-

Doxycycline (Dox); left) versus MYCN off (+Dox; right) in 4 independent experiments. F, Proliferation rate of SHEP-21N neuroblastoma cells with MYCN on (-Dox; green) versus MYCN off (+Dox; red). Viable cells were measured every 24 hours up to 120 hours with 30 technical replicates for each timepoint (56 for t = 0 h). Proliferation rates were established by calculating the fold-change in luminescence value at each timepoint using the following formula: [luminescence value at timepoint of interest] / [luminescence value at 0 h]. Dots indicate the average fold-increases \pm SD. The black dotted line box indicates the time window used to study adavosertib effects on cell viability and G2-M cell cycle arrest, with *a* = start of treatment and *b* = end of treatment. Tumor type key: brain tumors (BT), neuroblastoma (NB), sarcoma tumors (SAR), solid other (SO). SNVg = germline SNV. **P* < 0.05.



Supplementary Figure S7.

Glioma samples with PIK3R1 mutations are sensitive to trametinib. Heatmap of normalized copy number (CN) values for RAS-MAPK (top) and PI3K-AKT-mTOR genes (middle) and somatic SNVs in RAS-MAPK and PI3K-AKT-mTOR genes (bottom) in pediatric solid tumor samples without bona fide driver mutations in RAS-MAPK signaling (N=69). Trametinib AUC Z scores are represented by a color scale from red (low Z score; sensitive) to blue (high Z score; resistance); Normalized CN values of the genes are represented by a color scale from green (low normalized CN) to orange (high normalized CN). Middle annotations indicate the tumor purity and ploidy as estimated by WGS. Right annotations are the Pearson's correlation coefficient between trametinib AUC Z score and normalized copy number values or SNV presence (red; negatively correlated, blue; positively correlated). Tumor samples are ordered by trametinib AUC Z score value, with lowest values (most sensitive) on the left and highest values (most resistant) on the right. Top annotation indicates the tumor type. Tumor type key: brain tumors (BT), neuroblastoma (NB), sarcoma tumors (SAR), solid other (SO). Cancer subtype key: diffuse midline glioma (DMG), high-grade glioma (HGG), medulloblastoma (MB), Ewing's sarcoma (ES), osteosarcoma (OS), rhabdomyosarcoma (RMS), adrenocortical carcinoma (ACC), malignant rhabdoid tumor (MRT), and Wilms' tumor (WT).



