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Appendix Figure S1 - Association of DCAP-1-foci with mitochondria is age-dependent. A Representative images showing the localization of DCAP-1-foci relative to mitochondria in young versus older animals (red: DCAP-1, green: DiOC6(3), a mitochondrial-specific dye; n=2 independent experiments). B Quantification of the distances between DCAP-1-foci and mitochondria in young versus older animals (n=2 independent experiments with at least 40 animals/experiment; **P<0.01; two-tailed unpaired t-test). Data information: Scale bars, 10µm. Images were acquired using a X63 objective lens. Error bars denote SEM.



DCAP-2

DiOC6(3)

MERGE

С









Appendix Fig S2

Appendix Figure S2 - Association of DCAP-2-foci with mitochondria is age-dependent. Representative images showing the localization of DCAP-2-foci relative to mitochondria in **A**, young versus **B**, old animals (red: DCAP-2, green: DiOC6(3), a mitochondrial-specific dye; n=2 independent experiments). **C**, Quantification of the distances between DCAP-2-foci and mitochondria in young versus old animals (n=2 independent experiments with at least 40 animals/experiment; ****P<0.0001; two-tailed unpaired t-test). **D**, Representative image from Immunoblot analysis showing that DCAP-2 is less associated with mitochondria during ageing and **E**, Respective quantification showing DCAP-2 levels normalized to ATP-1 (n=3 independent experiments, **P<0.01; two-tailed unpaired t-test).

Data information: Scale bars, 10µm. Images were acquired using a X63 objective lens. Error bars denote SEM.









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Appendix Fig S3

Appendix Figure S3 - Association of CCF-1-foci with mitochondria is age-dependent. Representative images showing the localization of CCF-1-foci relative to mitochondria in **A**, young (1-day-old) versus **B**, old (7-day-old) animals (image in control is reused in Fig EV2E as conditions indicated in Appendix Fig S3A & Fig EV2E are part of the same experimental setup) (green: CCF-1, red: Mitotracker Deep Red FM, a mitochondrial-specific dye; n=2 independent experiments). **C**, Quantification of the distances CCF-1-foci obtain from mitochondria in young versus old animals (n=2 independent experiments with at least 40 animals/experiment; ****P<0.0001; two-tailed unpaired t-test). **D**, Representative image from Immunoblot analysis showing that CCF-1 is less associated with mitochondria in older animals compared to younger worms and **E**, Respective quantification of CCF-1 levels normalized to ATP-1 (n=3 independent experiments; *P<0.05; two-tailed unpaired t-test). Data information: Scale bars, 10µm. Images were acquired using a X63 objective lens. Error bars denote SEM.





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Appendix Figure S4 - NTL-2 and EDC-3 are coordinately associated with mitochondria in an age-dependent manner. Representative images showing the localization of NTL-2-foci and EDC-3-foci relative to mitochondria in **A**, young versus older animals (green: NTL-2, red: EDC-3, purple: Mitotracker Deep Red FM, a mitochondrial-specific dye). **B**, Quantification of the distances NTL-2- and **C**, EDC-3-foci obtain from mitochondria in young and older animals (n=3 independent experiments with at least 30 animals/experiment; ****P<0.0001; two-tailed unpaired t-test).

Data information: Scale bars, 20 µm. Images were acquired using a X63 objective lens. Error bars denote SEM.



Appendix Figure S5 - NTL-2 and EDC-3 become tightly associated with mitochondria upon *cyc-1* **genetic inhibition. A**, Representative images showing the localization of EDC-3-foci relative to mitochondria upon genetic inhibition of *cyc-1* (green: EDC-3, red: Mitotracker Deep Red FM, a mitochondria-specific dye). **B**, Quantification of the distances between EDC-3-foci and mitochondria upon genetic inhibition of *cyc-1* (n=3 independent experiments with at least 30 animals/experiment; ****P<0.0001; unpaired two-tailed t test). **C**, Representative images showing the localization of NTL-2-foci relative to mitochondria upon genetic inhibition of *cyc-1* (green: NTL-2, red: Mitotracker Deep Red FM, a mitochondria-specific dye). **D**, Quantification of the distances between NTL-2-foci and mitochondria upon genetic inhibition of *cyc-1* (n=3 independent experiments with at least 30 animals/experiment; ****P<0.0001; unpaired two-tailed t test). **C**, Representative images showing the localization of NTL-2-foci relative to mitochondria upon genetic inhibition of *cyc-1* (green: NTL-2, red: Mitotracker Deep Red FM, a mitochondria-specific dye). **D**, Quantification of the distances between NTL-2-foci and mitochondria upon genetic inhibition of *cyc-1* (n=3 independent experiments with at least 30 animals/experiment; ****P<0.0001; two-tailed unpaired *t*-test).

Data information: Scale bars, 20 µm. Images were acquired using a X63 objective lens. Error bars denote SEM.





Control

daf-2(RNAi)

nhr-49(RNAi)





EDC-3

В











MERGE



Ε

F

Appendix Fig S6

Appendix Figure S6 - Association of NTL-2- and EDC-3-foci with mitochondria in long- and short- lived animals deviates from their wild-type counterparts. **A**, Representative images showing the localization of NTL-2- and EDC-3-foci relative to mitochondria upon genetic inhibition of either *daf-2* or *nhr-49* (green: NTL-2, red: EDC-3, purple: Mitotracker Deep Red FM, a mitochondria-specific dye). **B**, Quantification of the distances NTL-2- and **C**, EDC-3-foci obtain from mitochondria upon genetic inhibition of either *daf-2* or *nhr-49* (n=3 independent experiments with at least 45 animals per experiment; *P<0.05, ***P<0.001; one-way analysis of variance (ANOVA)). **D**, Immunoblot analysis in isolated mitochondria of 1-day-old animals co-expressing EDC-3 and NTL-2. **E**, Quantification of EDC-3 and **F**, NTL-2 amount in the mitochondrial fraction obtained from animals subjected to the indicated genetic inhibitions compared to controls (n=4 independent experiments; *P<0.05, ***P<0.01; Welch's one-way analysis of variance (ANOVA) followed by Dunnett's T3 multiple comparisons test). All experiments in this Figure have been performed in animals that co-express NTL-2 and EDC-3.

Data information: Scale bars, 20 µm. Images were acquired using a X63 objective lens. Error bars denote SEM.

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Appendix Figure S7 - Components of the CCR4-NOT and the mRNA degradation complexes alter mtROS production. A, Mitochondrial ROS production is elevated in animals subjected to *edc-3*, *xrn-1*, *ccf-1* or *let-711* RNAi and **B**, Respective quantification is shown (n=3 independent experiments with at least 182 animals/experiment; *P<0.05, ****P< 0.0001; one-way analysis of variance (ANOVA)). Data information: Scale bars, 40µm. Images were acquired using a X5 objective lens. Error bars denote SEM.

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Appendix Figure S8 - Components of the CCR4-NOT and the mRNA degradation complexes alter mitochondrial membrane potential ($\Delta \psi$). A, Representative images showing fluorescence intensity of the mitochondrial membrane potential dye DiOC6(3) in 1-day-old animals fed bacteria expressing control, *edc-3* or *xrn-1* RNAi and **B**, Respective quantification (n=3 independent experiments with at least 97 animals/experiment; *P< 0.05, ***P< 0.001; one-way analysis of variance (ANOVA)). **C**, Representative images showing fluorescence intensity of the mitochondrial membrane potential dye DiOC6(3) in 1-day-old animals fed bacteria expressing control, *ccf-1* or *let-711* RNAi and **D**, Respective quantification (n=3 independent experiments with at least 124 animals/experiment; **P< 0.01, ***P< 0.001; one-way analysis of variance (ANOVA)).

Data information: Scale bars, 40µm. Images were acquired using a X4 objective lens. Error bars denote SEM.



Appendix Figure S9 - Components of the CCR4-NOT and the mRNA degradation complexes alter mitochondrial ATP levels. A, Representative fluorescent images showing mitochondrial ATP levels in young adult animals fed with bacteria expressing control, *edc-3*, *dcap-2* and *xrn-1* RNAis. **B**, Corresponding quantification is shown (n=3 independent experiments with at least 120 animals/experiment; ***P< 0.001; one-way analysis of variance (ANOVA)). **C**, Representative fluorescent images showing mitochondrial ATP levels in young adult animals fed with bacteria expressing control, *ntl-2*, *ccf-1* and *let-711* RNAis. **D**, Corresponding quantification is shown (n=3 independent experiment; ***P< 0.001; one-way analysis of variance (ANOVA)). BioTracker ATP-Red was used for *in vivo* monitoring of mitochondrial ATP levels.

Data information: Scale bars, 20µm. Images were acquired using a X4 objective lens. Error bars denote SEM.

Sodium Azide Control dcap-2(RNAi) edc-3(RNAi) xrn-1(RNAi) mev-1(kn-1)



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Appendix Figure S10 - Components of the mRNA degradation complex alter basal and maximal oxygen consumption rates (OCRs). A, Normalized OCRs profile plot versus time. **B**, Basal OCR per worm was measured at time points 3-5; before CCCP injection. **C**, Maximal OCR per worm was measured at time points 7-9 between CCCP and sodium azide injections. Seahorse XF analyzer was used to measure oxygen consumption rate (OCR) following a sequential injection of CCCP and sodium azide at the indicated time points. (n=2 independent experiments; **P< 0.01, ***P< 0.001, ****P<0.0001; one-way analysis of variance (ANOVA) followed by Dunnett's multiple comparison test). Data information: Error bars denote SEM.



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Appendix Figure S11 - Components of the CCR4-NOT and the mRNA degradation complexes alter intestinal mitochondrial abundance.

A, Representative images showing the effect of the indicated genetic inhibitions on the abundance of intestinal cell mitochondria in 1-day-old nematodes. **B**, Respective quantification is shown. (n=3 independent experiments with at least 194 animals/experiment; *P< 0.05, **P< 0.01, ***P< 0.001; one-way analysis of variance (ANOVA)).

Data information: Scale bars, 40µm. Images were acquired using a X4 objective lens. Error bars denote SEM.



p_{gst-4}GFP



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Control

dcap-2(RNAi) (1/2)



ntl-2(RNAi) (1/2)



ntl-2;skn-1(RNAi)

skn-1(RNAi) (1/2)

dcap-2;skn-1(RNAi)

С



Appendix Figure S12 - Components of the CCR4-NOT and the mRNA degradation complexes alter *gst-4* promoter expression in a SKN-1-dependent manner. **A**, Representative images corresponding to quantification shown in Fig 3C, Scale bars, 50µm. Images were acquired using a X5 objective lens. **B**, Representative images showing the expression levels of p_{gst-4} GFP in 1-day-old animals upon the indicated RNAi treatments diluted 1:1 and **C**, Respective quantification (n=3 independent experiments with at least 244 animals/experiment; **P<0,01, ***P< 0.001; one-way analysis of variance (ANOVA)).

Data information: Scale bars, 20µm. Images were acquired using X5 and X4 objective lenses. Error bars denote SEM.

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p_{gst-4}GFP

Control	, j	dcap-2(RNAi)	edc-3(RNAi)	xrn-1(RNAi)	ntl-2(RNAi)	ccf-1(RNAi)	let-711(RNAi)
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Appendix Figure S13 - Perturbation of the CCR4-NOT and the mRNA degradation complexes alters *gst-4* promoter expression. A, Measurement of the promoter activity of the SKN-1 target gene *gst-4* in 1-day-old animals upon the indicated RNAi treatments and **B**, Respective quantification is shown (n=3 independent experiments with at least 223 animals/experiment; **P<0,01, ***P< 0.001; one-way analysis of variance (ANOVA)).

Data information: Scale bars, 20µm. Images were acquired using a X4 objective lens. Error bars denote SEM.



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Appendix Figure S14 - Components of the CCR4-NOT and the mRNA degradation complexes localize in distinct foci. **A**, (top) Representative images showing the localization of CCF-1- (green) and DCAP-2- (red) foci in transgenic animals that co-express them, (middle) representative images showing the localization of NTL-2- (green) and DCAP-2- (red) labeled foci in transgenic animals that co-express them and (bottom) representative images showing the localization of EDC-3- (green) and DCAP-2- (red) labeled foci in transgenic animals that co-express them. **B**, Respective fluorescent intensity graphs of the foci marked with the yellow line in panel A. **C**, (top) Pearson's correlation coefficient values after measuring the correlation between CCF-1- and DCAP-2-foci, (middle) Pearson's correlation coefficient values after measuring the correlation between DCAP-2- and NTL-2-foci and (bottom) Pearson's correlation coefficient values after measuring the correlation between DCAP-2- and EDC-3-foci. (n=2 independent experiments with at least 15 animals for each strain/experiment).

Data information: Scale bar, 20µm. Images were acquired using the X63 lens. Error bars denote SEM.









Appendix Figure S15 - Genetic inhibition of the one type of body triggers an elevation in the protein abundance of components of the other type. A, Representative images showing DCAP-2 expression in the posterior half of young adult animals upon genetic inhibition of storage body components. Scale bar, 100µm. Images were acquired using the X40 lens. **B**, Quantification of DCAP-2 levels in whole animals (n=3 independent experiments with at least 86 animals/experiment; *P< 0.05, **P< 0.01, one-way analysis of variance (ANOVA)). **C**, Representative images showing NTL-2 levels in young adult animals upon genetic inhibition of degradation body components and **D**, Respective quantification is shown (n=3 independent experiments with at least 50 animals/experiment; ***P< 0.001; one-way analysis of variance (ANOVA)). Data information: Scale bar, 40µm. Images were acquired using the X4 lens. Error bars denote SEM.



Appendix Figure S16 - Quantification of total protein content in 1-day-old whole worm lysates under control conditions and upon inhibition of the indicated genes. (n=3 independent experiments with at least 400 animals/experiment; ***P<0.001; one-way analysis of variance (ANOVA)).

Data information: Error bars denote SEM.



0.0

2 3

1

5 6

4



3

4

2

5 6

0.0

0.0

1 2 3 4 5 6

1: unc-119(*ed3)III;Ex*[p_{ntl-2}NTL-2::GFP;UNC-119(+)] day 1

3: unc-119(ed3)III;Ex[p_{ntl-2}NTL-2::GFP;UNC-119(+)] day 7

2: unc-119(ed3)III day 1

2 3 4 5 5

1

0.0

4: unc-119(ed3)// day 7

5: unc-119(ed3)III;Ex[p_{ntl-2}NTL-2::GFP;UNC-119(+)] day1+CCCP

6: unc-119(ed3)III;Ex[p_{ntl-2}NTL-2::GFP;UNC-119(+)] day 1+ dcap-2(RNAi)

Appendix Figure S17 - NTL-2 preferentially binds MTPTs and their association is affected by age, mitochondrial stress and genetic inhibition of *dcap-2*. **A**, Analysis of expression of mRNAs encoding nuclear (FIB-1, NPP-22), ER (SPCS-1) and cytoplasmic (RHI-1) proteins by real time quantitative RT-PCR following RNA immunoprecipitation (RIP) in anti-GFP isolates from NTL-2::GFP transgenic animals, *unc-119(ed3)III* and p_{*gst-4*}GFP counterparts (used as negative controls) (n=2 independent experiments; ***P< 0.001; one-way analysis of variance (ANOVA)). **B**, Analysis of expression of select MTPTs by real time quantitative RT-PCR following RNA immunoprecipitation (RIP) in anti-GFP isolates from NTL-2::GFP transgenic and *unc-119(ed3)III* animals during ageing, upon CCCP treatment and genetic inhibition of *dcap-2* (n=2 independent experiments; ***P< 0.001; one-way analysis of variance (ANOVA)).

Data information: Error bars denote SEM. Numbers at the x axis represent sample names described after the last bar plot in the right part of each figure panel.



Appendix Figure S18 - Association of storage and degradation bodies with mitochondria is oppositely affected by genetic inhibition of *dcap-2*. **A**, Representative images showing the localization of NTL-2/storage bodies relative to mitochondria upon genetic inhibition of *dcap-2* (green: NTL-2, red: Mitotracker Deep Red FM, a mitochondrial specific dye). **B**, Quantification of the distances between NTL-2/storage bodies and mitochondria upon genetic inhibition of *dcap-2* (n=3 independent experiments with at least 30 animals/experiment; **P<0.01; two-tailed unpaired *t*-test). **C**, Representative images showing the localization of EDC-3/degradation bodies relative to mitochondria upon genetic inhibition of *dcap-2* (n=3 independent experiments with at least 30 animals/experiment; **P<0.01; two-tailed unpaired *t*-test). **C**, Representative images showing the localization of EDC-3/degradation bodies relative to mitochondria upon genetic inhibition of *dcap-2* (n=3 independent experiments with at least 30 animals/experiment; **P<0.01; two-tailed unpaired *t*-test). **C**, Representative images showing the localization of EDC-3/degradation bodies relative to mitochondria upon genetic inhibition of *dcap-2* (n=3 independent experiments with at least 30 animals per experiment; ***P<0.0001; two-tailed unpaired *t*-test). **E**, Representative image from Immunoblot analysis showing that EDC-3 is less associated with mitochondria upon genetic inhibition of *dcap-2* and **F**, Respective quantification of EDC-3 levels normalized to CTC-1 is shown (n= 3 independent experiments; ****P<0.0001; two-tailed unpaired *t*-test).

Data information: Scale bar, 20µm. Images were acquired using the X63 lens. Error bars denote SEM.



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Appendix Figure S19 - TOMM-20 levels increase upon genetic inhibition of storage body components. A, Genetic inhibition of either *ntl-2* or *let-711* increases the expression levels of TOMM-20::mRFP expressed in body wall muscle cells. B, Genetic inhibition of either *ntl-2* or *let-711* increases the expression levels of TOMM-20::mKate2 ubiquitously expressed in all animal tissues. *tomm-20(RNAi)* is used as a control. (n=2 independent experiments with at least 141 animals/experiment (for A) and 147 animals/experiment (for B); ***P< 0.001; one-way analysis of variance (ANOVA)).

Data information: Error bars denote SEM.



Appendix Figure S20 - Knockdown of ntl-2 increases MTPT protein levels in a TOMM-20- and AKAP-1-dependent manner. A, (left) Immunoblot analysis in 1-day-old whole animal extracts showing the protein levels of ATP-1 in control conditions and upon the indicated genetic inhibitions and (right) respective quantification (n= at least 3 independent experiments; *P<0.05; one-way analysis of variance (ANOVA) followed by Dunnett's T3 multiple comparisons test). B, (left) Immunoblot analysis in 1-day-old whole animal extracts showing the protein levels of F46B6.6 in control conditions and upon the indicated genetic inhibitions and (right) respective guantification (n= at least 3 independent experiments; *P<0.05; one-way analysis of variance (ANOVA) followed by Dunnett's T3 multiple comparisons test). C, (left) Representative images showing F46B6.6 protein levels in 1-day-old animals fed bacteria expressing control or ntl-2 RNAi. Scale bars, 40µm; and respective quantification is shown in right (n=2 independent experiments with at least 40 animals/experiment; ****P< 0.05; two-tailed unpaired t-test). D, (left) Immunoblot analysis in 1-day-old whole animal extracts showing the protein levels of MRPL-13 in control conditions and upon the indicated genetic inhibitions and respective quantification is shown in right (n= 4 independent experiments; *P<0.05; Welch's one-way analysis of variance (ANOVA) followed by Dunnett's T3 multiple comparisons test). E. (left) Immunoblot analysis in 1-day-old whole animal extracts showing the protein levels of T20H4.5 in control conditions and upon the indicated genetic inhibitions (n= at least 3 independent experiments; *P<0.05; Welch's one-way analysis of variance (ANOVA) followed by Dunnett's T3 multiple comparisons test).

Data information: Error bars denote SEM.



Appendix Figure S21 - Balance of storage and degradation body components regulates lifespan. A, Knockdown of storage and degradation body components oppositely affect longevity. B, Knockdown of degradation body components further extends the lifespan of CCF-1 overexpressing animals. C, Knockdown of degradation body components extends the lifespan of NTL-2 overexpressing animals. D, Knockdown of storage body components shortens the lifespan of transgenic animals overexpressing DCAP-2. E, Knockdown of storage body components shortens the lifespan of transgenic animals overexpressing DCAP-2. E, Knockdown of storage body components shortens the lifespan of transgenic animals overexpressing DCAP-2. E, Knockdown of storage body components shortens the lifespan of transgenic animals overexpressing DCAP-2. E, Knockdown of storage body components shortens the lifespan of transgenic animals overexpressing DCAP-2. E, Knockdown of storage body components shortens the lifespan of transgenic animals overexpressing DCAP-2. E, Knockdown of storage body components shortens the lifespan of transgenic animals overexpressing DCAP-2. E, Knockdown of storage body components shortens the lifespan of transgenic animals overexpressing EDC-3.

Data information: Lifespan assays were performed at 20°C; detailed lifespan data are given in Table EV4.





- Controlxrn-1(RNAi)
- ntl-2;xrn-1(RNAi)
- ntl-2(RNAi)

Appendix Figure S22 - Effects of concomitant knockdown of storage and degradation body components on longevity. **A**, Concomitant knockdown of *let-711* and *dcap-2* reverses *dcap-2* longevity. **B**, Concomitant knockdown of *ntl-2* and *dcap-2* reverses *dcap-2* longevity. **C**, Concomitant knockdown of *edc-3* and *let-711* reverses *edc-3* longevity. **D**, Concomitant knockdown of *edc-3* and *ntl-2* reverses *edc-3* longevity. **E**, Concomitant knockdown of *ntl-2* and *xrn-1* reverses *xrn-1* longevity.

Data information: Lifespan assays were performed at 20°C; detailed lifespan data are given in Table EV4.



Appendix Figure S23 - *tomm-20* knockdown extends lifespan.

Data information: Lifespan assays were performed at 20°C; detailed lifespan data are given in Table EV4.