Supplementary Material

Functional characterization of VirB/VirD4 and Icm/Dot type IV secretion systems from the plant-pathogenic bacterium *Xanthomonas euvesicatoria*

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Strain or plasmid	Relevant characteristics ¹	Reference or source
Xanthomonas euvesicatoria		
85-10	Pepper-race 2: wild type: Rif ^r	Canteros, 1990;
		Kousik and Ritchie, 1998
85-10∆ <i>virB4</i>	virB4 deletion mutant of strain 85-10	This study
85-10∆ <i>virD4</i>	virD4 deletion mutant of strain 85-10	This study
$85-10\Delta virG$	<i>virG</i> deletion mutant of strain 85-10	This study
85-10Δ <i>virA</i>	virA deletion mutant of strain 85-10	This study
$85-10\Delta virB11\Delta virB1$	virB11 and virB1 deletion mutant of strain 85-10	This study
$85-10\Delta virD4\Delta virB11\Delta virB1$	virD4, virB11 and virB1 deletion mutant of strain 85-10	This study
85-10	Derivative of strain 85-10 deleted in <i>virD4</i> , <i>virB11</i> ,	This study
$\Delta virD4\Delta virB11\Delta virB1\Delta virB4$	virB1 and virB4	
85-10	Derivative of strain 85-10 deleted in virB6, virB8, virB9	This study
$\Delta virB6\Delta virB8\Delta virB9\Delta virB10$	and virB10	
85-10	Derivative of strain 85-10 deleted in virB6, virB8, virB9,	This study
$\Delta virB6\Delta virB8\Delta virB9\Delta virB10$	virB10, icmE, dotC and dotB	
∆icmE∆dotCB		
$85-10\Delta dotCB$	Derivative of strain 85-10 deleted in <i>dotC</i> and <i>dotB</i>	This study
85-10∆ <i>icmE</i>	<i>icmE</i> deletion mutant of strain 85-10	This study
$85-10\Delta dotCB\Delta icmE$	Derivative of strain 85-10 deleted in <i>dotC</i> , <i>dotB</i> and <i>icmE</i>	This study
$85-10\Delta virB4\Delta icmE$	Derivative of strain 85-10 deleted in virB4 and icmE	This study
$85-10\Delta dotCB\Delta icmE\Delta virB4$	Derivative of strain 85-10 deleted in <i>dotC</i> , <i>dotB</i> , <i>icmE</i> and <i>virB4</i>	This study
85-10:: <i>kan^R</i>	Insertion mutant of 85-10 carrying <i>gfp</i> and the kanamycin resistance gene under control of the	This study
	gentamycin resistance promoter of pBRM on pXCV183; Rif ^r , Km ^r	
85-10Δ <i>xopQ</i>	<i>xopQ</i> deletion mutant of strain 85-10	Adlung et al., 2016
82-8	Pepper-race 1; wild type; Rif ^r	Bonas et al., 1989
85-10:: <i>gent</i> ^R	Insertion mutant of 85-10 carrying the gentamycin resistance gene under control of the <i>lac</i> promoter in the	This study
	hpaFG region; Rif ^r , Gm ^r	
$85-10\Delta virD4::gent^R$	Insertion mutant of $85-10\Delta virD4$ carrying the	This study
	gentamycin resistance gene under control of the <i>lac</i> promoter in the <i>hpaFG</i> region; Rif ^t , Gm ^r	
85-10 $\Delta virB4::gent^R$	Insertion mutant of $85-10\Delta virB4$ carrying the	This study
	gentamycin resistance gene under control of the lac	
	promoter in the <i>hpaFG</i> region; Rif ^r , Gm ^r	
E. coli		
OneShot®TOP10	F^- mcrA Δ(mrr-hsdRMS-mcrBC) φ80lacZΔM15 ΔlacX74 recA1 araΔ139 Δ(ara-leu)7697 galU galK	Invitrogen
Dh5a) nir	$\frac{1}{1} \frac{1}{1} \frac{1}$	Manard at al 1002
	$\frac{F}{F} = \frac{F}{recA} \frac{hsaR1}{(f_k, m_k)} \frac{\phi \delta \delta a a a CZ}{\phi \delta a a a CZ} \frac{DM13}{[\lambda p lr]}$	Strate con c
BL21(DE3) Besette DE2 #LvieS	F Ompt hsa520(Fb Fb) gat	Stratagene
Kosella DE5 pLyss	E^{-} trad D^{2} by $A + B + lasta A(las 7)M15/A(lastron AB)$	Vaniaah Daman at al
	glnV44 e14- gyrA96 recA1 relA1 endA1 thi hsdR17	1985
BTH101	F ⁻ , cya-99, araD139, galE15, galK16, rpsL1 (StrR), hsdR2, mcrA1, mcrB1	Euromedex; Battesti and Bouveret, 2012
Plasmids		
pRK2013	ColE1 replicon, TraRK ⁺ Mob ⁺ ; Km ^r	Figurski and Helinski, 1979
pBBR1MCS-5	Broad-host-range vector; <i>lac</i> promoter; Gm ^r	Kovach et al., 1995
pBRM	Golden Gate-compatible derivative of pBBR1MCS-5	Szczesny et al., 2010
pBRM-P	Golden Gate-compatible derivative of pBBR1MCS-5	Szczesny et al., 2010
	without promoter	

Table S1: Bacterial strains and plasmids used in this study

pBRM-P(stop)	Golden Gate-compatible derivative of pBRM-P with a stop codon in the 3' region	Hausner et al., 2013
pBRNM	Golden Gate-compatible derivative of pBBR1MCS-5 with 3 x c-Myc-encoding sequence in the 5' region of the inserted gene and a stop codon in the 3' region; Gm ¹	This study
pBRM-P+T	Golden Gate-compatible derivative of pBBR1MCS-5 without promoter upstream of the cloning sites and an <i>rrnb T1</i> terminator	Helm and Schmidtke, unpublished
pDSK602	Broad-host-range vector containing a <i>lacUV5</i> promoter; Sm ^r	Murillo et al., 1994
pICH41021	Derivative of pUC19 with mutated <i>Bsal</i> site; Ap ^r	Hausner et al., 2019
pUC57	Classical cloning vector with mutated <i>BsaI</i> site; Ap ^r	Thermo Fisher Scientific
pLAND	Derivative of pOK1 containing fragments of the <i>hpaFG</i> region, the <i>lac</i> promoter and a triple <i>c-Myc</i> epitope-encoding sequence; Sm ^r	Lorenz et al., 2012
pLAND-P	Derivative of pOK1 containing fragments of the <i>hpaFG</i> region, a triple <i>c-Myc</i> epitope-encoding sequence and lacking a promoter upstream of the <i>Bsa</i> I sites; Sm ^r	Lorenz et al., 2012
pOGG2	Golden Gate-compatible derivative of suicide vector pOK1, <i>sacB sacQ mobRK2 oriR6K</i> ; Sm ^r	Schulze et al., 2012
pAGM9121	pUC19-derived vector, <i>lacZa</i> fragment flanked by <i>Bpil</i> sites (CTCA/CGAG); GGAG/CGCT <i>Bsal</i> fusion sites; Sm ^r	Addgene #51833; Weber et al., 2011
pICH47732	Derived from pUC19, level 1 for promoters; <i>lacZa</i> fragment flanked by <i>BsaI</i> sites (GGAG/CGCT); TGCC/GCAA <i>BpiI</i> fusion sites; Ap ^r	Addgene #48000; Weber et al., 2011
pICH79631	Derived from pICH50251, containing dTALE-2 with stop-codon for position 2 in level M; GCAA/ACTA <i>BpiI</i> fusion sites; Ap ^r	Gift from S. Marillonnet
pICH50122	Derived from pUC19, end linker for position 3 contains a terminator sequence; ACTA/ GGGA <i>BpiI</i> fusion sites; Ap ^r	Gift from S. Marillonnet
pICH77739	Level 2 vector derived from pBIN19, RK2 ori, $lacZ\alpha$ fragment flanked by <i>BpiI</i> sites; Km ^r	Weber et al., 2011
pKT25 _{GG}	Golden Gate-compatible derivative of pKT25 encoding the T25 fragment downstream of a <i>lac</i> promoter and in frame with a C-terminal FLAG epitope-encoding sequence; contains <i>lacP-eforRed</i> flanked by <i>Bsa</i> I sites downstream of the <i>T25-FLAG</i> fragment; Km ^r	Otten and Büttner, 2021
pKNT25 _{GG}	Golden Gate-compatible derivative of pKNT25, encodes the T25 fragment in frame with an N-terminal FLAG epitope-encoding sequence downstream of the <i>lac</i> promoter, contains <i>lacP-eforRed</i> flanked by <i>Bsa</i> I sites upstream of the <i>T25-FLAG</i> fragment; Km ^r	Otten and Büttner, 2021
pUT18 _{GG}	Golden Gate-compatible derivative of pUT18 containing <i>lacP-eforRed</i> flanked by <i>Bsa</i> I sites upstream of the <i>FLAG-T18</i> fragment; Gm ^R	Otten and Büttner, 2021
pUT18C _{GG}	Golden Gate-compatible derivative of pUT18C containing <i>lacP-eforRed</i> flanked by <i>Bsa</i> I sites downstream of the <i>T18-FLAG</i> fragment; Gm ^R	Otten and Büttner, 2021
pICH41021-virD4 ₁₋₉₅₀	Derivative of pICH41021 containing bp 1 - 950 of <i>virD4</i> flanked by <i>BsaI</i> sites with TATG and GCCT overhangs	This study
pICH41021-virD4951-1450	Derivative of pICH41021 containing bp 951 - 1450 of <i>virD4</i> flanked by <i>BsaI</i> sites with GCCT and GTCA overhangs	This study
pICH41021-virD4 ₁₄₅₁₋₁₆₇₁	Derivative of pICH41021 containing bp 1451 – 1671 of <i>virD4</i> flanked by <i>BsaI</i> sites with GTCA and GGTG overhangs	This study

pUC57-XCV4361prom	Derivative of pUC57 containing the putative promoter of XCV4361 flanked by <i>BsaI</i> sites with ATTC and TATG overhangs	This study
pICH41021-tral	Derivative of pICH41021 containing <i>tral</i> flanked by <i>Bsal</i> sites with TATG and GGTG fusion sites	This study
pUC57-XCV0332	Derivative of pUC57 containing XCV0332 flanked by <i>Bsal</i> sites with TATG and GGTG fusion sites	This study
pICH41021-XCV3751	Derivative of pICH41021 containing <i>XCV3751</i> flanked by <i>BsaI</i> sites with TATG and GGTG fusion sites	This study
pUC57-XCV1120	Derivative of pUC57 containing <i>XCV1120</i> flanked by <i>Bsal</i> sites with TATG and GGTG fusion sites	This study
pICH41021-virB10	Derivative of pICH41021 containing <i>virB10</i> flanked by <i>Bsa1</i> sites with TATG and GGTG fusion sites	This study
pICH41021-virB11	Derivative of pICH41021 containing <i>virB11</i> flanked by <i>Bsa1</i> sites with TATG and GGTG fusion sites	This study
pICH41021- virB7	Derivative of pICH41021 containing <i>virB7</i> flanked by <i>Bsal</i> sites with TATG and GGTG fusion sites	This study
pICH41021-dotB	Derivative of pICH41021 containing <i>dotB</i> flanked by <i>Bsal</i> sites with TATG and GGTG fusion sites	This study
pICH41021-dotO ₁₋₁₀₉	Derivative of pICH41021 containing bp 1 - 109 of <i>dotO</i> flanked by <i>BsaI</i> sites with TATG and AGAC overhangs	This study
pICH41021-dotO ₁₁₀₋₃₀₄₈	Derivative of pICH41021 containing bp 110 - 3048 of <i>dotO</i> flanked by <i>BsaI</i> sites with AGAC and GGTG overhangs	This study
pICH41021-icmE ₁₋₆₄₅	Derivative of pICH41021 containing bp 1 - 645 of <i>icmE</i> flanked by <i>BsaI</i> sites with TATG and ACGA overhangs	This study
pICH41021-icmE646-796	Derivative of pICH41021 containing bp 646 - 796 of <i>icmE</i> flanked by <i>BsaI</i> sites with ACGA and GACG overhangs	This study
pICH41021-icmE ₇₉₇₋₁₂₄₈	Derivative of pICH41021 containing bp 797 - 1248 of <i>icmE</i> flanked by <i>BsaI</i> sites with GACG and GGTG overhangs	This study
pICH41021-dotL ₁₋₁₄₆₀	Derivative of pICH41021 containing bp 1 - 1460 of <i>dotL</i> flanked by <i>Bsal</i> sites with TATG and ACCG overhangs	This study
pICH41021-dotL ₁₄₆₁₋₂₀₀₁	Derivative of pICH41021 containing bp 1461 - 2001 of <i>dotL</i> flanked by <i>BsaI</i> sites with ACCG and GCGG overhangs	This study
pICH41021-dotL2002-2568	Derivative of pICH41021 containing bp 2002 - 2568 of <i>dotL</i> flanked by <i>Bsal</i> sites with GCGG and GGTG overhangs	This study
pXCV38 ₈₂₋₈ ∷spec ^R	Derivative of the native plasmid pXCV38 from <i>X.</i> <i>euvesicatoria</i> strain 82-8 containing a spectinomycin resistance gene inserted in <i>avrBs3</i>	Bonas et al., 1989

Expression constructs and sur	cide vectors	
pDSF300	Derivative of pDSK602 encoding AvrBs3	Van den Ackerveken
		et al., 1996
pBRNM-XCV3751	Derivative of pBRNM encoding c-Myc-XCV3751 under	This study
	control of the <i>lac</i> promoter	
pBRNM-XCV1120	Derivative of pBRNM encoding c-Myc-XCV1120 under	This study
	control of the <i>lac</i> promoter	
pBRNM-XCV0332	Derivative of pBRNM encoding c-Myc-XCV0332 under	This study
	control of the <i>lac</i> promoter	
pBRM-P(stop)-XCV4361prom-	Derivative of pBRM-P(stop) encoding c-Myc-TraI under	This study
c-Myc-traI	control of the putative XCV4361 promoter	
pLAND-virB4	Derivative of pLAND containing virB4	This study
pLAND-P-XCV0160prom-	Derivative of pLAND-P containing icmE under the	This study
icmE	control of the XCV0160 promoter	
pLAND-gent ^R	Derivative of pLAND containing the gentamycin	This study
	resistance gene cassette from vector pBRM	-

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pOGG2-kan ^R gfp(pXCV183)	Derivative of pOGG2 carrying the flanking sequences of	This study
	nucleotide position 2831 in pXCV183 and containing the	
	kanamycin resistance and the <i>gfp</i> gene	
pOGG2-∆virB4	Derivative of pOGG2 carrying flanking sequences of $virB4$	This study
pOGG2-AvirD4	Derivative of pOGG2 carrying flanking sequences of	This study
poolo Anida	virD4	This study
pOGG2-AvirG	Derivative of pOGG2 carrying flanking sequences of	This study
I	virG	
pOGG2-∆virA	Derivative of pOGG2 carrying flanking sequences of <i>virA</i>	This study
pOGG2- Δ virB11 Δ virB1	Derivative of pOGG2 carrying flanking sequences of	This study
1	virB11 and virB1	5
pOGG2-	Derivative of pOGG2 carrying flanking sequences of	This study
Δ virB6 Δ virB8 Δ virB9 Δ virB10	virB6 and virB10	2
pOGG2-∆icmE	Derivative of pOGG2 carrying flanking sequences of	This study
-	icmE	·
pOGG2-∆dotCB	Derivative of pOGG2 carrying flanking sequences of	This study
-	dotC and $dotB$	
BACTH constructs		
pUT18 _{GG} -virD4	Derivative of pUT18 _{GG} encoding VirD4-FLAG-T18	This study
pUT18C _{GG} -virD4	Derivative of pUT18CGG encoding T18-FLAG-VirD4	This study
pKNT25 _{GG} -virD4	Derivative of pKNT25-GG encoding VirD4-FLAG-T25	This study
pKT25 _{GG} -virD4	Derivative of pKT25 _{GG} encoding T25-FLAG-VirD4	This study
pUT18 _{GG} -dotL	Derivative of pUT18 _{GG} encoding DotL-FLAG-T18	This study
pUT18C _{GG} -dotL	Derivative of pUT18C _{GG} encoding T18-FLAG-DotL	This study
pKNT25 _{GG} -dotL	Derivative of pKNT25-GG encoding DotL-FLAG-T25	This study
pKT25 _{GG} -dotL	Derivative of pKT25 _{GG} encoding T25-FLAG-DotL	This study
pUT18 _{GG} -XCV3751	Derivative of pUT18 _{GG} encoding XCV3751-FLAG-T18	This study
pUT18C _{GG} -XCV3751	Derivative of pUT18C _{GG} encoding T18-FLAG-	This study
-	XCV3751	•
pKNT25 _{GG} -XCV3751	Derivative of pKNT25-GG encoding XCV3751-FLAG-	This study
	T25	
pKT25 _{GG} -XCV3751	Derivative of pKT25 _{GG} encoding T25-FLAG-XCV3751	This study
pUT18 _{GG} -XCV0332	Derivative of pUT18 _{GG} encoding XCV0332-FLAG-T18	This study
pUT18C _{GG} -XCV0332	Derivative of pUT18CGG encoding T18-FLAG-	This study
	XCV0332	
pKNT25 _{GG} -XCV0332	Derivative of pKNT25-GG encoding XCV0332-FLAG-	This study
	T25	
pKT25 _{GG} -XCV0332	Derivative of pKT25 _{GG} encoding T25-FLAG-XCV0332	This study
_pUT18 _{GG} -XCV1120	Derivative of pUT18 _{GG} encoding XCV1120-FLAG-T18	This study
pUT18C _{GG} -XCV1120	Derivative of pUT18C _{GG} encoding T18-FLAG-	This study
	XCV1120	
pKNT25 _{GG} -XCV1120	Derivative of pKNT25-GG encoding XCV1120-FLAG-	This study
pK125 _{GG} -XCV1120	Derivative of pKT25 _{GG} encoding T25-FLAG-XCV1120	This study
pUT18 _{GG} -tral	Derivative of pUT18 _{GG} encoding Tral-FLAG-T18	This study
pUT18C _{GG} -tral	Derivative of pUT18C _{GG} encoding T18-FLAG-Tral	This study
pKN125 _{GG} -tral	Derivative of pKN125-GG encoding Tral-FLAG-125	This study
pK125 _{GG} -tral	Derivative of pK125 _{GG} encoding 125-FLAG-Iral	This study
pUT18 _{GG} -virB10	Derivative of pUT18 _{GG} encoding VirB10-FLAG-T18	This study
pUT18C _{GG} -virB10	Derivative of pUT18C _{GG} encoding T18-FLAG-VirB10	This study
pKNT25 _{GG} -virB10	Derivative of pKN125-GG encoding VirB10-FLAG- T25	This study
pKT25 _{GG} -virB10	Derivative of pKT25 _{GG} encoding T25-FLAG-VirB10	This study
pUT18 _{GG} -virB11	Derivative of pUT18 _{GG} encoding VirB11-FLAG-T18	This study
pUT18C _{GG} -virB11	Derivative of pUT18C _{GG} encoding T18-FLAG-VirB11	This study
pKNT25 ₆₆ -virB11	Derivative of pKNT25-GG encoding VirB11-FLAG-	This study
1	T25	- ,
pKT25 _{GG} -virB11	Derivative of pKT25 _{GG} encoding T25-FLAG-VirB11	This study
pUT18 _{GG} -virB7	Derivative of pUT18 _{GG} encoding VirB7-FLAG-T18	This study

pUT18C _{GG} -virB7	Derivative of pUT18C _{GG} encoding T18-FLAG-VirB7	This study
pKNT25 _{GG} -virB7	Derivative of pKNT25-GG encoding VirB7-FLAG-T25	This study
pKT25 _{GG} -virB7	Derivative of pKT25 _{GG} encoding T25-FLAG-VirB7	This study
pUT18 _{GG} -dotB	Derivative of pUT18 _{GG} encoding DotB-FLAG-T18	This study
pUT18C _{GG} -dotB	Derivative of pUT18C _{GG} encoding T18-FLAG-DotB	This study
pKNT25gg-dotB	Derivative of pKNT25-GG encoding DotB-FLAG-T25	This study
pKT25 _{GG} -dotB	Derivative of pKT25 _{GG} encoding T25-FLAG-DotB	This study
pUT18 _{GG} -dotO	Derivative of pUT18 _{GG} encoding DotO-FLAG-T18	This study
pKNT25 _{GG} -dotO	Derivative of pKNT25-GG encoding DotO-FLAG-T25	This study
pKT25 _{GG} -dotO	Derivative of pKT25 _{GG} encoding T25-FLAG-DotO	This study
pUT18gg-icmE	Derivative of pUT18 _{GG} encoding IcmE-FLAG-T18	This study
pUT18Cgg-icmE	Derivative of pUT18C _{GG} encoding T18-FLAG-IcmE	This study
pKNT25cg-icmE	Derivative of pKNT25-GG encoding IcmE-FLAG-T25	This study
nKT25 cc-icmE	Derivative of pKT25 _{CC} encoding T25-FLAG-IcmF	This study
Constructs for promoter_rep	Derivative of pre12566 encoding 125 1 EAG teme	This study
pICH/1021_virB2	Derivative of pICH/1021 containing the putative	This study
prem41021-virb2prom	nemotor of vive 2 flowled by <i>BagLaitos</i> with ATTC and	This study
	TATC every heres for insertion into pPDM D	
alCII41021 viaD5	Derivative of nICII/1021 containing the nutative	This study
piCH41021-VIID3prom	nemotor of vive 5 florked by <i>BagLaites</i> with ATTC and	This study
	TATC events and for incortion into pDBM D	
"ICU41021	Derivative of a ICH41021 containing the material	This starler
pICH41021-VIrD4prom	Derivative of pICH41021 containing the putative	This study
	TATC such as a far insertion into a DBM D	
	Device the second secon	This set of the
pICH41021-VIrGprom	Derivative of pICH41021 containing the putative	This study
	TATC events and for incortion into a DDM D	
"ICH41021 J-44	Derivative of a ICH41021 containing the material	This starler
pICH41021-dotAprom	Derivative of pICH41021 containing the putative	This study
	TATC events and for incortion into a DBM D	
"ICU41021 J-4D	Derivative of a ICH41021 containing the material	This study
pICH41021-dotDprom	Derivative of pICH41021 containing the putative f_{1}	This study
	TATC such and for insertion into a DDM D	
	TATG overnangs for insertion into pBRM-P	T1.'
pICH41021-1cmL _{prom(229)}	Derivative of pICH41021 containing 229 bp upstream of	This study
	the annotated start site of <i>lcmL</i> flanked by <i>Bsal</i> sites	
	with ATTC and TATG overnangs for insertion into	
	Defection of 10111021 and initial 1201 and the	This set of the
pICH41021-1cmLprom(129)	Derivative of pICH41021 containing 129 bp downstream	This study
	of the annotated start site of <i>icmL</i> flanked by <i>Bsal</i> sites	
	with ATTC and TATG overnangs for insertion into	
		II. (1. 2010
pICH41021-hrpB1prom	Derivative of pICH41021 containing the promoter of	Hausner et al., 2019
	hrpB1 flanked by Bsa1 sites with ATTC and TATG	
	overhangs for insertion into pBRM-P	II. 1D#4
pICH41021-stgtp	Derivative of pICH41021 containing sfgfp flanked by	Hausner and Buttner,
	Bsal sites with TATG and GGTG overhangs for	unpublished
	insertion into pBRM	
pBRM-P+T-virB2prom-stgfp	Derivative of pBRM-P+T encoding sfGFP under control	This study
	of the putative <i>virB2</i> promoter	
pBRM-P+T-virB5 _{prom} -sfgfp	Derivative of pBRM-P+T encoding sfGFP under control	This study
	of the putative <i>virB5</i> promoter	
pBRM-P+T-virD4prom-sfgfp	Derivative of pBRM-P+T encoding sfGFP under control	This study
	of the putative <i>virD4</i> promoter	
pBRM-P+T-virG _{prom} -sfgfp	Derivative of pBRM-P+T encoding sfGFP under control	This study
	of the putative virG promoter	
pBRM-P+T-dotAprom-sfgfp	Derivative of pBRM-P+T encoding sfGFP under control	This study
	of the putative <i>dotA</i> promoter	
pBRM-P+T-dotDprom-sfgfp	Derivative of pBRM-P+T encoding sfGFP under control	This study
	of the nutative datD promoter	

pBRM-P+T-icmL _{prom(229)} -sfgfp	Derivative of pBRM-P+T encoding sfGFP under control of the putative <i>icmL</i> promoter upstream of the annotated start codon	This study
pBRM-P+T-icmL _{prom(129)} -sfgfp	Derivative of pBRM-P+T encoding sfGFP under control of the putative <i>icmL</i> promoter downstream of the annotated start codon	This study
pBRM-P+T-hrpB1prom-sfgfp	Derivative of pBRM-P+T encoding sfGFP under control of the <i>hrpB1</i> promoter	This study
pBRM-sfgfp _{plac}	Derivative of pBRM encoding sfGFP under control of the <i>lac</i> promoter	Hausner and Büttner, unpublished
pICH41021-virB2prom	Derivative of pICH41021 containing the putative promoter of <i>virB2</i> flanked by <i>BpiI</i> sites with CTCA and CGAG overhangs for fusion with <i>dTALE-2</i>	This study
pICH41021-virB5 _{prom}	Derivative of pICH41021 containing the putative promoter of <i>virB5</i> flanked by <i>BpiI</i> sites with CTCA and CGAG overhangs for fusion with <i>dTALE-2</i>	This study
pICH41021-virD4prom	Derivative of pICH41021 containing the putative promoter of <i>virD4</i> flanked by <i>BpiI</i> sites with CTCA and CGAG overhangs for fusion with <i>dTALE-2</i>	This study
pICH41021-virG _{prom}	Derivative of pICH41021 containing the putative promoter of <i>virG</i> flanked by <i>BpiI</i> sites with CTCA and CGAG overhangs for fusion with <i>dTALE-2</i>	This study
pICH41021-dotAprom	Derivative of pICH41021 containing the putative promoter of <i>dotA</i> flanked by <i>BpiI</i> sites with CTCA and CGAG overhangs for fusion with <i>dTALE-2</i>	This study
pICH41021-dotD _{prom}	Derivative of pICH41021 containing the putative promoter of <i>dotD</i> flanked by <i>BpiI</i> sites with CTCA and CGAG overhangs for fusion with <i>dTALE-2</i>	This study
pICH41021-icmL _{prom(229)}	Derivative of pICH41021 containing a putative promoter 229 bp upstream of the annotated start codon of <i>icmL</i> flanked by <i>BpiI</i> sites with CTCA and CGAG overhangs for fusion with <i>dTALE-2</i>	This study
pICH41021-icmL _{prom(129)}	Derivative of pICH41021 containing a putative promoter 129 bp downstream of the annotated start codon of <i>icmL</i> flanked by <i>BpiI</i> sites with CTCA and CGAG overhangs for fusion with <i>dTALE-2</i>	This study
pICH41021-hrpB1prom	Derivative of pICH41021 containing the putative promoter of <i>hrpB1</i> flanked by <i>BpiI</i> sites with CTCA and CGAG overhangs for fusion with <i>dTALE-2</i>	This study
pAGM9121-virB2prom	Derivative of pAGM9121 containing the putative promoter of <i>virB2</i> flanked by <i>BsaI</i> sites with GGAG and CGCT overhangs	This study
pAGM9121-virB5 _{prom}	Derivative of pAGM9121 containing the putative promoter of <i>virB5</i> flanked by <i>BsaI</i> sites with GGAG and CGCT overhangs	This study
pAGM9121-virD4 _{prom}	Derivative of pAGM9121 containing the putative promoter of <i>virD4</i> flanked by <i>BsaI</i> sites with GGAG and CGCT overhangs	This study
pAGM9121-virG _{prom}	Derivative of pAGM9121 containing the putative promoter of <i>virG</i> flanked by <i>BsaI</i> sites with GGAG and CGCT overhangs	This study
pAGM9121-dotAprom	Derivative of pAGM9121 containing the putative promoter of <i>dotA</i> flanked by <i>BsaI</i> sites with GGAG and CGCT overhangs	This study
pAGM9121-dotD _{prom}	Derivative of pAGM9121 containing the putative promoter of <i>dotD</i> flanked by <i>Bsal</i> sites with GGAG and CGCT overhangs	This study
pAGM9121-icmL _{prom(229)}	Derivative of pAGM9121 containing the putative promoter 229 bp upstream of the annotated start codon of <i>icmL</i> flanked by <i>BsaI</i> sites with GGAG and CGCT overhangs	This study

pAGM9121-icmL _{prom(129)}	Derivative of pAGM9121 containing the putative promoter 129 bp downstream of the annotated start codon of <i>icmL</i> flanked by <i>Bsal</i> sites with GGAG and CGCT overhangs	This study
pAGM9121-hrpB1prom	Derivative of pAGM9121 containing the putative promoter of <i>hrpB1</i> flanked by <i>BsaI</i> sites with GGAG and CGCT overhangs	This study
pICH47732-virB2prom	Derivative of pICH47732 containing the putative promoter of <i>virB2</i> flanked by <i>BpiI</i> sites with TGCC and GCAA overhangs	This study
pICH47732-virB5 _{prom}	Derivative of pICH47732 containing the putative promoter of <i>virB5</i> flanked by <i>BpiI</i> sites with TGCC and GCAA overhangs	This study
pICH47732-virD4 _{prom}	Derivative of pICH47732 containing the putative promoter of <i>virD4</i> flanked by <i>BpiI</i> sites with TGCC and GCAA overhangs	This study
pICH47732-virG _{prom}	Derivative of pICH47732 containing the putative promoter of <i>virG</i> flanked by <i>BpiI</i> sites with TGCC and GCAA overhangs	This study
pICH47732-dotAprom	Derivative of pICH47732 containing the putative promoter of <i>dotA</i> flanked by <i>BpiI</i> sites with TGCC and GCAA overhangs	This study
pICH47732-dotD _{prom}	Derivative of pICH47732 containing the putative promoter of <i>dotD</i> flanked by <i>BpiI</i> sites with TGCC and GCAA overhangs	This study
pICH47732-icmL _{prom(229)}	Derivative of pICH47732 containing the putative promoter 229 bp upstream of the annotated start codon of <i>icmL</i> flanked by <i>BpiI</i> sites with TGCC and GCAA overhangs	This study
pICH47732-icmL _{prom(129)}	Derivative of pICH47732 containing the putative promoter 129 bp downstream of the annotated start codon of <i>icmL</i> flanked by <i>BpiI</i> sites with TGCC and GCAA overhangs	This study
pICH47732-hrpB1prom	Derivative of pICH47732 containing the putative promoter of <i>hrpB1</i> flanked by <i>BpiI</i> sites with TGCC and GCAA overhangs	This study
pICH77739-virB2 _{prom} - dTALE2	Derivative of pICH77739 encoding dTALE-2 under control of the putative <i>virB2</i> promoter	This study
pICH77739-virB5 _{prom} - dTALE2	Derivative of pICH77739 encoding dTALE-2 under control of the putative <i>virB5</i> promoter	This study
pICH77739-virD4 _{prom} - dTALE2	Derivative of pICH77739 encoding dTALE-2 under control of the putative <i>virD4</i> promoter	This study
pICH77739-virG _{prom} -dTALE2	Derivative of pICH77739 encoding dTALE-2 under control of the putative <i>virG</i> promoter	This study
pICH77739-dotAprom-dTALE2	Derivative of pICH77739 encoding dTALE-2 under control of the putative <i>dotA</i> promoter	This study
pICH77739-dotDprom-dTALE2	Derivative of pICH77739 encoding dTALE-2 under control of the putative <i>dotD</i> promoter	This study
pICH77739-icmL _{prom(229)} - dTALE2	Derivative of pICH77739 encoding dTALE-2 under control of the putative <i>icmL</i> promoter	This study
pICH77739-icmL _{prom(129)} - dTALE2	Derivative of pICH77739 encoding dTALE-2 under control of the putative <i>icmL_{new}</i> promoter	This study
plCH77739-hrpB1 _{prom} - dTALE2	Derivative of pICH77739 encoding dTALE-2 under control of the <i>hrpB1</i> promoter	This study

¹ Ap, ampicillin; Gm, gentamycin; Km, kanamycin; R, resistant; Rif, rifampicin; Sm, spectinomycin; Str, streptomycin.

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Name	Sequence ¹
Primers for the generation of exp	ression constructs
virD4-Bsa-for	TTT GGTCTCT TATG TCCAAACCCAAAATCATCG
virD4-950-rev	TTT GGTCTCT AGGC GAAGGTTTCATCCGCCTG
virD4-950-for	TTT GGTCTCT GCCT CGATCATGGGCACCTTG
virD4-1450-rev	TTT GGTCTCT TGAC GTTCTTCTTCTTGATGG
virD4-1450-for	TTT GGTCTCT GTCA CCCGAGGGCGGGAAACCAC
virD4-Bsa-rev	TTT GGTCTCT CACC CACTTTGATGGTAAGC
traI-pB-for	TTT GGTCTCT TATG ATGCTCAATGTGACGCCGATTCG
traI-pB-rev	TTT GGTCTCT CACC GCCAAAGCGTCCGTACTCGGACG
3751-Bsa-for	TTT GGTCTCT TATG CCGACGAACTACTCAC
3751-Bsa-rev	TTT GGTCTCT CACC ACCGACGCTGCGCCCCTGGC
XCV0332-for	TTT GGTCTCT TATG ACCGACGCACCGGTCATC
XCV0332-rev	TTT GGTCTCT CACC GCGCGGCGGCAAACGCAG
XCV1120-for	TTT GGTCTCT TATG GTGGAAGTCCGGCAAATC
XCV1120-rev	TTT GGTCTCT CACC CATTGTCATCCTTG
icmE-F1_for	TTT GGTCTCT TATG ATGGCTCAGCAGGAAAGCGATATC
icmE-F1-rev	TTT GGTCTCT TCGT CGGAGGCCGTACTTGCGAC
icmE-F2-for	TTT GGTCTCT ACGA CATGGCGGAAACCACCAG
icmE-F2-rev	TTT GGTCTCT CGTC CCGCGCATGACGCTTC
icmE-F3-for	TTT GGTCTCT GACG ATCGAAACCAGCGAGTTCG
icmE-F3-rev	TTT GGTCTCT CACC TTTGCCGAATGCGGATGC
dotB-for	TTT GGTCTCT TATG AACGCGGTGCTCTCCGACCAG
dotB-rev	TTT GGTCTCT CACC CTCACCCTCCCCGGCCAG
dotO-F1-for	TTT GGTCTCT TATG AAGATCGTCGAGCAAATCGCAGATATTG
dotO-F1-rev	TTT GGTCTCT GTCT TCGGTGTATTCGATGTCCAGGTAG
dotO-F2-for	TTT GGTCTCT AGAC GAAACCACCTTCGTGCTCAAG
dotO-F2-rev	TTT GGTCTCT CACC TGCACGGCGGTCACCTTG
dotL-F1-for	TTT GGTCTCT TATG AGCATCCAGGGCGTCGACAAGC
dotL-F1-rev	TTT GGTCTCT CGGT GGGGTCCTCAACCTTCCCAATG
dotL-F2-for	TTT GGTCTCT ACCG AAACCTGGGAGATGATCCAAGCAAT
	GGGCGGCGAAG
dotL-F2-rev	TTT GGTCTC T <i>CCGC</i> CGGCGCTGCCGCCACCAGATACAG
dotL-F3-for	TTT GGTCTCT GCGG TGGACTCAATGCTCAGCCGCAG
dotL-F3-rev	TTT GGTCTCT CACC AAGCTCATTCGGCGCTCCTGAACTG
virB10-for	TTT GGTCTCT TATG AGCGATCAGACCAACAAGCCGCAG
virB10-rev	TTT GGTCTCT CACC CGGGATGAACGTCACGCCGTCG
virB11-for	TTT GGTCTCT TATG TCGCAGATTGCCTCCTTC
virB11-rev	TTT GGTCTCT CACC CGCACCTGCATAGATG
XCVc0034-for	TTT GGTCTCT TATG CCTAAGGCCCTCTGGACATG
XCVc0034-rev	TTT GGTCTCT CACC GTGTCGCCATTGGTTGTCATCC
prom4361-for	TTT GGTCTCT ATTC AGGACAACGCGGTTCGTG
prom4361-rev	TTT GGTCTCT CATA GCGCGTGGCCTATGCG
Bsa-prom0160-for	TTT GGTCTCT ATTC TGATCGAAGATGCTGCTG
Bsa-prom0160-rev	TTT GGTCTCT GATG TGGACAGCGGCGCAG
Bsa-icmE-for-pLAND	TTT GGTCTCT CATC ATGATGGCTCAGCAGGAAAGC
Gent-Bsa-for	TTT GGTCTCT TATG TTACGCAGCAGCAAC
Gent-Bsa-Stop-rev	TTT GGTCTCT CACC TTAGGTGGCGGTACTTG

Table S2:	Primers	used in	this	study.
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Primers for colony PCR	
dotC-detect-for	TTGATTTCGGCTCCTTGCTC
dotC-detect-rev	ATCTGGTCTGCCTGCTGCTC
dotB-detect-for	CTTCTGGTCCATGCCAAAGC
dotB_detect_rev	CGCGGAGGATGATCGAGATA
icmE-F1-for	TTT GGTCTCT TATG ATGGCTCAGCAGGAAAGCGATATC
icmE-F3-rev	TTT GGTCTCT CACC TTTGCCGAATGCGGATGC
virB11-detect-for	GAATCTCGATGACGGCGTTC
virB11-detect-rev	GAATCTCGCCCATGATGAC
virB4-detect-for	GTTCCAGCGCTCCCACGGTC
virB4-detect-rev	GTGCGCGTTGTCGAACACG
virBl0-for	TTT CCTCTCT CACCCCCCATCA ACCTCACCCCCCCCC
virD4-detect-for	
virD4-detect-rev	GTCCGTGAGCAGGAAGTCAT
avrBs3-Bsa-for	TTT GGTCTC T <i>TATG</i> GATCCCATTCGTTC
avrBs3-Bsa-50 rev	TTT GGTCTCT GATC GGACATCGTCCGCCGAGC
hpaF-seq-for	CTTCAATTGACGATGGTC
hpaG-seq-rev	CTGCATGAGGCCGATGTTTTCAG
pKT25/pUT18-for	ACTGGAAAGCGGGCAGTGAG
pKT25-rev	GCTGCGCAACTGTTGGGAAG
pUT18-rev	GGTGTCGGGGCTGGCTTAAC
pBRM-1180-for	ATGCATGCGCCCAATACGCAAACCG
pBRM-endlacZ-rev	CACTCATCGCAGTCGGCCTATTG
M13-for	GTAAAACGACGGCCAGTG
M13-rev	GGAAACAGCTATGACCATG
pOK-mcs-for	
pOK-mcs-rev	CTTGTCCTTTAACGAGGATTGTTAC
Primers for the generation of del	etion constructs
pXCV183-F1-for	TTT GGTCTCT CGAC CCGGACAGCTCGGTTCCGGCCTTG
	TTT GGTCTCT ACTG ATTGCACCATGCAACTAGCCCCG
pXCV183-F1-rev	AGCTGAAG
pXCV183-F2-prom-for	TTT GGTCTCT CAGT TGACATAAGCCTGTTCGGTTCG
pXCV183-F2-prom-rev	TTT GGTCTC T <i>CGTT</i> GCTGCTCCATAACATCAAACATCG
pAC v 183-F 3-kan-lor	
aVCV192 E2 log agy	
pAC v 183-F 3-kan-rev	CATACAAC
nVCV192 E4 momboo7 for	
pXCV183-F4-promacZ-for pXCV183-F4-GFP-rev/	
nXCV183-F5-for	TTT GCTCTCT GGTG TCAGGGCGCATCGGTATCGCCACGAG
nXCV183-F5-rev	TTT GCTCTCT ATGG GCCGCAGCCGATACTA AGCACCTAGAG
virG-pOGG2-1-for	
vii0-p0002-1-101	CGGCAC
virG-pOGG2-1-rev	TTT GGTCTCT GGAA TGCAGCTTGATCATCGACTCTCTATC
virG-pOGG2-2-for	TTT GGTCTCT TTCC GAGCACGCTCGATGACACAGCCGCC
	GGCGCGAGC
	Geoegade
virG-pOGG2-2-rev	TTT GGTCTCT ATGG AGCAGCACGGCGCTGAAGATCGTC
virG-pOGG2-2-rev pOGG2-virA/C-for	TTT GGTCTCT ATGG AGCAGCACGGCGCTGAAGATCGTC TTT GGTCTCT CGAC TGGAACCAGCAGTTCCAGCACG
virG-pOGG2-2-rev pOGG2-virA/C-for pOGG2-virA/C-rev	TTT GGTCTCT ATGG AGCAGCACGGCGCTGAAGATCGTC TTT GGTCTCT CGAC TGGAACCAGCAGTTCCAGCACG TTT GGTCTCT GTGC CTAGAAGTTACAACGCAGGATG
virG-pOGG2-2-rev pOGG2-virA/C-for pOGG2-virA/C-rev pOGG2-virA/N-for	TTT GGTCTCT ATGG AGCAGCACGGCGCTGAAGATCGTC TTT GGTCTCT CGAC TGGAACCAGCAGTTCCAGCACG TTT GGTCTCT GTGC CTAGAAGTTACAACGCAGGATG TTT GGTCTCT GCAC AGGCTGAAGAAGATCAGC
virG-pOGG2-2-rev pOGG2-virA/C-for pOGG2-virA/C-rev pOGG2-virA/N-for pOGG2-virA/N-rev	TTT GGTCTCT ATGG AGCAGCACGGCGCTGAAGATCGTC TTT GGTCTCT CGAC TGGAACCAGCAGTTCCAGCACG TTT GGTCTCT GTGC CTAGAAGTTACAACGCAGGATG TTT GGTCTCT GCAC AGGCTGAAGAAGATCAGC TTT GGTCTCT ATGG CGAGCCAGGCCAGACCATCG
virG-pOGG2-2-rev pOGG2-virA/C-for pOGG2-virA/C-rev pOGG2-virA/N-for pOGG2-virA/N-rev virB6-10-pOGG2-1-for	TTT GGTCTCT ATGG AGCAGCACGGCGCTGAAGATCGTC TTT GGTCTCT CGAC TGGAACCAGCAGTTCCAGCACG TTT GGTCTCT GTGC CTAGAAGTTACAACGCAGGATG TTT GGTCTCT GCAC AGGCTGAAGAAGATCAGC TTT GGTCTCT ATGG CGAGCCAGGCCAGACCATCG TTT GGTCTCT ATGG TTGCAACAGCAGTTTCAGCAG

virB6-10-pOGG2-2-for	TTT GGTCTCT ACCA CGTGACGTTCATCCCGTGAG
virB6-10-pOGG2-2-rev	TTT GGTCTCT CGAC GGCCAACTTCTCAAACGGCAGC
virB4-pOGG2-Bsa-FIfor	TTT GGTCTCT CGAC CGCTTGGACATAGCTGTG
virB4-pOGG2-Bsa-FIrev	TTT GGTCTCT GTCA GTCGAACTTCTCTTTTAAAACCTC
virB4-pOGG2-Bsa-FIIfor	TTT GGTCTCT TGAC CATGAAAAAGACCC
virB4-pOGG2-Bsa-FIIrev	TTT GGTCTCT ATGG CTATACCGCGAGCGCAAG
virD4-FII-BsaI-rev	TTT GGTCTCT CGAC CGTCGCGGGTACTTGAG
virD4-FII-BsaI-for	TTT GGTCTCT TGAT TGATGTCACATTTGCAG
virD4-FI-BsaI-for	TTT GGTCTCT ATCA GCGCCCTTCTCCTTAGG
virD4-FI-BsaI-rev	TTT GGTCTCT ATGG TGGCATGTGTCAGTTCG
virB11-pOGG2-Bsa-FII-for	TTT GGTCTCT CGAC CATGAAGTCGGCCATCATC
virB11-pOGG2-Bsa-FII-rev	TTT GGTCTCT GTCA GGCTCGGCCCTCACGGGATG
virB1-pOGG2-Bsa-FI-for	TTT GGTCTCT TGAC CGGCTTTTCTCGCATGC
virB1-pOGG2-Bsa-FI-rev	TTT GGTCTCT ATGG GCCATCGTGTTGCGAACCTT
pOGG2-icmE-F1-for	TTT GGTCTCT CGAC CCTGCCGGTACCACTGTGC
pOGG2-icmE-FI-rev	TTT GGTCTCT TTTC CTGCTGAGCCATCACG
pOGG2-icmE-FII-for	TTT GGTCTCT GAAA TAAAGGCTCCCCATGAAC
pOGG2-icmE-FII-rev	TTT GGTCTCT ATGG CGCTTTCAGTTGGATAC
pOGG2-dotCB-F1-for	TTT GGTCTCT CGAC ATGTTTGCGCCCGATAGC
pOGG2-dotCB-FI-rev	TTT GGTCTCT TCAC AAACCGCCATTGCTG
pOGG2-dotCB-FII-for	TTT GGTCTCT GTGA GAGGGTGAGTGATGTCAG
pOGG2-dotCB-FII-rev	TTT GGTCTCT ATGG GCGATCTGCGGTTACCAG
Primers for the amplification of p	oredicted promoter regions
promvirB2-for	TTT GGTCTCT ATTC ACTATGCGGTTTATCTGGTTG
promvirB2-rev	TTT GGTCTCT CATA TCTGAAACCTCTCAGATTGTTAG
promvirB5-for	TTT GGTCTCT ATTC ATTGCCGGACTGACGCACATCTATCG
promvirB5-rev	TTT GGTCTCT CATA ATCGAAGGTCGGGATGGGGGGTTCG
promvirD4-for	TTT GGTCTCT ATTC GATATCGTGTTGTGGCGATTACG
promvirD4-rev	TTT GGTCTCT CATA GCGCCCTTCTCCTTAGGCTTG
promvirG-for	TTT GGTCTCT ATTC GTCGGATCGGGCTGGCGACG
promvirG-rev	TTT GGTCTCT CATA CGACTCTCTATCCCGGCATG
promdotA-for	TTT GGTCTCT ATTC GCCTGATCTGGCCCGGTCATC
promdotA-rev	TTT GGTCTCT CATA ATTGACGAGCGCGCCACAAACG
promdotD-for	TTT GGTCTCT ATTC TAGCGACACATATACGTGACCGTG
promdotD-rev	TTT GGTCTCT CATA CGGGGCACGTTGCCTTTCGTTG
promicmL-for	TTT GGTCTCT ATTC CCGGCATAGGTTGGACACAGG
promicmL-rev	TTT GGTCTCT CATA GAACTCTGCCGGTCACTCG
promicmL(129)-for	TTT GGTCTC T <i>ATTC</i> GTGCAATCGGTGTCCAACTATGC
promicmL(129)-rev	TTT GGTCTCT CATA TCGTCGCGCCACAACTGTTTG
dTALE-pvirB2-for	GAAGAC AA CTCA GGAG ACTATGCGGTTTATCTGGTTG
dTALE-pvirB2-rev	GAAGAC AA CTCG AGCG TCTGAAACCTCTCAGATTGTTAG
dTALE-pvirB5-for	GAAGAC AA <i>CTCA</i> GGAG
	ATTGCCGGACTGACGCACATCTATCG
dTALE-pvirB5-rev	GAAGAC AA CTCG AGCG ATCGAAGGTCGGGGATGGGGGGTTCG
dTALE-virD4-for	GAAGAC AA CTCA GGAG GATATCGTGTTGTGGCGATTACG
dTALE_virD4_rev	GAAGAC AA CTCG AGCG GCGCCCTTCTCCTTAGGCTTG
dTALE-pvirG-for	GAAGAC AA CTCA GGAG GTCGGATCGGGCTGGCGACG
dIALE-pvirG-rev	GAAGAC AA CTCG AGCG CGACTCTCTATCCCGGCATG
dIALE-pdotA-tor	GAAGACAA CICA GGAG GCCIGAICIGGCCCGGTCAIC
dIALE-pdotA-rev	GAAGAC AA CTCC AGCG ATTGACGAGCGCGCCACAAACG
dIALE-pdotD-tor	GAAGACAA CICA GGAG IAGCGACACATATACGTGACCGTG
dIALE-pdotD-rev	GAAGAC AA <i>CTCG</i> AGCG CGGGGGCACGTTGCCTTTCGTTG

dTALE-picmL-for	GAAGAC AA CTCA GGAG CCGGCATAGGTTGGACACAGG
dTALE-picmL-rev	GAAGAC AA CTCG AGCG GAACTCTGCCGGTCACTCG
dTALE-picmL(129)_for	GAAGAC AA CTCA GGAG GTGCAATCGGTGTCCAACTATGC
dTALE-picmL(129)-rev	GAAGAC AA CTCG AGCG TCGTCGCGCCACAACTGTTTG
dTALE-phrpB1-for	GAAGAC AA CTCA GGAG GAATGTGTGGATGGCCT
	GGCGAAG
dTALE_phrpB1-rev	GAAGAC AA CTCG AGCG GCGGACCGTACACGTGGGGAACG
¹ BsaI and BpiI recognition sites are	shown in bold, resulting overhangs in italics.

Protein	A. tumefacien	ıs	X. axonopodis pv.	. citri	X. axonopodis p	v. citri	Putative function/
	strain GV310	1	strain 306		strain 306		localization ²
	[NZ KY000036	5.1]	Chromosome [AE00	08923]	pXAC64 [AE00	8925]	
	accession number ¹	aa	accession number1	aa	accession	aa	
					number ¹		
VirB1	WP 010974915.1	245	AAM37466.1	280	AAM39282.1	292	lytic transglycosylase
VirB2	WP 010891502.1	121	AAM37465.1	136	AAM39293.1	125	pilin subunit
VirB3	WP 010891501.1	108	AAM37464.1	103	AAM39292.1	99	inner membrane
	_						platform
VirB4	WP_010974916.1	789	AAM37463.1	817	AAM39291.1	877	ATPase
VirB5	WP_010891499.1	220	AAM37462.1	275	AAM39290.1	220	pilus-tip protein
VirB6	WP_010974917.1	295	AAM37461.1	350	AAM39287.1	288	inner membrane
							platform
VirB7	WP_010891497.1	168	AAM37471.1	139	AAM39288.1	131	lipoprotein, outer
							membrane core
							complex
VirB8	WP_010891496.1	237	AAM37470.1	348	AAM39286.1	224	inner membrane
							platform
VirB9	WP_010891495.1	393	AAM37469.1	255	AAM39285.1	260	outer membrane core
							complex
VirB10	WP_010891494.1	377	AAM37468.1	389	AAM39284.1	406	inner/outer
							membrane core
							complex
VirB11	WP_010974918.1	344	AAM37467.1	346	AAM39283.1	340	ATPase
VirD4	WP_010974920.1	668	AAM37472.1	557	-	-	ATPase
	WP_010891492.1	253	AAM36537.1	124	-	-	two-component
VirG							system response
							regulator
VirA	WP_010974914.1	833	AAM36536.1	676	-	-	two-component
• 117 1							system sensor kinase
TraG	-	-	-	-	AAM39276.1	524	conjugative coupling
1140						<u> </u>	protein TrwB
TraI	-	-	-	-	AAM39277.1	991	conjugative relaxase
1101				1			TrwC

Table S3: Accession numbers and predicted functions of VirB/VirD4 proteinsfrom A. tumefaciens and x. axonopodis pv. citri strain 306.

¹ These proteins were used for the pairwise sequence alignments with corresponding proteins from *X. euvesicatoria* strain 85-10. The corresponding amino acid identities between homologous proteins are given in Fig. 1. ² See main text for details and references.

Table S4: 3	Sequences of	predicted vir and	icm/dot promoters	from X. eu	vesicatoria.
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Predicted	Sequence $(5' - 3')^1$
promoter of:	
virB2	actatgcggtttatctggttgaggcggagcacgagagcagccggtggtggtggcggatgacggtgaagttacccacgcacaacaaag cttatgaggtcgttaccgcgcttggtaaaaccaagctctggaaacgcttggacatagctgtgg <mark>atttca</mark> tcaaggagagctgcccgc acacaaagaccgtgttcgtagtttttgccccgtagcacacctaacaatctgagaggtttcaga
virB5	attgccggactgacgcacatctatcgctactttgcctcacatcgccaattggcaacccaagggcctttacgcagacaaggtcctaag attgggagaaatgatcagtgcccatgtggcagtgggcggaagtacaagcattgctgtgctaccagcgctccgacattt <mark>cattg</mark> atac cgg <mark>cttaac</mark> tgacttccctgttc <mark>cattgc</mark> tccccgaaccccatcccgaccttcgat
virD4	gatatcgtgttgtggcgattacgcgtgacgaggtgtatgcacccgacgcggtcgtggcatacgccgtcgtgaccgatgcgggtacg cggatcacgccggatttgtctctggaccaggccaaggtctggatcgattcgctggtggagagcggaggcggacgcaagtcc gagcttgtcgaccataagccggtcgtgcgtcgctagcccgccatcgtgatgcaagcctaaggagagggggcgc
virG	gtcggatcgggctggcgacgggatcggtccctcacccgtgttgtcgcagagcgagtggctggtcacaagatcgaacaaaagtcgt catcgaccacggcacgg
dotA	gcctgatctggcccggtcatccggcggccgagcggaccaggcgatcggctaagagtggctcccgatccacagtccagttctcg gccctatccatgacgctgaccaagtccccagttccgggggcataactacgcc <mark>agaac</mark> tcgacgaggtgggggggggggggggggggggggg
dotD	tagcgacacatatacgtgaccgtgcacagagccggcctgtcacattcgttcg
icmL ₂₂₉	ccggcataggttggacacaggccgccgaacagctgatttctagtgattttgcgcggtgcttgccgacgtccatatcacttggtgcggt aagtcagtgaagtga
icmL ₁₂₉	gtgcaatcggtgtccaactatgccgttcttcccacggccaaaggccacattcgttggcatatatc <mark>gttga</mark> cccgcct <mark>gtta</mark> tggtgtcgt atcctcg <mark>tgaac</mark> gatttccaaacagttgtggcgcgacga

¹ Predicted *vir* boxes are indicated in yellow.

Protein	L. pneumophila strain	n Philadelphia1	Putative function/ localization ²
	[NC00294	2.5]	
	accession number ¹	aa	
DotA	WP_010948386.1	1048	inner membrane protein
DotB	WP_011947587.1	369	ATPase
DotC	WP_010948375.1	303	lipoprotein, outer membrane core complex
DotD	WP_010948374.1	163	lipoprotein, outer membrane core complex
IcmT	WP_010946190.1	86	inner membrane protein
IcmW	WP_010948388.1	151	cytoplasmic chaperon
IcmP/DotM	WP_011214699.1	376	inner membrane protein
IcmO/DotL	WP_010946195.1	783	ATPase
IcmL/DotI	WP_010946198.1	212	inner membrane protein
IcmK/DotH	WP_015444826.1	360	periplasmic
IcmE/DotG	WP_010946200.1	1048	inner membrane protein
IcmG/DotF	WP_010946201.1	269	inner/outer membrane core complex
IcmC/DotE	WP_010946202.1	194	inner membrane protein
IcmD/DotP	WP_011214707.1	132	inner membrane protein
IcmJ/DotN	WP_011214708.1	208	cytoplasmic
IcmB/DotO	WP_010946205.1	1009	cytoplasmic

Table S5: Accession numbers and predicted functions of Icm/Dot proteins from L. pneumophila.

¹ These proteins were used for the pairwise sequence alignments with corresponding proteins from *X. euvesicatoria* strain 85-10. The corresponding amino acid identities between homologous proteins are given in Fig. 2. ² See main text for details and references.

Table S6: Transfer of plasmid $pXCV38_{82-8}$:: spec^R from strain 82-8 into the recipientstrain 85-10:: gent^R.

Donor		Recipient		Transconjugant ¹		
Strain	CFU	Strain	CFU	Strain	CFU	% of
						recipient
82-8	5×10^{8}	85-10:: <i>gent^R</i>	5×10^{8}	85-10:: <i>gent^R</i>	3.59 ×	0.072
pXCV38 ₈₂₋₈ ::spec ^R				pXCV38 ₈₂₋₈ :: <i>spec</i> ^{<i>R</i>}	105	
	6.45×10^{8}		6.45×10^{8}		1.343 ×	2.08
					10^{7}	
	7.05×10^{8}		7.05×10^{8}		8.23 ×	1.17
					10^{6}	
85-10::gent ^R	5.55×10^{8}	85-10:: <i>kan^R</i>	5.55×10^{8}	85-10:: <i>kan^R</i>	44.000	0.008
pXCV38 ₈₂₋₈ :: <i>spec</i> ^{<i>R</i>}				pXCV38 ₈₂₋₈ :: <i>spec</i> ^{<i>R</i>}		
	5.52×10^{8}		5.52×10^{8}		41.500	0.008
	4.56×10^{8}		4.56×10^{8}		35.600	0.008

¹ Transconjugants were selected on NYG plates containing rifampicin, gentamycin and spectinomycin. The presence of plasmid pXCV38₈₂₋₈::*spec^R* was confirmed by agarose gel electrophoresis and PCR (see text). Numbers represent the numbers of transconjugants obtained in three independent conjugation experiments using the listed donor and recipient strains.

Table S7:	XVIPCDs in	T4S candida	te substrates	from X.	euvesicatoria.
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XCV0332 MTDAPVIHQGPVGPDHPEHPDHYLFAQIREAVSVLDAELGKPVDQASERMAARLLPLA KQHGFDQVDYVVLSRHLGEVGENV <mark>FL</mark> WR <mark>G</mark> ELS <mark>DPAH</mark> LRAHITTQEAMETSVDASLAQ LEEINRRLMLRLPPR
KQHGFDQVDYVVLSRHLGEVGENV <mark>F</mark> L <mark>V</mark> R <mark>G</mark> ELS <mark>DPAH</mark> LRAHITTQEAMETSVDASLAQ LEEINRRLMLRLPPR
LEEINRRLMLRLPPR
XCV1120 MEVRQIGKGRAGQPDIEPQAVASNEAVLADNPAHPDHSTYQQIHSWVRGTGNWNEEE
SKNVTASLYKQQTEDPLLKRVDQVTGGLGRDGAHNVFAVYAPHGMGVAPMFHA <mark>HV</mark>
DGRQAAQEPAQQNLQQAEAIQQTQVRQQVLEQTQQNVQQEQGPRMTM
XCV3751 MPTNYSRAQVLDIIEREAAERNIPRDDFLRFAHVETGGAFDELASRGPGGAKGLFQFTP
PTAQQYGIAGRELDAVANTDAAARLYLDNRTSLVNRHERDGRPYLSGKPEPDGLDMY
LAHQQGAGGYRSLQTAIATGSFGLESTRANILNNVGEKELKSLTGVDSATFRRMSDKD
MAQTFVQYWDTKMDRIRIPEKGIEPISTTQQAPSHAAPAHSAPAQPEKPAAQGIALHAA
YDLTNKYDHVKYGLGAKDPDKGRVDCSGWVVEMQNATMDEINKGAGKAIFTKDEKF
SPGFDSASELLRKAELRSGVLIQGKDVTAQTLKEGMIIGEDNGPQSWDKGRYKGIDHIT
MVVRDPKDGQLKISQSRGGEGVELSSLDSYLERKHAKGVKLYASDPLSEARDLLQERS
QNKQQSHAATEHKPSQAHAAADAPGVLRENARGAEVRTLQQTLQQLGYKDAGGNEL
KADGAYGQRTSEAVKAFQRAHGLQDDGVVGRDTQAALKQAEKTPLLSEKTNPDHPLF
NQAVNKLEQLGPNAFANRQQLVNAAGHMTFEAKVSGMQRIDMVAQSKDGNGLFAV
QGQPIDPAHQKIYIDKATAAEKPLEQSSNAVKQEAQIQIQVQDQHEQQKSQGKSVG
ITAI MILNVIPIKUNNŲ Y AAAHY FSAADDY Y AKEHPUE WUGUUAUAUULI UPVEUAUUSKL
RALEQVERLAEAKKKVKGK5YKEKIANNIVIGKFKHEMISKAKDPQLHIHAVVLNMIQ
KADUA W KALSNEDIFK V QTE V DAL I KAELAKULQALU I AIKL V DDQUNFELDTISKD
ULAFSAKSKVILEALANEUKI KATATILEKUIISLATKYKKDESDKELVKUTWVEKSKE I GIDVGI DSOL DGDTVE AGDSEGTDGDEDGDAGDDIAATSI DASI TDAGAVWOVAINU
GDALSSAGWKSVI OELKGWSDKOAOOVVGNAIKOGSI VDAEKDVTTOKALADEKAI
AIERTGPGAIEPIMTAAAVKTALEGTALSAGORSAVETIVSTKNREVGIOGDAGTGKTV
TVNOAVALIKOASAVSEGVRTVALAPVGNOVKALKNEGI FAHTLASELRTKDKPIDGK
TIIVI DESGVVGAROMEOVMRVVEK AGARMVI I GDTKOTEAIEAGKPEAOLOODGM
OTARISFIOROK DHFI KTAVFHAAFGRVTPSI AHIKHVFFI KFPIFRHRAIVNDYIOI TE
PERRETI IVAGTNEARREINRMVROSI DI TGKGREFETI TRVDMTOAORREAPSYOPG
MVIOPEKDDOKAGLTRGETYRVKDALPGNALVLORODGTTTTINPRKATOLSVYRLER
AELSVGDTVRINRNDPGRDLTNGDRMRVAGVIGGVVKLESVEORDGRPARALELPTN
RPLHLEHAYASTVHSAOGLTNDRALIALDTNSRTTSMNLYYVAISRACHEARVYTNSV
KELPAAIARRFDKTTALAIOREROLORRDAGMOPKGAADGKOALOROOFOOORKOPA
SGKKPSEYGRFG

¹Conserved amino acid sequences of the XVIPCD are indicated as follows: GLxRIDHV in yeqllow, FAVQGxxDPAHxRAHV in green and the glutamine-rich C-terminal tail in purple.



Figure S1 Drehkopf et al. Figure S1: Synthesis and immunological detection of T18 and T25 fusion proteins in *E. coli* strain JM109.

A) Immunological detection of VirB fusion proteins. Cell extracts from *E. coli* strain JM109 with expression constructs encoding T18 and T25 fusions of VirB7, VirB10, VirB4, VirB11 and VirD4 as indicated were analysed by immunoblotting, using a FLAG epitope-specific antibody. Asterisks mark signals at the expected size of the fusion proteins. Additional signals likely correspond to degradation products or result from unspecific binding of the antibody to proteins in the cell extracts.

B) Analysis of T18 and T25 fusions of candidate T4S substrates and TraG. Cell extracts of *E. coli* strain JM109 containing expression constructs for the synthesis of T18 and T25 fusions of XCV0332, XCV1120, XCV3571, TraI and TraG as indicated were analysed by immunoblotting as described in (A). Proteins at the expected size are indicated by asterisks. Note that XCV0332-T18, XCV0332-T25 and XCV3751-T25 were not detectable.

C) Immunological detection of T18 and T25 fusions of Icm and Dot proteins. JM109 cell extracts containing expression constructs for the synthesis of T18 and T25 fusions of DotL, DotB, DotO and IcmE were analysed by immunoblotting as described in (A). Proteins at the expected size of the fusions are indicated with asterisks. Additional signals likely correspond to degradation products or protein complexes in case of DotB, which were not dissolved during SDS-PAGE. Note that T18-DotO and IcmE-T18 were not detectable.



Figure S2 Drehkopf et al.

Figure S2: Results of BACTH assays with TraG from X. euvesicatoria.

T18 and T25 fusions of TraG, the predicted ATPase VirB11, the structural component VirB10 and the putative relaxase TraI were analysed in *E. coli* BTH101 cells as indicated and bacterial cultures were grown on indicator plates containing X-gal and IPTG. As control, fusion proteins were tested against the T18 or T25 domain alone as indicated. All proteins were stably synthesized as was shown by immunoblot analysis of bacterial cell extracts, using a FLAG epitope-specific antibody (see Fig. S1). Interaction studies were performed at least three times with similar results. One representative colony is shown. Results are summarized in Table 2.