

Supplementary Information

ROR γ t-Raftlin1 complex regulates the pathogenicity of Th17 cells and colonic inflammation.

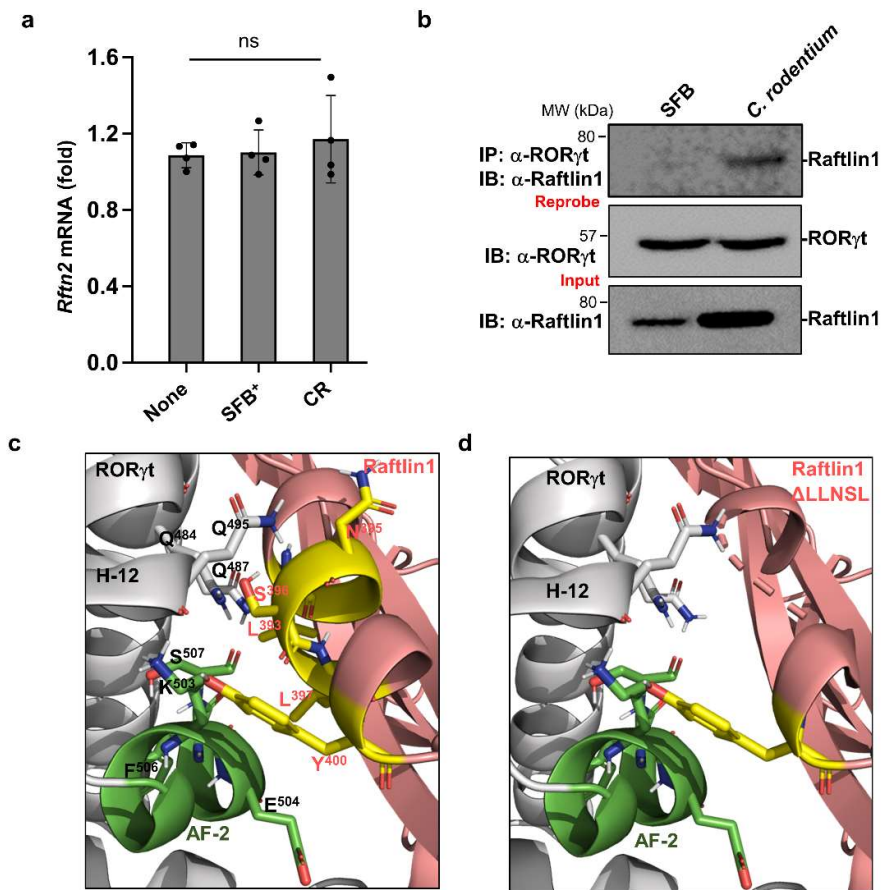
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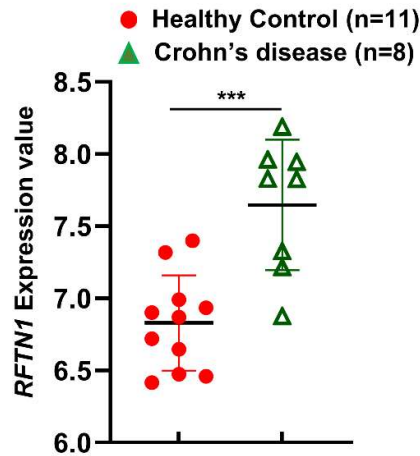
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Uncropped western blots and PCR agarose gels of Supplementary figures

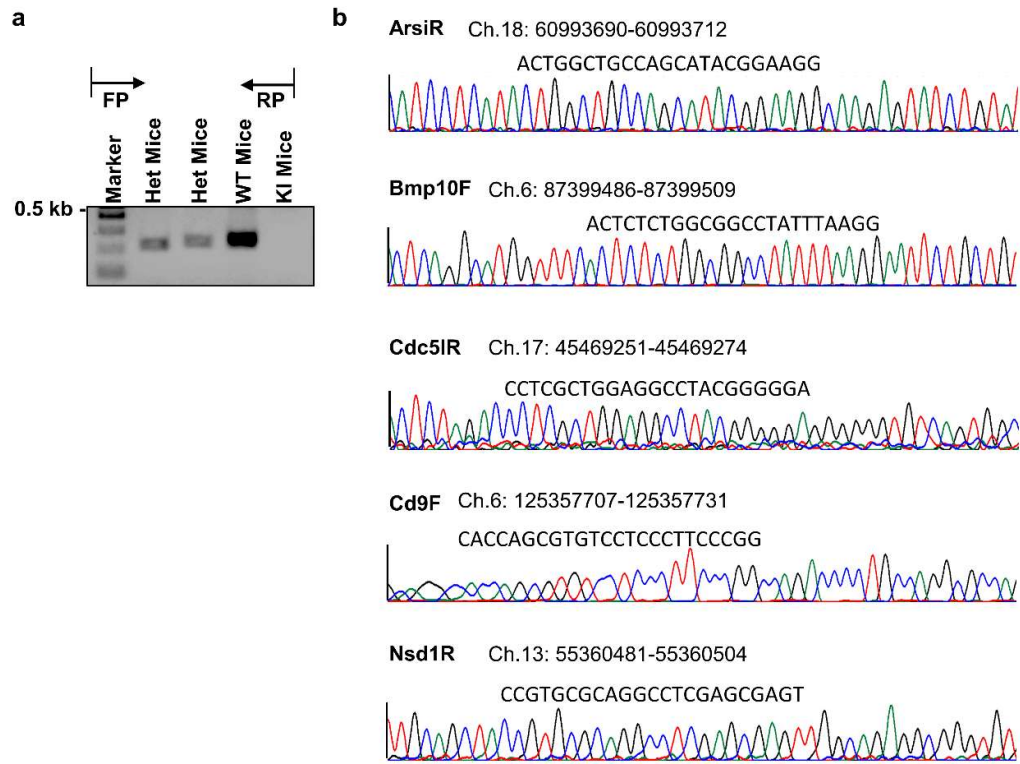


Supplementary Fig. 1: ROR γ t interacts with Raftlin1. **a**, Expression of *Rftn2* in CD4⁺ T cells isolated from colonic lamina propria of *C. rodentium* infected, SFB colonized, and uninfected WT mice; n=4 mice per group, p=0.51. **b**, Immunoassay in the lysates of CD4⁺ T cells isolated from colonic lamina propria of *C. rodentium* infected and SFB colonized WT mice were subjected to immunoprecipitation with anti-ROR γ t antibody and immunoblot analysis with anti-Raftlin1 or anti-ROR γ t antibody. Immunoblots are from one experiment representative of three independent experiments with similar results. **c**, The LBD domain of ROR γ t (PDB: 5X8U) and Raftlin1 (AlphaFold2 predicted structure) are docked using HADDOCK2.4. The docked interface is shown as a cartoon diagram. The LLNSL motif of Raftlin1 makes non-covalent interaction with the AF2 domain and α -helix 12 of ROR γ t. The amino acids involved in non-covalent interactions are shown in the stick model. ROR γ t Helix-12 is shown in color grey, AF2 domain in color green, Raftlin1 in color salmon, and LLNSL motif of Raftlin1 in color yellow. **d**,

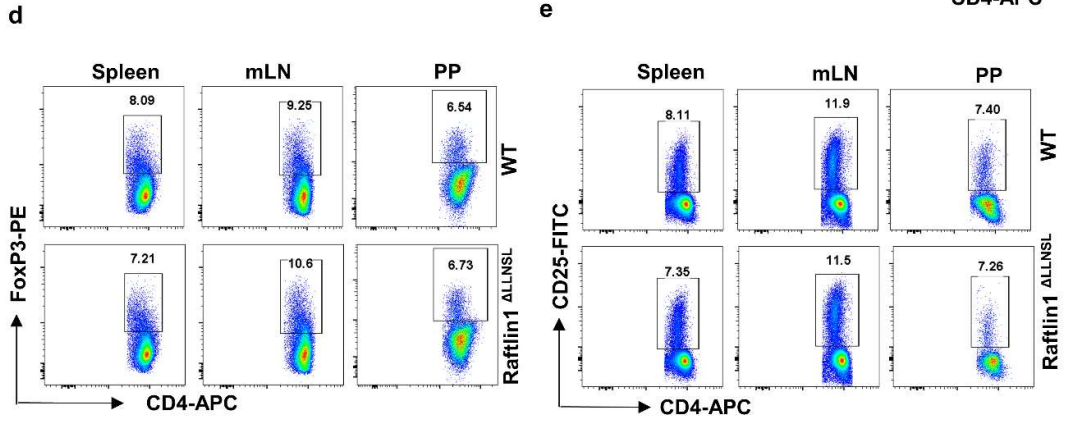
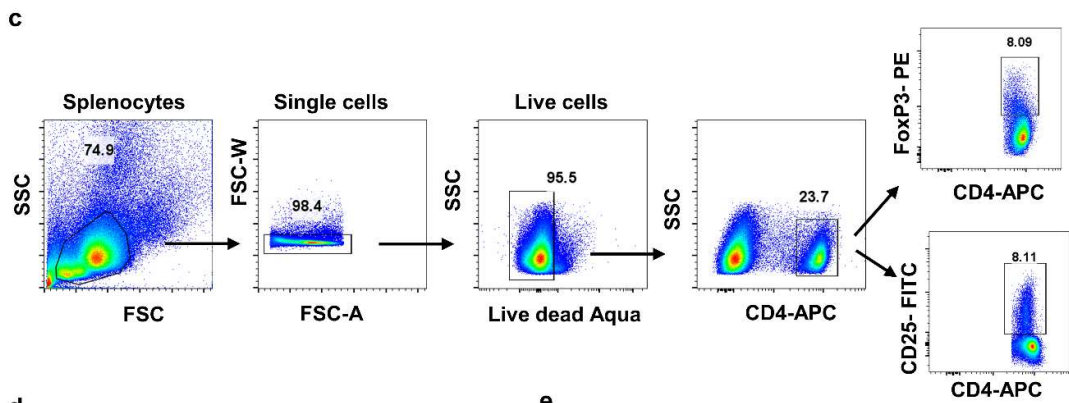
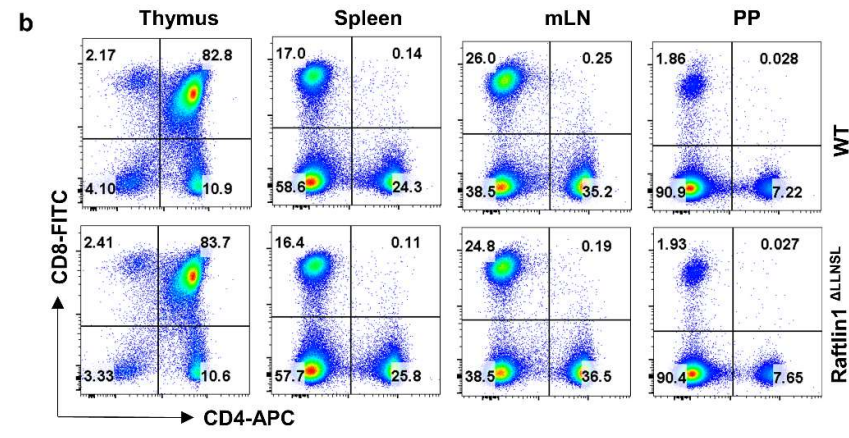
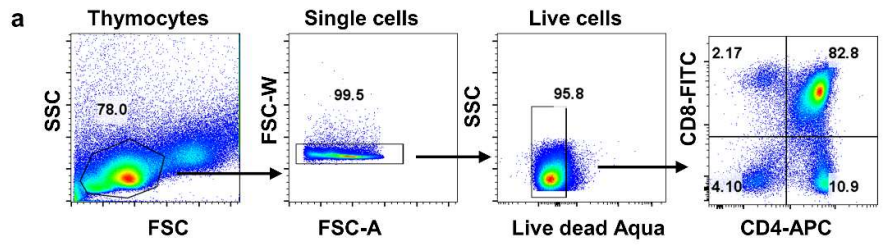
the protein-protein interface between ROR γ t and Raftlin1 is disrupted after deletion of the LLNSL motif in Raftlin1. Statistical significance (a) was determined by unpaired Student's t-test (two-tailed) with $p < 0.05$ considered statistically significant. ns- not significant, error bars are mean \pm SD. Source data are provided as a Source data file.

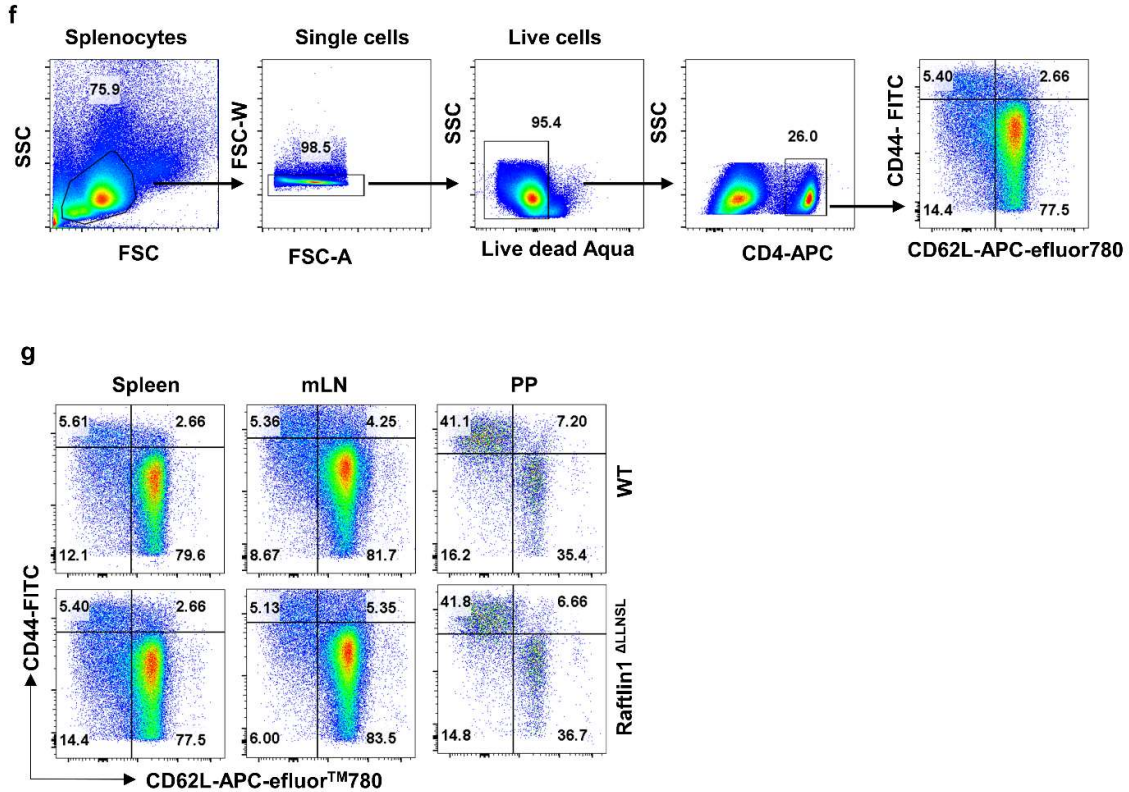


Supplementary Fig. 2: Raftlin1 expression in Crohn's disease patients. GEO dataset (GSE59071) was analyzed, and the normalized expression value of *RFTN1* is shown (n=11 healthy control and n=8 Crohn's disease patient's samples, p=0.0003). Statistical significance was determined by unpaired Student's t-test (two-tailed) with $p < 0.05$ considered statistically significant. *** $p < 0.001$, error bars are mean \pm SD. Source data are provided as a Source data file.

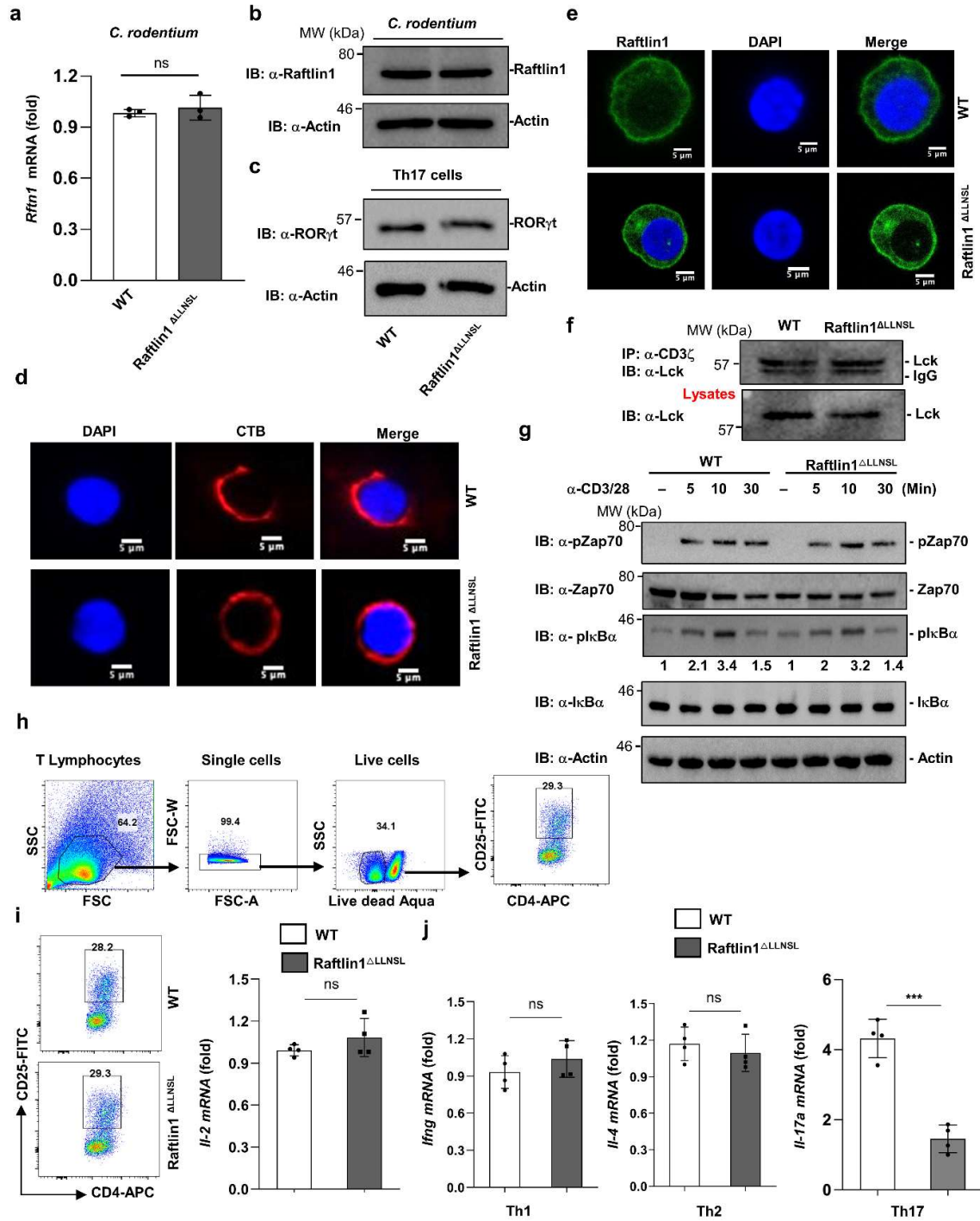


Supplementary Fig. 3: Genotyping and off-target gRNA analysis in the *Raftlin1*^{ΔLNSL} mice. **a**, Genotyping of *Raftlin1*^{ΔLNSL} mice by PCR. **b**, Representative Sanger sequencing chromatograms to check the potential off-target sites of gRNA in *Raftlin1*^{ΔLNSL} mice. No off-target cleavage sites were detected.





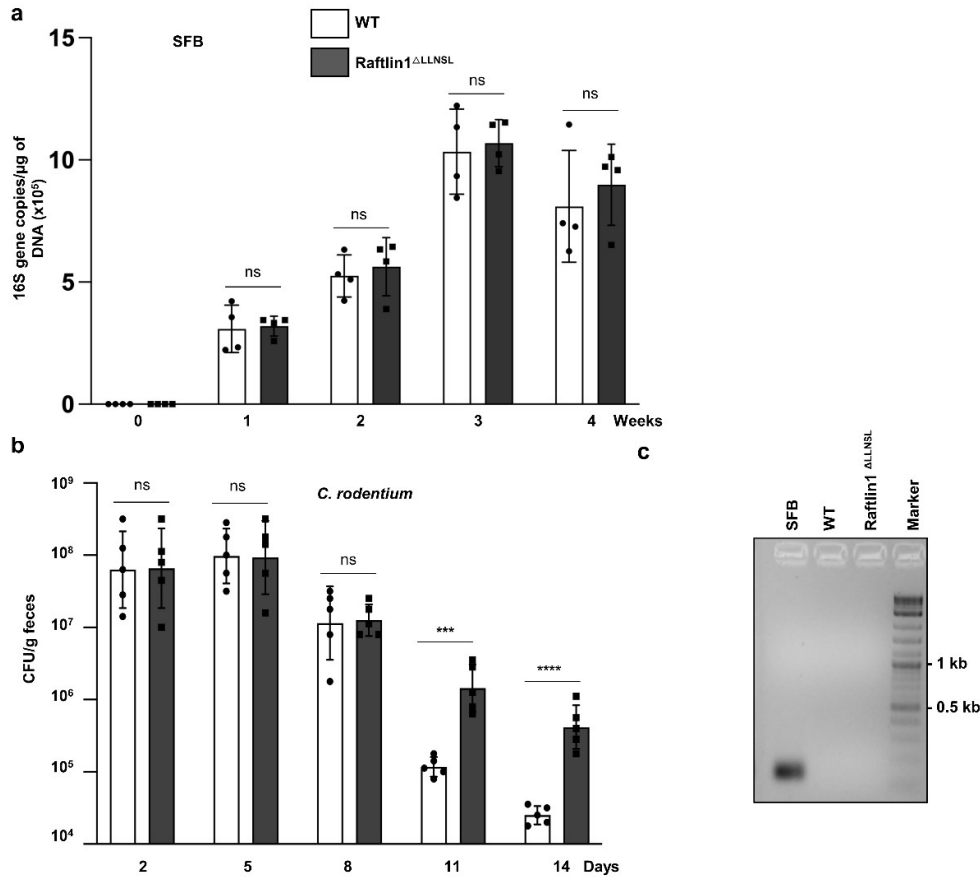
Supplementary Fig. 4: Flow cytometric analysis of T cells in the spleen, mesenteric lymph node (mLN), Peyer's patches (PP), and thymus of WT and *Raf^{flin1}^{ALLNSL}* mice. a, Gating strategy of cells from thymus of WT and *Raf^{flin1}^{ALLNSL}* mice stained with Live-Dead Aqua, anti-CD4, and anti-CD8 antibodies. **b,** Representative images of cells from thymus, spleen, mLN, and PP of WT and *Raf^{flin1}^{ALLNSL}* mice were stained with antibodies against CD4 and CD8 and analyzed by flow cytometry. **c,** Gating strategy of cells from the spleen of WT and *Raf^{flin1}^{ALLNSL}* mice stained with Live-Dead Aqua, anti-CD4, anti-FoxP3, and anti-CD25 antibodies. **d-e,** Representative image of cells from spleen, mLN, and PP of WT and *Raf^{flin1}^{ALLNSL}* mice were stained with antibodies against CD4, FoxP3 and CD25, and analyzed by flow cytometry. **f,** Gating strategy of cells from the spleen of WT and *Raf^{flin1}^{ALLNSL}* mice stained with Live-Dead Aqua, anti-CD4, anti-CD44, and anti-CD62L antibodies. **g,** Representative image of cells from spleen, mLN, and PP of WT and *Raf^{flin1}^{ALLNSL}* mice stained with antibodies against CD4, CD44, and CD62L, and analyzed by flow cytometry. b, d, e, g n=5 mice per group.



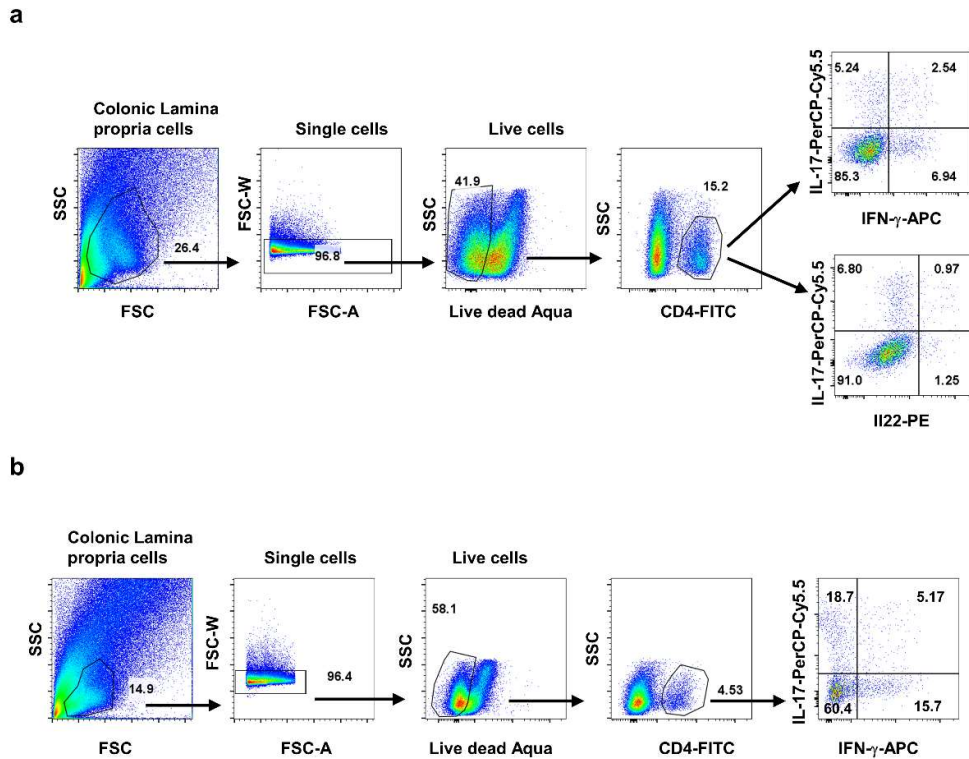
Supplementary Fig. 5: Lipid raft composition and TCR signaling in *Raftlin1*^{ALLNSL} T cells.

a-b, Expression of *Raftlin1* in *C. rodentium* infected WT and *Raftlin1*^{ALLNSL} mice by real-time PCR and western blotting, respectively. $n = 3$ mice per group, $p = 0.508$. **c**, Expression of *ROR γ t* in WT and *Raftlin1*^{ALLNSL} Th17 cells. **d**, *CD4*⁺ T cells were purified from WT and *Raftlin1*^{ALLNSL}

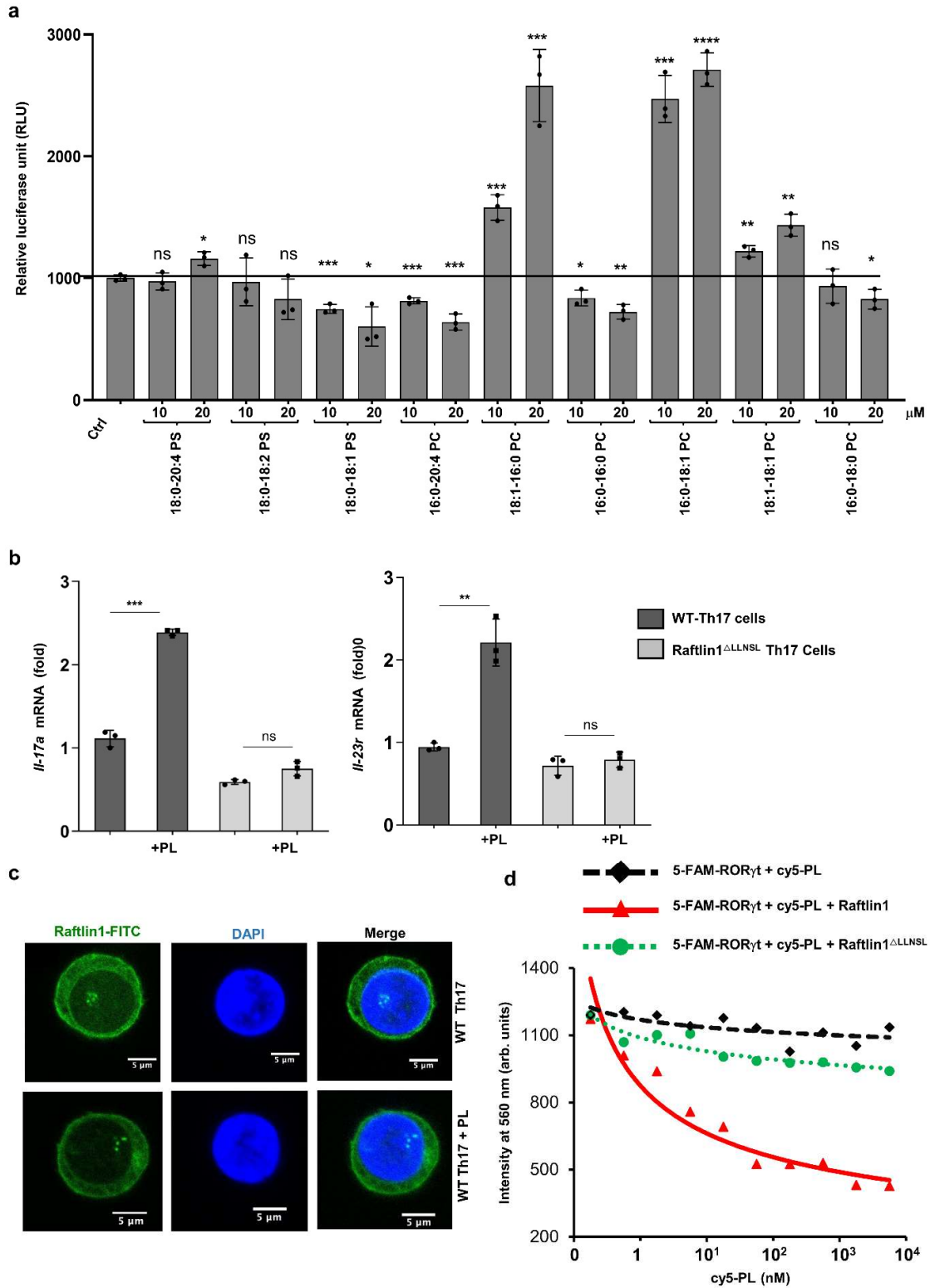
mice and stimulated with anti-CD3 and anti-CD28 antibodies for 48 hours. The membrane GM1 was stained with cholera toxin B (CTB) to check lipid raft formation by microscopy. The image is representative of three independent experiments. Scale bars, 5 μ m. **e**, Confocal microscopy analysis of the localization of Raftlin1 in WT and Raftlin1^{ALLNSL} in CD4⁺ T cells. The image is representative of three independent experiments. Scale bars, 5 μ m. **f**, Lysates of activated CD4⁺ T cells were isolated from WT and Raftlin1^{ALLNSL} mice, immunoprecipitated with anti-CD3 ζ antibody, and immunoblotted with anti-Lck antibody. **g**, CD4⁺ T cells from WT and Raftlin1^{ALLNSL} mice were stimulated with anti-CD3/anti-CD28 antibody for the indicated time. The cell lysates were prepared and analyzed by immunoblot with the indicated antibodies. **h**, Gating strategy for flow cytometry of CD4⁺ T cells from WT and Raftlin1^{ALLNSL} mice stained with Live-Dead Aqua, anti-CD4, and anti-CD25 antibodies. **i**, Expression of CD25 and *Il-2* on activated CD4⁺ T cells from WT and Raftlin1^{ALLNSL} mice; n=4 mice per group, p=0.244. **j**, CD4⁺ T cells isolated from WT and Raftlin1^{ALLNSL} mice and differentiated under Th1, Th2 and Th17 polarizing conditions. The expression of *Ifng*, *Il-4*, and *Il-17a* was analyzed by real-time PCR; n=4 mice per group, p=0.327 (*Ifng*), p=0.494 (*Il-4*), p=0.0001 (*Il-17a*). **b**, **c**, **f**, **g** Immunoblots are from one experiment representative of three independent experiments with similar results. Statistical significance was determined by unpaired Student's t-test (two-tailed) in **a**, **i**, **j** with p < 0.05 considered statistically significant. ***p < 0.001, ns- not significant, error bars are mean \pm SD. Source data are provided as a Source data file.



Supplementary Fig. 6: Analysis of SFB colonization and *C. rodentium* load in WT and Raftlin1^{ΔLLNSL} mice. **a**, Relative abundance of SFB in the feces of WT and Raftlin1^{ΔLLNSL} mice after gavage of feces from SFB monocolonized mice by quantifying SFB specific 16S using real-time PCR; n=4 biologically independent mice; p=0.83 (week 1), p=0.62 (week 2), p=0.74 (week 3), p=0.55 (week 4) **b**, Quantification of *C. rodentium* loads (CFU/g) in the stool of WT and Raftlin1^{ΔLLNSL} mice after the indicated time of the post-infection; n=5 mice per group; p=0.95 (day 2), p=0.94 (day 5), p=0.87 (day 8), p=0.0001 (day 11), p=0.00003 (day 14). **c**, the absence of SFB in our mice colony was confirmed by PCR using SFB-specific 16s primers. Data are from one experiment representative of three independent experiments with similar results. **a**, **b** Statistical significance was determined by unpaired Student's t-test (two groups) with p < 0.05 considered statistically significant. ***p < 0.001, ****p < 0.0001, ns- not significant, error bars are mean±SD. Source data are provided as a Source data file.

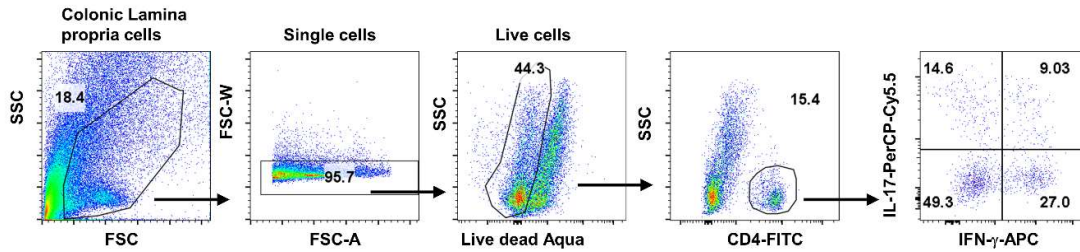


Supplementary Fig. 7: Flow cytometry gating strategy for the analysis of IL-17, IL-22, and IFN- γ in CD4⁺ T cells of colonic lamina propria cells (Fig. 3e, f, 4g). **a, Gating strategy of colonic lamina propria cells from WT and Raftlin1 ^{Δ LLNSL} mice stained with Live-Dead Aqua, anti-CD4, anti-IL-17, anti-IL22, and anti-IFN- γ antibodies, and analyzed by flow cytometry. **b**, Gating strategy of colonic lamina propria cells from Rag1^{-/-} mice adoptively transferred CD4⁺CD25⁻CD45RB^{hi} cells from WT and Raftlin1 ^{Δ LLNSL} mice stained with Live-Dead Aqua, anti-CD4, anti-IL-17 and anti-IFN- γ antibodies and analyzed by flow cytometry. Data are from one experiment representative of three independent experiments with similar results.**



Supplementary Fig. 8: IL-17 promoter-luciferase reporter activity induced by lipids in Th17 cells. **a**, Jurkat T cells were cultured in lipid-depleted conditional media and transfected

with various combinations (below plot) of IL-17-promoter-driven luciferase plasmid (pGL4-mIL17p), the plasmid encoding Flag-ROR γ t along with Raftlin1 and cultured in the presence of various lipids (10 μ M or 20 μ M) for 16 h. The relative luciferase activity was measured to quantify the ROR γ t transcriptional activity. Results are presented in relative luciferase units (RLU). n= 3 biological replicates; (18:0-20:4 PS, 10 μ M p=0.57, 20 μ M p= 0.01), (18:0-18:2 PS, 10 μ M p=0.80, 20 μ M p= 0.15), (18:0-18:1 PS, 10 μ M p=0.0007, 20 μ M p= 0.014), (16:0-20:4 PS, 10 μ M p=0.0009, 20 μ M p= 0.0009), (18:1-16:0 PS, 10 μ M p=0.0008, 20 μ M p= 0.0008), (16:0-18:1 PS, 10 μ M p=0.0002, 20 μ M p<0.0001), (18:1-18:1 PS, 10 μ M p=0.002, 20 μ M p= 0.0014), (16:0-18:0 PS, 10 μ M p=0.46, 20 μ M p= 0.02). **b**, Naive CD4⁺ T cells were isolated from WT or Raftlin1^{ALLNSL} mice and differentiated under Th17 polarizing conditions followed by the addition of phospholipids (PL) 18:1-16:0-PC, 16:0-18:1-PC, and 18:1-18:1-PC at 48 h in lipid-depleted fresh media. The expression of *Il-17a* and *Il-23r* was measured by real-time PCR at 96 h. n= 3 biological replicates; p=0.00003 (*Il-17a*), p=0.0016 (*Il-23r*). **c**, Localization of Raftlin1 in WT Th17 cells cultured in the presence or absence of phospholipid by confocal microscopy. The image is representative of three independent experiments. Scale bars, 5 μ m. **d**, Fluorescence emission intensity of 5-FAM labeled ROR γ t (5-FAM-ROR γ t) at 520 nm (Excitation = 470 nm) at increasing concentration of Cy5 labeled phospholipids (Cy5-PL) in the presence of GST-Raftlin1 (red triangles), GST-Raftlin1^{ALLNSL} (green dots), and control GST only (black dots). The concentrations of Cy5-PL (0-10⁴ nM), 5-FAM-ROR γ t (5 nM), GST-Raftlin1 (10 nM), and GST-Raftlin1^{ALLNSL} (10 nM) were used. Data are from one experiment representative of three independent experiments with similar results. **a**, **b** Statistical significance was determined by unpaired Student's t-test (two-tailed) with p < 0.05 considered statistically significant. *p < 0.05, **p < 0.01, ***p < 0.001, ****p < 0.0001, ns- not significant, error bars are mean \pm SD. Source data are provided as a Source data file.



Supplementary Fig. 9: Flow cytometry gating strategy for the analysis of IL-17 and IFN- γ in CD4⁺ T cells of colonic lamina propria cells (Fig. 5h). Gating strategy of colonic lamina propria cells from *Rag1*^{-/-} mice adoptively transferred phospholipid treated Th17 cells from WT and *Raftlin1* ^{Δ LLNSL} mice stained with Live-Dead Aqua, anti-CD4, anti-IL-17, and anti-IFN- γ antibodies, and analyzed by flow cytometry. Data are from one experiment representative of three independent experiments with similar results.

Supplementary Table 1: List of lipids detected in Raftlin1 and ROR γ t immunoprecipitated samples from *C. rodentium* infected WT mice:

S. No	Lipids	IP: α-Raftlin1 (pmol/mg)	IP: α-RORγt (pmol/mg)	IP: IgG
1	1-stearoyl-2-arachidonoyl-sn-glycero-3-phospho-L-serine (16:0-20:4-PS)	56.41	50.93	0
2	1-stearoyl-2-linoleoyl-sn-glycero-3-phospho-L-serine (18:0-18:2-PS)	32.78	16.42	0
3	1-stearoyl-2-oleoyl-sn-glycero-3-phospho-L-serine (18:0-18:1-PS)	21.68	4.25	0
4	1-palmitoyl-2-arachidonoyl-sn-glycero-3-phosphocholine (16:0-20:4-PC)	40.39	21.14	0
5	1-oleoyl-2-palmitoyl-sn-glycero-3-phosphocholine (18:1-16:0-PC)	27.06	8.69	0
6	1,2-di-palmitoyl-sn-glycero-3-phosphocholine (16:0-16:0-PC)	21.85	5.21	0
7	1-Palmitoyl-2-oleoyl-sn-glycero-3-phosphocholine (16:0-18:1-PC)	20.22	18.54	0
8	1,2-dioleoyl-sn-glycero-3-phosphatidylcholine (18:1-18:1-PC)	63.19	11.51	0
9	1-palmitoyl-2-stearoyl-sn-glycero-3-phosphocholine (16:0-18:0-PC)	20.22	18.54	0

Supplementary Table 2: List of predicted Off-target genes

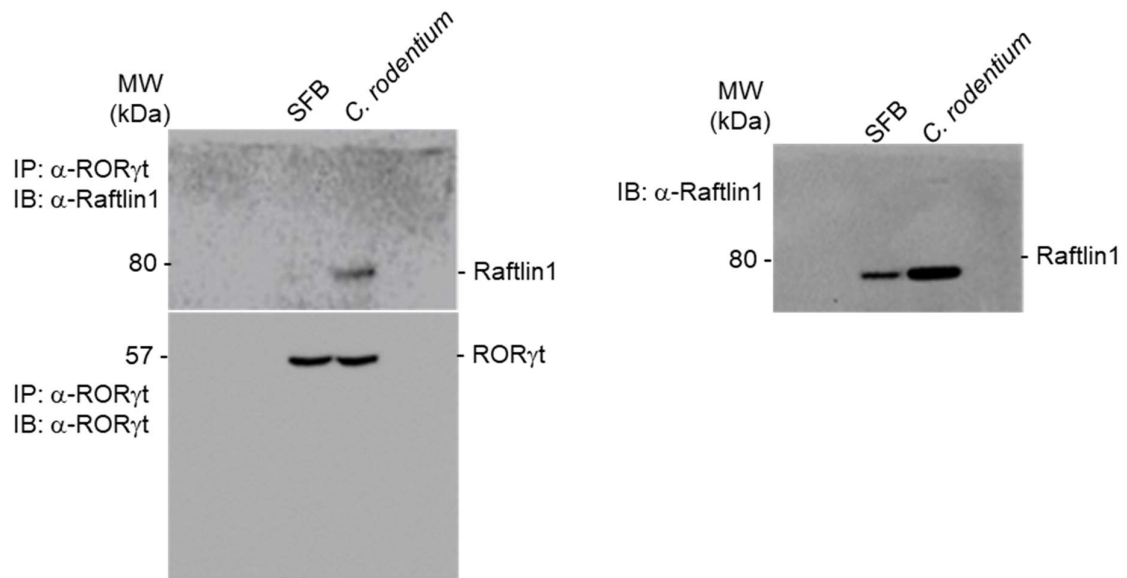
S.No	Genes	CFD Score
1	<i>Ccl9</i>	0.63
2	<i>Septin5</i>	0.13
3	<i>Cd9</i>	0.09
4	<i>Arsi</i>	0.09
5	<i>Tcaf3</i>	0.09
6	<i>Prx</i>	0.08
7	<i>Bmp10</i>	0.06
8	<i>Nsd1</i>	0.04
9	<i>Cdc5l</i>	0.03
10	<i>Prnp/Prn</i>	0.01
11	<i>Pdgfb</i>	0.00
12	<i>Myo16</i>	0.00
13	<i>Tns3</i>	0.00
14	<i>Gdnf</i>	0.00
15	<i>Kctd17</i>	0.00

Supplementary Table 3: PCR primers sequence of the predicted off-target genes

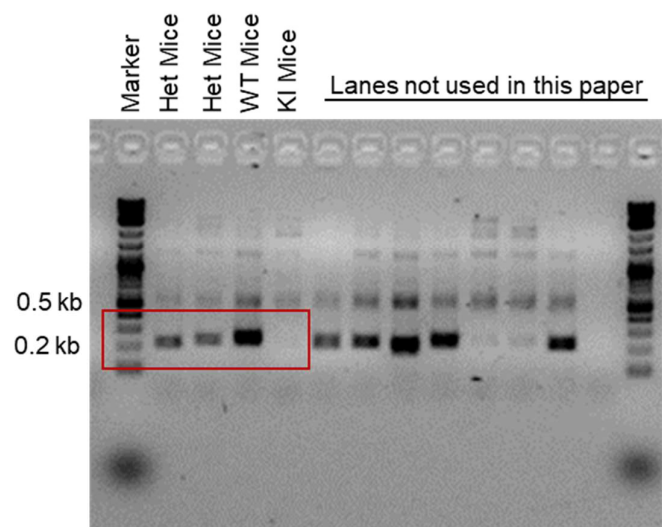
S.no	Genes	Primers
1	<i>Ccl9</i>	Fw:5'-CGTTTGATCCAGATCTTGAGGC-3' Rv:5'-GGCTCAGCAGTTAAGAGCACTG-3'
2	<i>Septin5</i>	FW:5'-CCTCTTCTGCCTCGGGTTTT-3' Rv:5'-CGGGTGAAGAGCAAGGAGAG-3'
3	<i>Cd9</i>	Fw:5'-AGCAATCTGTCACCTGGTTCG-3' Rv:5'-CAGTCTGTGTAGCCCTTGGG-3'
4	<i>Arsi</i>	Fw:5'-TGATCACCAGCACACCAGTC-3' Rv:5'-GGCCACCGACCATTGTTAGA-3'
5	<i>Tcaf3</i>	FW: 5'-TGGCAGAAGTTGTGGAAGAGG-3' Rv:5'-CTGCACAGACGCTATAGGGG-3'
6	<i>Prx</i>	Fw:5'-CACTCGACCTCTCTGGCAAG-3' Rv:5'-GGGCTGACTCCAAGATCTTGT-3'
7	<i>Bmp10</i>	Fw:5'-ACCTCGGAGTAGCACCCCTAG-3' Rv:5'-GGGAGGGGAGAGTCTTCAGT-3'
8	<i>Nsd1</i>	Fw:5'-GCAGCAGCCATGTTTGTCAA-3' Rv:5'-CGGGAAAAGTTCTCCAGGCT-3'
9	<i>Cdc5l</i>	Fw:5'-CAGATGGTTGGCTCAGTGGT-3' Rv:5'-ATGGTGTCATCCCTCTGCAC-3'

Uncropped western blots and PCR agarose gels of Supplementary figures

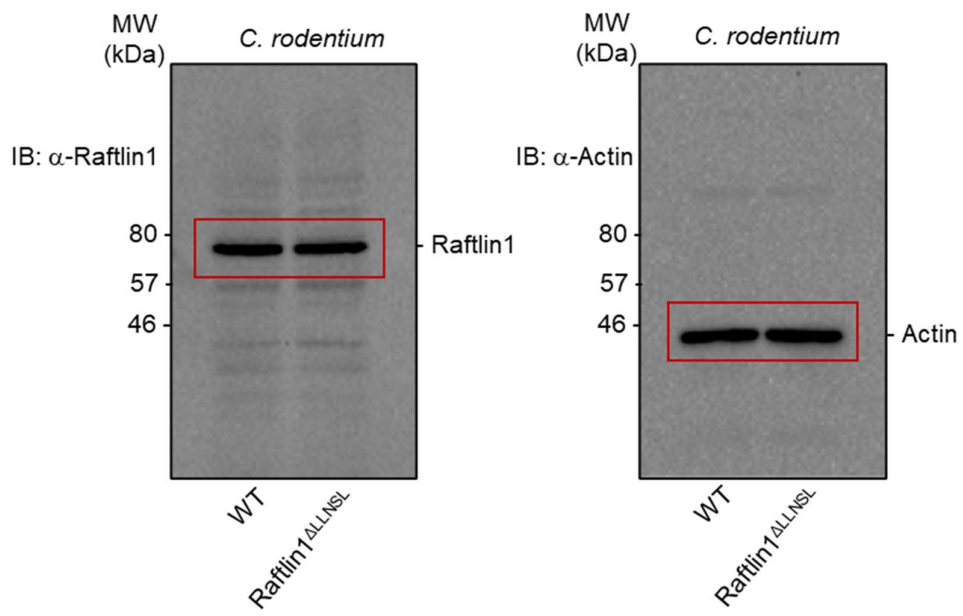
Supplementary Fig. 1b



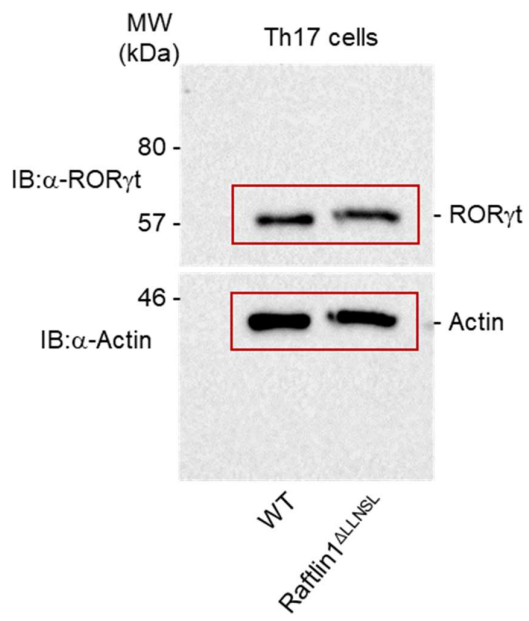
Supplementary Fig. 3a



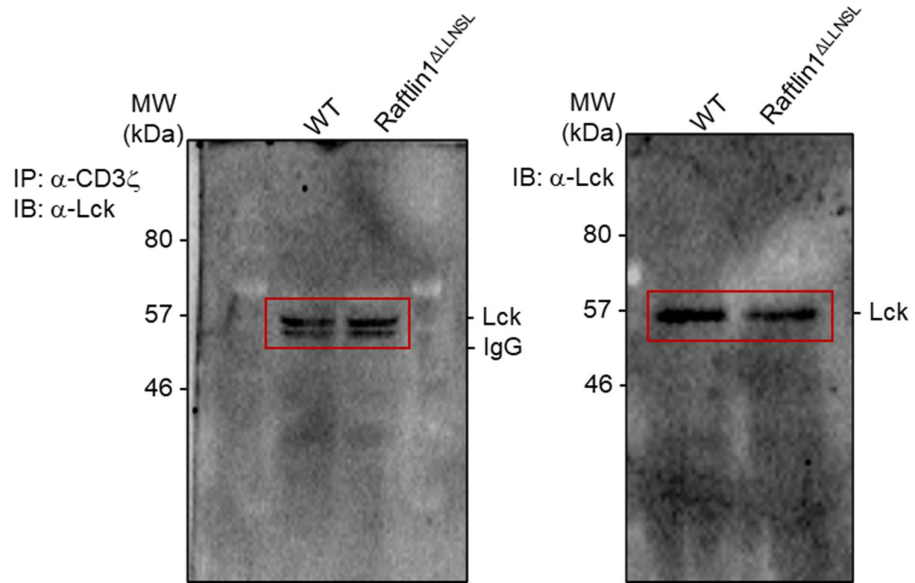
Supplementary Fig. 5b



Supplementary Fig. 5c



Supplementary Fig. 5f



Supplementary Fig. 5g

