

Figure S1. Monocyte subset gating strategy by flow cytometry. Left panel, whole population of monocytes from human blood (isolated by elutriation); right panel, subset determination by staining with antibodies anti-CD14 and CD16. P1, whole population; P2, classical CD14<sup>++</sup>/CD16<sup>-</sup> monocytes; P3, intermediate CD14<sup>++</sup>/CD16<sup>+</sup> monocytes; P4, non-classical CD14<sup>+</sup>/CD16<sup>++</sup> monocytes.

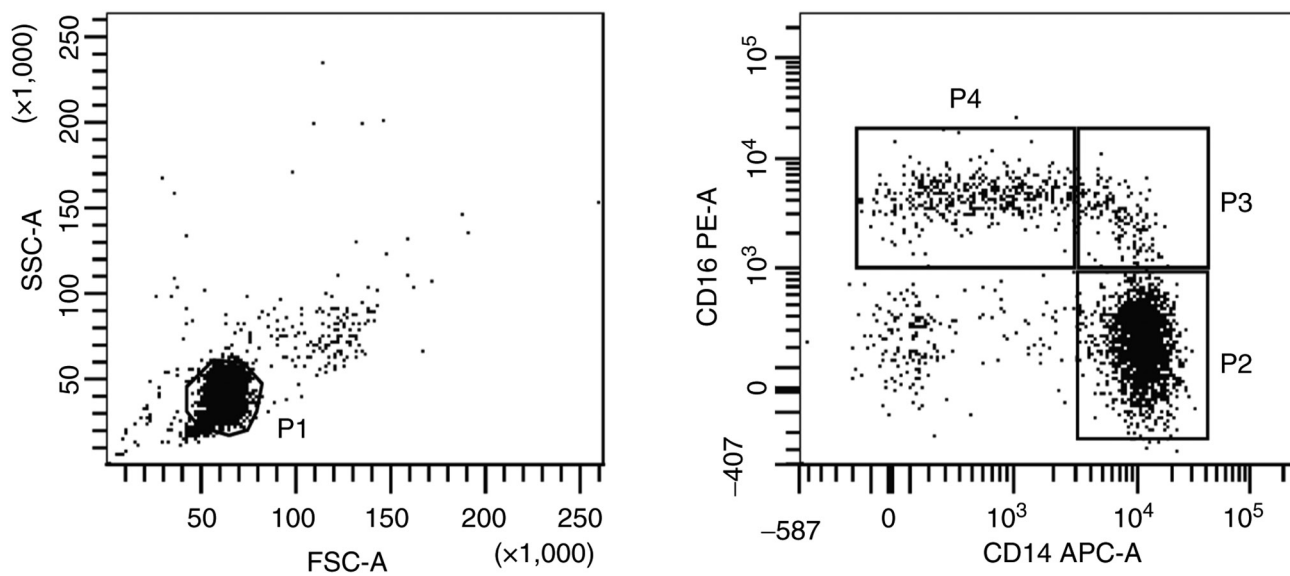
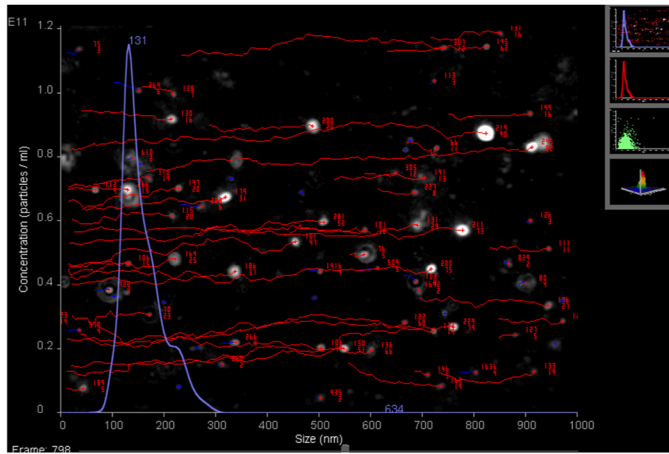


Figure S2. Representative nanoparticle tracking analysis measurement of TEVs<sub>HCT116</sub>. (A and B) Sample video screenshots from the NTA measurement obtained with NanoSight LM10HS equipped with the LM14 488-nm laser module (Malvern Instruments, Ltd.).

A



B

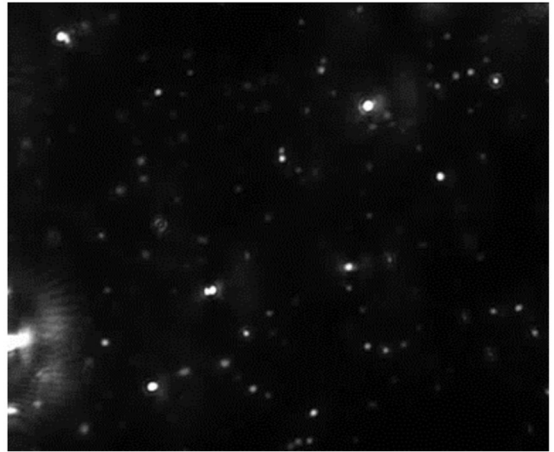


Figure S3. Efficiency of TEVs<sub>HCT116</sub> labeling with SYTO RNaselect dye by NTA and flow cytometry. (A) Representative NTA size profile of TEVs<sub>HCT116</sub> without dye. (B) Representative NTA size profile of SYTO RNA-labeled TEVs<sub>HCT116</sub>. (C) Representative histogram of TEVs<sub>HCT116</sub> in FL1 channel without labeling. (D) Representative histogram of SYTO RNA-labeled TEVs<sub>HCT116</sub> in FL1 channel. NTA, nanoparticle tracking analysis; TEVs, tumor-derived extracellular vesicles.

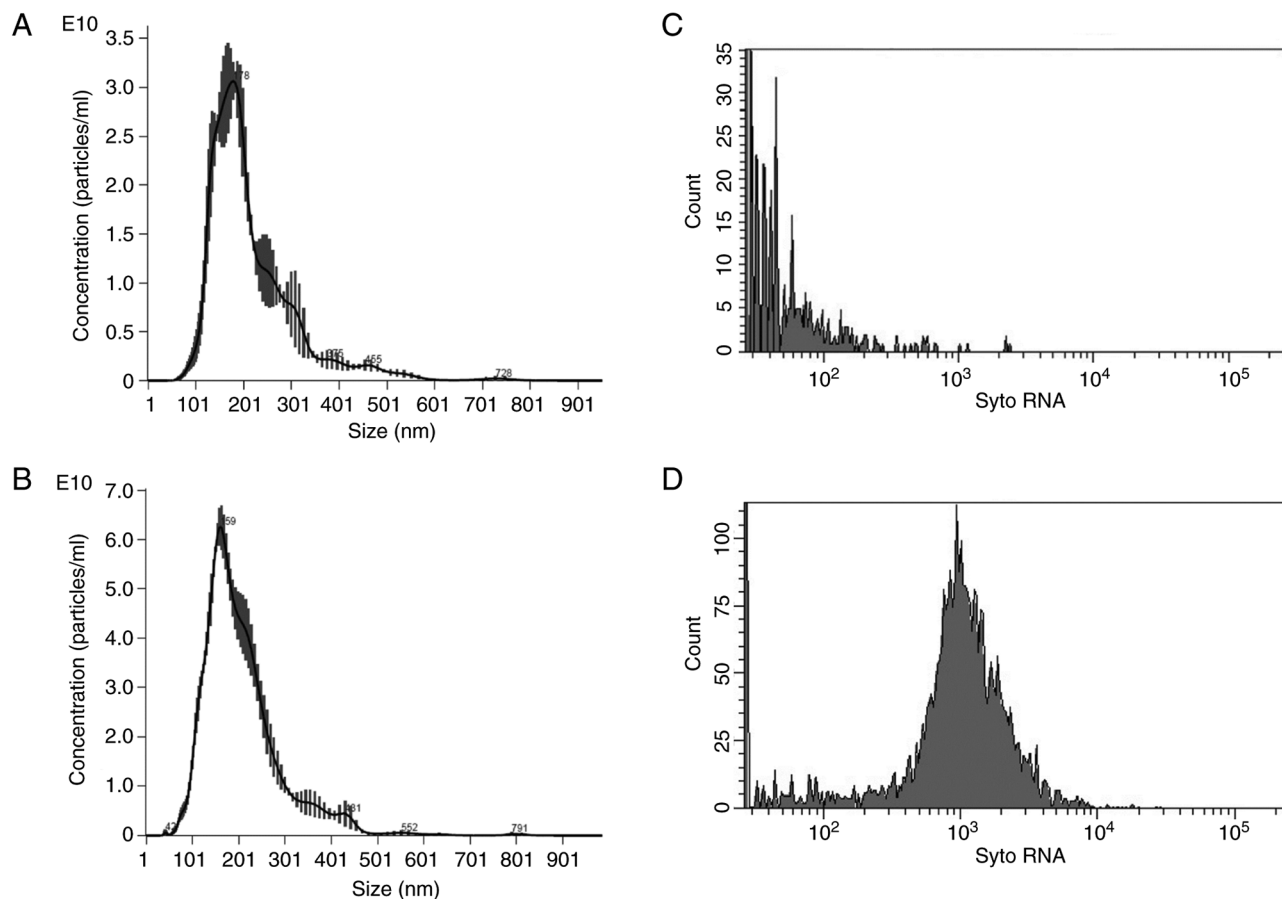


Figure S4. Representative CD44 expression on monocytes after 0, 2 and 18 h incubation at 37°C.

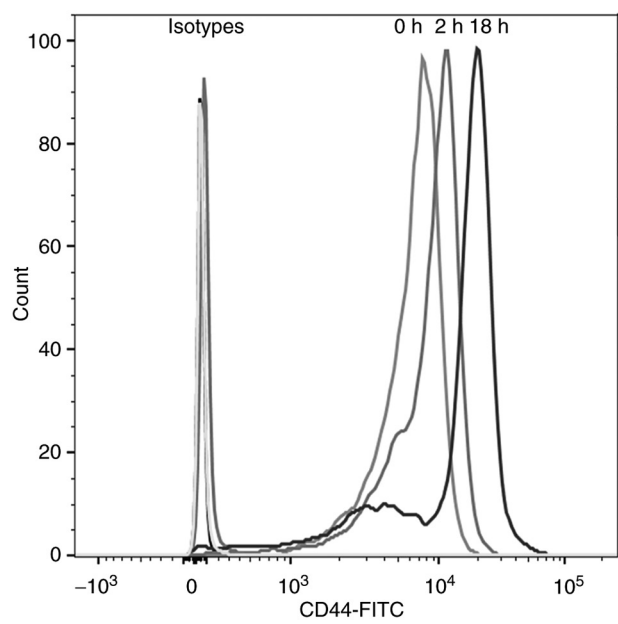


Figure S5. Kinetics of TEV<sub>HCT116</sub> and TEV<sub>SW1116</sub> attachment to monocytes. Data are presented as the mean  $\pm$  SEM of three experiments. TEVs, tumor-derived extracellular vesicles; MFI, mean fluorescence intensity.

