

1 **Supplementary Figure Legends**

2 **S1 Fig. K_{ATP} channels are important for HPV gene expression in cervical cancer cells.**

3 **A)** Expression levels of *E6* and *E7* mRNA in HeLa cells treated with tolbutamide (200 μ M)
4 measured by RT-qPCR. Samples were normalised against *U6* mRNA levels. **B)**
5 Representative western blots of E6 and E7 expression in HeLa cells treated with increasing
6 doses of tolbutamide. GAPDH served as a loading control. **C)** Mean DiBAC₄(3) fluorescence
7 levels in HeLa cells treated with increasing dose of tolbutamide. Samples were normalised to
8 DMSO control. Data represent means \pm SD of three biological replicates. *P<0.05, **P<0.01,
9 ***P<0.001, ****P<0.0001 (Student's t-test).

10

11 **S2 Fig. Depletion of SUR2 has no impact on HPV gene expression or proliferation in** 12 **cervical cancer cells.**

13 **A)** Relative expression of *ABCC9B* mRNA in HeLa and SiHa cells transfected with a pool of
14 SUR2-specific siRNA measured by RT-qPCR. Samples were normalised against *U6* mRNA
15 levels. **B)** Relative mean DiBAC₄(3) fluorescence levels in HeLa and SiHa cells transfected
16 with SUR2 siRNA. **C)** Relative expression of *E6* and *E7* mRNA in HeLa and SiHa cells
17 transfected with SUR2 siRNA measured by RT-qPCR. Samples were normalised against *U6*
18 mRNA levels. **D)** Representative western blots of E6 and E7 expression in HeLa and SiHa
19 cells transfected with SUR2 siRNA. GAPDH served as a loading control. **E-G)** Growth curve
20 analysis (**E**), colony formation assay (**F**) and soft agar assay (**G**) of HeLa and SiHa cells after
21 transfection of SUR2-specific siRNA. Data represent means \pm SD of three biological replicates
22 with individual data points displayed. *Ns* not significant, *P<0.05, **P<0.01, ***P<0.001
23 (Student's t-test).

24

25 **S3 Fig. Depletion of Kir6.2 reduces HPV gene expression and proliferation in cervical** 26 **cancer cells.**

27 **A)** Relative expression of *KCNJ11* mRNA in HeLa and SiHa cells transfected with a pool of
28 Kir6.2-specific siRNA measured by RT-qPCR. Samples were normalised against *U6* mRNA
29 levels. **B)** Relative mean DiBAC₄(3) fluorescence levels in HeLa and SiHa cells transfected
30 with Kir6.2 siRNA. **C)** Relative expression of *E6* and *E7* mRNA in HeLa and SiHa cells
31 transfected with Kir6.2 siRNA measured by RT-qPCR. Samples were normalised against *U6*
32 mRNA levels. **D)** Representative western blots of E6 and E7 expression in HeLa and SiHa
33 cells transfected with Kir6.2 siRNA. GAPDH served as a loading control. **E-G)** Growth curve
34 analysis (**E**), colony formation assay (**F**) and soft agar assay (**G**) of HeLa and SiHa cells after
35 transfection of Kir6.2-specific siRNA. Data shown is means ± SD of three biological replicates
36 with individual data points displayed where appropriate. *P<0.05, **P<0.01, ***P<0.001,
37 ****P<0.0001 (Student's t-test).

38

39 **S4 Fig. Stable suppression of SUR1 expression decreases the proliferation of cervical**
40 **cancer cells.**

41 Growth curve analysis (**A**), colony formation assay (**B**) and soft agar assay (**C**) of monoclonal
42 HeLa cell lines stably expressing either a non-targeting (shNTC) or a SUR1-specific shRNA.
43 Data shown is means ± SD of three biological replicates with individual data points displayed
44 where appropriate. *P<0.05, **P<0.01, ***P<0.001, ****P<0.0001 (Student's t-test).

45

46 **S5 Fig. K_{ATP} channel inhibition does not impact upon the survival of cervical cancer**
47 **cells.**

48 **A)** Representative western blots of PARP and caspase 3 cleavage in HeLa and SiHa cells
49 treated with DMSO or glibenclamide (10 µM) for the indicated durations. Staurosporine
50 treatment (STS, 1 µM for 6 hours) served as a positive control for apoptosis induction. GAPDH
51 served as a loading control. **B)** Flow cytometry analysis of Annexin V assay using HeLa and

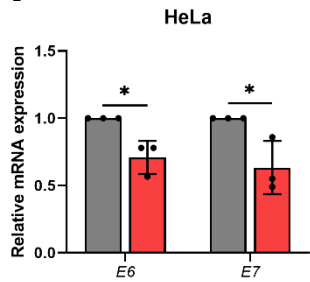
52 SiHa cells treated with DMSO or glibenclamide (10 μ M) for the indicated durations. Bars
53 represent means \pm SD of three biological replicates. *Ns* not significant (Student's t-test).

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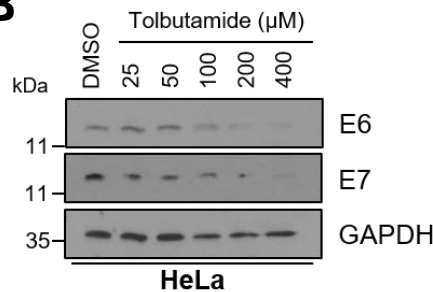
55 **S1 Table. List of primers used for RT-qPCR in this study.**

Supplementary Figure 1

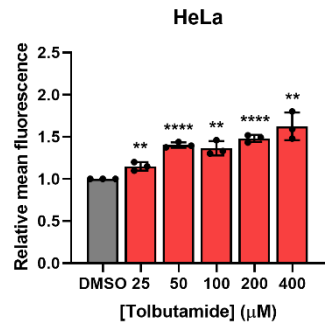
A



B

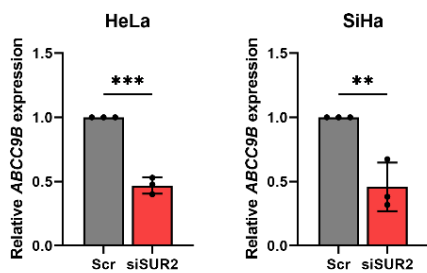


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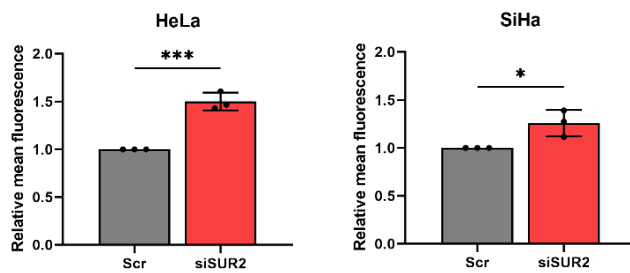


Supplementary Figure 2

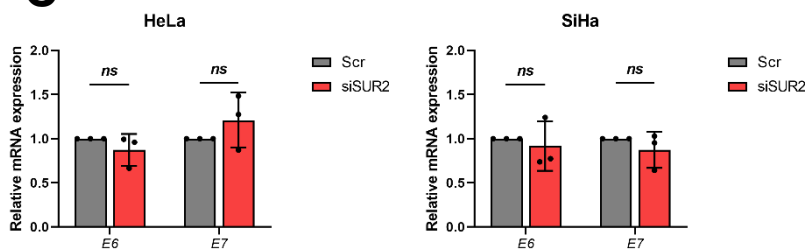
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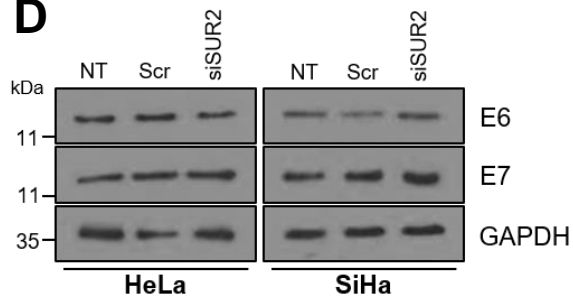
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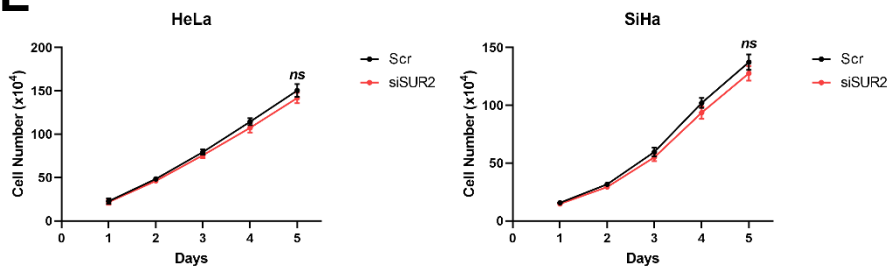
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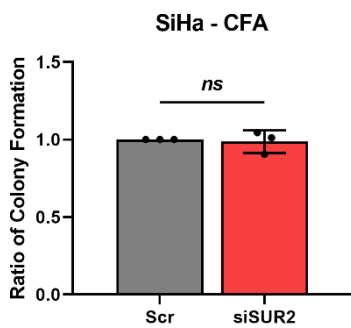
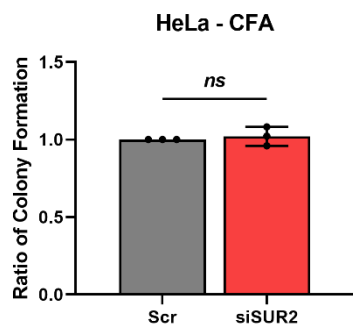
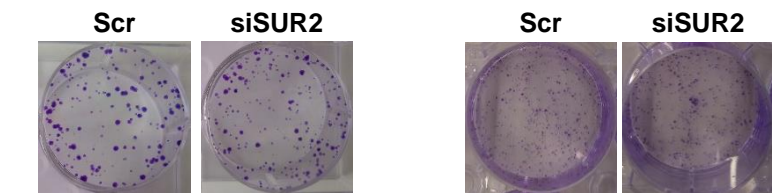
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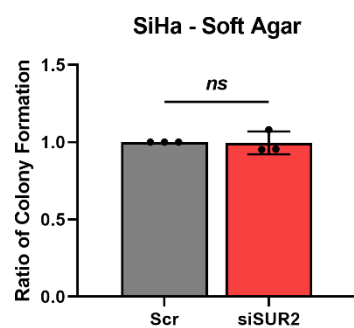
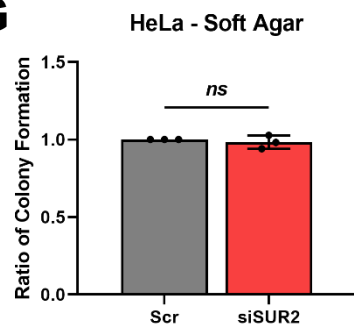
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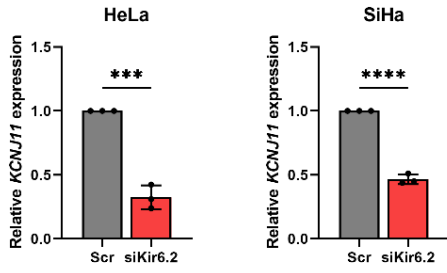


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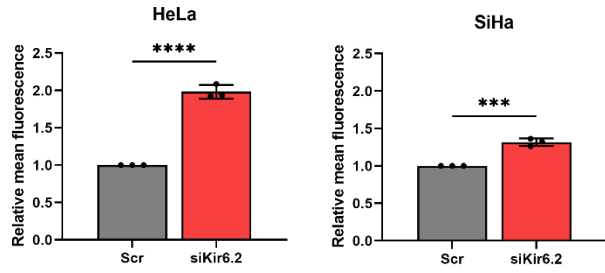


Supplementary Figure 3

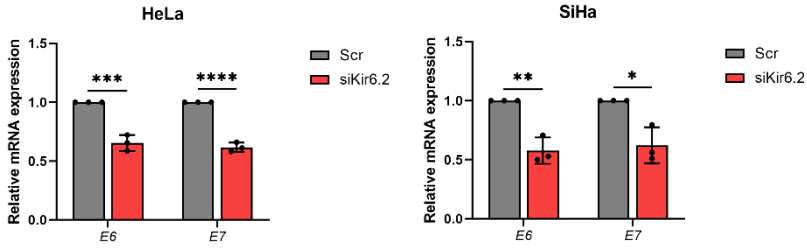
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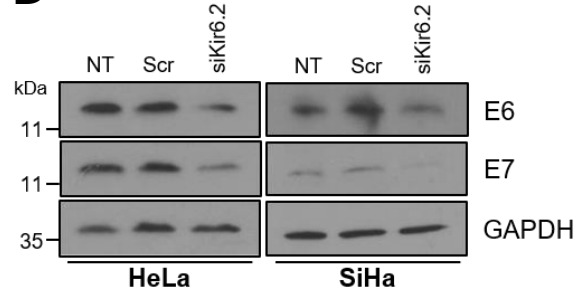
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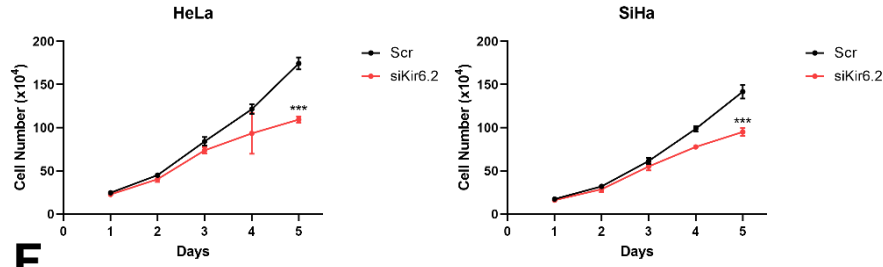
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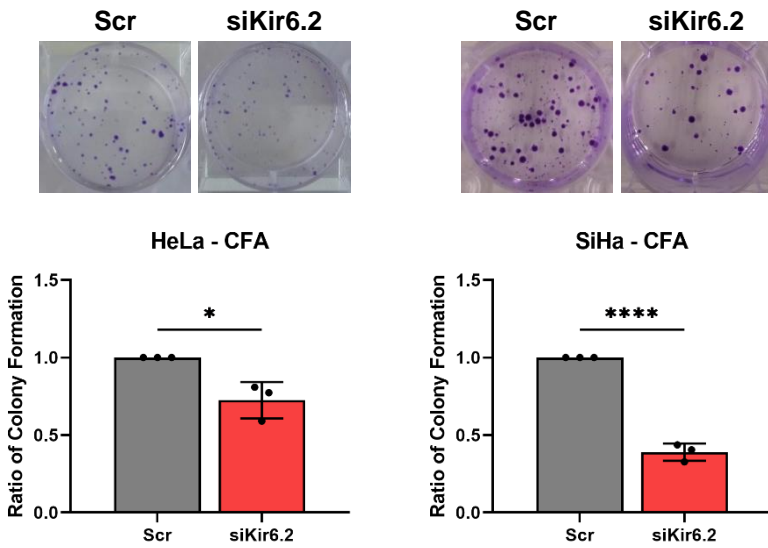
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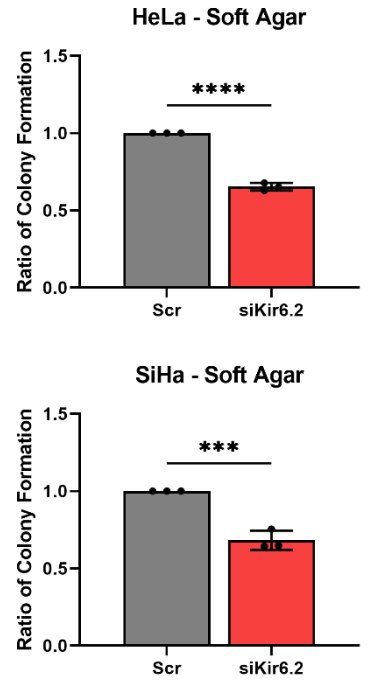
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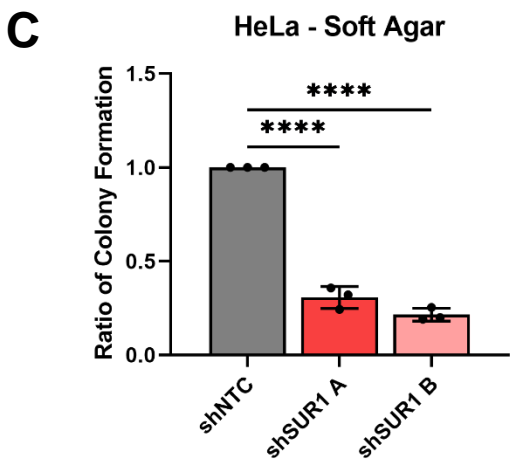
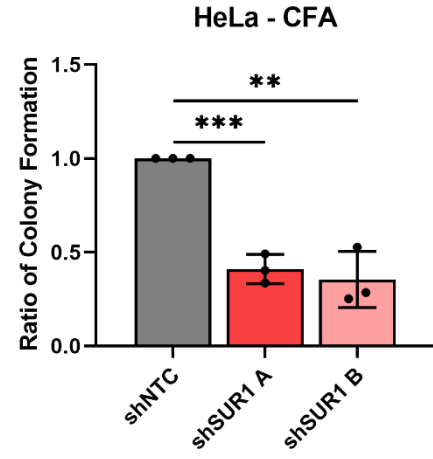
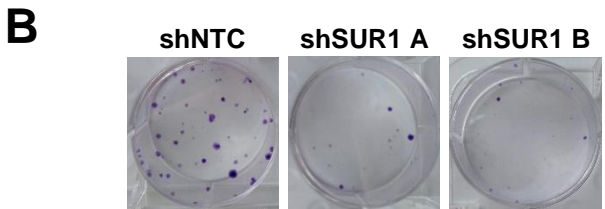
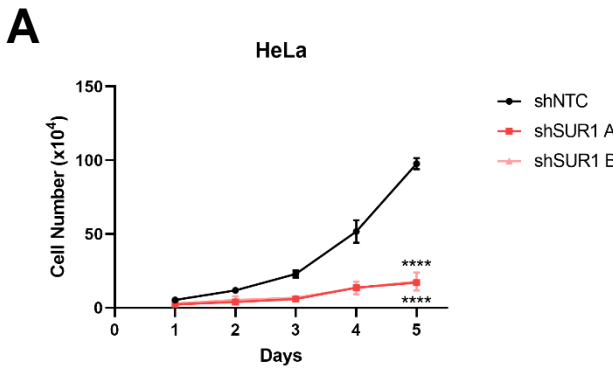
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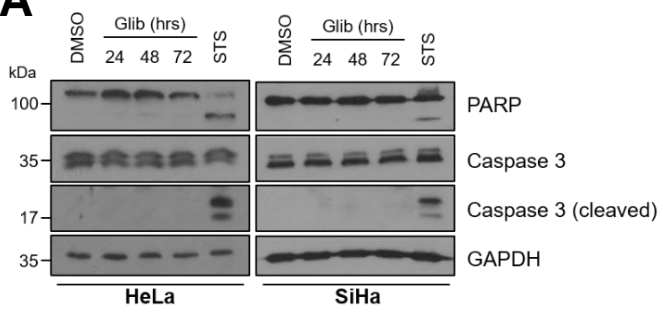


Supplementary Figure 4

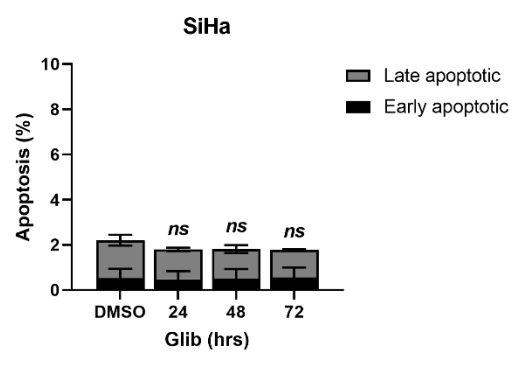
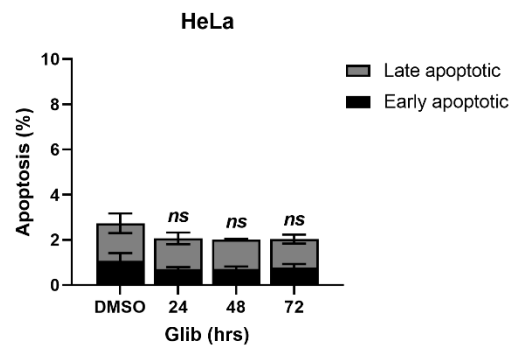


Supplementary Figure 5

A



B



Supplementary Table 1

Transcript	Forward primer (5'-3')	Reverse primer (5'-3')
HPV16 <i>E6</i>	CTGCAATGTTTCAGGACCCAC	GTTGTTTGCAGCTCTGTGCAT
HPV16 <i>E7</i>	ATTAAATGACAGCTCAGAGGA	GCTTTGTACGCACAACCGAAGC
HPV18 <i>E6</i>	TGGCGCGCTTTGAGGA	TG TTCAGTTCCGTGCACAGATC
HPV18 <i>E7</i>	GACCTAAGGCAACATTGCA	GCTCGTGACATAGAAGGTC
<i>KCNJ8</i>	CTGGCTGCTCTTCGCTATC	AGAATCAAAACCGTGATGGC
<i>KCNJ11</i>	CCAAGAAAGGCAACTGCAACG	ATGCTTGCTGAAGATGAGGGT
<i>ABCC8</i>	GGTGACCGAATCCCACCATC	CAGGGCAATTAGCAGCTTGG
<i>ABCC9A</i>	CTGGCTTTCTTCAGAATGGT	AAATACCCTCAGAAAAGACTAAAAC
<i>ABCC9B</i>	TGTGATGAAGCGAGGAAATA	TGACACTTCCATTCTGAGAGA
<i>GFP</i>	ACGTAAACGGCCACAAGTTC	AAGTCGTGCTGCTTCATGTG
<i>CCNA2</i>	TGGAAAGCAAACAGTAAACAGCC	GGGCATCTTCACGCTCTATTT
<i>CCNB1</i>	AAGAGCTTTAACTTTGGTCTGGG	CTTTGTAAGTCCTTGATTTACCATG
<i>CCND1</i>	CCGCTGGCCATGAACTACCT	ACGAAGGTCTGCGCGTGTT
<i>CCNE1</i>	GCCAGCCTTGGGACAATAATG	CTTGACGTTGAGTTTGGGT
<i>U6</i>	CTCGCTTCGGCAGCACA	AACGCTTCACGAATTTGCGT