## **Supplementary Information**

Akbari et al., A genome-wide association study of blood cell morphology identifies cellular proteins implicated in disease aetiology

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**Supplementary Fig. 1 | Histograms of technically adjusted ncCBC traits stratified by sex.** Probability density histograms of the 63 ncCBC traits stratified by sex (orange=female, blue=male). The data are from the technically adjusted traits (Methods) restricted to the participants who contribute data to the GWAS of the respective trait, the numbers of which are given in Supplementary Data 2.























**Supplementary Fig. 2 | Covariation between technically adjusted ncCBC traits and age stratified by sex.** Covariation between the 63 nCBC traits and participant age stratified by participant sex. Parameters of the stratified trait distributions were estimated in bins corresponding to years of age. The linearly interpolated coloured points show estimates of the within strata-means and the underlying coloured ribbons show the corresponding 95% confidence intervals. The dashed lines show estimates of the upper and lower quartiles. The data are from the technically adjusted traits (Methods) restricted to the participants who contribute data to the GWAS of the respective trait, the numbers of which are given in Supplementary Data 2.























**Supplementary Fig. 3** | **Covariation between technically adjusted ncCBC traits and BMI stratified by sex.** Covariation between the 63 nCBC traits and participant body mass index (BMI) stratified by participant sex. Parameters of the stratified trait distributions were estimated in bins corresponding to 1kg m<sup>-2</sup> of BMI. The linearly interpolated coloured points show estimates of the within strata-means and the underlying coloured ribbons show the corresponding 95% confidence intervals. The dashed lines show estimates of the upper and lower quartiles. The data are from the technically adjusted traits (Methods) restricted to the participants who contribute data to the GWAS of the respective trait, the numbers of which are given in Supplementary Data 2.



**Supplementary Fig. 4 | Genetic correlation within ncCBC traits.** A heatmap of estimates of genetic correlation between ncCBC phenotypes, made by applying LD Score regression to GWAS summary statistics<sup>1,2</sup>. The transparent rows and columns correspond to phenotypes (NE-FSC-DW, NE-SSC-DW, MO-FSC-DW) for which it was not possible to estimate genetic correlation due to an insufficiency of genetic association signals. The absolute genetic correlation between pairs of phenotypes of red cells is generally higher than the absolute genetic correlation between pairs of phenotypes of subtypes of white cells. There is limited evidence of strong absolute correlations between pairs of phenotypes of different cell-types. The starred cells are those for which a (two-sided) test of the null-hypothesis that the corresponding correlation is equal to zero is rejected at a Bonferroni corrected significance threshold (*P*-value < 1.28x10<sup>-5</sup>).



**Supplementary Fig. 5 | Phenotypic correlation within ncCBC traits.** A heatmap of the Pearson correlations between the ncCBC phenotypes in the sub-sample of the INTERVAL cohort used for the GWAS analysis. Correlations between pairs of phenotypes of red cells and pairs of phenotypes of platelets are stronger on average than correlations between pairs of phenotypes of white cells. There is low correlation between pairs of phenotypes.



**Supplementary Fig. 6 | Genetic correlation between ncCBC traits and cCBC traits.** A heatmap of estimates of genetic correlation between the ncCBC traits and the haematological traits studied by Vuckovic et al.<sup>3</sup> made by applying LD Score regression to GWAS summary statistics generated from the INTERVAL cohort<sup>4</sup>. There is limited genetic correlation between ncCBC phenotypes of white cells and previously studied standard haematological traits. The starred cells are those for which a (two-sided) test of the null-hypothesis that the corresponding correlation is equal to zero is rejected at a Bonferroni corrected significance threshold (*P*-value <  $2.27 \times 10^{-5}$ ).



**Supplementary Fig. 7 | Phenotypic correlation between ncCBC traits and cCBC traits.** A heatmap of the Pearson correlation between the ncCBC phenotypes and the haematological traits studied by Vuckovic et al.<sup>3</sup> in the sub-sample of the INTERVAL cohort used for the GWAS analysis<sup>4</sup>. There is limited Pearson correlation between ncCBC phenotypes of white cells and previously studied standard haematological traits.



Supplementary Fig. 8 | The joint distribution of the number of LD clumps by cell-type. A bar plot counting LD clumps by the cell-types of the ncCBC traits with which the variants they contain are associated. For all but 64 of the clumps, the variants in each clump were associated with traits of a single cell-type. The green fraction of each bar corresponds to the LD clumps that contain no variant in LD ( $r^2$ >0.8) with a variant identified by Vuckovic et al. or Chen et al.<sup>3,5</sup>



**Supplementary Fig. 9** | **The joint distribution of the number of LD clumps by traits within cell-types.** Venn diagrams showing the count distribution of LD clumps cross-classified by association with selected ncCBC (-SSC, -SFL, -FSC, -SSC-DW, -SFL-DW, -FSC-DW) traits and CBC cell count traits of basophils (a), neutrophils (**b**), eosinophils (**c**), monocytes (**d**) and lymphocytes (**e**). The suffix '-DW' absent a trait type (i.e. absent -SSC, -SFL, -FSC) means any of the distribution width traits of the given cell-type. (e.g. the region labelled NE-DW counts clumps containing at least one variant associated with one of NE-SSC-DW, NE-SFL-DW or NE-FSC-DW).



Supplementary Fig. 10 | ATAC-seq enrichment analysis for neutrophils (a), eosinophils (b), monocytes (c), and lymphocytes (d). Bar plots showing -log<sub>10</sub>(P-value)s for tests (performed by g-chromVAR) of the null hypothesis that there is no enrichment of associated genetic-variants in regions of open chromatin adjusting for GC content and average ATAC-seq peak intensity. Each sub-panel corresponds to associations with a different flow cytometry phenotype. Each bar corresponds to regions of open chromatin measured in a different cell-type. No adjustments have been made for multiple testing. e A legend for panels a-d indicating the cell-type corresponding to the colour of each bar. **f** Diagram of a haematopoietic tree with cell-types coloured according to the scheme displayed in panel e. We thank Joanna Westmoreland for the artwork in panels a-d; panel f is reproduced with permission from Ulirsch, J. C. et al. Interrogation of human hematopoiesis at single-cell and single-variant resolution. Nat. Genet. 51, 683-693 (2019), Spring-

HSC

mDC

Gran

Mono

CMF

80

Ery

MPP

Mega

MEP



Supplementary Fig. 11 | Distribution of effect sizes of genetic associations between rs6993770-T and plasma protein concentrations by protein location. Histograms of estimates of the additive allelic effect size (measured in SD of phenotypic variance per allele) of rs69933770-T on the mean concentration of 2,928 plasma proteins, taken from Sun et al.<sup>7</sup> The mean effect size amongst proteins expressed in MKs (bottom and middle panel) is lower than the mean effect size amongst proteins which are not expressed in MKs (top panel). Within MKs, the mean effect size amongst proteins localised to a-granules (bottom panel) is lower than the mean effect size amongst proteins expressed in MKs but not localised to  $\alpha$ -granules (middle panel). The difference in averages, adjusted for gene expression level measured by RNA-seq of MKs, was estimated to be 0.038 phenotypic standard deviations per allele (*P*-value 5.6x10<sup>-10</sup>), implying that rs6993770-T differentially reduces the plasma concentration of proteins expressed in MKs according to whether or not they are stored in  $\alpha$ -granules. The vertical red lines indicate the means of the corresponding histograms. The red error bars represent the 95% confidence intervals of the estimates of the means.



**Supplementary Fig. 12** [Effect of ablation of ZFPM2 on megakaryopoiesis. a Confirmation by Sanger sequencing of the genotypes of, and the out-of-frame deletions in, the ZFPM2 knockout cell lines (KO1 and KO2). Red shadings highlight mutations and deletions with respect to the reference sequence, which is that of the wild type (WT) cell line. b Flow cytometry plots from arbitrarily selected technical replicates of the differentiated WT, KO1 and KO2 cell lines (FSC = forward scatter; SSC = side scatter). The gated regions correspond to the MK populations. The overlying numbers indicate the percentage of cells in each plot that are gated. c Box plot showing, for each biological replicate, the distribution of the proportion of cells in each nested technical replicate (three per cell line) that are MKs relative to the greatest such proportion amongst the wild type technical replicates. The centre line and the lower and upper hinges of each box plot correspond respectively to the median and the first and third quartiles of the data. The upper whiskers extend to the greatest data point no greater than the third quartile plus 1.5 times the interquartile range. The lower whiskers are defined symmetrically.

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