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Supplemental information

Targeting phenylpyruvate restrains excessive

NLRP3 inflammasome activation and pathological

inflammation in diabetic wound healing

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PPA (µg/g)

Figure S1. Phenylpyruvate is associated with delayed wound healing and prolonged

inflammation. Related to Figure 1. (A) Correlation analysis among differential metabolites in the phenylalanine, tyrosine and tryptophan biosynthesis pathways from comparison of DFU group and NDW group. The colors indicate the values of Pearson r. (B-E) Correlation analysis between differential metabolites in the phenylalanine, tyrosine and tryptophan biosynthesis pathways and NLRP3 expression in DFUs (n=12). (F) Schematic representation of targeted metabolite analysis in diabetic wounds and nondiabetic wounds. (G) Targeted analysis of phenylalanine pathway metabolites. The numbers indicate the fold change in diabetic wounds compared to nondiabetic wounds. Values larger than one suggest upregulation, whereas values smaller than one suggest downregulation in diabetic wounds. White indicates no detection. (H-K) Statistical analysis of collagen volume fraction, Ki-67, α-SMA, and CD31 staining in cutaneous wounds (n=3). (L, M) ELISA analysis of inflammatory factors, including IL1β and TNFα, in cutaneous wounds on day 6 (n=3). Data are shown as mean ± SD. *P<0.05, **P<0.01.



Figure S2. Dietary phenylalanine restriction increases wound healing but has no obvious effect on the general physiological condition *in vivo*. Related to Figure 2. (A, B) Peripheral blood glucose levels and food intake of diabetic mice fed different phenylalanine diets (n=12). (C-F) Statistical analysis of collagen volume fraction, Ki-67, α -SMA, and CD31 staining in cutaneous wounds from different groups (n=3). (G, H) ELISA analysis of inflammatory factors, including IL1 β and TNF α , in cutaneous wounds on day 8 (n=3). (I) Representative images of H&E staining of various organs from mice fed different phenylalanine diets. Scale bar: 100 µm. Data are shown as mean ± SD. *P<0.05, **P<0.01; n.s., not significant.



Figure S3. The effect of phenylpyruvate on macrophages. Related to Figure 3. (A) Experimental diagram of macrophage acquisition and polarization. Flow cytometry data showing the identification of macrophage acquisition. **(B-D)** BMDMs were pretreated with DMSO (control) or increasing concentrations of phenylpyruvate for 4 h and then stimulated with media or IL4 (20 ng/mL) for 24 h. The black triangle indicates increasing phenylpyruvate concentrations (PPA) starting from the left: 200, 400, and 800 μM. The RT–qPCR results showing relative mRNA expression of *Arg1, Chil3*, and *Retnla*

normalized to Actb (n=3). (E) Immunostaining and statistical analysis of Mrc1 in BMDMs after pretreatment with phenylpyruvate and stimulation with IL4 (n=3). Scale bar: 150 µm. (F) BMDMs were treated as in (B). Flow cytometry data for the M2 surface marker CD163, showing mean fluorescence intensity (MFI) (left) and representative histograms of CD163 (right) (n=3). (G) RTqPCR assay showing the relative NLRP3 mRNA expression after treatment with phenylpyruvate (n=3). (H) Immunoblot analysis of BMDMs pretreated with increasing phenylpyruvate concentrations and then stimulated with media or LPS (100 ng/mL) with antibodies recognizing phosphorylated p65, total p65, phosphorylated ERK1/2, total ERK1/2, phosphorylated Akt, total Akt, phosphorylated p38, total p38, GAPDH and ACTB (n=3). (I) Immunostaining and statistical analysis showing the uptake of phenylpyruvate into BMDMs, which were pretreated with phenylpyruvate (400 μ M) with or without cytochalasin D (10μ M) (n=3). Scale bar: 50 μ m. (J-L) BMDMs were pretreated with dimethyl sulfoxide (DMSO, control) or increasing concentrations of phenylalanine for 4 h and then stimulated with media or LPS (100 ng/mL) for 24 h. The black triangle indicates increasing phenylalanine concentrations starting from the left: 200, 400, and 800 µM. The RT-qPCR results showing relative mRNA expression of Nos, Tnf, and Il6 normalized to Actb (n=3). (M) Immunoblot analysis of NLRP3 protein levels in BMDMs pretreated with increasing phenylalanine concentrations (n=3). Data are shown as mean \pm SD. **P<0.01; n.s., not significant.



Figure S4. Identification of PPT1 as a target of phenylpyruvate. Related to Figure 4. (A)

Immunostaining and statistical analysis of LysoTracker in BMDMs treated with phenylpyruvate (n=3). Scale bar: 50 µm. **(B)** Immunostaining and statistical analysis of LysoSensor in BMDMs treated with phenylpyruvate (n=3). Scale bar: 50 µm. **(C)** RT–qPCR results of Psap, M6pr, and Ppt1 mRNA expression in BMDMs after knockdown of Psap, M6pr, and Ppt1, respectively (n=3). (**D**) Immunoblot results of Psap, M6pr, and Ppt1 mRNA expression in BMDMs after knockdown of Psap, M6pr, and Ppt1, respectively (n=3). (**E**) RT–qPCR results showing Nlrp3 mRNA expression in BMDMs after knockdown of Ppt1 (n=3). (**F**) A schematic showing the PPT1 protein structure. (**G**) A mutation of the binding site between PPT1 and phenylpyruvate was established based on the autodocking results. (**H**) Enzymatic activity measurement of PPT1 MUT protein coincubated with increasing phenylpyruvate concentrations (n=3). (**I**) RT–qPCR results showing Ppt1 mRNA expression in BMDMs treated with increasing phenylpyruvate concentrations (n=3). (**J**) Immunoblot analysis of PPT1 expression in BMDMs treated with increasing phenylpyruvate concentrations (n=3). (**J**) Immunoblot analysis of PPT1 expression in BMDMs treated with increasing phenylpyruvate concentrations (n=3). Data are shown as mean ± SD. **P<0.01; n.s., not significant.

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	ID	Position	Peptide	Score	Cutoff	ID	Position	Peptide	Score	Cutoff
	NLRP3_MOUSE	6	**MTSVRCKLAQYLE	39.746	4.222	NLRP3_RAT	8	MKMMSVRCKLAQYLE	37.94	4.222
	NLRP3_MOUSE	405	NEVLFTMCFIPLVCW	6.612	4.222	NLRP3_RAT	407	NEILFTMCFIPLVCW	6.432	4.222
	NLRP3_MOUSE	783	RCGLSHQCCFDISSV	4.135	3.419	NLRP3_RAT	786	CGLTHQCCFNISSVL	4.275	4.222
	NLRP3_MOUSE	784	CGLSHQCCFDISSVL	4.522	4.222	NLRP3_RAT	836	QKLWLVSCCLTSACC	7.581	3.419
	NLRP3_MOUSE	834	QKLWLVSCCLTSACC	7.581	3.419	NLRP3_RAT	837	KLWLVSCCLTSACCQ	4.674	4.222
	NLRP3_MOUSE	835	KLWLVSC <mark>C</mark> LTSACCQ	4.498	4.222	NLRP3_RAT	842	SCCLTSACCQDLALV	7.106	3.419
	NLRP3_MOUSE	840	SCCLTSACCQDLALV	7.106	3.419	NLRP3_RAT	843	CCLTSAC <mark>C</mark> QDLALVL	6.242	4.222
	NLRP3_MOUSE	841	CCLTSACCQDLALVL	6.242	4.222					
	ID	Position	Pentide	Score	Cutoff	ID	Position	Pentide	Score	Cutoff
	NLRP3 HUMAN	8	MKMASTRCKLARVLE	35 677	4 222	NLRP3 MACMU	8	MKMASTRCKLARVLE	35.68	4 222
	NLRP3 HUMAN	2.79	LGDLIMSCCPDPNPP	10.37	3.419	NLRP3 MACMU	279	LGDLIMSCCPDPNPP	10.42	3.419
	NLRP3 HUMAN	280	GDLIMSCCPDPNPPI	6.36	4.222	NLRP3_MACMU	280	GDLIMSCCPDPNPPI	6.264	4.222
	NLRP3 HUMAN	409	NEVLFTMCFIPLVCW	4,409	4.222	NLRP3 MACMU	409	NEVLFTMCFIPLVCW	4,409	4.222
	NLRP3 HUMAN	786	RCGLSHECCFDISLV	4.182	3.419	NLRP3 MACMU	786	CGLSHECCFDISLVL	4.443	4.222
	NLRP3 HUMAN	787	CGLSHECCFDISLVL	4.412	4.222	NLRP3 MACMU	836	KKLWLVSCCLTSACC	6.046	3.419
	NLRP3 HUMAN	837	KKLWLVSCCLTSACC	5.748	3.419	NLRP3 MACMU	837	KLWLVSC <mark>C</mark> LTSACCQ	4.768	4.222
	NLRP3 HUMAN	838	KLWLVSC <mark>C</mark> LTSACCQ	4.768	4.222	NLRP3 MACMU	842	SCCLTSACCQDLASV	6.048	3.419
	NLRP3 HUMAN	843	SCCLTSACCQDLASV	6.111	3.419	NLRP3 MACMU	843	CCLTSACCODLASVL	6.215	4.222
	NLRP3 HUMAN	844	CCLTSACCODLASVL	6.215	4.222	NLRP3 MACMU	956	NCNLTSHCCWDLSTL	4.338	3.419
	NLRP3 HUMAN	957	NCNLTSHCCWDLSTL	4.338	3.419	NLRP3 MACMU	957	CNLTSHCCWDLSTLL	4.791	4.222
	NLRP3_HUMAN	958	CNLTSHCCWDLSTLL	4.791	4.222	NLRP3_MACMU	989	DLGVMMFCEVLKQQS	4.501	4.222
	NLRP3_HUMAN	990	DLGVMMFCEVLKQQS	4.501	4.222					
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	ID	Position	Peptide	Score	Cutoff					
	NLRP3_BOVIN	8	MRMVSVRCKLARYLE	37.395	4.222					
	NLRP3_BOVIN	276	LGDLIASCCPGPNPP	10.332	3.419					
	NLRP3_BOVIN	277	GDLIASCCPGPNPPI	4.773	4.222					
	NLRP3_BOVIN	406	NEILFI MCFIPL VCW	5.028	4.222					
	NLRP3_BOVIN	791	RKLWLGQUCLSHQUU	0.694	3.419					
	NLRP3_BOVIN	/81	VELSHQUEFNISSV	4.063	2 419					
	NLRP3_BOVIN	832	KKLWLVSCCLISASC	4.729	3.419					
	NLRP2 DOVIN	052	NOSI TSHCOWDI STI	4.775	3 410					
	NLRP3 BOVIN	953	CSUTSHCCWDUSTU	4 402	4 222					
	NLRP3_BOVIN	985	DLGVMLLCEVLKOOG	4.298	4.222					
В	3 ABE palmitoylation assay									
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Figure S5. Identification of the palmitoylation site of NLRP3. Related to Figure 5. (A) Alignment of NLRP3 sequences containing predicted palmitoylation sites among different species. **(B)** A schematic illustrating the general process of the ABE palmitoylation assay. **(C)** MEFs were transfected

with wild-type (WT) NLRP3 or the indicated NLRP3 mutants for 24 h with or without knockdown of Ppt1. ABE assay and immunoblot analysis showing palmitoylation levels of the indicated NLRP3 mutants (n=3). Data are shown as mean \pm SD. **P<0.01.



Figure S6. BMDMs were treated with phenylpyruvate (400 μ M) for 4 h and LPS (100 ng/mL) for 24 h and then stimulated with ATP (3 mM) for 45 min. Cell lysates were collected to determine the interaction between NLRP3 and HSC70 by immunoprecipitation (IP) and immunoblot analysis (n=3), Related to Figure 6.



Figure S7. Identification of CD36-mediated phenylpyruvate uptake in macrophages. Related to Figure 7. (A) RT–qPCR assay showing relative mRNA expression of CD36 and LRP1 in BMDMs with knockdown of CD36 and LRP1, respectively (n=3). (B) Immunoblot analysis of CD36 and LRP1 expression in BMDMs with knockdown of CD36 and LRP1, respectively (n=3). (C) Immunostaining and statistical analysis showing the uptake of phenylpyruvate in BMDMs with or without knockdown of LRP1. The FITC antibody indicated phenylpyruvate, and the lipid raft antibody indicated plasma membrane (n=3). Scale bar: 50 μ m. (D) Immunoblot analysis of total LRP1 protein expression in BMDMs treated with increasing phenylpyruvate concentrations. The black triangle indicates increasing phenylpyruvate concentrations starting from the left: 200, 400, and 800 μ M (n=3). Data are shown as mean \pm SD. **P<0.01; n.s., not significant.