Chlamydia trachomatis in acute salpingitis

J. PAAVONEN,* P. SAIKKU,+ E. VESTERINEN,* AND K. AHO±

From the *I-II Departments of Obstetrics and Gynaecology, Helsinki University Central Hospital; †Department of Virology, University of Helsinki; and the ‡Central Public Health Laboratory, Helsinki, Finland

SUMMARY In a study to evaluate the possible role of *Chlamydia trachomatis* and *Neisseria gonorrhoeae* in acute salpingitis, 26% of 106 patients with severe symptoms had positive culture results for *C. trachomatis*; 43% of the 72 patients from whom paired sera were obtained had either positive culture results for or seroconversion in the single antigen immunofluorescence test to *C. trachomatis*. Twenty-six per cent of patients harboured *N. gonorrhoeae* and 14% had gonococcal complement-fixing antibody titres ≥ 8 . Intrauterine devices were used by 48% of patients, no difference being found in the frequency of use between patients harbouring *C. trachomatis* or *N. gonorrhoeae*. The possible role of *C. trachomatis* should be considered in the treatment of acute salpingitis.

Introduction

Acute salpingitis is a common disease and seems to be increasing. In 99% of such cases the infection ascends from the lower genital tract, usually the cervix (Falk, 1946, 1965; Sweet, 1977). The remaining cases are caused by tuberculous salpingitis or by infections which spread from nearby pelvic structures, such as the appendix. Despite the high incidence (Widholm and Kallio, 1965; Weström and Mårdh, 1977) and morbidity (Weström, 1975) of acute salpingitis, the cause of the tubal infection remains uncertain in a large proportion of cases. Acute salpingitis has been associated with Neisseria gonorrhoeae (Falk, 1965; Sweet, 1977; World Health Organisation, 1978), with Mycoplasma hominis and T-mycoplasma (Mårdh and Weström 1970), and with mixed aerobic and anaerobic bacterial pathogens (Chow et al., 1975; Eschenbach et al., 1975). A significant decrease in the percentage of cases of salpingitis attributed to N. gonorrhoeae has been noted, however, during the past decade in Scandinavia (Kallings and Moberg, 1977; Weström and Mårdh, 1977). Recently, Chlamydia trachomatis has attracted attention as an important infective agent (Schachter, 1978). A relationship between C.

trachomatis infection and acute salpingitis was suggested by Dunlop and his co-workers in 1966 (Dunlop et al., 1966). More recently, Mårdh et al. (1977a) recovered C. trachomatis, during laparoscopy, from six out of 20 specimens from the Fallopian tubes of patients with acute salpingitis compared with zero out of 12 specimens from patients with no signs of genital infection.

The present work is a prospective study of the frequency of urogenital *C. trachomatis* and *N. gonorrhoeae* infections in patients admitted to hospital with acute salpingitis and evaluates the possible role of chlamydiae in this disease using antibody studies.

Material and methods

PATIENTS

The study population consisted of 106 women with acute pelvic symptoms who attended the outpatient clinics of the first and second departments of gynaecology and obstetrics at the Helsinki University Central Hospital from October 1977 to June 1978.

The clinical diagnosis of acute salpingitis was based on common criteria: pelvic pain of short duration, tender adnexal masses, increased erythrocyte sedimentation rate, and, usually, fever. All the patients were admitted to hospital because of the severe symptoms. Ten patients had received antimicrobial treatment during the two weeks before admission.

Address for reprints: Dr J. Paavonen, I Department of Obstetrics and Gynaecology, University Central Hospital Hartmaninkatu 2, SF-00290 Helsinki 29, Finland

Received for publication 19 September 1978

The mean age of the patients was 27.3 years and the range, 15-49 years. Fifty-one (48%) patients were fitted with an intrauterine device (IUD), 10 (9%) were taking oral contraceptives, and the remainder used other methods or none at all.

CULTURE TECHNIQUES AND SEROLOGICAL METHODS

Cervical and urethral specimens were cultured for C. trachomatis in irradiated McCoy cells (Paavonen et al., 1978b). Serum antibodies were determined by the single antigen immunofluorescence test (IFAT) using C. trachomatis L2 (434 Bu) inclusions in dog kidney cells as antigen (Saikku and Paavonen, 1978). Acute and convalescent phase sera (obtained one to four weeks later) were obtained from 72 patients. Cervical and urethral specimens were cultured for N. gonorrhoeae by the conventional method (Lennette et al., 1974). The gonococcal microcomplement fixation (CF) test was performed using a mixture of 20 recently isolated strains as the test antigen (Aho and Sievers, 1972). Statistical significance was evaluated by the χ^2 test with Yates's correction.

Results

Of the 106 patients with acute salpingitis, 27 (26%) harboured C. trachomatis in their urogenital tract, 12 in the cervix and urethra, 13 in the cervix alone, and two in the urethra alone. In paired sera 10 (46%) out of 22 chlamydia-positive patients and nine (18%) out of 50 chlamydia-negative patients showed significant (≥ fourfold) change in IFAT titres (Figure). Thus, seroconversion occurred in 19 (26%) out of 72 patients with salpingitis from whom paired sera were obtained. The geometric mean titre (GMT) of titres ≥ 8 was 219 in chlamydia-positive patients and 73 in chlamydia-negative patients. The 10 patients in whom antimicrobial treatment was begun before culture was attempted were all chlamydia-negative. Five of them, however, showed significant change in IFAT titres in paired sera (Figure).

Of the 27 chlamydia-positive patients, eight (30%) also had gonorrhoea compared with 19 (24%) out of 79 chlamydia-negative patients (statistically not significant, $P > 0 \cdot 1$). A total of 15 (14%) of the 106 patients had gonococcal CF titres of ≥ 8 . Of the 27 patients with positive culture results for N. gonorrhoeae, nine (33%) had titres of ≥ 8 compared with six (8%) of the 79 culture-negative patients (statistically significant, $P < 0 \cdot 01$). The highest CF titre was 16, and even this occurred in only five patients, all of whom were culture-positive.

Of the 51 patients using an IUD, nine were chlamydia-positive, eight were N. gonorrhoeae-positive, and two harboured both organisms.

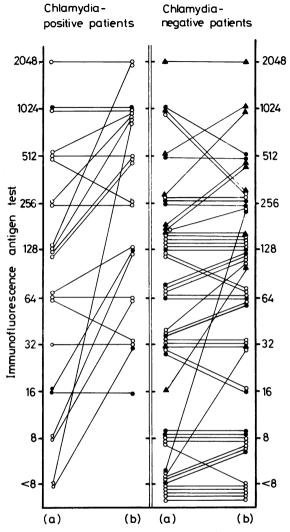


Figure Antibody titres in IFAT against C. trachomatis in paired sera from patients with acute salpingitis. (Acute phase (a) and convalescent phase (b) sera are connected with a line)

Negative gonococcal isolation; ● positive gonococcal isolation; ● antibiotic treatment begun before culture

Discussion

The results of the present study provide additional support for the concept that acute salpingitis may in many cases be due to infection with C. trachomatis; 26% of the patients harboured the organism, and 26% showed seroconversion in the single antigen immunofluorescence test. Seroconversion occurred more often ($\chi^2_1 = 4.60$; P < 0.025) among

chlamydia-positive patients than among chlamydianegative patients. The average GMT in patients with acute salpingitis was high compared with that (GMT 27) in our earlier findings in women of fertile age (mean 29.5 years) representing hospital outpatients (Paavonen et al., 1978b) or in asymptomatic but chlamydia-positive sexual partners of men with nongonococcal urethritis (GMT 80) (Saikku and Paavonen, 1978), thus suggesting systemic chlamydial infection.

Similarly, Eschenbach et al. (1975) noted rising micro-IF antibody titres to C. trachomatis in 24% of women with nongonococcal pelvic inflammatory disease. In addition, Lycke et al. (1976) demonstrated chlamydial antibodies in the haemolysis-in-gel test in sera from all seven chlamydia-positive patients with salpingitis and in four of 17 chlamydia-negative patients with salpingitis. Hamark et al. (1976) observed a significant rise in CF antibody titre in four patients with salpingitis from whom C. trachomatis was isolated from the cervix. In one of these women the organism was isolated from the Fallopian tubes as well.

The frequency of gonococcal infection was as low in our study as in the recent findings in Sweden (Danielsson et al., 1975; Weström and Mårdh, 1977). The role of mycoplasmas as genital tract pathogens is under debate (Lancet, 1970; Mårdh and Weström, 1970; Eschenbach et al., 1975; Vaughan-Jackson et al., 1977; Paavonen et al., 1978a). Furthermore, the isolation of anaerobes from the Fallopian tubes has not been successful (Weström and Mårdh, 1975, Mårdh et al., 1977b).

It is still possible that the frequent finding of C. trachomatis in the urogenital tract reflects only the sexual mode of transission of pathogens responsible for salpingitis or that chlamydial cervical infection allows other pathogens to ascend. Seroconversions and high antibody titres, however, provide evidence to the contrary; C. trachomatis seems to participate actively in this systemic infection. The fact that even chlamydia-negative patients had high IFAT titres (GMT 73) needs explaining. High titres may be due to a past chlamydial infection. Technical failure as well as recent treatment with antimicrobial agents might be responsible for the negative culture results in some cases.

In the present series the overall frequency with which IUDs were used was high (48%). This may be compared with the frequency of 29% among 202 sexually active partners of men with nongonococcal urethritis examined in the same clinic during the same period (Paavonen et al., unpublished data). No difference was found between chlamydia-positive and N. gonorrhoeae-positive patients in the use of IUDs. Other investigators have found a frequency rate of 24-81% in patients with salpingitis (Sweet, 1977). Weström et al. (1976) reported a significantly increased risk of acquiring acute salpingitis in patients using an IUD compared with non-users.

The late sequelae of salpingitis are well known: infertility, increased frequency of ectopic pregnancies, and chronic abdominal pain. The risk of spread of cervical infection to the Fallopian tubes must be considered in the treatment of cervical chlamydiae. The role of C. trachomatis in the aetiology of acute salpingitis necessitates a reexamination of present treatment guidelines. In addition, the isolation of C. trachomatis in many cases (30% in the present study) concomitantly with N. gonorrhoeae should influence the choice of treatment for gonococcal disease. A treatment regimen which is not totally effective against C. trachomatis may give rise to persistent, latent chlamydial infection which later requires further treatment for chronic salpingitis and its long-term sequelae. At the moment, the role of N. gonorrhoeae in the aetiology of salpingitis may well be overestimated.

This study was aided by a grant from the Finnish Cultural Foundation.

References

Aho, K., and Sievers, K. (1972). Screening for gonococcal arthritis. Scandinavian Journal of Rheumatology, 1, 84-86. Chow, A. W., Malkasian, K. L., Marshall, J. R., and Guze, L. B.

(1975). The bacteriology of acute pelvic inflammatory disease. American Journal of Obstetrics and Gynecology, 12, 876-879. Danielsson, D., Falk, V., and Forslin, L. (1975). Acute salpingitis and gonorrhoea on a gynaecological ward. In Genital Infections

and their Complications, p. 151. Edited by D. Danielsson, L. Julin, and P-A. Mårdh, Almovist Wiksell International: Stockholm.

Dunlop, E. M. C., Al-Hussaini, M. K., Freedman, A., Garland, J. A., Harper, I. A., Jones, B. R., Race, J. W., duToit, M. S., Treharne, J. D., and Wright, D. J. M. (1966). Infection by TRIC agent and other members of the Bedsonia group; with a note on Reiter's disease. Genital infection and disease of the eye. Transactions of the Ophthalmological Societies of the United Kingdom, 86, 321-334.

Eschenbach, D. A., Buchanan, T. M., Pollock, H. M., Forsyth, P. S., Alexander, E. R., Lin, J-S., Wang, S-P., Wentworth, B. B., McCormack, W. M., and Holmes, K. K. (1975). Polymicrobial etiology of acute pelvic inflammatory disease. New England Journal of Medicine, 293, 166-171.

Falk, H. C. (1946). Interpretation of the pathogenesis of pelvic infection as determined by cornual resection. American Journal of Obstetrics and Gynecology, 52, 66-73

Falk, V. (1965). Treatment of acute nontuberculous salpingitis with antibiotics alone and in combination with glucocorticoids. Acta

Obstetricia et Gynecologica Scandinavica, 44, Supplement 6. Hamark, B., Brorsson, J-E., Eilard, T., and Forssman, L. (1976).

Salpingitis and chlamydiae subgroup A. Acta Obstetricia et Gynecologica Scandinavica, 55, 377-378.
Kallings, L. O. and Moberg, I. (1977). Epidemiology of gonorrhoea. In Gonorrhoea: Epidemiology and Pathogenesis, p. 3. Edited by F. A. Skinner, P. D. Walker, and H. Smith. Academic Press; London

Lancet (1970). Editorial: Mycoplasma and the urogenital tract. Lancet, 2, 876-877.

Lennette, E. H., Spaulding, E. H., and Truant, J. P. (1974). Manual of Clinical Microbiology, second edition, pp. 124-129. American Society for Microbiology; Washington DC. Lycke, E., and Peterson, M. (1976). Hemolysis-in-gel test for

demonstration of chlamydia antibodies. Journal of Clinical

Microbiology, 4, 450-452. Mårdh, P-A., Ripa, T., Svensson, L., and Weström, L. (1977a). Chlamydia trachomatis infection in patients with acute salpingitis. New England Journal of Medicine, 269, 1377-1379. Mårdh, P-A., Ripa, T., Wang, S-P., and Weström, L. (1977b).

Chlamydia trachomatis as an etiological agent in acute salpingitis. In Non-gonococcal Urethritis and Related Infections, p. 77. Edited by D. Hobson and K. K. Holmes, American Society for Microbiology, Washington DC. Mårdh, P-A., and Weström, L. (1970). Tubal and cervical cultures

in acute salpingitis with special reference to Mycoplasma hominis and T-strain mycoplasmas. British Journal of Venereal Diseases,

46, 179-186.

Paavonen, J., Kousa, M., Saikku, P., Vesterinen, E., Jansson, E. and Lassus, A. (1978a). Examination of men with non-gonococcal urethritis and their sexual partners for Chlamydia trachomatis and Ureaplasma urealyticum. Sexually Transmitted Diseases, 5, 93-96.

Paavonen, J., Saikku, P., Vesterinen, E., Meyer, B., Vartiainen, E., and Saksela, E. (1978b). Genital chlamydial infections in patients

and Saksela, E. (1978b). Genital chlamydial infections in patients attending a gynaecological outpatient clinic. British Journal of Venereal Diseases, 54, 257-261.
Saikku, P., and Paavonen, J. (1978). Single-antigen immunofluorescence test for chlamydial antibodies. Journal of Clinical Microbiology, 8, 119-122.
Schachter, J. (1978). Chlamydial infections. New England Journal of Medicine, 298, 428-435; 490-495; 540-549.

Sweet, R. L. (1977). Diagnosis and treatment of acute salpingitis.

 Sweet, R. L. (1977). Diagnosis and treatment of acute saminguis.
 Journal of Reproductive Medicine, 19, 21-30.
 Vaughan-Jackson, J. D., Dunlop, E. M. C., Darougar, S., Treharne, J. D., and Taylor-Robinson, D. (1977). Urethritis due to Chlamydia trachomatis. British Journal of Venereal Diseases, 53, 180-183.

Weström, L. (1975). Effect of acute pelvic inflammatory disease on fertility. American Journal of Obstetrics and Gynecology, 121,

707-713.

Weström, L., Bengtsson, L. P., and Mårdh, P-A. (1976), The risk of pelvic inflammatory disease in women using intrauterine contraceptive devices as compared to non-users. Lancet, 2, 221-224.

Weström, L., and Mårdh, P-A. (1975). Acute salpingitis. Aspects on etiology, diagnosis and prognosis. In Genital Infections and their Complications, p. 157. Edited by D. Danielsson, L. Julin and P-A. Mårdh, Almqvist & Wiksell International: Stockholm.

Weström, L., and Mardh, P-A. (1977). Epidemiology, etiology, and prognosis of acute salpingitis: a study of 1457 laparoscopically verified cases. In Nongonococcal Urethritis and Related Infections, p. 84. Edited by D. Hobson and K. K. Holmes, American Society for Microbiology: Washington DC. Widholm, O., and Kallio, H. (1965). Suppurative adnexal in-

fections. Report of 830 patients treated in 1947-61. Gynecologia,

160, 321-332.

World Health Organisation (1978). Neisseria gonorrhoeae and gonococcal infections. WHO Technical Report Series No. 616, 59-60. WHO: Geneva.