1 Supporting information for

PI3K block restores age-dependent neurovascular coupling defects associated with cerebral small vessel disease

- Pratish Thakore^{1†}, Evan Yamasaki^{1†}, Sher Ali^{1†}, Alfredo Sanchez Solano¹, Cassandre
 Labelle-Dumais², Xiao Gao^{3,4}, Myriam M. Chaumeil^{3,4}, Douglas B. Gould², and Scott
 Earley^{1*}
- ⁷ ¹Department of Pharmacology, Center for Molecular and Cellular Signaling in the
- 8 Cardiovascular System University of Nevada, Reno School of Medicine; Reno, Nevada,
- 9 89557-0318, USA
- ²Department of Ophthalmology and Anatomy, Institute for Human Genetics, University
- 11 of California San Francisco School of Medicine; San Francisco, CA 94143, USA
- ³Department of Physical Therapy and Rehabilitation Science, UCSF, San Francisco
 USA
- ⁴Department of Radiology and Biomedical Imaging, UCSF, San Francisco USA
- 15 *Corresponding author: Email: <u>searley@med.unr.edu</u>
- [†]These authors contributed equally to this work.

17	*Address Correspondence To:	Scott Earley, Ph.D.
18		University of Nevada, Reno School of Medicine
19		Manville Health Sciences Building, Room 8
20		MS-0318
21		Reno, NV, 89557-0318, USA
22		Phone: (775) 784-4117
23		Fax: (775) 784-1620
24		Email: <u>searley@med.unr.edu</u>



Fig. **S1** – *supplement 1. Kir2.1 currents in cerebral artery ECs.* (A) Illustration of the cerebral artery EC isolation procedure. Scale bar = 10 μ m. (B) Representative I-V traces and summary data showing Kir2.1 current densities in freshly isolated cerebral artery ECs from 3 M-old *Col4a1*^{+/+} and *Col4a1*^{+/G394V} mice (n = 5–6 cells from 5 animals per group, ns = not significant, unpaired t-test). (C) Representative I-V traces and summary data showing Kir2.1 current densities in freshly isolated cerebral artery ECs from 12 M-old *Col4a1*^{+/+} and *Col4a1*^{+/G394V} mice (n = 6–10 cells from 3 to 4 animals per

- 34 group; *p<0.05, unpaired t-test). (D and E) Representative traces (D) and summary data
- 35 (E) demonstrating the vasodilator response to raising extracellular [K+] in cerebral
- 36 arteries from 12 M-old $Col4a1^{+/+}$ and $Col4a1^{+/G394V}$ mice (n = 9–13 preparations from 6
- to 10 animals per group, *p<0.05, non-repeated measures two-way ANOVA). (F)
- 38 Summary data showing the contractile response to 30 and 60 mM [K+] in cerebral
- 39 arteries from 12 M-old $Col4a1^{+/+}$ and $Col4a1^{+/G394V}$ mice (n = 7–12 preparations from 5
- 40 to 10 animals per group, ns = not significant, unpaired t-test).



42 Fig. S2 – supplement 2. High field magnetic resonance imaging. (A) Representative

- 43 susceptibility weighted (SWI) and T2 weighted (T2W) MRI images from 12 M-old
- 44 *Col4a1*^{+/+} and *Col4a1*^{+/G394V} mice. (B) Quantification of ventricle/brain ratio for 12 M-old
- 45 $Col4a1^{+/+}$ and $Col4a1^{+/G394V}$ mice (n = 7 animals per group, ns = not significant, unpaired
- 46 t-test).





- 59 (N) the increase in blood flow following 20 s contralateral WS in 12 M-old $Col4a1^{+/+}$ and
- $Col4a1^{+/G394V}$ mice (n = 5 animals per group, *p<0.05, unpaired t-test).



Fig. S3 – supplement 2. Ipsilateral whisker stimulation. (A to C) Summary data
showing no change in blood flow following ipsilateral whisker stimulation for 1 s (A), 2 s
(B), and 5 s (C) in 3 M- and 12 M-old *Col4a1^{+/+}* and *Col4a1^{+/-G394V}* mice (n = 6 animals
per group, ns = not significant, unpaired t-test). (D) Summary data showing no change

- 66 in blood flow following ipsilateral whisker stimulation for 20 s in 12 M-old *Col4a1*^{+/+} and
- $Col4a1^{+/G394V}$ mice (n = 5 animals per group, ns = not significant, unpaired t-test).





Fig. S3 – supplement 3. Whisker stimulation response in the presence of BaCl₂. (A 69 70 and B) Representative traces (A) and summary data (B) showing the increase in blood 71 flow following 1 s contralateral whisker stimulation (WS) in 12 M-old Col4a1^{+/+} and 72 $Col4a1^{+/G394V}$ mice before and after topical BaCl₂ (100 µM) application to the cranial 73 surface (n = 5 animals per group, *p<0.05, ns = not significant, repeated measures two-74 way ANOVA). (C to F) Latency (C), duration (D), rise rate (E) and decay rate (F) were 75 also analyzed (n = 5 animals per group, *p<0.05, ns = not significant, repeated 76 measures two-way ANOVA). (G) No change in blood flow was detected following 1 s 77 ipsilateral whisker stimulation (n = 5 animals per group, p < 0.05, ns = not significant, 78 repeated measures two-way ANOVA). (H and I) Representative traces (H) and 79 summary data (I) showing the increase in blood flow following 2 s contralateral whisker stimulation (WS) in 12 M-old Col4a1^{+/+} and Col4a1^{+/G394V} mice before and after topical 80 81 BaCl₂ (100 μ M) application to the cranial surface (n = 5 animals per group, *p<0.05, ns 82 = not significant, repeated measures two-way ANOVA). (J to M) Latency (J), duration

83 (K), rise rate (L) and decay rate (M) were also analyzed (n= 5 animals per group,

84 *p<0.05, ns = not significant, repeated measures two-way ANOVA). (N) No change in

85 blood flow was detected following 2 s ipsilateral whisker stimulation (n= 5 animals per

group, *p<0.05, ns = not significant, repeated measures two-way ANOVA). (O and P)

87 Representative traces (O) and summary data (P) showing the increase in blood flow

- following 5 s contralateral whisker stimulation (WS) in 12 M-old Col4a1^{+/+} and
- 89 $Col4a1^{+/G394V}$ mice before and after topical BaCl₂ (100 µM) application to the cranial

90 surface (n = 5 animals per group, *p<0.05, ns = not significant, repeated measures two-

91 way ANOVA). (Q to T) Latency (Q), duration (R), rise rate (S) and decay rate (T) were

92 also analyzed (n= 5 animals per group, *p<0.05, ns = not significant, repeated

93 measures two-way ANOVA). (U) No change in blood flow was detected following 5 s

94 ipsilateral whisker stimulation (n= 5 animals per group, *p<0.05, ns = not significant,

```
95 repeated measures two-way ANOVA).
```



97 Fig. S3 – supplement 4. Cerebral blood flow response to application of KCI to the

98 *cranial surface.* (A and B) Representative traces (A) and summary data (B) showing

99 the increase in blood flow in 12 M-old $Col4a1^{+/+}$ and $Col4a1^{+/G394V}$ mice following topical

100 application of KCI (10 mM) onto the cranial surface, in the absence and presence of

101 BaCl₂ (100 μ M) (n= 5 animals per group, *p<0.05, ns = not significant, repeated

102 measures two-way ANOVA).



103

104 *Fig.* **S4** – *supplement* **1**. *ATP levels in brain capillary* **ECs**. (A) PIP₂ synthesis

pathway. (B) Summary data showing normalized [ATP] in brain capillary ECs from 12

106 M-old $Col4a1^{+/+}$ and $Col4a1^{+/G394V}$ mice. (n = 9 sets of ~500 cells from three mice per

107 group, ns = not significant, unpaired t-test). (C) PIP₂ depletion pathway. (D)

108 Representative I-V traces and summary data showing Kir2.1 current densities in freshly

- 109 isolated brain capillary ECs treated with the PLC inhibitor U73122 (10 µM) from 12 M-
- old $Col4a1^{+/+}$ and $Col4a1^{+/G394V}$ mice (n = 4 cells from 3 animals per group, *p<0.05,
- 111 unpaired t-test).





- 125 and $Col4a1^{+/G394V}$ mice (n = 6 preparations from 3 to 5 animals per group, ns = not
- 126 significant, unpaired t-test).



128 Fig. S5 – supplement 1. Functional hyperemic response following 2 s and 5 s 129 whisker stimulation in GSK1059615-treated animals. (A and B) Representative 130 traces (A) and summary data (B) showing the increase in blood flow following 2 s 131 contralateral whisker stimulation (WS) in 12 M-old Col4a1^{+/+} and Col4a1^{+/-}G394V mice 132 treated with vehicle (saline) or GSK1059615 (10 mg/kg) for 28 days, s.c. (n = 5 animals 133 per group, *p<0.05, ns = not significant, non-repeated measures two-way ANOVA). (C 134 to F) Latency (C), duration (D), rise rate (E) and decay rate (F) were also analyzed (n= 5 135 animals per group, *p<0.05, ns = not significant, non-repeated measures two-way 136 ANOVA). (G and H) Representative traces (G) and summary data (H) showing the 137 increase in blood flow following 5 s contralateral WS in 12 M-old Col4a1^{+/+} and 138 Col4a1^{+/G394V} mice treated with vehicle (saline) or GSK1059615 (10 mg/kg) for 28 days, 139 s.c. (n = 5 animals per group, *p<0.05, ns = not significant, non-repeated measures two-140 way ANOVA). (I to L) Latency (I), duration (J), rise rate (K) and decay rate (L) were also 141 analyzed (n= 5 animals per group, *p<0.05, ns = not significant, non-repeated measures 142 two-way ANOVA).





144 Fig. S5 – supplement 2. Ipsilateral whisker stimulation in GSK1059615 treated

145 *animals.* (A to C) Summary data showing no change in blood flow following ipsilateral

146 whisker stimulation for 1 s (A), 2 s (B), and 5 s (C) in 12 M-old *Col4a1*^{+/+} and

- 147 Col4a1^{+/G394V} mice treated with vehicle (saline) or GSK1059615 (10 mg/kg) for 28 days,
- s.c. (n = 5 animals per group, ns = not significant, non-repeated measures two-way
- 149 ANOVA).



Fig. S5 – supplement 3. Myogenic response in GSK1059615 treated animals. (A)
 Typical recordings of the inner diameter of isolated cerebral arteries from 12 M-old

153 $Col4a1^{+/+}$ and $Col4a1^{+/G394V}$ mice treated with GSK1059615 (10 mg/kg) for 28 days

154 showing the myogenic response to increases in intraluminal pressure (active) and the

155 dilation of the same arteries when extracellular Ca²⁺ has been removed (passive). (B)

156 Summary data showing the effects of GSK1059615 treatment on myogenic tone (n= 5

preparations from 3 animals per group, ns = not significant, repeated measures two-wayANOVA).