- 1386 Supplementary materials for this manuscript include the following:
- 1387 7 figures
- 1388 4 videos

Figure 1-S1 Time (min:sec)



1390	Figure 1-figure supplement 1. B-cells spread and contract on Fab'-coated-planar lipid
1391	bilayers. Splenic B-cells were pre-warmed to 37°C and incubated with planar lipid bilayers
1392	coated with monobiotinylated Fab' fragment of goat anti-mouse IgG+M (Fab'-PLB) and imaged
1393	live at 37°C by interference reflection microscopy (IRM). Shown are individual frames from a
1394	time-lapse IRM image of one B-cell. The plasma membrane area contacting with Fab'-PLB (B-
1395	cell contact zone) visualized by IRM increased between 0-1 min after landing, indicating
1396	spreading, and decreased after 1 min 30 sec, indicating contraction. Scale bar, 2 μ m.



Figure 1-figure supplement 2. CK-666 significantly decreases Arp2/3 recruitment to the B-1398 cell contact zone. WT splenic B-cells were treated with CK-689 or CK-666 (50 µM) before (0 1399 min) and after maximal spreading (2 min) during incubation with Fab'-PLB at 37°C. Cells were 1400 1401 fixed at 3 and 7 min, permeabilized, stained for Arp2, and imaged using IRM and TIRF. Shown are representative images (A) and the MFI of Arp2 in the contact zone at 3 min and 7 min 1402 compared between B-cell treated with CK-689 (black dots), CK-666 from 0 min (red dots), and 1403 1404 CK-666 from 2 min (purple dots) (B). Data points represent individual cells from 3 independent 1405 experiments with ~20 cells per condition per experiment. Scale bar, 2 µm. *** p<0.001, by non-1406 parametric student's *t*-test.

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Figure 1-S3



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Figure 1-figure supplement 3. CK-666 treatment before but not after maximal B-cell 1408 spreading decreased the spreading kinetics. WT splenic B-cells were treated with CK-689 or 1409 1410 CK-666 (50 µM) before (0 min) and after maximal spreading (2 min) during incubation with Fab'-1411 PLB and imaged live at 37°C by IRM. The area occupied by the B-cell contact zone was 1412 measured using IRM images and custom codes made in MATLAB. The mean spreading rate of each cell during its early spreading phase was quantified using the contact area versus the time 1413 1414 curve of that cell by linear regression. The averaged spreading rates (±SEM) were generated from 3 independent experiments with ~15 cells per condition per experiment. * p>0.05, ** 1415 1416 *p*<0.01, by non-parametric student's *t*-test.

Figure 1-S4





Figure 3-S1



1428

1429 Figure 3-figure supplement 1. Emerging of Inner F-actin foci from lamellipodia. Splenic B-1430 cells from LifeAct-GFP transgenic mice were treated with DMSO, imaged live using TIRF and IRM during incubation with Fab'-PLB at 37°C, and analyzed using kymographs that were 1431 randomly generated from each cell. Shown are six examples of the kymographs used for 1432 analysis. Arrows indicate the emergence of inner F-actin foci near the lamellipodia. 1433 Lamellipodia-derived inner F-actin foci were identified by their LifeAct-GFP FI ≥2 fold of their 1434 1435 nearby region, inside location in the contact zone, migrating away the lamellipodial F-actin, and trackable for ≥ 8 sec. 1436



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1439 Figure 6-figure supplement 1. Fab'-PLB, but not Tf-PLB, induces BCR clustering and

1440 **phosphorylation.** WT splenic B-cells were pre-labeled with Cy3-Fab fragment of goat anti-

1441 mouse IgM+G at a concentration of 2.5 µg per 10⁶ cells at 4°C for 30 min, followed by

1442 incubation with Fab'-PLBs or Tf-PLBs for 5 min at 37°C. Cells were fixed, permeabilized,

stained for pCD79a, and imaged using IRM and TIRF. Shown are representative IRM and TIRF

1444 images from three independent experiments. Scale bar, 2 μm.

Figure 6-S2





1446 Figure 6-figure supplement 2. Tracking and analyzing AF546-Fab' clusters in the B-cell 1447 contact zone. WT splenic B-cells were incubated with AF546-Fab'-PLB at 37°C and imaged live using IRM and TIRF. Shown are individual frames from time-lapse images of IRM (A) and 1448 1449 TIRF (**B**), showing AF546-Fab' clusters within the contact zone of one DMSO-treated (vehicle 1450 control for Wisko) B-cell for 140 sec since the beginning of contraction. Fab' FI is shown as heat 1451 maps using NIH ImageJ. The boundary of the contact zone, detected using IRM images by a 1452 custom MATLAB script, is shown in yellow dashed lines. Arrows point to three representative 1453 clusters among the other clusters detected in the contact zone using custom MATLAB codes. 1454 Cluster detection masks for the three representative clusters are shown (C). Moving tracks for the three AF546-Fab' clusters are shown alongside the initial (black dashed lines) and final state 1455 (gray dashed lines) of the contact zone (**D**). Tracks were generated by following the peak of 1456 1457 AF546 FI in each cluster as it moved. AF546-Fab' peak FI versus time curves for the three 1458 representative clusters are plotted over the duration that each cluster could be detected (E). Surface plots (2.5-D plots) of AF546-Fab' FI show a zoomed-in region consisting of each of the 1459 1460 three AF546-Fab' clusters (F). Colors in (B) and (F) are scaled to AF546-Fab' FI values. Scale 1461 bars, 1 µm.

1462 Video Legends

1463 Figure 1-Video 1. Effects of CK-666, Wiskostatin, conditional N-WASP knockout, and WASP knockout on B-cell contraction. Splenic B-cells were incubated with planar lipid 1464 1465 bilayers coated with monobiotinylated Fab' fragment of goat anti-mouse IgG+M (Fab'-PLB) in 1466 the absence and presence of various inhibitors and imaged live at 37°C at one frame per 2 1467 seconds by interference reflection microscopy (IRM). Shown are representative IRM time-lapse images of WT B-cells treated with CK-689 or CK-666 (50 µM) before (0 min) and after maximal 1468 1469 spreading (2 min) (A), WT B-cells treated with DMSO or Wiskostatin (Wisko, 10 µM) 10 min 1470 before and during incubation with Fab'-PLB (B), B-cells from flox control and B-cell-specific N-WASP knockout (cNKO) mice (C), and B-cells from WT or WASP knockout mice (WKO) (D). 1471 The frame in which the contact zone first appears was considered time 0. The videos are sped 1472 1473 up by 20x compared to real time. Scale bar, 2 µm. 1474

Figure 2-Video 1. Effects of CK-666 and WKO on F-actin foci formation. Splenic B-cells 1475 1476 from LifeAct-GFP transgenic mice were incubated with Fab'-PLB in the absence and presence of CK-689 or CK-666 and imaged live at 37°C at one frame per 2 seconds by IRM and total 1477 1478 internal reflection fluorescence microscopy (TIRF). Shown are representative IRM (A and C) and TIRF (**B** and **D**) time-lapse images of WT B-cells treated with CK-689 or CK-666 (50 µM) 1479 1480 before and after maximal spreading (A and B) and WT and WKO B-cells expressing LifeAct-GFP (**C** and **D**). The video is sped up by 20x compared to real time. Scale bar, 2 μm. 1481 1482 1483 Figure 5-Video 1. Wiskostatin treatment inhibits NMII ring-like structure formation. B-cells from mice expressing GFP fusion of non-muscle myosin IIA (GFP-NMIIA) and LifeAct-RFP 1484

transgenes were treated with DMSO or Wisko (10 μ M) 10 min before and during incubation with

1486 Fab'-PLB. The B-cell contact zones were imaged live at one frame per 2 seconds using IRM

and TIRF. Shown are representative IRM and TIRF time-lapse images. The video is sped up by
20x compared to real time. Scale bar, 2 µm.

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1490 Figure 6-Video 1. Inhibition of B-cell contraction reduces the molecular density within 1491 BCR clusters. Splenic B-cells were incubated with AF546-Fab'-PLB in the absence and presence of various inhibitors and imaged live at 37°C at one frame per 2 seconds by IRM and 1492 1493 TIRF. Shown are representative IRM (A, C, and E) and TIRF (B, D, and F) time-lapse images of WT B-cells treated with CK-689 or CK-666 (50 µM) before and after maximal spreading (A and 1494 **B**), WT B-cells treated with DMSO or Wisko (10 µM) 10 min before and during incubation with 1495 Fab'-PLB (C and D), and flox control or cNKO B-cells (E and F). TIRF images (B, D, and F) are 1496 shown as AF546-Fab' FI maps. The video is sped up by 20x compared to real time. Scale bar, 2 1497 1498 μm.