Supplementary Information

Functionally distinct cancer-associated fibroblast subpopulations establish a tumor promoting environment in squamous cell carcinoma

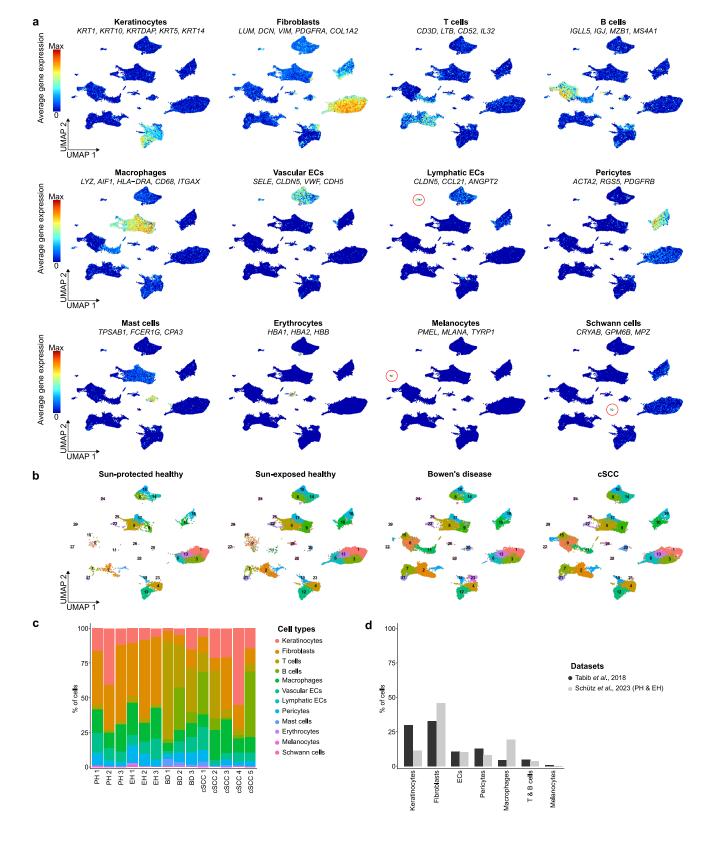
Sabrina Schütz¹, Llorenç Solé-Boldo¹, Carlota Lucena-Porcel^{2,3}, Jochen Hoffmann⁴, Alexander Brobeil^{2,3}, Anke S. Lonsdorf⁴, Manuel Rodríguez-Paredes^{1*} & Frank Lyko^{1*}

¹Division of Epigenetics, DKFZ-ZMBH Alliance, German Cancer Research Center, 69120 Heidelberg, Germany; ²Institute of Pathology, Ruprecht-Karls University of Heidelberg, 69120 Heidelberg, Germany; ³Tissue Bank of the National Center for Tumor Diseases (NCT), 69120 Heidelberg, Germany; ⁴Department of Dermatology, University Hospital, Ruprecht-Karls University of Heidelberg, 69120 Heidelberg, Germany

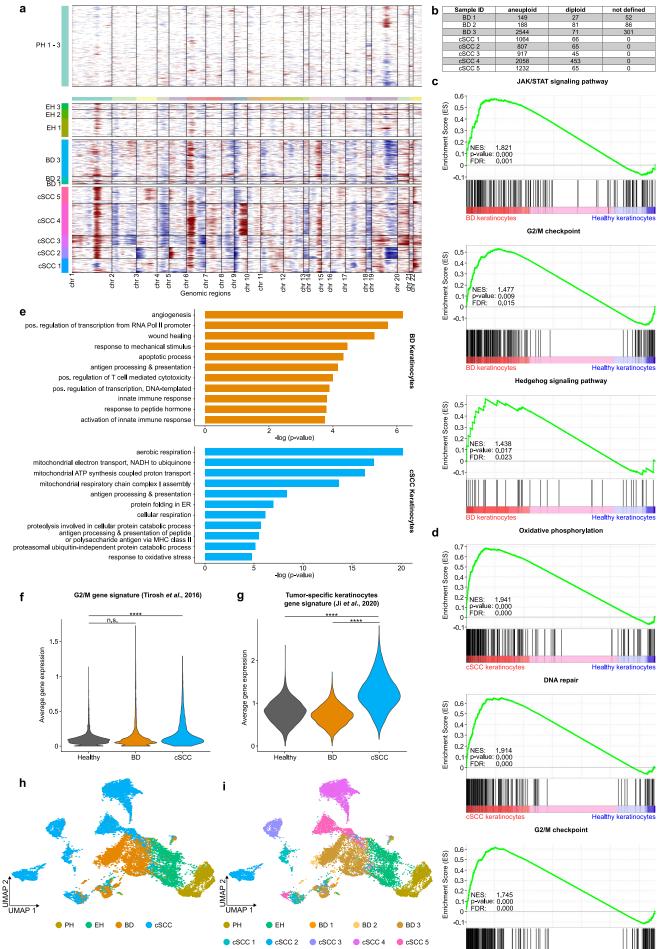
*Equal contribution.

Corresponding authors: Manuel Rodríguez-Paredes (m.rodriguez@dkfz.de, +49 6221424631)

Frank Lyko (<u>f.lyko@dkfz.de</u>, +49 6221423800)



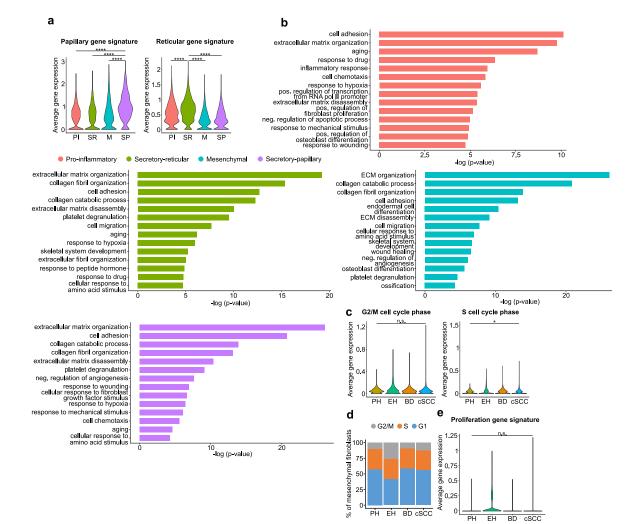
Supplementary Figure 1. scRNA-seq analysis identifies skin cell types. (a) UMAP plots visualizing the average gene expression of three to five known marker genes for typical cutaneous cell types. Red indicates maximum gene expression, while blue indicates low or no expression. **(b)** UMAP plots showing the integrated dataset with all clusters for UVR-protected healthy skin, chronically UVR-exposed healthy skin, BD and cSCC samples separately. **(c)** Bar plot depicting cell type proportions for all scRNA-seq samples. **(d)** Bar plot showing cell type proportions of the human skin scRNA-seq study from Tabib *et al.*, 2018¹ and the present study. UVR: ultraviolet radiation, PH: UVR-protected healthy, EH: UVR-exposed healthy, BD: Bowen's disease, cSCC: cutaneous squamous cell carcinoma. Source data are provided as a Source Data file.



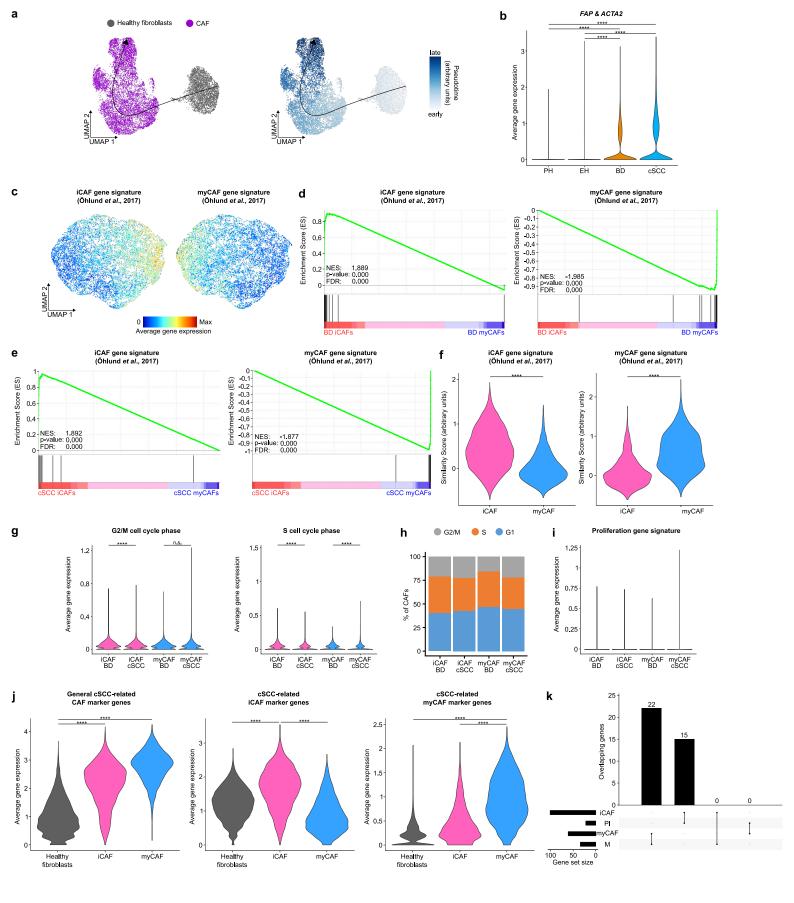
Healthy keratin

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Supplementary Figure 2. Characterization of malignant keratinocytes derived from BD and cSCC samples. (a) Heatmaps visualizing the distribution of copy number variation (CNV) events in keratinocytes from all scRNA-seg samples across the chromosomes. Each row represents a single cell and columns correspond to genes, ordered by chromosome position. Gene amplifications are indicated in red and deletions are marked in blue. (b) Table including the total numbers of keratinocytes from BD and cSCC samples that were either classified as aneuploid or diploid, and non-defined keratinocytes. (c) & (d) Gene Set Enrichment Analysis (GSEA) plots for different pro-tumorigenic pathways that are activated in BD keratinocytes (c) or cSCC keratinocytes (d) compared to healthy keratinocytes. Normalized Enrichment Scores (NES), false discovery rates (FDR) and p-values are given for all plots, based on the Kolmogorov Smirnov test. (e) Top 11 terms from Gene Ontology (GO) analyses performed with the most upregulated genes in BD keratinocytes or cSCC keratinocytes compared to healthy keratinocytes. Terms are ordered according to p-value determined by the Fisher's Exact test. (f) Violin plot displaying the average gene expression of proliferation-related genes important in the G2/M cell cycle phase² in keratinocytes from healthy samples, BD and cSCC. Healthy vs. BD: p-value = 0.76, Healthy vs. cSCC: p-value < 2.2e-16. (g) Violin plot showing the average gene expression of a publicly available cSCC-derived keratinocyte gene signature³ in keratinocytes from healthy samples, BD and cSCC. Healthy vs. cSCC: p-value < 2.2e-16, BD vs. cSCC: p-value < 2.2e-16. (h) UMAP plot displaying sample type-specific clusters of keratinocytes after merging, but not integrating, all datasets. (i) UMAP plot showing patient-specific clusters of keratinocytes from BD and cSCC samples after merging, but not integrating, all datasets. For violin plots, statistical analyses were performed using a two-sided Wilcoxon Rank Sum test (****: p-value < 0.0001). chr: chromosome, UVR: ultraviolet radiation, PH: UVR-protected healthy, EH: UVR-exposed healthy, BD: Bowen's disease, cSCC: cutaneous squamous cell carcinoma, n.s.: not significant. Source data are provided as a Source Data file.

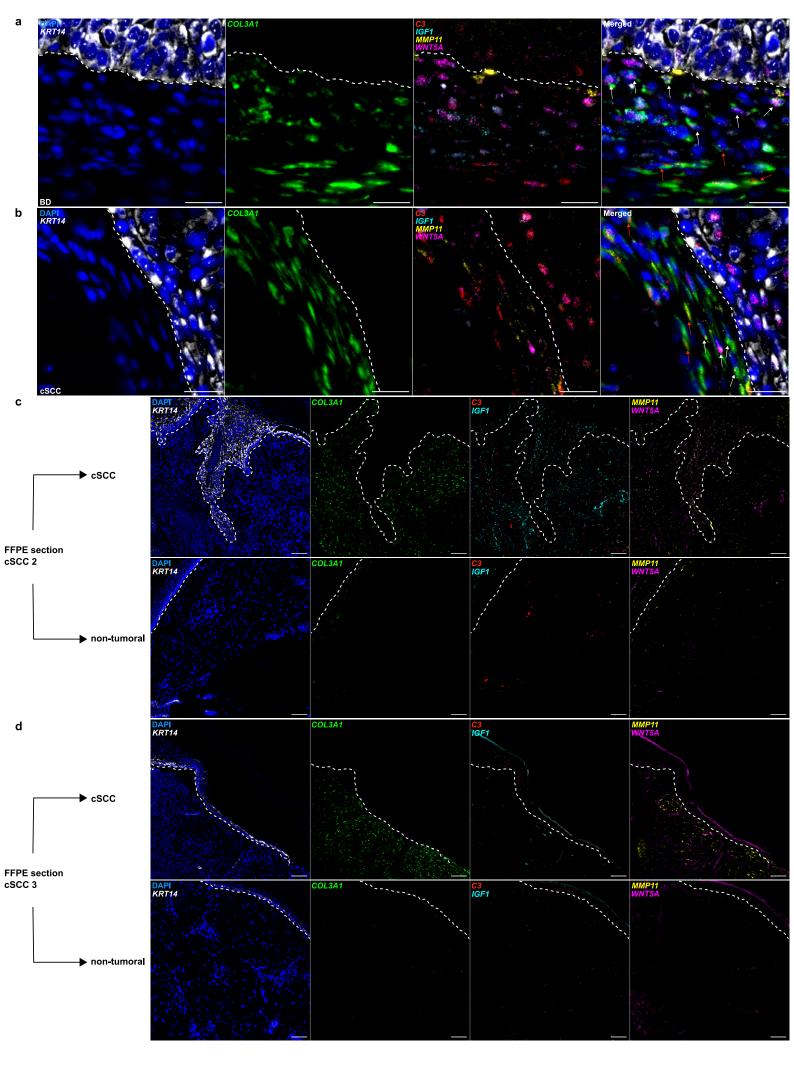


Supplementary Figure 3. Previously defined fibroblast subpopulations are maintained during cSCC development. (a) Violin plots showing the average gene expression of known marker genes for papillary and reticular fibroblasts respectively⁴ in all fibroblasts analyzed. All indicated comparisons: p-value < 2.2e-16. (b) Top 14 terms from Gene Ontology (GO) analyses performed with the most representative genes expressed in each fibroblast subpopulation. Terms are ordered according to *p*-value, determined by the Fisher's Exact test. (c) Violin plots visualizing the average gene expression of cell cycle-related genes² in mesenchymal fibroblasts from all sample types. G2/M phase PH vs. cSCC: p-value = 0.399, S phase PH vs. cSCC: p-value = 0.0271. (d) Bar plot depicting cell cycle phase ratios for mesenchymal fibroblasts detected in the different sample types. (e) Violin plot showing the average gene expression of known genes related to cell proliferation⁵ for mesenchymal fibroblasts from all sample types. PH vs. cSCC: p-value = 0.879. For violin plots, statistical analyses were performed using a two-sided Wilcoxon Rank Sum test (*: p-value < 0.05, ****: p-value < 0.0001). PH: UVR-protected healthy, EH: UVR-exposed healthy, BD: Bowen's disease, cSCC: cutaneous squamous cell carcinoma, PI: pro-inflammatory, SR: secretoryreticular, M: mesenchymal, SP: secretory-papillary, n.s.: not significant. Source data are provided as a Source Data file.

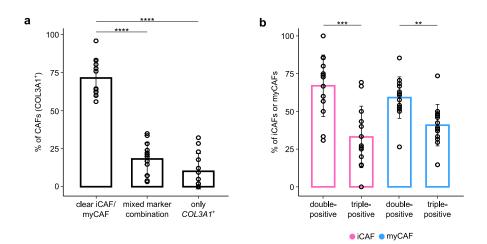


Supplementary Figure 4. Detection of iCAFs and myCAFs in BD and cSCC. (a) UMAP plots visualizing the progression trajectory of healthy fibroblasts from UVR-protected skin towards CAFs from BD and cSCC samples. White and light blue indicate an earlier time point during this process, while dark blue indicates later time points on an arbitrary pseudotime axis. (b) Violin plot showing the average gene expression of FAP and ACTA2 in fibroblasts from UVR-protected healthy and chronically UVR-exposed healthy skin samples as well as BD and cSCC. All indicated comparisons: p-value < 2.2e-16. (c) UMAP plots depicting the average gene expression of pancreatic iCAF- and myCAF-related gene signatures^{6, 7}. Red indicates maximum gene expression, while blue indicates low or no expression. (d) & (e) Gene Set Enrichment Analysis (GSEA) plots for gene signatures of pancreatic iCAFs and myCAFs^{6, 7} that are enriched in BD- (d) and cSCC-derived (e) iCAFs and myCAFs, respectively. For all plots, the normalized Enrichment Score (NES), false discovery rate (FDR) and p-value are given, based on the Kolmogorov Smirnov test. (f) Violin plots visualizing similarity scores in arbitrary units between BD/cSCC-derived CAFs and gene signatures of pancreatic iCAFs and myCAFs^{6, 7}. Both comparisons: p-value < 2.2e-16. (g) Violin plots showing the average gene expression of cell cycle-related genes² in iCAFs and myCAFs from BD and cSCC, G2/M phase iCAF BD vs. iCAF cSCC: p-value = 6.63e-05, myCAF BD vs. myCAF cSCC: p-value = 0.1, S phase iCAF BD vs. iCAF cSCC: p-value = 2.81e-09, myCAF BD vs. myCAF cSCC: p-value = 3.31e-05. (h) Bar plot depicting cell cycle phase ratios for iCAFs and myCAFs, in BD and cSCC. (i) Violin plot visualizing the average gene expression of known genes related to cell proliferation⁵ for iCAFs and myCAFs from BD and cSCC. (j) Violin plots showing the average gene expression of general CAF marker genes and specific iCAF/myCAF marker genes defined in cSCC (Supplementary Data 4) for healthy fibroblasts (PH & EH) and CAFs (BD & cSCC). All indicated comparisons: p-value = 0. (k) Bar plot depicting the overlap of the most representative genes of iCAFs, myCAFs, pro-inflammatory fibroblasts and mesenchymal fibroblasts. For violin plots, statistical analyses were performed using a two-sided Wilcoxon Rank Sum test (****: p-value < 0.0001). UVR: ultraviolet radiation, PH: UVR-protected healthy, EH: UVR-exposed healthy, BD: Bowen's disease, cSCC: cutaneous squamous cell carcinoma,

iCAF: inflammatory cancer-associated fibroblast, myCAF: myofibroblastic cancer-associated fibroblast, PI: pro-inflammatory, M: mesenchymal, n.s.: not significant. Source data are provided as a Source Data file.

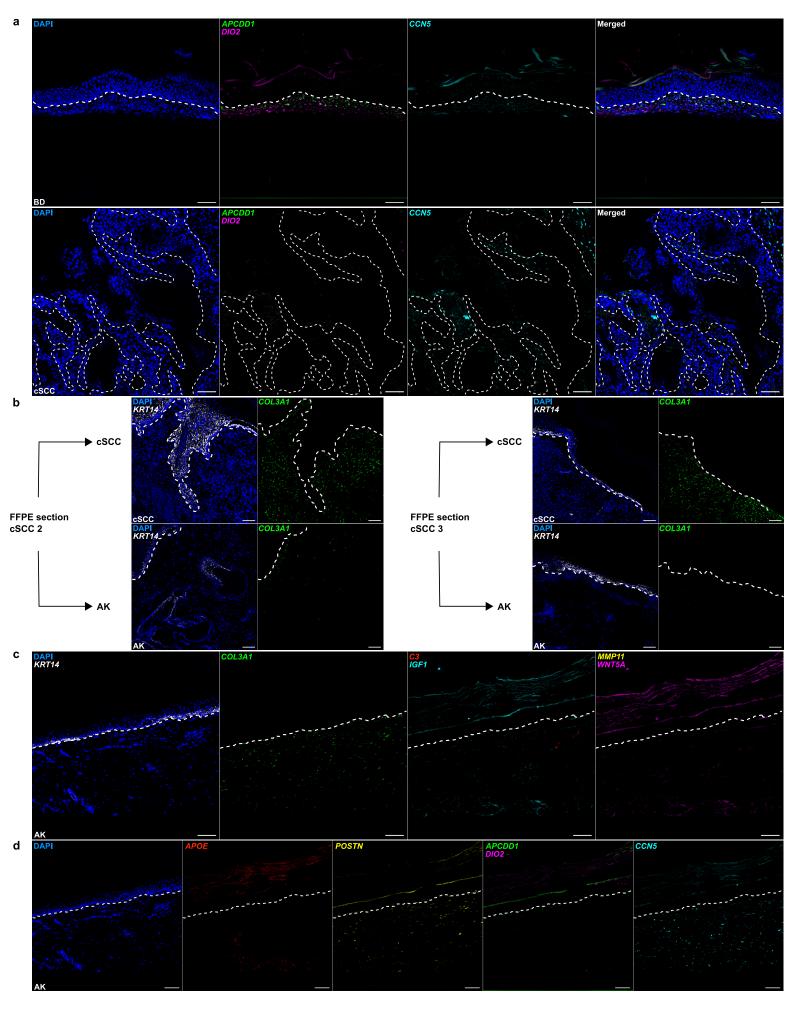


Supplementary Figure 5. Multiplexed RNA FISH detects iCAFs and myCAFs in BD and cSCC but not in adjacent non-tumoral skin. Exemplary microscopy images of FFPE BD (a) and cSCC (b) samples as well as a direct comparison of a tumor area and a non-tumoral but chronically UVR-exposed skin area of the same FFPE sections cSCC 2 (c) and cSCC 3 (d). All images show the respective channels visualizing the hybridization with fluorescent probes against the mRNA of *KRT14* (white) as keratinocyte marker, *COL3A1* (green) as general CAF marker, *C3* (red) and *IGF1* (cyan) as iCAF markers, as well as *MMP11* (yellow) and *WNT5A* (magenta) as myCAF markers. Nuclei were counterstained with DAPI. Dashed lines separate epidermal keratinocytes from dermal cells. Scale bars: 25μ m for (a) and (b) or 100μ m for (c) and (d). White arrows indicate myCAFs whereas red arrows indicate iCAFs. BD: n = 3 patients, cSCC: n = 3 patients. FISH: fluorescence *in situ* hybridization, UVR: ultraviolet radiation, FFPE: formalin-fixed paraffin embedded, iCAF: inflammatory cancer-associated fibroblast, myCAF: myofibroblastic cancer-associated fibroblast, BD: Bowen's disease, cSCC: cutaneous squamous cell carcinoma, EH: UVR-exposed healthy skin.

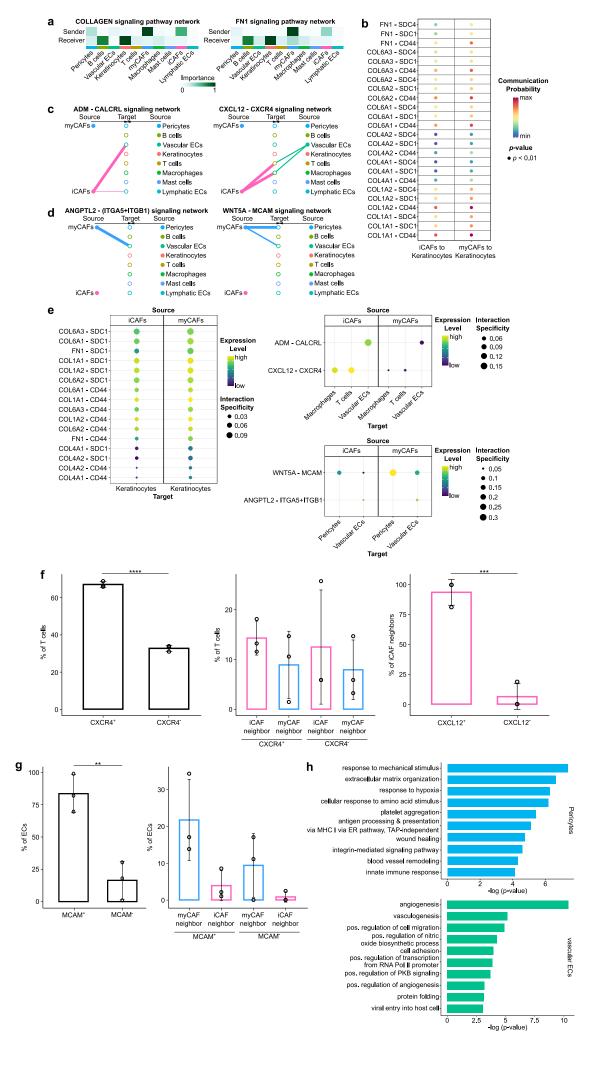


Supplementary Figure 6. Conditions for quantification of multiplexed RNA FISH assays.

(a) Bar plot showing the proportions of CAFs (COL3A1+) that were unambiguously classified as iCAFs or myCAFs in cSCC due to the expression of either C3 and/or IGF1, or MMP11 and/or WNT5A, respectively. CAFs showing mixed combinations for iCAF and myCAF marker genes, as well as cells expressing only COL3A1 but no specific iCAF/myCAF markers, were not considered for following analyses and guantifications. Clear iCAFs/myCAFs vs. mixed marker: p-value = 1.215e-011, clear iCAFs/myCAFs vs. only COL3A1+: p-value = 1.487e-012. (b) Bar plot visualizing the ratios of all iCAFs and myCAFs detected in cSCC with respect to the expression of marker gene combinations. Double-positive cells showed co-expression of COL3A1 as general CAF marker and one subpopulation-specific gene (C3 or IGF1 for iCAFs, and MMP11 or WNT5A for myCAFs). Triple-positive cells showed co-expression of COL3A1 and both of the respective subpopulation-specific genes. iCAFs double vs. triple: p-value = 0.000288, myCAFs double vs. triple: p-value = 0.00240. Statistical analyses were performed using two-sided t-tests (**: *p*-value < 0.01, ***: *p*-value < 0.001, ****: *p*-value < 0.0001) and error bars show the standard deviation. Bars indicate mean values and each dot represents a dermal region with around 100 COL3A1⁺ cells/CAFs (cSCC: n = 13 dermal regions). Source data are provided as a Source Data file.

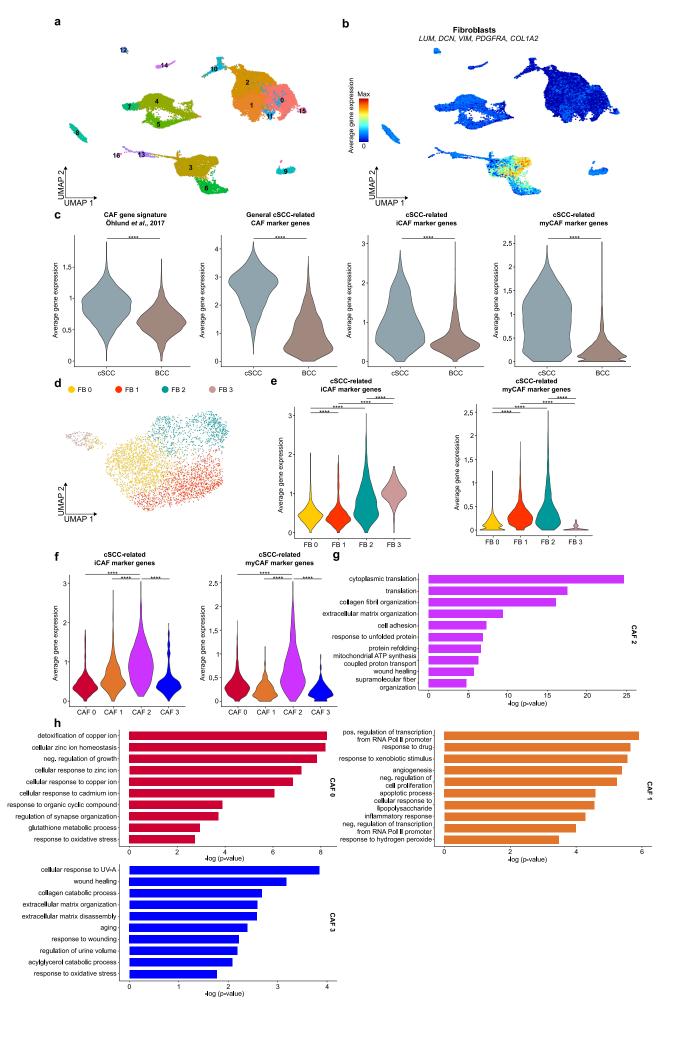


Supplementary Figure 7. Multiplexed RNA FISH detects fibroblast subpopulation marker genes in the cSCC disease continuum. (a) Representative microscopy images of FFPE BD and cSCC samples hybridized with fluorescent probes against the mRNA of APCDD1 (green) and DIO2 (magenta) as secretory-papillary fibroblast marker genes, and CCN5 (cyan) as secretory-reticular marker. Nuclei were counterstained with DAPI. Dashed lines separate epidermal keratinocytes from dermal cells. Scale bar: 100µm. (b) Illustrative microscopy images of tumor areas and AK regions in the same FFPE sections cSCC 2 and cSCC 3 hybridized with fluorescent probes against the mRNA of KRT14 (white) as keratinocyte marker and COL3A1 (green) as general cSCC-CAF marker. Nuclei were counterstained with DAPI. Dashed lines separate epidermal keratinocytes from dermal cells. Scale bar: 100µm. (c) Exemplary microscopy image of a FFPE AK sample hybridized with fluorescent probes against the mRNA of *KRT14* (white) as keratinocyte marker, *COL3A1* (green) as general CAF marker, C3 (red) and IGF1 (cyan) as iCAF markers as well as MMP11 (yellow) and WNT5A (magenta) as myCAF markers. Nuclei were counterstained with DAPI. Dashed lines separate epidermal keratinocytes from dermal cells. Scale bar: 100µm. (d) Representative microscopy image of a FFPE AK sample hybridized with fluorescent probes against the mRNA of APOE (red) as proinflammatory fibroblast marker, POSTN (yellow) as mesenchymal fibroblast marker, APCDD1 (green) and DIO2 (magenta) as secretory-papillary fibroblast marker genes as well as CCN5 (cyan) as secretory-reticular marker. Nuclei were counterstained with DAPI. Dashed lines separate epidermal keratinocytes from dermal cells. Scale bar: $100\mu m$. AK: n = 3 patients, BD: n = 3 patients, cSCC: n = 3 patients. FISH: fluorescence *in situ* hybridization, FFPE: formalinfixed paraffin embedded, iCAF: inflammatory cancer-associated fibroblast, myCAF: myofibroblastic cancer-associated fibroblast, AK: actinic keratosis, BD: Bowen's disease, cSCC: cutaneous squamous cell carcinoma.



Supplementary Figure 8. CAFs affect cancer cells and tumor microenvironment via common and specific signaling pathways. (a) Heatmaps visualizing predictions (CellChat) of the most important "sender" and "receiver" cell types within the cSCC tissue for different signaling pathway networks. White color or light green correlate with no or low involvement, while dark green represents high importance. (b) Dot plot showing significant interactions (CellChat) between CAF-derived ligands and receptors expressed in malignant keratinocytes in cSCC. Red indicates high interaction probability and blue represents low interaction probability. The dot size correlates with the *p*-value of the interaction, based on the method trimean. (c) Plots depicting signaling networks (CellChat) with iCAFs as the most important "sender" cells secreting the ligands ADM (left) and CXCL12 (right). Line thickness correlates with interaction strength. Thicker lines indicate stronger signals. (d) Plots visualizing signaling networks (CellChat) with myCAFs as the most important "sender" cells secreting the ligands ANGPTL2 (left) and WNT5A (right). (e) Dot plots showing significant (p-value < 0.01) interactions (LIANA) between CAF-derived ligands and receptors expressed in keratinocytes or different non-malignant cell types in cSCC, based on the robust rank aggregation method. Purple color correlates with low expression levels and yellow indicates high expression levels. Interaction specificity is depicted by the dot size. (f) Bar plots visualizing signal quantification for iCAF interactions in RNA FISH assays with respect to receptor expression (CXCR4) in T cells (p-value = 0.0000115, left), their iCAF/myCAF proximity (center), and ligand expression (CXCL12) in iCAFs (p-value = 0.000584, right). (g) Bar plots depicting signal quantification for myCAF interactions in RNA FISH assays with respect to receptor expression (MCAM) in vascular ECs (p-value = 0.00508, left), and their iCAF/myCAF proximity with myCAFs expressing the ligand WNT5A (right). (h) Top 10 terms from Gene Ontology (GO) analyses with the most upregulated genes from chronically UVR-exposed healthy skin to cSCC tumors in pericytes and vascular ECs. Terms are ordered according to p-value, determined by the Fisher's Exact test. For quantifications in (f) and (g), statistical analyses were performed using two-sided t-tests (**: *p*-value < 0.01, ***: *p*-value < 0.001, ****: *p*-value < 0.0001) and error bars show the standard deviation. Bars indicate mean values and each dot represents a dermal

region with around 100 T cells or vascular ECs (cSCC: n = 3 dermal regions). UVR: ultraviolet radiation, iCAF: inflammatory cancer-associated fibroblast, myCAF: myofibroblastic cancer-associated fibroblast, EC: endothelial cell, cSCC: cutaneous squamous cell carcinoma. Source data are provided as a Source Data file.



Supplementary Figure 9. Assessment of cSCC-related iCAF and myCAF marker gene expression in BCC CAFs. (a) UMAP plot visualizing unsupervised clustering of nine BCC samples from Yerly et al., 2022⁸ and Guerrero-Juarez et al., 2022⁹. (b) UMAP plot depicting the average gene expression of known fibroblast marker genes. Red indicates maximum gene expression, while blue indicates low or no expression. (c) Violin plots displaying the average gene expression levels of previously described PDAC-related CAF signatures⁶, as well as our newly defined cSCC-related CAF signatures in cSCC-derived CAFs and BCC-derived fibroblasts. All indicated comparisons: p-value < 2.2e-16. (d) UMAP plot of re-clustered fibroblasts from all BCC samples. (e) Violin plots showing the average gene expression of cSCC-related iCAF (FB 0 vs. FB 1: p-value = 9.09e-07, FB 0 vs. FB 2: p-value < 2.2e-16, FB 1 vs. FB 3: p-value < 2.2e-16, FB 2 vs. FB 3: p-value = 3.46e-15, left) and myCAF (all indicated comparisons: p-value < 2.2e-16, right) marker genes in BCC fibroblasts. (f) Violin plots showing the average gene expression of cSCC-related iCAF (left) and myCAF (right) marker genes in BCC CAFs. All indicated comparisons: p-value < 2.2e-16. (g) Top 10 terms from a Gene Ontology (GO) analysis with the most representative genes of BCC CAF cluster 2. Terms are ordered according to *p*-value, determined by the Fisher's Exact test. (h) Top 10 terms from GO analyses with the most representative genes of BCC CAF clusters 0, 1 and 3. Terms are ordered according to *p*-value, determined by the Fisher's Exact test. For violin plots, statistical analyses were performed using a two-sided Wilcoxon Rank Sum test (****: p-value < 0.0001). FB: fibroblast, iCAF: inflammatory cancer-associated fibroblast, myCAF: myofibroblastic cancer-associated fibroblast, cSCC: cutaneous squamous cell carcinoma, BCC: basal cell carcinoma, PDAC: pancreatic ductal adenocarcinoma. Source data are provided as a Source Data file.

Sample ID	Gender	Age (years)	Localization	Chronic UVR exposure	Immune status	Diagnosis	Grading & tumor thickness	Number of cells
PH 1*	male	53 – 70	inguinal	no	IC	healthy	N/A	2,592
PH 2*	male		inguinal	no	IC	healthy	N/A	1,833
PH 3*	male		inguinal	no	IC	healthy	N/A	4,386
EH 1	male		temple	yes	IC	healthy	N/A	13,439
EH 2	male	47 – 87	cheek	yes	IC	healthy	N/A	8,757
EH 3	male		head	yes	IC	healthy	N/A	7,380
BD 1	male		head	yes	IC	BD	N/A	10,047
BD 2**	male	65 – 85	upper arm	yes	IC	BD	early invasive	7,183
BD 3**	male		shoulder	yes	IC	BD	early invasive	18,988
cSCC 1	male	72 – 93	head	yes	IC	cSCC	G3-4 / 6.0 mm	18,336
cSCC 2	male		head	yes	IC	cSCC	G2 / 3.6 mm	4,054
cSCC 3	male		head	yes	IC	cSCC	G2 / 6.0 mm	4,533
cSCC 4	male		head	yes	IC	cSCC	G1 / 8.0 mm	4,536
cSCC 5	male		head	yes	IC	cSCC	G1-2 / 3.2 mm	8,989

Supplementary Table 1. Overview of the samples used for single-cell RNA sequencing.

UVR: ultraviolet radiation, PH: UVR-protected healthy, EH: chronically UVR-exposed healthy, BD: Bowen's disease, cSCC: cutaneous squamous cell carcinoma,

IC: immunocompetent, * raw data obtained from Solé-Boldo *et al.*, 2020¹⁰, ** samples obtained from the same patient.

Sample type	Total fibroblast number	PI fibroblasts	SR fibroblasts	M fibroblasts	SP fibroblasts
PH	3,982	1,547	1,437	376	622
EH	13,637	5,998	3,557	2,285	1,797
BD	3,807	1,279	1,152	764	612
cSCC	5,956	1,318	1,236	2,455	947

Supplementary Table 2. Total numbers of fibroblasts and their subpopulations in each sample type analyzed.

PH: UVR-protected healthy, EH: chronically UVR-exposed healthy, BD: Bowen's disease, cSCC: cutaneous squamous cell carcinoma, PI: pro-inflammatory, SR: secretory-reticular, M: mesenchymal, SP: secretory-papillary.

Sample ID	Gender	Age (years)	Localization	Immune status	Grading & tumor thickness	Biopsy
EH 1	male		face	IC	N/A	incisional
EH 2	male	65 – 83	head	IC	N/A	incisional
EH 3	male		face	IC	N/A	incisional
AK 1	male		head	IC	N/A	incisional
AK 2	male	82 – 88	face	IC	N/A	shave
AK 3	male		head	IC	N/A	shave
BD 1	male		head	IC	N/A	shave
BD 2	male	77 – 82	head	IC	N/A	shave
BD 3	male		head	IC	N/A	shave
cSCC 1	male		head	IC	G2 / 4.0 mm	incisional
cSCC 2	male		head	IC	G1 / 2.0 mm	incisional
cSCC 3	male	71 – 85	head	IC	G2 / 1.2 mm	incisional
cSCC 4	male		face	IC	G2 / 5.0 mm	incisional
cSCC 5	male		face	IC	G2 / 3.6 mm	incisional

Supplementary Table 3. Overview of the FFPE samples used for RNA FISH.

EH: chronically UVR-exposed healthy, UVR: ultraviolet radiation, AK: actinic keratosis, BD:

Bowen's disease, cSCC: cutaneous squamous cell carcinoma, IC: immunocompetent.

References

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