

1 ***Bacteroides Fragilis* in the gut microbiomes of Alzheimer’s Disease activates microglia and**
2 **triggers pathogenesis in neuronal C/EBP β transgenic mice**

3 **Supplementary Information files**

4 **By**

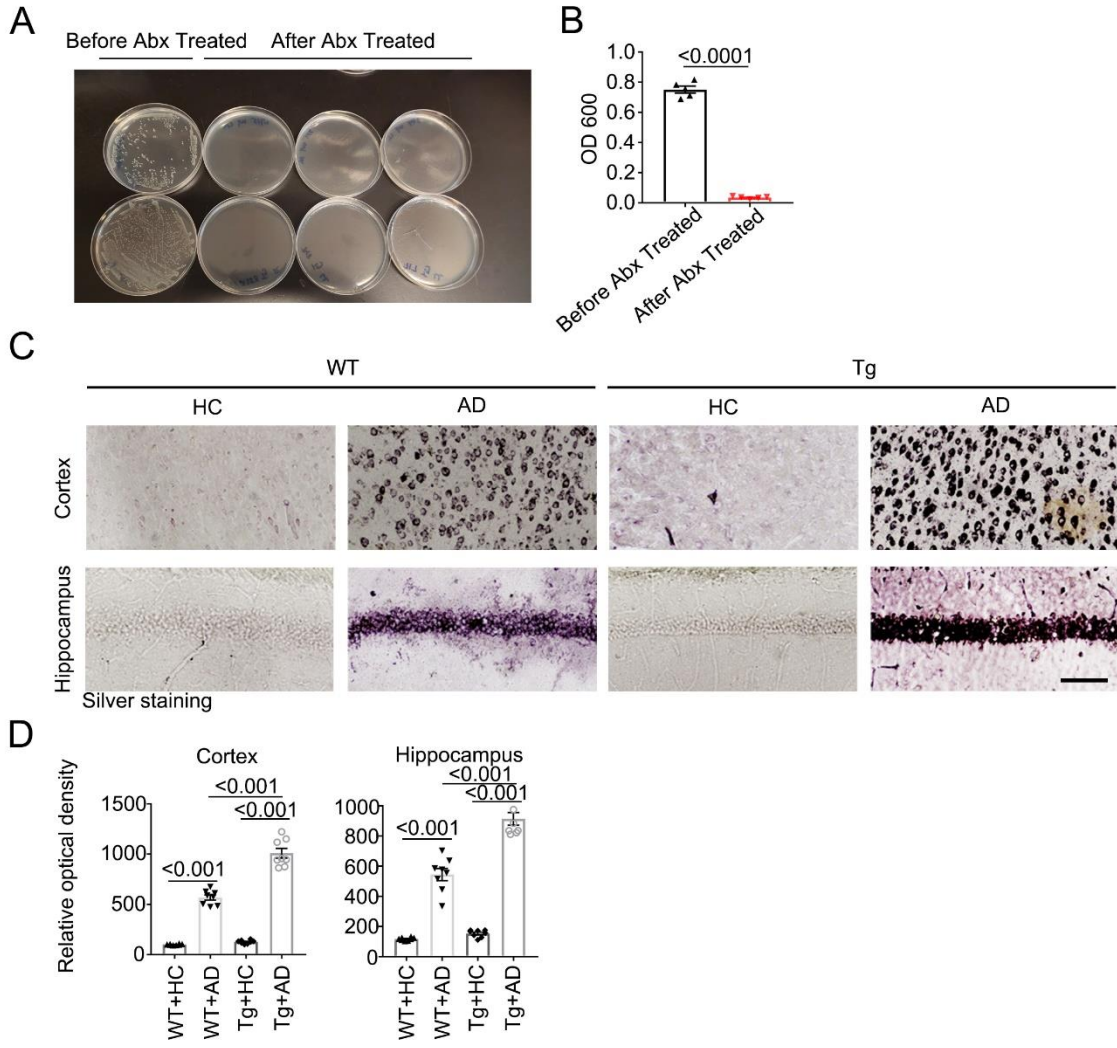
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6 Haran^{4,5,6}, Beth A. McCormick^{5,6}, Xiji Shu^{2,*}, Xiaochuan Wang^{7,8,*}, and Keqiang Ye^{1,9,*}

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10 **This file contains Supplementary figures 1-16.**



1 **Figure S1. AD humanized Abx-treated mice display increased aggregated proteins and**
2 **apoptotic neurons compared with HC humanized Abx-treated mice in WT and C/EBP β Tg**
3 **mice.**

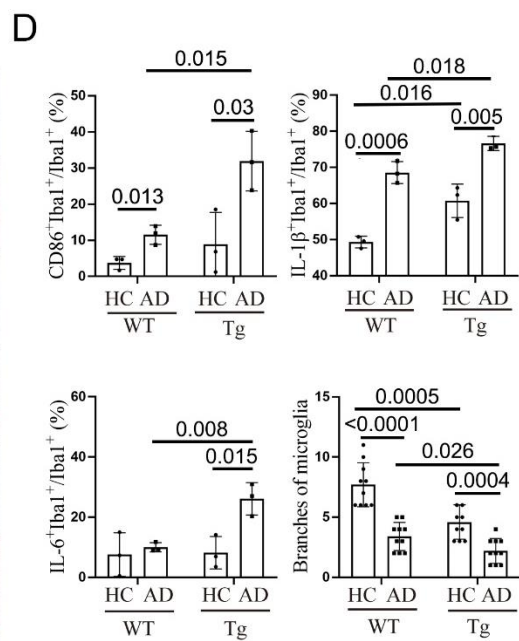
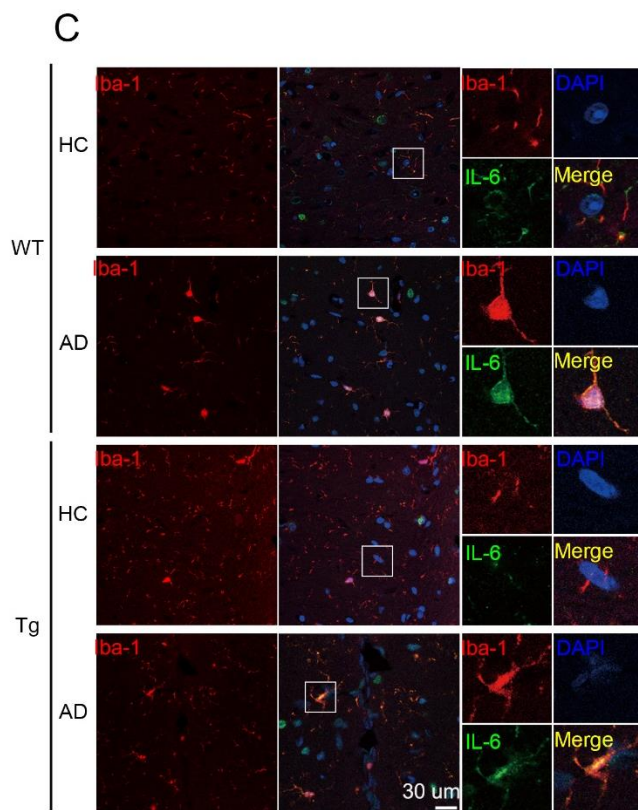
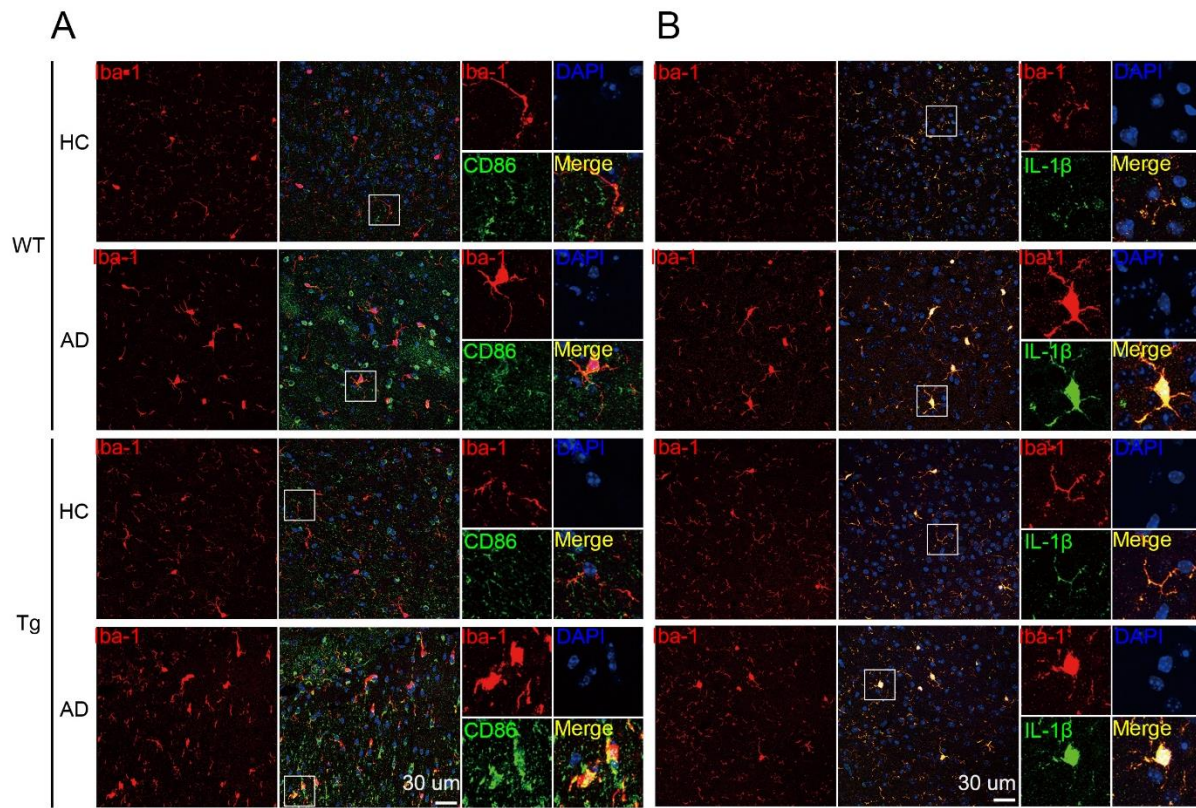
4 (A-B) In vitro cultures of bacteria from fecal pellets with/without Abx treated 3-month-old mice,
5 showed that antibiotics treatment was effective in killing the bacteria. (n = 5 biologically
6 independent samples in each group, data are shown as mean \pm SEM, two-tailed Student's t test).

7 (C) Silver staining in the cortex and the hippocampus. Scale bar: 50 μ m. (D) Quantitative

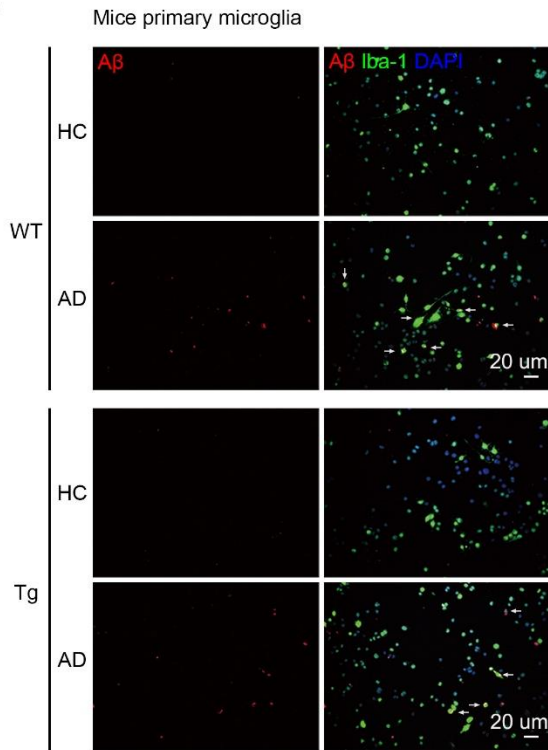
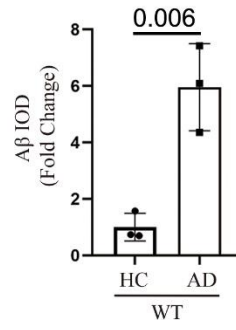
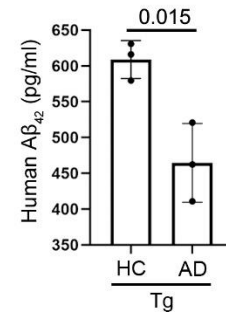
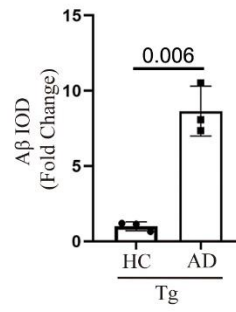
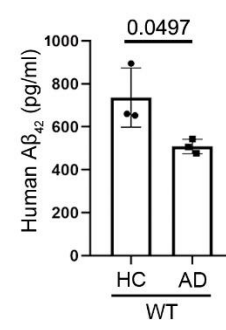
8 analysis of Silver staining. Aggregated proteins were significantly increased in AD humanized

9 Abx-treated mice. (n = 8 biologically independent samples in each group, data are shown as

10 mean \pm SEM, one-way ANOVA and Bonferroni's multiple comparison test).



1 **Figure S2. AD humanized Abx-treated mice display increased cytokines in microglia**
2 **compared with HC humanized Abx-treated mice in WT and C/EBP β Tg mice.**
3 (A) Immunofluorescent staining of Iba-1 (red) and CD86 (green) in the cortex region of the
4 brains from AD or HC humanized Abx-treated 6-month-old mice. Scale bar: 30 μ m. (B)
5 Immunofluorescent staining of Iba-1 (red) and IL-1 β (green) in the cortex region of the brains
6 from AD or HC humanized Abx-treated 6-month-old mice. Scale bar: 30 μ m. (C)
7 Immunofluorescent staining of Iba-1 (red) and IL-6 (green) in the cortex region of the brains
8 from AD or HC humanized Abx-treated 6-month-old mice. Scale bar: 30 μ m. (D) Quantitative
9 analysis of Iba-1⁺ and CD86⁺/IL-1 β ⁺/IL-6⁺ percentage in Iba-1⁺ microglia, and branches of Iba-
10 1⁺ microglia (n = 3 biologically independent samples in each group, data are shown as mean \pm
11 SEM, one-way ANOVA and Bonferroni's multiple comparison test).

A**B****C**

1 **Figure S3. AD humanized Abx-treated mouse primary microglia display better ability to**
2 **engulf A β than HC.**

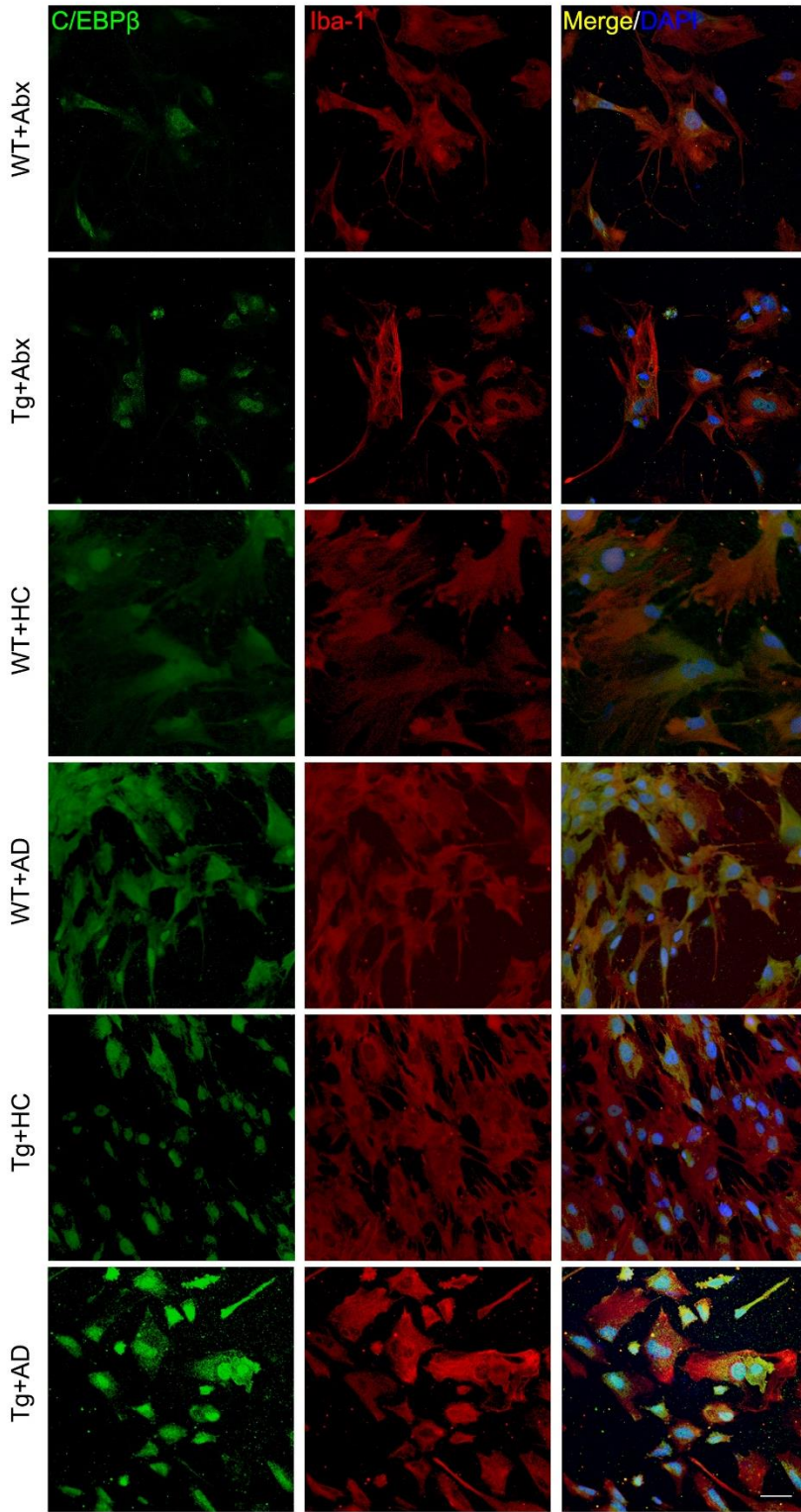
3 The primary cultures of microglia cells from AD or HC humanized Abx-treated 6-month-old
4 mice, then treated with human A β_{42} fibrils for 24 hours. (A) Immunofluorescent staining of
5 human A β (red) and Iba-1 (green) in the primary cultured microglia cells. Phagocytosed
6 microglia cells with engulfed A β_{42} was designed with white arrows. Scale bar: 20 μ m. (B)

7 Quantitative analysis A β IOD (Integrated optical density) in Iba-1⁺ microglia (n = 3 biologically
8 independent samples in each group, data are shown as mean \pm SEM, two-tailed Student's t test).

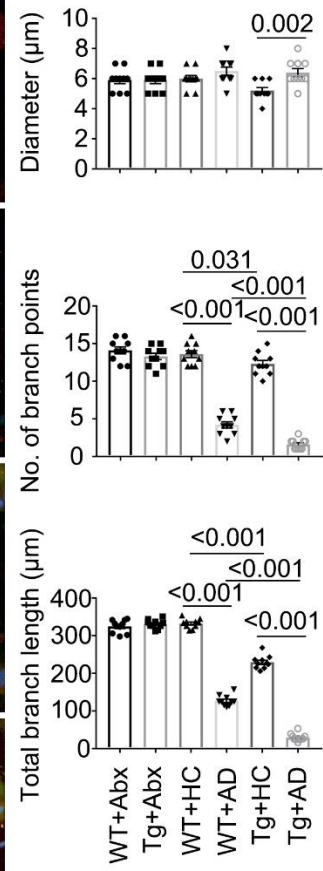
9 (C) Human A β_{42} concentration in the medium were detected with ELISA (n = 3 biologically

10 independent samples in each group, data are shown as mean \pm SEM, two-tailed Student's t test).

A
Rat primary microglia treated with Mice serum



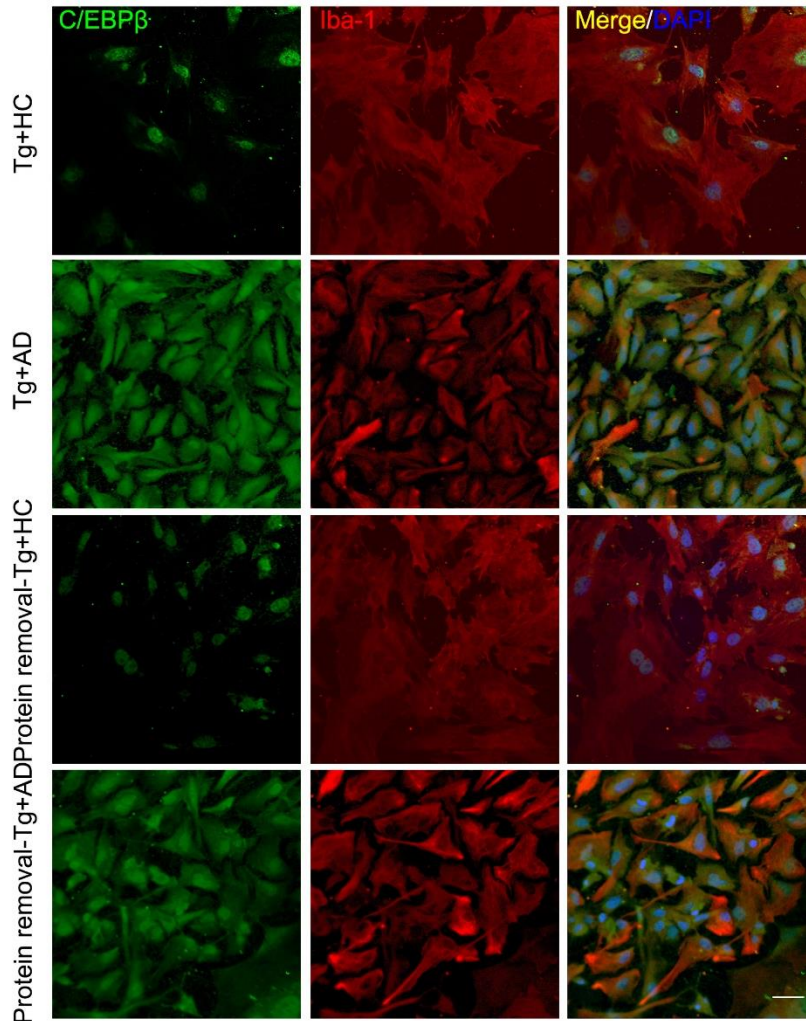
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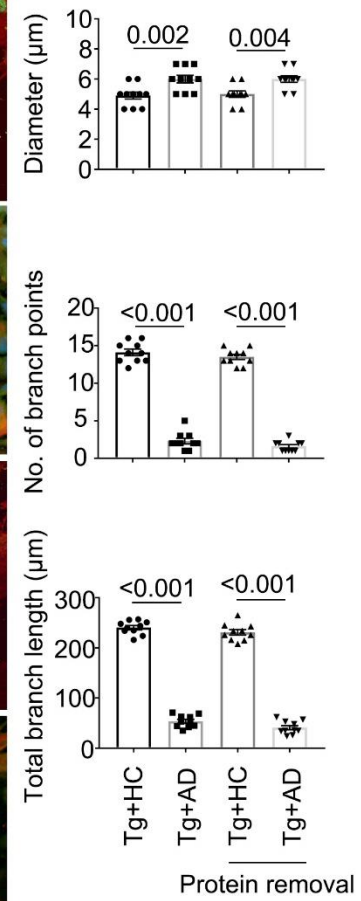
1 **Figure S4. AD humanized Abx-treated mouse serum strongly activated rat primary**
2 **microglia compared with HC humanized Abx-treated mouse serum.**
3 (A) Rat primary microglia treated with HC or AD humanized Abx-treated 6-month-old mice
4 serum (20 $\mu\text{g}/\text{ml}$), 24 h later, the microglia were fixed and staining with C/EBP β (green) and Iba-
5 1 (red). Scale bar: 10 μm . (B) Quantitative analysis of diameter, number of branch points, and
6 total branch length in Iba-1 (red) positive microglia. (n = 10 biologically independent samples in
7 each group, data are shown as mean \pm SEM, one-way ANOVA and Bonferroni's multiple
8 comparison test).

A

Rat primary microglia treated with Mice serum



B

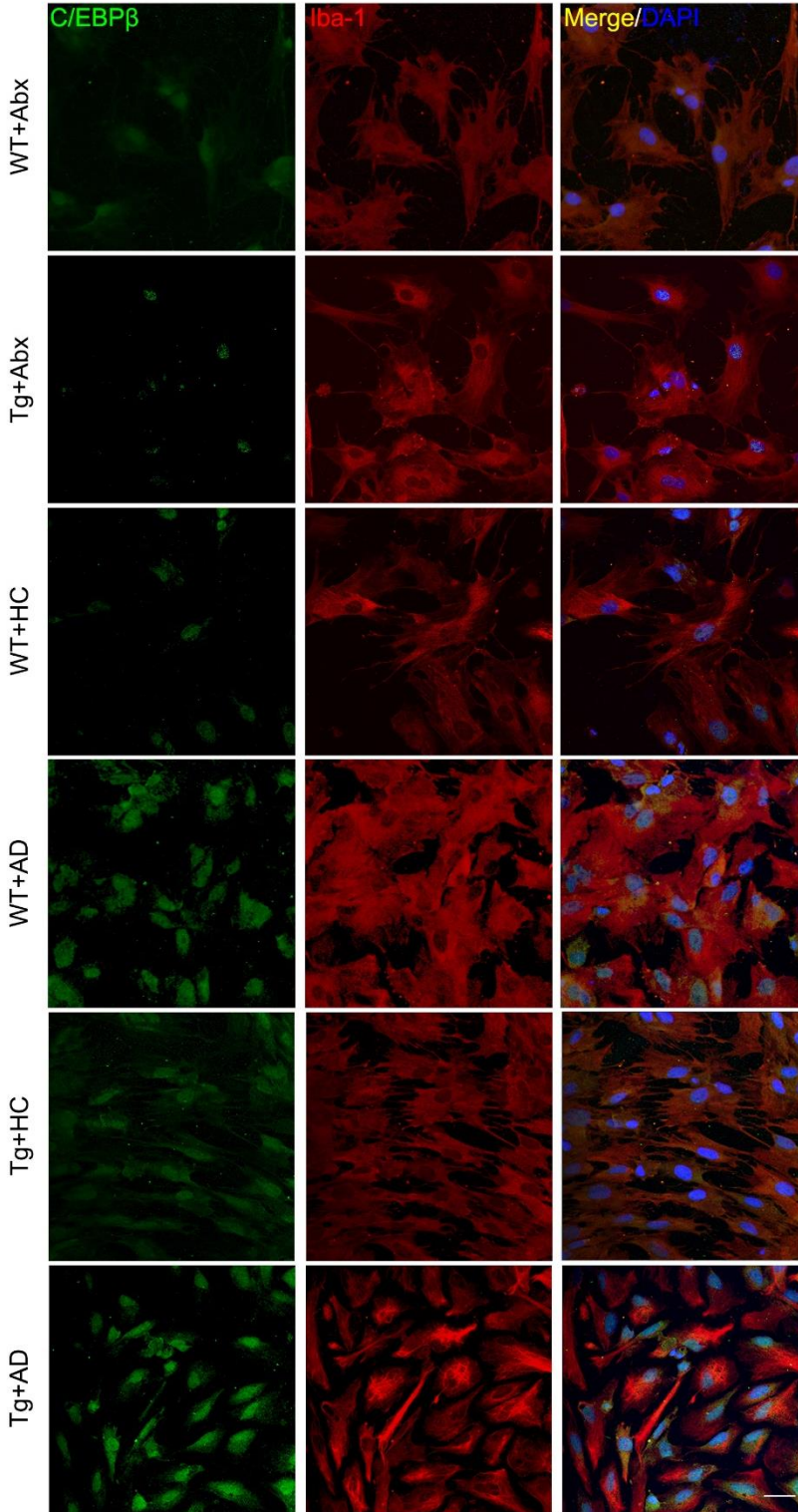


1 **Figure S5. Protein depletion from AD humanized Abx-treated mouse serum did not affect**
2 **rat primary microglia activation.**

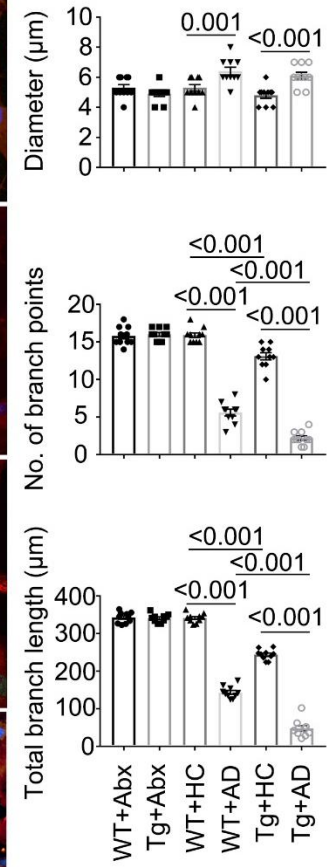
3 (A) Rat primary microglia treated with HC or AD humanized Abx-treated 6-month-old C/EBP β
4 Tg mice serum (20 μ g/ml) or TCA treated protein removal serum, 24 h later, the microglia were
5 fixed and staining with C/EBP β (green) and Iba-1 (red). Scale bar: 10 μ m. (B) Quantitative
6 analysis of diameter, number of branch points, and total branch length in Iba-1 (red) positive
7 microglia. (n = 10 biologically independent samples in each group, data are shown as mean \pm
8 SEM, one-way ANOVA and Bonferroni's multiple comparison test).

A

Rat primary microglia treated with Mice brain lysis



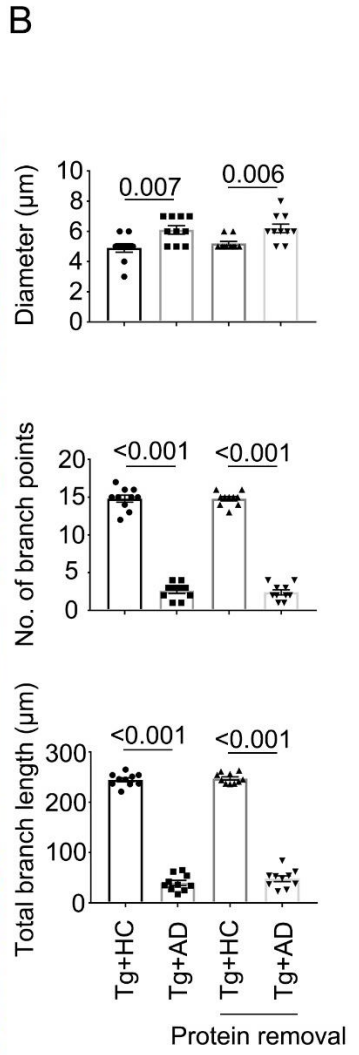
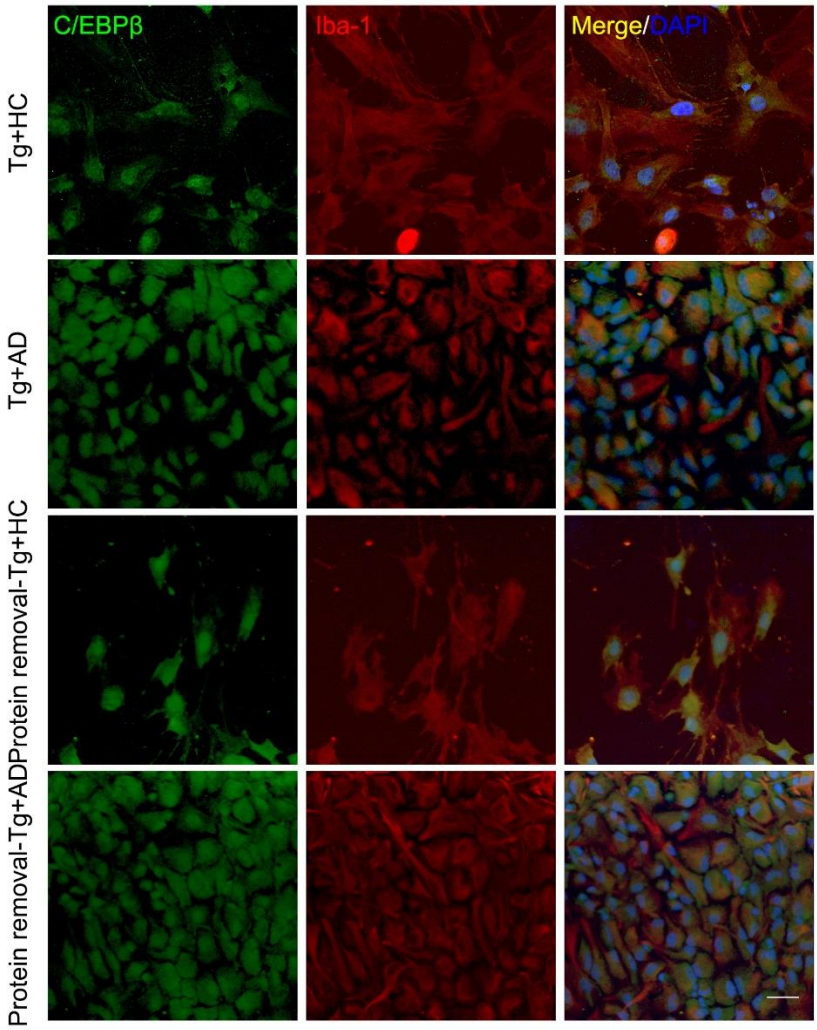
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1 **Figure S6. AD humanized Abx-treated mouse brain lysates activated rat primary microglia**
2 **more robust than HC.**

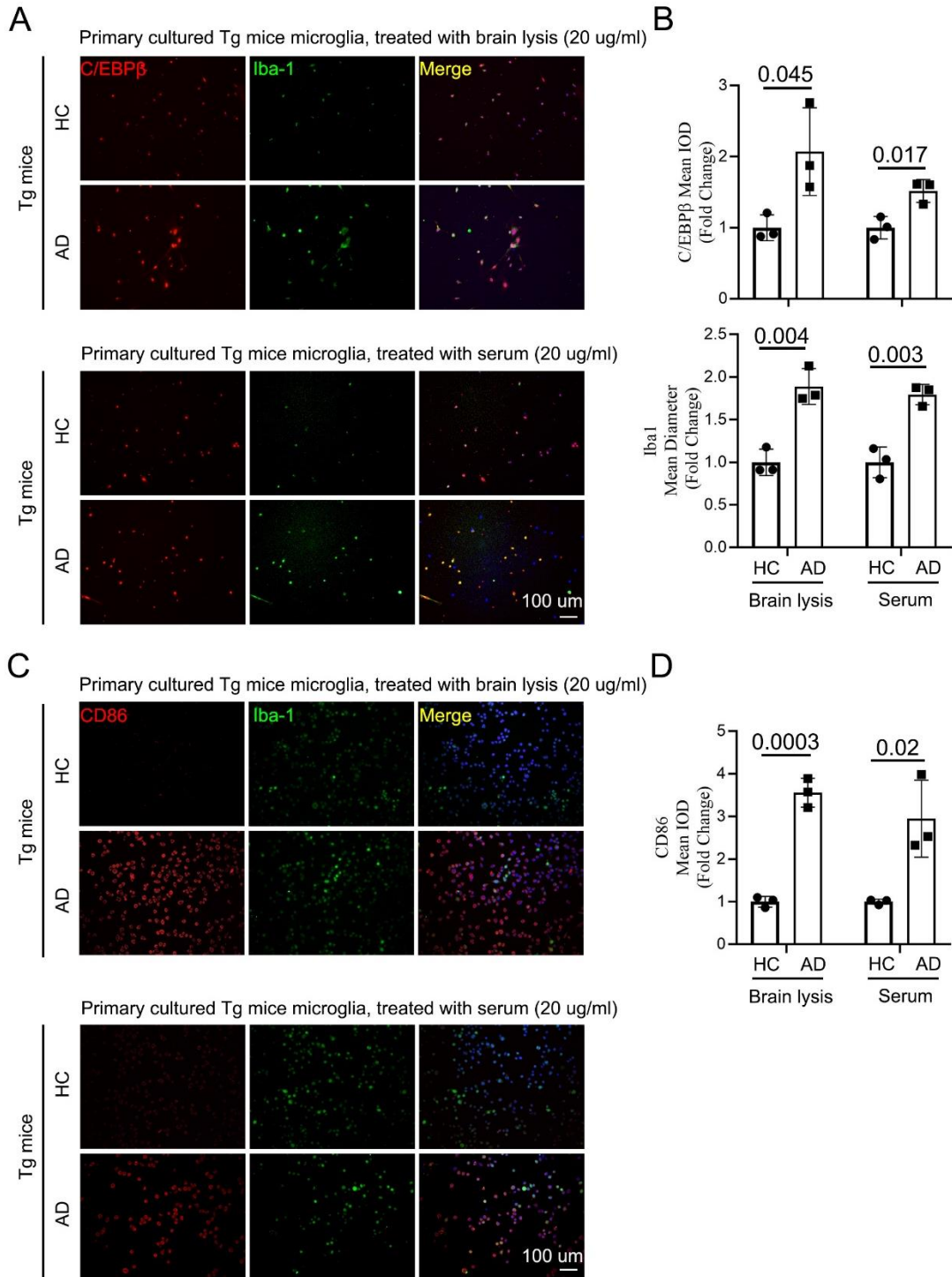
3 (A) Rat primary microglia treated with HC or AD humanized Abx-treated 6-month-old mice
4 brain lysis (20 µg/ml), 24 h later, the microglia were fixed and staining with C/EBPβ (green) and
5 Iba-1 (red). Scale bar: 10 µm. (B) Quantitative analysis of diameter, number of branch points,
6 and total branch length in Iba-1 (red) positive microglia. n=10 biologically independent samples
7 in each group, data are shown as mean ± SEM, one-way ANOVA and Bonferroni's multiple
8 comparison test.

A
Rat primary microglia treated with Mice brain lysis



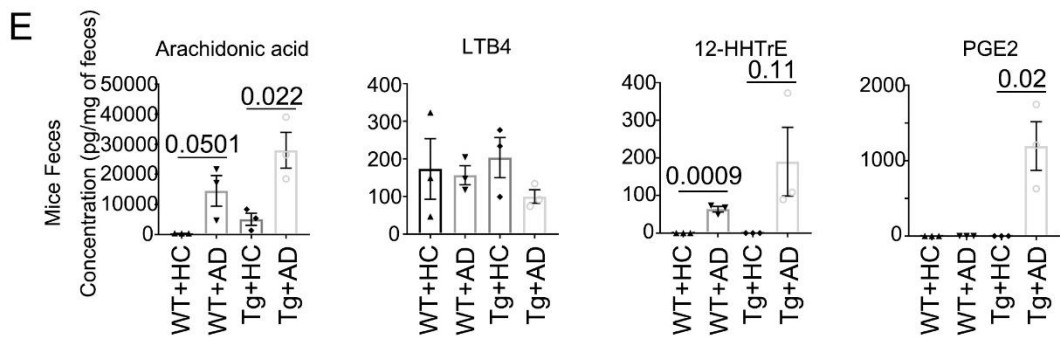
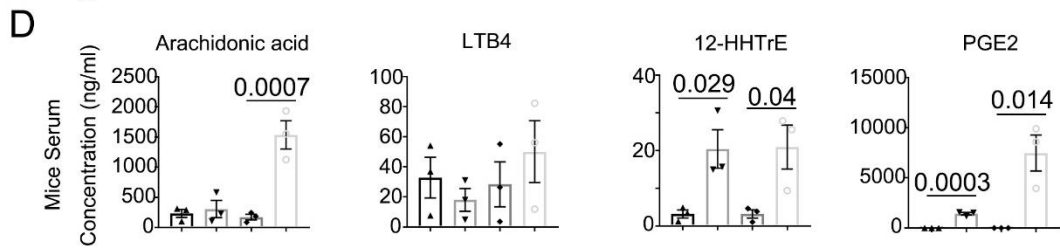
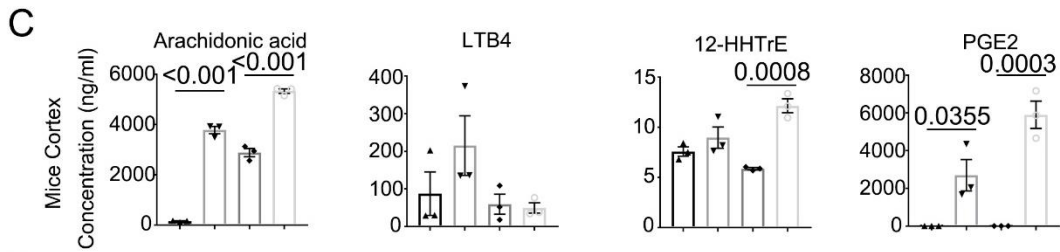
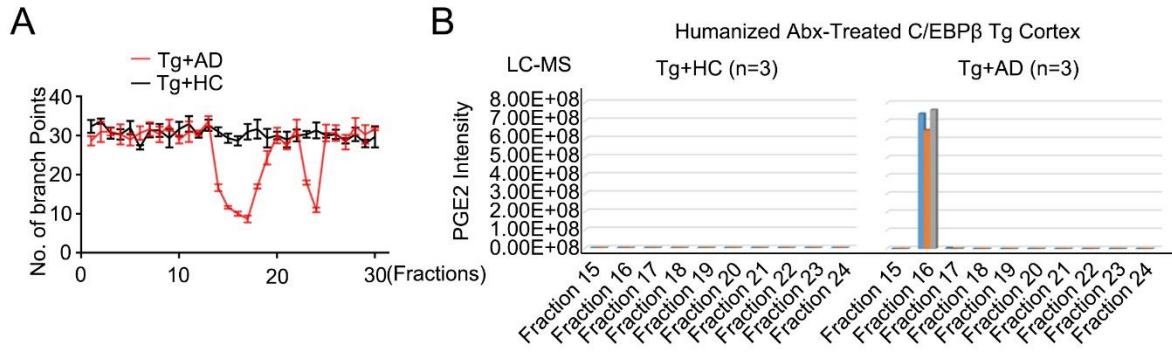
1 **Figure S7. Protein depletion from AD humanized Abx-treated mice brain lysates did not**
2 **affect rat primary microglia.**

3 (A) Rat primary microglia treated with HC or AD humanized Abx-treated 6-month-old C/EBP β
4 Tg mice brain lysis (20 μ g/ml) or TCA treated protein removal brain lysis, 24 h later, the
5 microglia were fixed and staining with C/EBP β (green) and Iba-1 (red). Scale bar: 10 μ m. (B)
6 Quantitative analysis of diameter, number of branch points, and total branch length in Iba-1 (red)
7 positive microglia. (n = 10 biologically independent samples in each group, data are shown as
8 mean \pm SEM, one-way ANOVA and Bonferroni's multiple comparison test).



1 **Figure S8. AD humanized Abx-treated mice brain lysates/serum activated Tg mice primary**
2 **microglia more robust than HC.**

3 The primary cultured microglia from C/EBP β Tg mice, treated with HC/AD humanized Abx-
4 treated 6-month-old mice brain lysis or serum for 24 h. (A) The microglia were fixed and
5 staining with C/EBP β (red) and Iba-1 (green). Scale bar: 100 μ m. (B) Quantitative analysis of
6 C/EBP β mean IOD and Iba-1⁺ microglia soma mean diameter (n = 3 biologically independent
7 samples in each group, data were shown as mean \pm SEM, two-tailed Student's t test). (C) The
8 microglia were fixed and staining with CD86 (red) and Iba-1 (green). Scale bar: 100 μ m. (D)
9 Quantitative analysis of CD86 mean IOD (n = 3 biologically independent samples in each group,
10 data were shown as mean \pm SEM, two-tailed Student's t test).

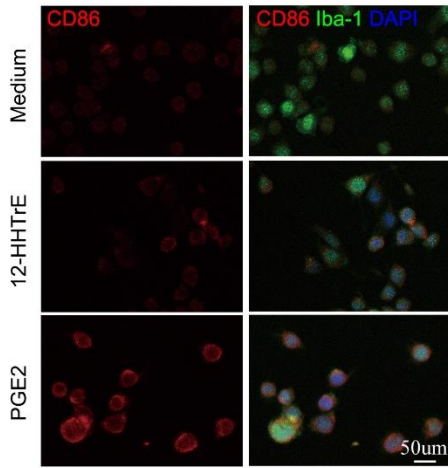


1 **Figure S9. AD humanized Abx-treated mice display increased arachidonic acid, 12-**
2 **HHTrE, and PGE2 compared with HC.**

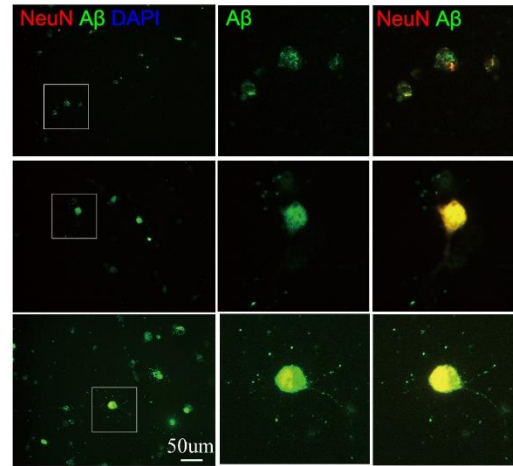
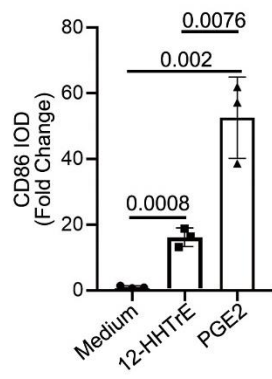
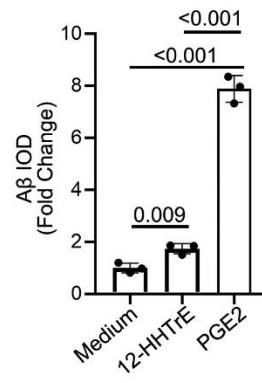
3 (A) TCA treated protein removal from HC or AD humanized Abx-treated 6-month-old C/EBP β
4 Tg mice brain lysates were separated into 30 fractions with HPLC. 96-well planted rat primary
5 microglia were treated with each fraction for 10 ul/well. 24 h later, the number of branch points
6 was counted for the activity of microglia. Only AD humanized Abx-treated C/EBP β Tg mice
7 showed changes in the activity, including fractions 12-20, 22-25. (B) After LC-MS analysis of
8 fractions 15-24, PGE2 was found increased in AD humanized Abx-treated mice compared with
9 HC humanized Abx-treated 6-month-old mice. Source data are provided as a Source Data file.
10 (C-E) HPLC analysis the concentrations of arachidonic acid (AA) and its metabolites in HC or
11 AD humanized Abx-treated 6-month-old WT or C/EBP β Tg mice brain lysis (C), serum (D), and
12 feces samples (E). (n=3 biologically independent samples in each group, data are shown as mean
13 \pm SEM, one-way ANOVA and Bonferroni's multiple comparison test).

A

Primary cultured Tg mice microglia

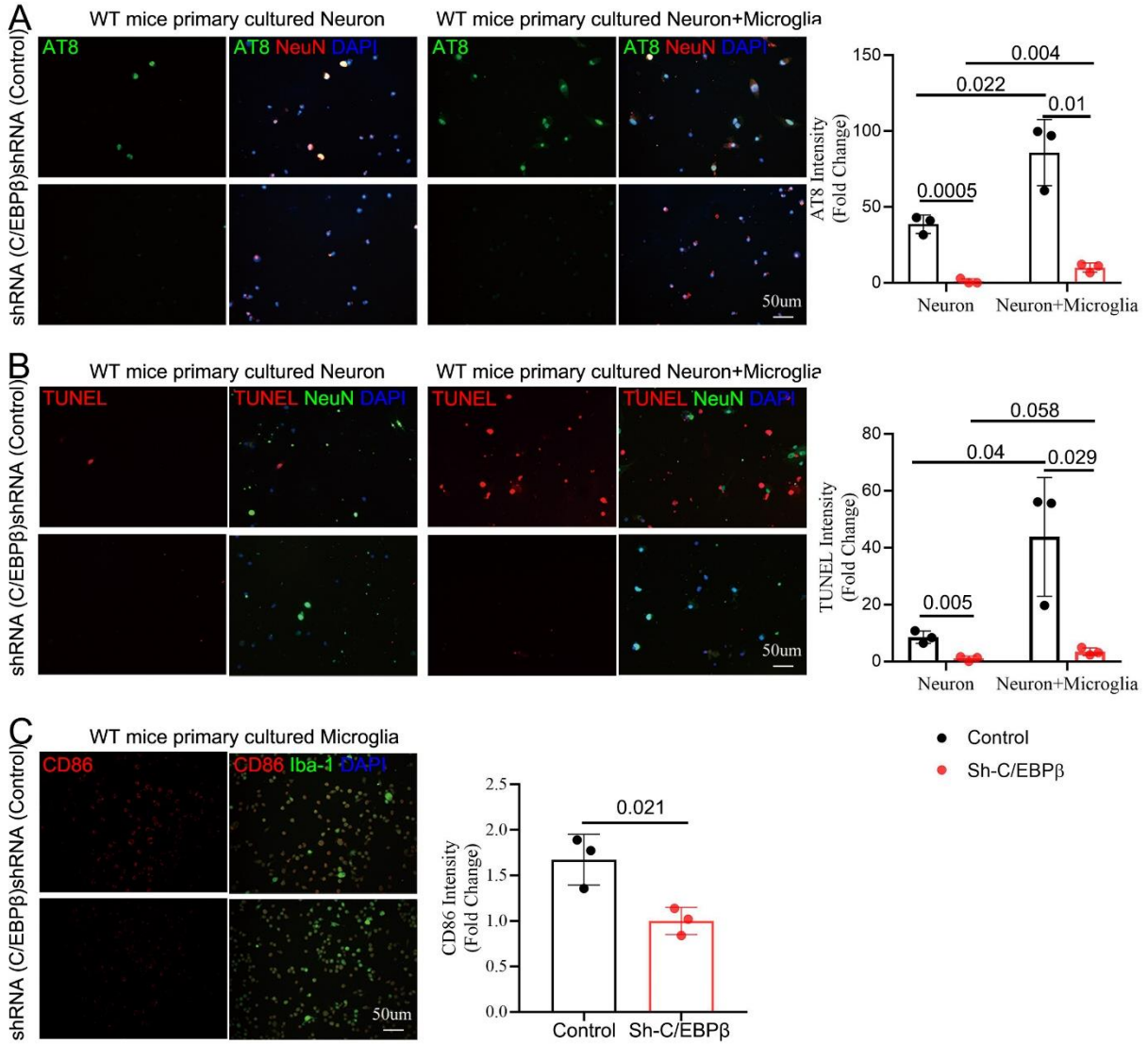
**B**

Primary cultured Tg mice neuron

**C****D**

1 **Figure S10. 12-HHTrE and PGE2 enhanced the activation of microglia and neuronal A β ₄₂**
2 **deposition.**

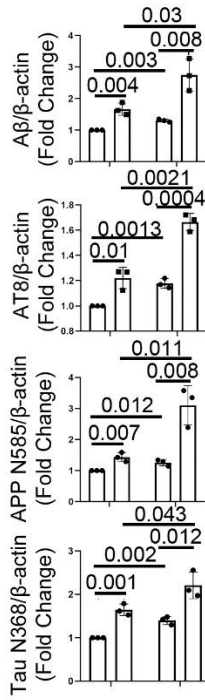
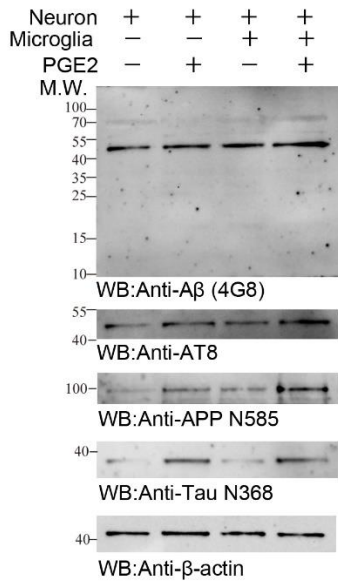
3 In vitro cultured microglia and neurons from Tg mice were employed for the analysis of
4 microglia activation and A β ₄₂ deposition. (A) The primary cultured microglia from C/EBP β Tg
5 mice, treated with 12-HHTrE or PGE2 (20 ug/ml) for 24 h. Then fixed for immunofluorescent
6 staining of CD86 (red) and Iba-1 (green). (B) The primary cultured neuron from C/EBP β Tg
7 mice, treated with 12-HHTrE or PGE2 (20 ug/ml) for 24 h. Then fixed for immunofluorescent
8 staining of NeuN (red) and A β (4G8, green). (C-D) Quantitative analysis of CD86/A β mean IOD
9 (n = 3 biologically independent samples in each group, data were shown as mean \pm SEM, one-
10 way ANOVA and Bonferroni's multiple comparison test).



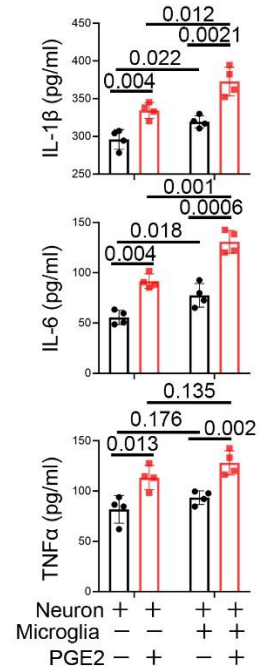
1 **Figure S11. Deletion of C/EBP β from primary neuronal cultures or microglia alleviate the**
2 **detrimental effect of *B. Fragilis* metabolites.**

3 The primary cultured neurons and neurons + microglia mixture from WT mice, followed w/wo
4 AAV-sh-C/EBP β knockdown, then treated with PGE2 (20 ug/ml) for 24 h. (A) Immunofluorescent
5 staining of AT8 (green) and NeuN (red) in the primary cultured neurons or neurons + microglia
6 mixture. Scale bar: 50 μ m. Quantitative analysis showed AT8 intensity was significantly decreased
7 after C/EBP β knockdown (n=3 biologically independent samples in each group, data were shown
8 as mean \pm SEM, one-way ANOVA and Bonferroni's multiple comparison test). (B)
9 Immunofluorescent staining of NeuN (green) and TUNEL (red) in the primary cultured neuron or
10 neurons + microglia mixture. Scale bar: 50 μ m. Quantitative analysis showed TUNEL intensity
11 was significantly decreased after C/EBP β knockdown (n = 3 biologically independent samples in
12 each group, data were shown as mean \pm SEM, one-way ANOVA and Bonferroni's multiple
13 comparison test). (C) Immunofluorescent staining of Iba-1 (green) and CD86 (red) in the primary
14 cultured microglia. Scale bar: 50 μ m. Quantitative analysis showed CD86 intensity was
15 significantly decreased after C/EBP β knockdown (n = 3 biologically independent samples in each
16 group, data were shown as mean \pm SEM, two-tailed Student's t test).

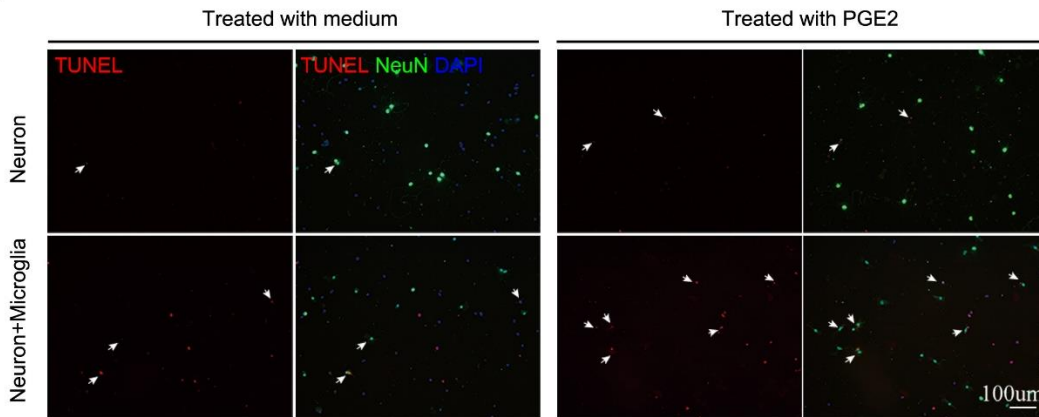
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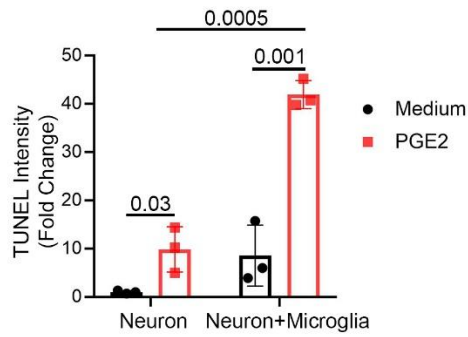
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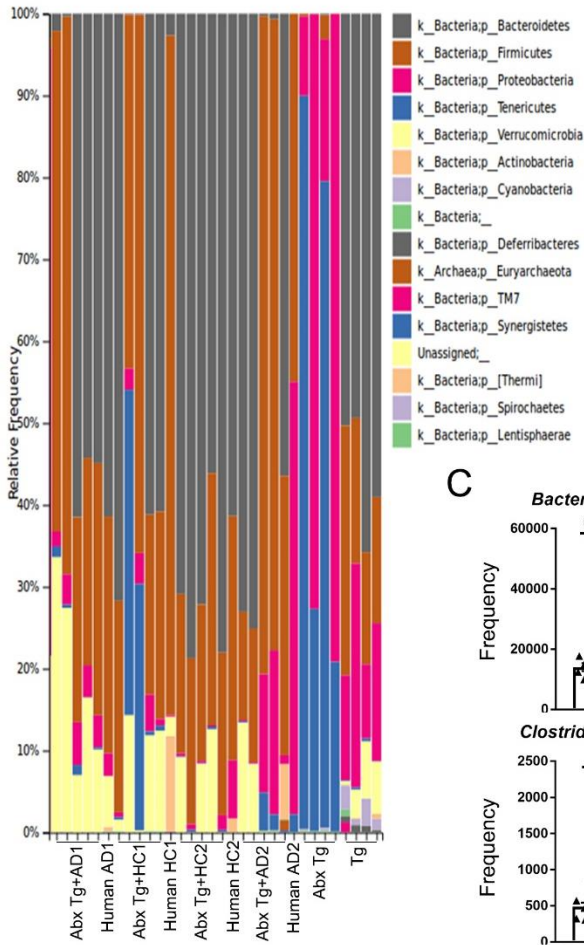
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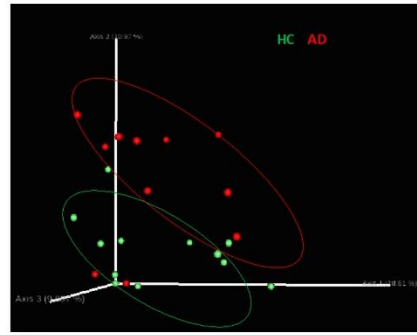
1 **Figure S12. Mixed cultures of neurons and microglia showed higher inflammatory**
2 **cytokines, A β ₄₂, AT8 levels, and apoptosis than neurons alone.**

3 The primary cultured neurons, mixed cultures of neurons + microglia from Thy1- C/EBP β Tg mice.
4 Treated w/wo PGE2. (A) WB detecting the protein levels of A β , AT8, APP N585, and Tau N368.
5 Quantitative analysis the protein fold changes between neurons, and mixed cultured neurons +
6 microglia (n = 3 biologically independent samples in each group, data were shown as mean \pm SEM,
7 one-way ANOVA and Bonferroni's multiple comparison test). (B) inflammatory cytokines
8 including IL-1 β , IL-6, and TNF- α were tested by ELISA (n = 3 biologically independent samples
9 in each group, data were shown as mean \pm SEM, one-way ANOVA and Bonferroni's multiple
10 comparison test). (C) Immunofluorescent staining of NeuN (green) and TUNEL (red) in the
11 primary cultured neurons or neurons + microglia mixture. Scale bar: 100 μ m. Arrows showed the
12 NeuN positive TUNEL deposited. (D) Quantitative analysis showed TUNEL intensity was
13 significantly increased after PGE2 treatment or microglia addition (n = 3 biologically independent
14 samples in each group, data were shown as mean \pm SEM, one-way ANOVA and Bonferroni's
15 multiple comparison test).

A

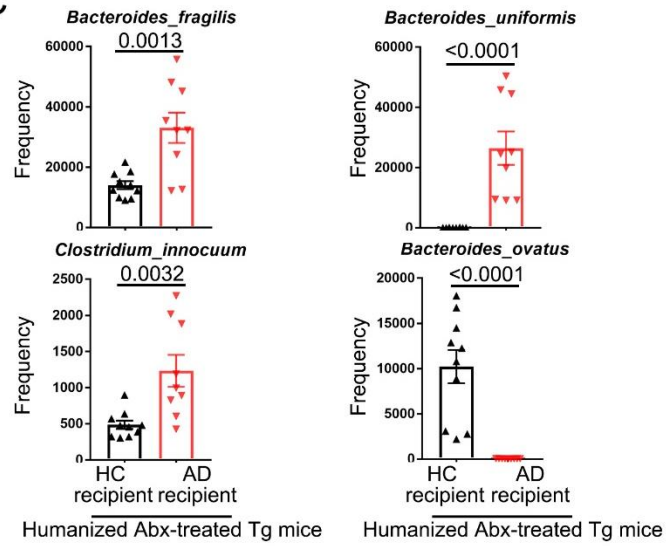


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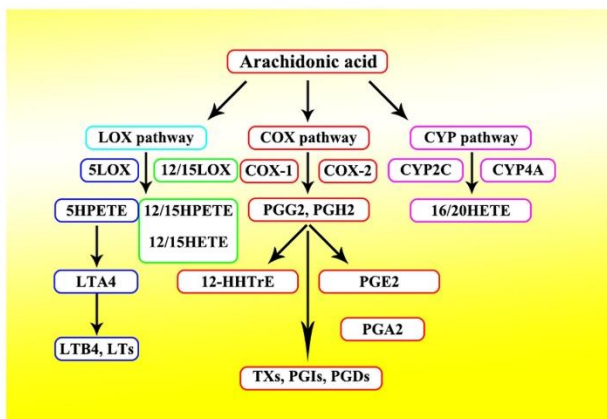


HC = HC donor and HC microbiota engraftment in Abx-treated Tg mice
AD = AD donor and AD microbiota engraftment in Abx-treated Tg mice

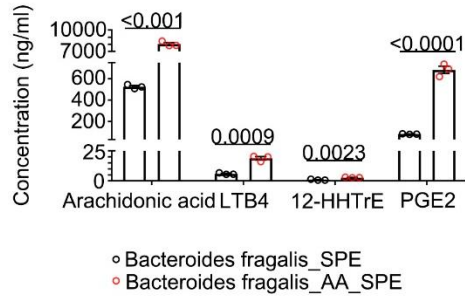
C



D



E



1 **Figure S13. Microbiome analysis of humanized Abx-treated WT and C/EBP β Tg mice fecal**
2 **samples, revealing Bacteroides elevation in the feces from human AD fecal inoculated Abx-**
3 **treated mice.**

4 (A) Relative abundance of bacterial phyla determined by high throughput sequencing analysis.

5 (B) Principal coordinate plot (PcoA) of microbial community structure. (C) Mean frequency of

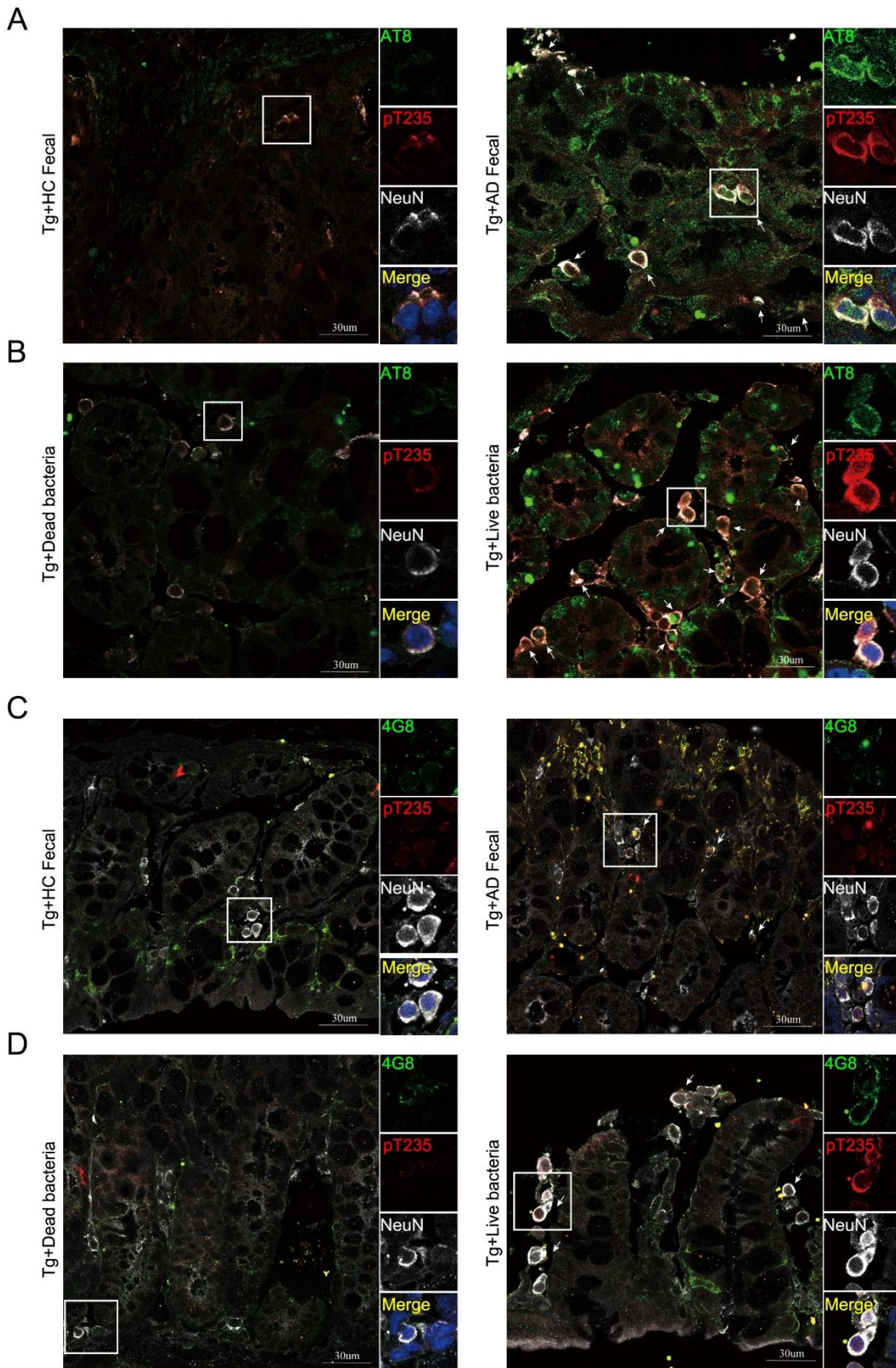
6 bacterial species. (n = 9-10 biologically independent samples in each group, data represent the

7 means \pm SEM, two-tailed Student's t test). (D) KEGG arachidonic acid metabolism pathway of

8 *Bacteroides fragilis* (1.11.1.9). (E) Concentrations of arachidonic acid (AA) and its metabolites

9 in culture medium from *in vitro* cultures of *Bacteroides fragilis* (n = 3 biologically independent

10 samples in each group, data were shown as mean \pm SEM, two-tailed Student's t test).



1 **Figure S14. The enteric neuronal A β ₄₂ and AT8 signals were increased after human AD**
2 **fecal or live *B. fragilis* treatment.**

3 (A-B) Immunofluorescent staining of AT8 (green) and active C/EBP β pT235 (red) in the
4 intestine region of gut from human AD fecal or *B. Fragilis* treated C/EBP β Tg mice
5 (representative of 3 mice). Scale bar: 30 μ m. Arrows showed the AT8 and pT235 positive

6 neurons. (C-D) Immunofluorescent staining of A β 4G8 (green) and C/EBP β pT235 (red) in the
7 intestine region of gut from human AD fecal or *B. Fragilis* treated C/EBP β Tg mice
8 (representative of 3 mice). Scale bar: 30 μ m. Arrows showed the 4G8 and pT235 positive
9 neurons.

Lipid Metabolism

A

		Welch's Two-Sample t-Test			
		A:D-CERPs Tg HC-CERPs Tg			
Feces		Colored by:			
Sub Pathway	Biochemical Name	Statistics	Fold of Change	Statistics	Fold of Change
Long Chain Saturated Fatty Acid	myristate (14:0)	1.60	1.60		
	palmitate (16:0)	1.74	1.74		
	stearate (18:0)	1.59	1.59		
	arachidate (20:0)	1.64	1.64		
Long Chain Monounsaturated Fatty Acid	palmitoleate (16:1n7)	1.79	1.79		
	oleate/vaccenate (18:1)	2.04	2.04		
	eicosanoate (20:1)	2.74	2.74		
	erucate (22:1n9)	2.02	2.02		
Long Chain Polyunsaturated Fatty Acid (n3 and n6)	hexadecatrienoate (16:3n3)	1.58	1.58		
	stearidonate (18:4n3)	1.52	1.52		
	eicosapentaenoate (EPA, 20:5n3)	2.17	2.17		
	heneicosapentaenoate (21:5n3)	1.77	1.77		
	docosapentaenoate (n3 EPA, 22:5n3)	2.31	2.31		
	docosahexaenoate (DHA, 22:6n3)	2.23	2.23		
	risinate (24:6n3)	3.51	3.51		
	hexadecadienoate (16:2n6)	1.57	1.57		
	linoleate (18:2n6)	1.76	1.76		
	linolenate (alpha or gamma, 18:3n3 or n6)	1.75	1.75		
	dihomo-linoleate (20:2n6)	1.45	1.45		
	dihomo-linolenate (20:3n3 or n6)	1.61	1.61		
	arachidonate (20:4n6)	2.27	2.27		
	docosatrienoate (22:3n6)*	0.65	0.65		
adrenate (22:4n6)	1.72	1.72			
docosapentaenoate (n6 EPA, 22:5n6)	1.44	1.44			
docosadienoate (22:2n6)	1.72	1.72			

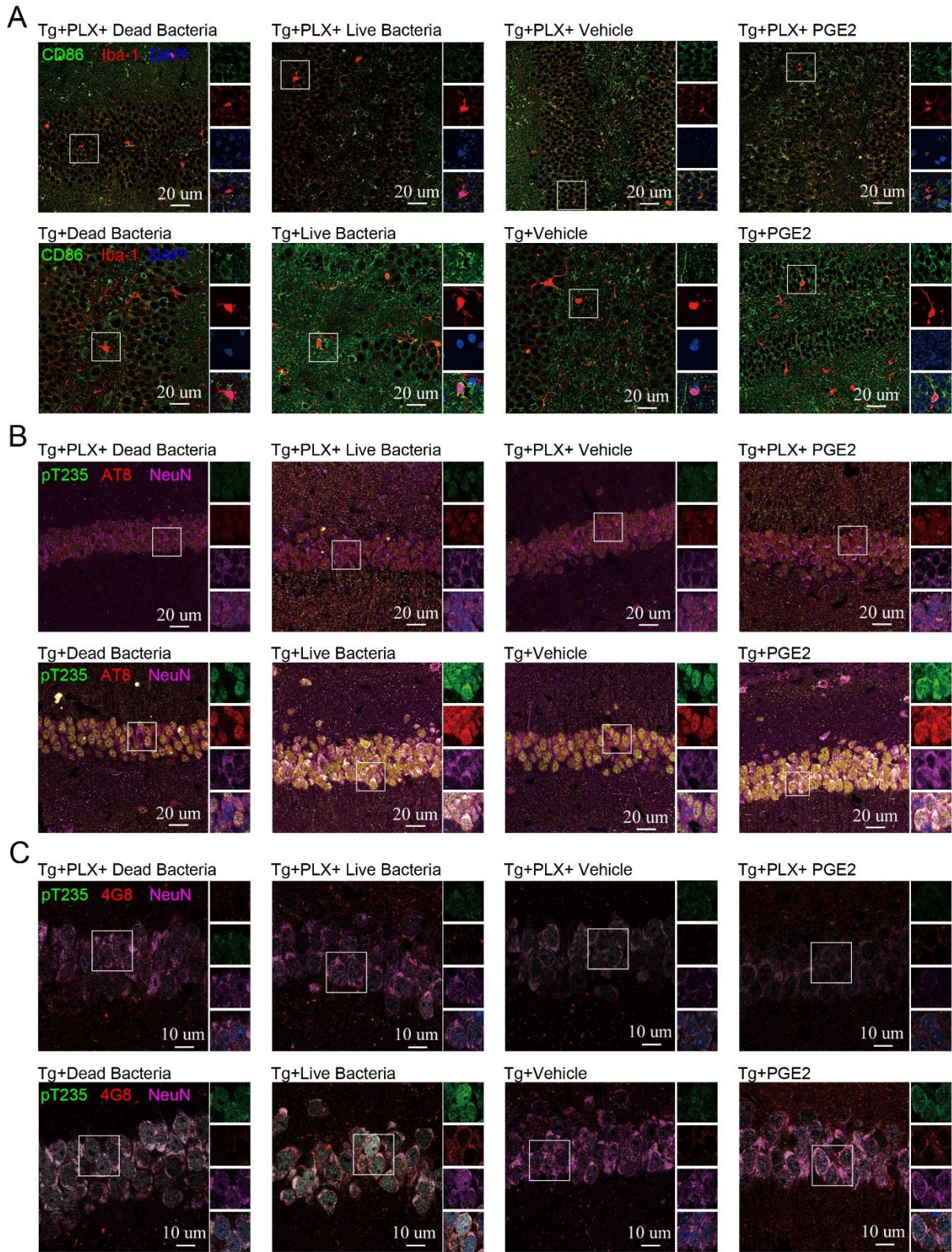
		Welch's Two-Sample t-Test			
		A:D-CERPs Tg HC-CERPs Tg			
Feces		Colored by:			
Sub Pathway	Biochemical Name	Statistics	Fold of Change	Statistics	Fold of Change
Monoacylglycerol	1-myristoylglycerol (14:0)	1.68	1.68		
	1-palmitoylglycerol (16:0)	2.58	2.58		
	1-palmitoleoylglycerol (16:1)*	1.89	1.89		
	1-oleoylglycerol (18:1)	2.43	2.43		
	1-linoleoylglycerol (18:2)	2.07	2.07		
	1-linolenoylglycerol (18:3)	1.92	1.92		
	1-dihomo-linolenoylglycerol (20:3)	2.58	2.58		
	1-docosahexaenoylglycerol (22:6)	1.91	1.91		
	2-myristoylglycerol (14:0)	1.35	1.35		
	2-palmitoylglycerol (16:0)	1.39	1.39		
	2-diacylglycerol (18:1)	2.72	2.72		
	2-linoleoylglycerol (18:2)	1.47	1.47		
	2-docosahexaenoylglycerol (22:6)*	1.61	1.61		
	1-heptadecanoylglycerol (17:1)*	1.62	1.62		
Diacylglycerol	diacylglycerol (16:1/18:2 [2], 18:0/18:2)	0.66	0.66		
	palmitoyl-oleoylglycerol (16:0/18:1) [2]	0.62	0.62		
	palmitoyl-linolenoylglycerol (16:0/18:2)	0.74	0.74		
	palmitoyl-linoleoylglycerol (16:0/18:2)	0.64	0.64		
	oleoyl-linoleoylglycerol (18:1/18:2) [2]	0.72	0.72		
	linoleoyl-linoleoylglycerol (18:2/18:2)	0.78	0.78		
	linoleoyl-linolenoylglycerol (18:2/18:3)	0.67	0.67		
	linoleoyl-linolenoylglycerol (18:2/18:3)	0.72	0.72		
	linoleoyl-linolenoylglycerol (18:2/18:3)	0.62	0.62		
	linolenoyl-linolenoylglycerol (18:3/18:3)	0.51	0.51		
	linoleoyl-docosahexaenoylglycerol (1)	0.86	0.86		

B

		Welch's Two-Sample t-Test			
		CERPs Tg WT		A:D-CERPs Tg HC-CERPs Tg	
Brain		Colored by:			
Sub Pathway	Biochemical Name	Statistics	Fold of Change	Statistics	Fold of Change
Long Chain Saturated Fatty Acid	myristate (14:0)	0.98	0.98	1.61	1.61
	palmitate (16:0)	1.14	1.14	1.77	1.77
	stearate (17:0)	1.19	1.19	2.00	2.00
	stearate (18:0)	1.16	1.16	1.66	1.66
Long Chain Monounsaturated Fatty Acid	nonadecanoate (19:0)	1.17	1.17	2.25	2.25
	arachidate (20:0)	1.26	1.26	2.10	2.10
	palmitoleate (16:1n7)	0.92	0.92	2.00	2.00
	10-heptadecanoate (17:1n7)	1.02	1.02	2.36	2.36
Long Chain Polyunsaturated Fatty Acid	oleate/vaccenate (18:1)	1.15	1.15	2.12	2.12
	10-monodienoate (19:1n9)	1.09	1.09	2.74	2.74
	eicosanoate (20:1)	1.20	1.20	2.52	2.52
	erucate (22:1n9)	1.27	1.27	3.13	3.13
Long Chain Polyunsaturated Fatty Acid (n3 and n6)	tetradecatrienoate (14:2)*	0.93	0.93	1.50	1.50
	hexadecatrienoate (16:3n3)	1.11	1.11	1.62	1.62
	stearidonate (18:4n3)	1.11	1.11	1.58	1.58
	eicosapentaenoate (EPA, 20:5n3)	1.13	1.13	2.10	2.10
	docosapentaenoate (n3 EPA, 22:5n3)	1.03	1.03	2.68	2.68
	docosahexaenoate (DHA, 22:6n3)	1.18	1.18	2.30	2.30
	docosatrienoate (22:3n3)	1.78	1.78	1.04	1.04
	risinate (24:6n3)	1.27	1.27	2.67	2.67
	hexadecadienoate (16:2n6)	1.17	1.17	1.55	1.55
	linoleate (18:2n6)	1.34	1.34	1.99	1.99
	linolenate (alpha or gamma, 18:3n3 or n6)	1.06	1.06	2.00	2.00
	dihomo-linoleate (20:2n6)	1.50	1.50	2.37	2.37
	dihomo-linolenate (20:3n3 or n6)	1.32	1.32	2.10	2.10
	arachidonate (20:4n6)	0.99	0.99	1.01	1.01
docosatrienoate (22:3n6)*	0.82	0.82	3.32	3.32	
adrenate (22:4n6)	1.12	1.12	2.28	2.28	
docosapentaenoate (n6 EPA, 22:5n6)	1.21	1.21	2.35	2.35	
docosadienoate (22:2n6)	1.46	1.46	3.01	3.01	
mead acid (20:3n6)	0.84	0.84	2.21	2.21	

		Welch's Two-Sample t-Test			
		CERPs Tg WT		A:D-CERPs Tg HC-CERPs Tg	
Brain		Colored by:			
Sub Pathway	Biochemical Name	Statistics	Fold of Change	Statistics	Fold of Change
Monoacylglycerol	1-myristoylglycerol (14:0)	0.92	0.92	1.42	1.42
	1-palmitoylglycerol (16:0)	1.06	1.06	2.06	2.06
	1-palmitoleoylglycerol (16:1)*	0.86	0.86	2.37	2.37
	1-oleoylglycerol (18:1)	1.04	1.04	1.67	1.67
	1-linoleoylglycerol (18:2)	1.34	1.34	2.53	2.53
	1-dihomo-linolenoylglycerol (20:3)	0.85	0.85	2.78	2.78
	1-arachidonylglycerol (20:4)	0.96	0.96	2.10	2.10
	1-docosahexaenoylglycerol (22:6)	1.00	1.00	1.97	1.97
	2-palmitoylglycerol (16:0)	1.03	1.03	2.23	2.23
	2-palmitoleoylglycerol (16:1)*	0.83	0.83	2.46	2.46
	2-oleoylglycerol (18:1)	0.90	0.90	2.62	2.62
	2-linoleoylglycerol (18:2)	1.32	1.32	2.47	2.47
	2-arachidonylglycerol (20:4)	0.97	0.97	1.93	1.93
	2-docosahexaenoylglycerol (22:6)*	0.93	0.93	1.84	1.84

1 **Figure S15. Metabolomics analysis of the brains, serum and feces samples from HC or AD**
2 **humanized Abx-treated C/EBP β Tg mice and the brains from WT and C/EBP β Tg mice.**
3 (A-B) The differences in microbiomes related metabolites from feces samples (A) and brain
4 lysates (B). Red and green shaded cells indicate $p \leq 0.05$ (red indicates the fold change values are
5 significantly higher for that comparison; green values significantly lower). Light red and light
6 green shaded cells indicate $0.05 < p < 0.10$ (light red indicates the fold change values trend higher
7 for that comparison; light green values trend lower). Source data are provided as Supplementary
8 Data 1.



1 **Figure S16. Deletion of microglia attenuated PGE2-induced AD pathologies in Thy1-**
2 **C/EBP β Tg mice.**
3 3-month-old Thy1-C/EBP β Tg mice feed w/wo PLX3397 (600 ppm in chow, 7 days), followed by
4 bacteria/PGE2 treatment (Tg + B.F (Dead), Tg + B.F (Live), Tg + Vehicle, Tg + PGE2) for 8
5 weeks, n = 3/group. (A) Immunofluorescent staining of CD86 (green) and Iba-1 (red) in the
6 hippocampus region of the brain from *B. fragilis* or PGE2 treated C/EBP β Tg mice. Scale bar: 20
7 μ m. (B) Immunofluorescent staining of pT235 (green) and AT8 (red) in the hippocampus region
8 of the brain from *B. fragilis* or PGE2 treated C/EBP β Tg mice. Scale bar: 20 μ m. (C)
9 Immunofluorescent staining of pT235 (green) and 4G8 (red) in the hippocampus region of the
10 brain from *B. fragilis* or PGE2 treated C/EBP β Tg mice. Scale bar: 20 μ m.