

Supplementary Table 1 - These species were found on GTDB taxonomy and include species representatives that passed quality checks. Each of these species originates from a genus that was the top performing microbial feature in the one-vs-all tumour models provided by Poore *et al.* [1] and was considered the most stringent approach to decontamination.

GTDB Taxonomy	Isolation Source Information
Leucothrix sp002747455	swab sample of gingival sulcus (mouth) from 29 year old lactating female Dolphin_Z https://www.ncbi.nlm.nih.gov/biosample/SAMN07757601/
Leucothrix mucor	seaweed, Friday Harbor, WA https://www.ncbi.nlm.nih.gov/biosample/SAMN02440559/
Leucothrix pacifica	Seawater - Pacific Ocean: South Pacific Gyre https://www.ncbi.nlm.nih.gov/biosample/SAMN09228920/
Leucothrix arctica	Seawater - Svalbard: coast of Kongsfjorden, Arctic https://www.ncbi.nlm.nih.gov/biosample/SAMN09228919/
Thalassomonas actiniarum	Sea anemone https://www.ncbi.nlm.nih.gov/biosample/SAMN03278135/
Thalassomonas viridans	Cultivated oysters https://www.ncbi.nlm.nih.gov/biosample/SAMN03278134/
Ignicoccus_A sp013154085	deepsea hydrothermal sulfide chimney https://www.ncbi.nlm.nih.gov/biosample/SAMN12405658/
Ignicoccus_A sp015521275	marine hydrothermal vent biome https://www.ncbi.nlm.nih.gov/biosample/SAMN12837294/
Ignicoccus_A hospitalis	<i>Ignicoccus hospitalis</i> Kin4/I . <i>Ignicoccus hospitalis</i> Kin4/I was isolated from gravel obtained from the shallow marine hydrothermal system of the Kolbeinsey Ridge, north of Iceland and will be used for comparative analysis with other thermophiles https://www.ncbi.nlm.nih.gov/bioproject/13914
Ignicoccus islandicus	“The genome sequence of this strain will provide information on the adaptation to high temperature environments and be used for evolutionary and comparative genomic analysis.” https://www.ncbi.nlm.nih.gov/bioproject/PRJNA61347/
Salinimicrobium xinjiangense	Soil sediment of a salt lake https://www.ncbi.nlm.nih.gov/biosample/SAMN02440840/
Salinimicrobium terrae	Saline soil https://www.ncbi.nlm.nih.gov/biosample/SAMN02440676/
Salinimicrobium sp012038135	marine sediment https://www.ncbi.nlm.nih.gov/biosample/SAMN14480397/
Salinimicrobium marinum	Unstated https://www.ncbi.nlm.nih.gov/biosample/SAMD00245570/
Salinimicrobium catena	Unstated https://www.ncbi.nlm.nih.gov/biosample/SAMN04488140/
Salinimicrobium sediminis	Unstated https://www.ncbi.nlm.nih.gov/biosample/SAMN06296241/

Supplementary Table 2 – Viruses as top features in one-vs-all models. These were identified by searching NCBI at genus level and investigating each individual species. Each of these species originates from a genus that was the top performing feature in one of the one-vs-all tumour models presented by Poore *et al.* under the most stringent decontamination conditions.

<u><i>Areca palm velarivirus 1</i></u>	Complete genome sequence of a novel Velarivirus infecting areca palm in China https://www.ncbi.nlm.nih.gov/nuccore/NC_027121.1/
<u><i>Cordyline virus 1</i></u> (velarivirus)	An assemblage of closteroviruses infects Hawaiian ti (<i>Cordyline fruticosa</i> L.) https://www.ncbi.nlm.nih.gov/nuccore/NC_038421.1/
<u><i>Cordyline virus 2</i></u> (velarivirus)	Differentiation, distribution, and elimination of closteroviruses infecting <i>Cordyline fruticosa</i> (L.) in Hawaii https://www.ncbi.nlm.nih.gov/nuccore/NC_043453.1/
<u><i>Cordyline virus 3</i></u> (velarivirus)	Differentiation, distribution, and elimination of closteroviruses infecting <i>Cordyline fruticosa</i> (L.) in Hawaii https://www.ncbi.nlm.nih.gov/nuccore/NC_043107.1/
<u><i>Cordyline virus 4</i></u> (velarivirus)	Differentiation, distribution, and elimination of closteroviruses infecting <i>Cordyline fruticosa</i> (L.) in Hawaii https://www.ncbi.nlm.nih.gov/nuccore/NC_043108.1/
<u><i>Grapevine leafroll-associated virus 7</i></u> (velarivirus)	Molecular characterization and taxonomy of grapevine leafroll-associated virus 7 https://www.ncbi.nlm.nih.gov/nuccore/NC_016436.1/
<u><i>Little cherry virus 1</i></u> <u><i>Malus domestica virus A</i></u> (velarivirus)	Complete genome structure and phylogenetic analysis of little cherry virus, a mealybug-transmissible closterovirus https://www.ncbi.nlm.nih.gov/nuccore/NC_001836.1/
<u><i>Malus domestica virus A</i></u> (velarivirus)	Genomic characterization of a novel velarivirus <i>Malus domestica virus A</i> (MdoVA) infecting apple https://www.ncbi.nlm.nih.gov/nuccore/NC_055599.1/
<u><i>Brome streak mosaic virus</i></u> (<i>Tritimovirus</i>)	The complete nucleotide sequence and genome organization of the mite-transmitted brome streak mosaic rymovirus in comparison with those of potyviruses https://www.ncbi.nlm.nih.gov/nuccore/NC_003501.1/
<u><i>Oat necrotic mottle virus</i></u> (<i>Tritimovirus</i>)	Complete nucleotide sequence of Oat necrotic mottle virus: a distinct Tritimovirus species (family Potyviridae) most closely related to Wheat streak mosaic virus https://www.ncbi.nlm.nih.gov/nuccore/NC_005136.1/
<u><i>Tall oatgrass mosaic virus</i></u> (<i>Tritimovirus</i>)	Tall oatgrass mosaic virus (TOgMV): a novel member of the genus Tritimovirus infecting <i>Arrhenatherum elatius</i> https://www.ncbi.nlm.nih.gov/nuccore/NC_022745.1/
<u><i>Wheat eglid mosaic virus</i></u> (<i>Tritimovirus</i>)	Analyses of the complete sequence of the genome of wheat Eglid mosaic virus, a novel species in the genus Tritimovirus https://www.ncbi.nlm.nih.gov/nuccore/NC_009805.1/
<u><i>Wheat streak mosaic virus</i></u> (<i>Tritimovirus</i>)	Phylogenetic relationships within the family potyviridae: wheat streak mosaic virus and brome streak mosaic virus are not members of the genus rymovirus https://www.ncbi.nlm.nih.gov/nuccore/NC_001886.1/
<u><i>Yellow oat grass mosaic virus</i></u> (<i>Tritimovirus</i>)	Genome sequence of two isolates of Yellow oatgrass mosaic virus, a new grass-infecting Tritimovirus https://www.ncbi.nlm.nih.gov/nuccore/NC_024471.1/
<u><i>Aedes pseudoscutellaris reovirus</i></u> (Dinovernavirus)	isolated from <i>Aedes pseudoscutellaris</i> mosquito cells https://www.ncbi.nlm.nih.gov/nuccore/NC_007666.1/ 10.1016/j.virol.2005.08.028

<u><i>Chaetoceros socialis forma radians RNA virus 1</i></u> (Bacillarnavirus)	Isolation and characterization of a single-stranded RNA virus infecting the bloom-forming diatom <i>Chaetoceros socialis</i> https://www.ncbi.nlm.nih.gov/nucleotide/NC_012212.1/
<u><i>Chaetoceros tenuissimus RNA virus O1</i></u> (Bacillarnavirus)	Isolation and characterization of a single-stranded RNA virus infecting the marine planktonic diatom <i>Chaetoceros tenuissimus</i> Meunier https://www.ncbi.nlm.nih.gov/nucleotide/NC_038321.1/
<u><i>Rhizosolenia setigera RNA virus O1</i></u> (Bacillarnavirus)	Complete nucleotide sequence of a single-stranded RNA virus infecting the bloom-forming diatom <i>Rhizosolenia setigera</i> https://www.ncbi.nlm.nih.gov/nucleotide/NC_018613.1/
<u><i>Agropyron mosaic virus</i></u> (Rymovirus)	Functional replacement of Wheat streak mosaic virus HC-Pro with the corresponding cistron from a diverse array of viruses in the family https://www.ncbi.nlm.nih.gov/nucleotide/NC_005903.1/
<u><i>Hordeum mosaic virus</i></u> (Rymovirus)	Functional replacement of Wheat streak mosaic virus HC-Pro with the corresponding cistron from a diverse array of viruses in the family Potyviridae https://www.ncbi.nlm.nih.gov/nucleotide/NC_005904.1/
<u><i>Ryegrass mosaic virus</i></u> (Rymovirus)	The complete nucleotide sequence of the Ryegrass mosaic virus https://www.ncbi.nlm.nih.gov/nucleotide/NC_001814.1/

Supplementary Table 3 – The proportion (and total number) of all primary tumour samples across sequencing centers as provided by Poore et al. in file: Metadata-TCGA-All-18116-Samples.csv. Any cells where at least 90% of the samples originate from one sequencing center are highlighted in red.

Primary Tumour Type	Sequencing Center								
	Baylor College of Medicine	Broad Institute of MIT and Harvard	Canada's Michael Smith Genome Sciences Centre	Harvard Medical School	MD Anderson - Institute for Applied Cancer Science	MD Anderson - RPPA Core Facility (Proteomics)	University of North Carolina	Washington University School of Medicine	
Adrenocortical Carcinoma	0	0	0	0	0	0	1 (76)	0	0
Bladder Urothelial Carcinoma	0	0.05 (27)	0	0.09 (50)	0.11 (60)	0	0.75 (406)	0	0
Brain Lower Grade Glioma	0	0.05 (30)	0	0.09 (51)	0	0	0.86 (505)	0	0
Breast Invasive Carcinoma	0	0	0	0.02 (18)	0 (1)	0	0.92 (1046)	0.07 (78)	0
Cervical and Endocervical	0	0	0	0.02 (6)	0.11 (41)	0	0.82 (293)	0.05 (18)	0

Cholangiocarcinoma	0	0	0	0	0	0	1 (36)	0
Colon Adenocarcinoma	0.05 (40)	0	0	0.11 (86)	0	0	0.84 (651)	0
Esophageal Carcinoma	0	0	0.69 (147)	0	0.24 (50)	0	0	0.07 (15)
Glioblastoma Multiforme	0	0.95 (304)	0	0	0	0	0	0.05 (17)
Head and Neck Squamous Cell Carcinoma	0.04 (25)	0.04 (25)	0	0.12 (81)	0.03 (21)	0	0.77 (512)	0
Kidney Chromophobe	0.32 (30)	0	0	0	0	0	0.68 (65)	0
Kidney Renal Clear Cell Carcinoma	0.03 (27)	0 (3)	0	0	0	0	0.97 (929)	0
Kidney Renal Papillary Cell Carcinoma	0.09 (26)	0	0	0	0	0	0.91 (277)	0
Liver Hepatocellular Carcinoma	0.1 (39)	0	0	0	0	0	0.9 (357)	0
Lung Adenocarcinoma	0	0.04 (23)	0	0.17 (108)	0	0	0.79 (503)	0
Lung Squamous Cell Carcinoma	0	0.06 (29)	0	0	0	0	0.94 (480)	0
Diffuse Large B-cell Lymphoma	0.1 (5)	0	0	0	0	0	0.9 (45)	0
Mesothelioma	0	0	0	0	0	0	1 (82)	0
Ovarian Serous Cystadenocarcinoma	0.04 (33)	0.02 (14)	0.93 (712)	0	0	0	0	0.01 (9)
Pancreatic Adenocarcinoma	0	0	0	0	0	0	1 (172)	0
Pheochromocytoma and Paranglioma	0	0	0	0	0	0	1 (174)	0
Prostate Adenocarcinoma	0	0.02 (14)	0	0.18 (108)	0	0	0.8 (493)	0
Rectum Adenocarcinoma	0.06 (18)	0	0	0.14 (42)	0	0	0.79 (231)	0

Sarcoma	0	0	0	0	0	0	0.89 (250)	0.11 (30)
Skin Cutaneous Melanoma	0	0.01 (1)	0	0.12 (14)	0	0	0.87 (102)	0
Stomach Adenocarcinoma	0	0.04 (28)	0.8 (573)	0.15 (110)	0.01 (5)	0	0	0
Testicular Germ Cell Tumors	0	0	0	0	0	0	1 (133)	0
Thymoma	0	0	0	0	0	0	1 (119)	0
Thyroid Carcinoma	0	0.05 (28)	0	0.11 (71)	0.04 (23)	0	0.8 (496)	0
Uterine Carcinosarcoma	0	0	0	0	0	0	1 (57)	0
Uterine Corpus Endometrial Carcinoma	0	0	0	0.12 (106)	0	0	0.83 (723)	0.04 (39)
Uveal Melanoma	0	0	0	0	0.4 (51)	0	0.6 (76)	0

[1] Poore GD, Kopylova E, Zhu Q, Carpenter C, Fraraccio S, Wandro S, et al. Microbiome analyses of blood and tissues suggest cancer diagnostic approach. Nature. 2020;579:567-74.