

Supplementary

Structures of CTCF-DNA complexes including all eleven zinc fingers

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Figures S1- S5

Tables S1-S2

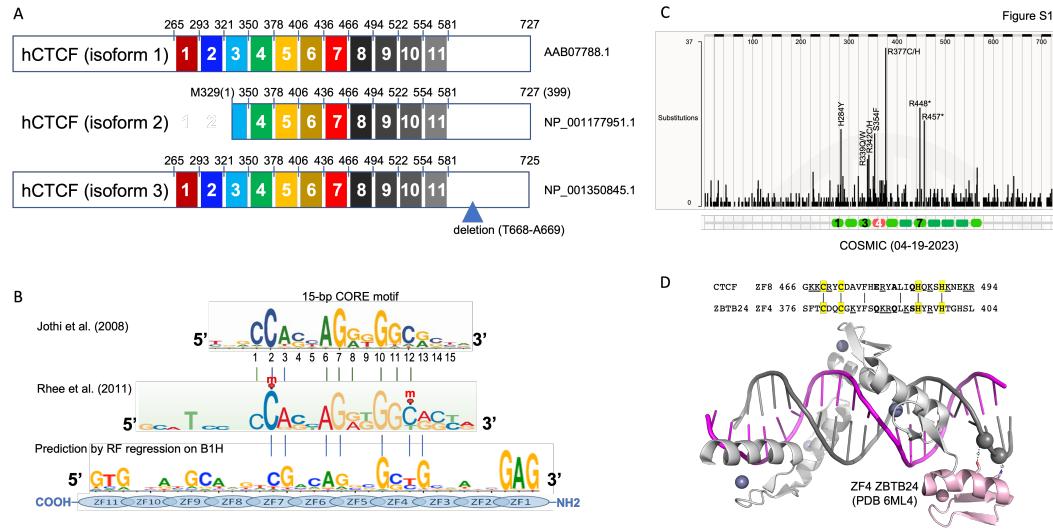


Figure S1. (A) Humans have three isoforms of CTCF, one of which is missing ZFs 1, 2, and half of 3. **(B)** Two examples of CTCF-binding consensus CORE sequence (base pairs 1-15) as determined by ChIP-seq (1) and ChIP-exo (2). DNA cytosine methylation (indicated by red circles and letter m) occurs at positions 2 and 12 of the consensus sequence in a subset of CTCF-binding sites (3). The experimentally determined consensus is aligned with a predicted CTCF-DNA-binding specificity of ZF3-7. **(C)** CTCF mutations found in the Catalogue of Somatic Mutations in Cancer (COSMIC). The highest rate of mutations is C>T substitutions at codons of H284, R339, R342, S354, R377, R448 and R457. H284 of ZF1 is a zinc ligand. R339 of ZF3 is a base-interacting residue (see Figure 2F). R342 of ZF3 interacts with a DNA phosphate group. Ser354 is next to zinc-ligand Cys353 of ZF4. R377 of ZF4 interacts with a DNA phosphate group. R448* and R457* result in two deletions which eliminate translation of ZF7 and beyond. **(D)** Comparison between CTCF ZF8 and ZBTB24 ZF4 which holds two large and changed residues at positions -6 and -5. A spacer of ZF4 of ZBTB24 across DNA major groove.

Figure S2. CTCF is highly conserved among vertebrates, especially in the ZF DNA binding Domain. Human CTCFL is shown for comparison. The green dots (●) indicate the two sites of sumoylation (4) – both lysine residues are highly conserved among CTCF orthologs, but both are missing in CTCFL. The maroon white dots (◐) indicate sites of poly-ADP-ribosylation (5). The magenta dots (●) indicate sites of phosphorylation during mitosis that reduce DNA binding (6). The insertions in *Danio rerio* CTCF are indicated by cyan triangles.

Numbering above the sequence indicates residues receiving particular attention in the text, and refer to the human ortholog. Letters in **bold red** are positions making base-specific contacts that are substituted in cases of human CTCF-related disorder (7); all are fully-conserved, even in CFCTL (shown for comparison). Grey shading is where ≥ 5 of the six sequences are identical. The ZFs are shaded yellow, with substitutions in cyan, with Zn-coordinating residues in bold. The highly-conserved Met in white on black (M) in ZF3 indicates the initiation codon for CTCF isoform 2 (NP_001177951.1), which in humans has a somewhat broader sequence specificity and competes with canonical CTCF, disrupts CTCF/cohesin binding, alters CTCF-mediated chromatin looping and promotes IFI6 activation and apoptosis (8). The white on black Thr-Ala (T-A) indicates two residues missing in CTCF isoform 3 (NP_001350845.1).

While all 11 CTCF ZFs are highly conserved among vertebrates, the number of substitutions is highest in ZFs 9-11. In particular, the whale shark *Rhincodon typus* has 12 substitutions relative to the human ortholog in ZFs 9-11, twice as many as the runner-up (*Danio rerio*). Three-quarters of the *R. typus* substitutions are fully conserved among other members of the class Chondrichthyes (not shown).

Figure. S2. CTCF is highly conserved among vertebrates, especially in the DBD. Human CTCFL is shown for comparison.

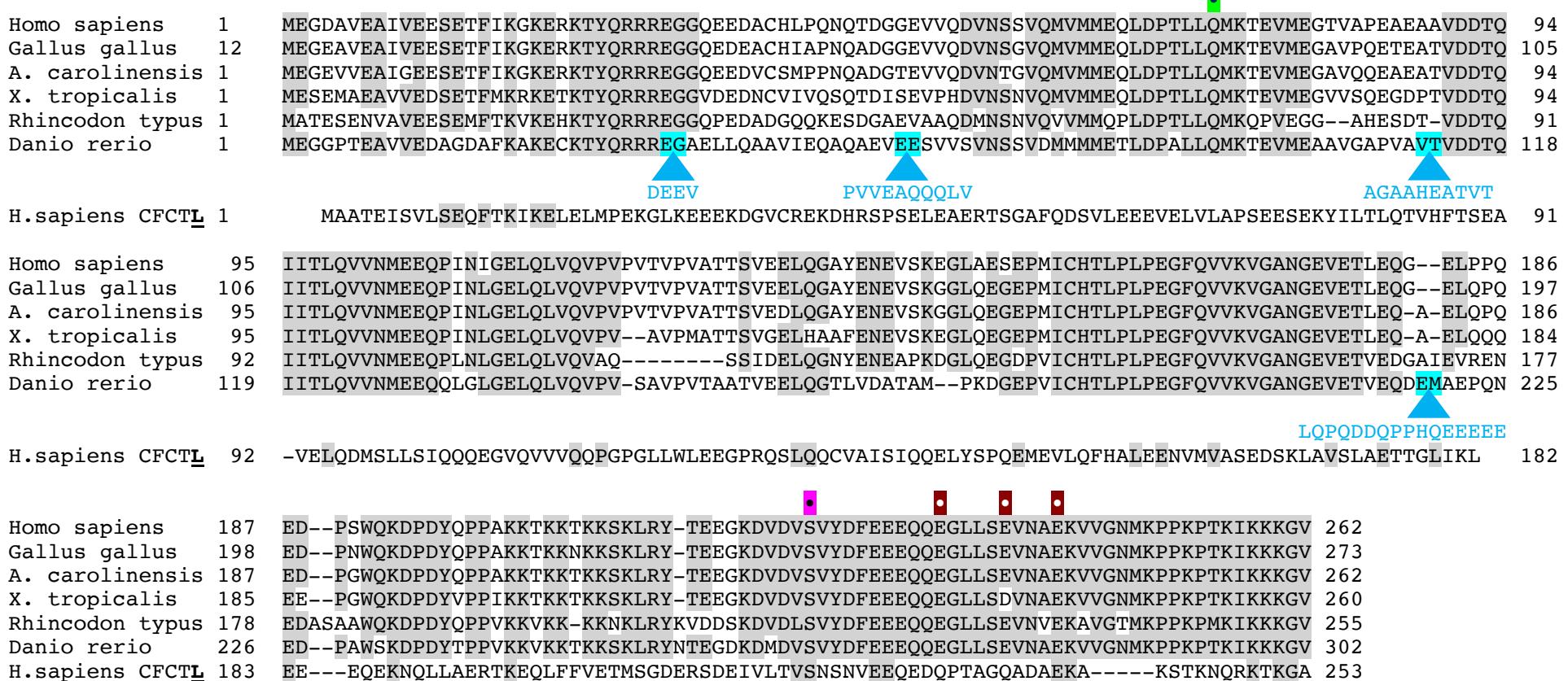
Description		Query cover	E value	Per. Ident	Len	Acc.	Accession
transcriptional repressor CTCF isoform X1 [Homo sapiens]		100%	0.0	100.00	727	AAB07788.1	Mammalia
transcriptional repressor CTCF isoform X1 [Gallus gallus]		100%	0.0	93.55	739	XP_015134605.1	Aves
PREDICTED: transcriptional repressor CTCF [Anolis carolinensis]		100%	0.0	92.08	731	XP_003225381.1	Reptilia
transcriptional repressor CTCF [Xenopus tropicalis]		100%	0.0	85.93	734	NP_001116268.1	Amphibia
transcriptional repressor CTCF [Rhincodon typus]		100%	0.0	76.55	720	XP_048462054.1	Chondrichthyes
transcriptional repressor CTCF [Danio rerio]		100%	0.0	71.94	798	NP_001001844.1	Osteichthyes
transcriptional repressor CTCFL isoform 1 [Homo sapiens]		85%	2e-167	47.51	663	NP_001373922.1	

The green dots (●) indicate the two sites of sumoylation – both lysines are highly conserved among CTCF orthologs, but both are missing in CTCFL.

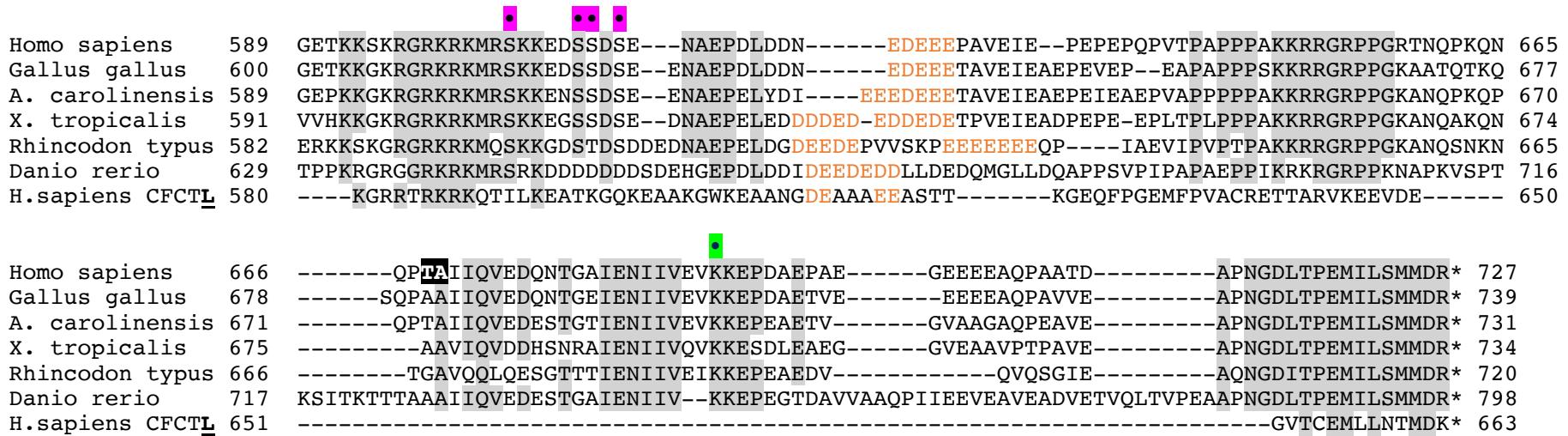
The maroon white dots (●) indicate sites of poly-ADP-ribosylation.

The magenta dots (●) indicate sites of phosphorylation during mitosis.

The insertions in Danio rerio CTCF are indicated by cyan triangles.



		283	284														
		ZF1		•		ZF2		•		ZF3		339	342				
Homo sapiens	263	KKT	QCELCSY	TCPRRSNLDR	HMKSH	TDERPHK	CHLCGRAF	RTVTLLRNH	LNTHTGTRP	HKCPDCDMAF	VTS	GELVR	HRRYKHT	347			
Gallus gallus	274	KKT	QCELCSY	TCPRRSNLDR	HMKSH	TDERPHK	CHLCGRAF	RTVTLLRNH	LNTHTGTRP	HKCPDCDMAF	VTS	GELVR	HRRYKHT	358			
A. carolinensis	263	KKT	QCELCSY	TCPRRSNLDR	HMKSH	TDERPHK	CHLCGRAF	RTVTLLRNH	LNTHTGTRP	HKCPDCDMAF	VTS	GELVR	HRRYKHT	347			
X. tropicalis	261	KKT	QCELCSY	TCPRRSNLDR	HMKSH	TDERPHK	CHLCGRAF	RTVTLLRNH	LNTHTGTRP	HKCPDCDMAF	VTS	GELVR	HRRYKHT	345			
Rhincodon typus	256	KKT	QCELCSY	TCPRRSNLDR	HMKSH	TDERPHK	CHLCGRAF	RTVTLLRNH	LNTHTGTRP	HKCPDCDMAF	VTS	GELVR	HRRYKHT	340			
Danio rerio	303	KKT	QCELCSY	TCPRRSNLDR	HMKSH	TDERPHK	CHLCGRAF	RTVTLLRNH	LNTHTGTRP	HKCPDCDMAF	VTS	GELVR	HRRYKHT	387			
H.sapiens CFCTL	254	KGT	FHCDVCMFTSSRMRM	SFNRMHMKSH	T	SEKPH	CHLC	LKT	RTVTLLRNH	VNTHTGTRP	YKC	DCNMAF	VTS	GELVR	HRRYKHT		
															338		
		354	362	365		377			392								
		ZF4			•				ZF5		•						
Homo sapiens	348	EKP	FKCSMCDYASVEVSKLKR	HIRSHTGERP	FQCSLCSYASRD	TYKLKR	HMRTH	SGEKP	YECYICHARFT	QSGTMKMHILQKHTEN						433	
Gallus gallus	359	EKP	FKCSMCDYASVEVSKLKR	HIRSHTGERP	FQCSLCSYASRD	TYKLKR	HMRTH	SGEKP	YECYICHARFT	QSGTMKMHILQKHTEN						444	
A. carolinensis	348	EKP	FKCSMCDYASVEVSKLKR	HIRSHTGERP	FQCSLCSYASRD	TYKLKR	HMRTH	SGEKP	YECYICHARFT	QSGTMKMHILQKHTEN						433	
X. tropicalis	346	EKP	FKCSMCDYASVEVSKLKR	HIRSHTGERP	FQCSLCSYASRD	TYKLKR	HMRTH	SGEKP	YECYICHARFT	QSGTMKMHILQKHTEN						431	
Rhincodon typus	341	EKP	FKCSMCDYASVEVSKLKR	HIRSHTGERP	FQCSLCSYASRD	TYKLKR	HMRTH	SGEKP	YECYICHARFT	QSGTMKMHILQKHTEN						426	
Danio rerio	388	EKP	FKCSMCDYASVEVSKLKR	HIRSHTGERP	FQCSLCSYASRD	TYKLKR	HMRTH	SGEKP	YECYICHARFT	QSGTMKMHILQKHTEN						473	
H.sapiens CFCTL	339	EKP	FKCSMCKYASVEASKLKR	HVRSH	TGERP	FQCQCQCSYASRD	TYKLKR	HMRTH	SGEKP	YECYICHARFT	QSGTMKMHILQKHTEN						424
		448	450	454.		•			467	470							
		ZF7				•			ZF8								
Homo sapiens	434	VAK	FHCPHCDTVIARKSDLGV	HLRKQHSYIEQGKK	CRYCDAVFH	ERYALIOHQKSH	KNEKRF	KCDCDYACRQER	HFMIMHKRT	HGT						519	
Gallus gallus	445	VAK	FHCPHCDTVIARKSDLGV	HLRKQHSYIEQGKK	CRYCDAVFH	ERYALIOHQKSH	KNEKRF	KCDCDYACRQER	HFMIMHKRT	HGT						530	
A. carolinensis	434	VAK	FHCPHCDTVIARKSDLGV	HLRKQHSYIEQGKK	CRYCDAVFH	ERYALIOHQKSH	KNEKRF	KCDCDYACRQER	HFMIMHKRT	HGT						519	
X. tropicalis	432	VAK	FHCPHCDTVIARKSDLGV	HLRKQHSYIEQGKK	CRYCDT	VFH	ERYALIOHQKSH	KNEKRF	KCDCDYACRQER	HFMIMHKRT	HGT					517	
Rhincodon typus	427	VAK	FHCPHCDTVIARKSDLGV	HLRKQHSYIEQGKK	CRYCDAVFH	ERYALIOHQKSH	KNEKRF	KCDCDYACRQER	HFMIMHKRT	HGT						512	
Danio rerio	474	VAK	FHCPHCDTVIARKSDLGV	HLRKQHSYIEQGKK	CRYCDAVFH	ERYALIOHQKSH	KNEKRF	KCDCDYACRQER	HFMIMHKRT	HGT						559	
H.sapiens CFCTL	425	VPK	YOCPHCATIIARKSDLRV	HVMRNL	HAYSAEL	KCRYC	SAV	FH	ERYALIOHQKTH	KNEKRF	KCHCS	YACK	QERHM	TAHIRT	HGT	510	
		536							566								
		ZF10							ZF11								
Homo sapiens	520	EKP	YACSHCDKTFRQKQLLD	MHF	KRYHD	DPNFVPAAF	VCSKGKTF	RTN	TMAR	HADNCAGPDG	VEGENG						588
Gallus gallus	531	EKP	YACSHCDKTFRQKQLLD	MHF	KRYHD	DPNFVPAAF	VCSKGKTF	RTN	TMAR	HADNC	SGLDGG	EGENG				599	
A. carolinensis	520	EKP	YACSHCDKTFRQKQLLD	MHF	KRYHD	DPNFVPAAF	VCSKGKTF	RTN	TMAR	HADNC	SGLDGG	EGENG				588	
X. tropicalis	518	EKP	YACSHCDKTFRQKQLLD	MHF	KRYHD	PSFVPAAF	VCSKGKTF	RTN	TMAR	HADNC	TGP	DG	VEGENGESE		590		
Rhincodon typus	513	EKP	YACSQCDKTFRQKQLLD	MHF	KRYHD	PSFVPAIFAT	VCTKCGKA	FT	TRN	TMAR	HADNC	TGP	DG	TEGEE	DAV	581	
Danio rerio	560	EKP	YACSQCEKTFRQKQLLD	MHF	R	PNFVPTSFV	C	CGKTF	TRN	TMAR	HADNC	TGMDSAD	GENG			628	
H.sapiens CFCTL	511	EKP	FTCLSCNKFRQKQLLNA	HFRKYHD	ANFIPTV	YKCS	CSKGKGF	SRWINL	HLRHSEK	C	GS	GEAK	SAASG			579	



Numbering above the sequence indicates residues receiving particular attention in the text, and refer to the human ortholog.

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Grey shading is where ≥5 of the six sequences are identical.

The ZFs are shaded yellow, with substitutions in cyan, with Zn-coordinating residues in bold.

The highly-conserved Met in white on black (**M**) in ZF3 indicates the initiation codon for CTCF isoform 2 (NP_001177951.1)

The white on black Thr-Ala (**T-A**) indicates two residues missing in CTCF isoform 3 (NP_001350845.1).

Figure S3

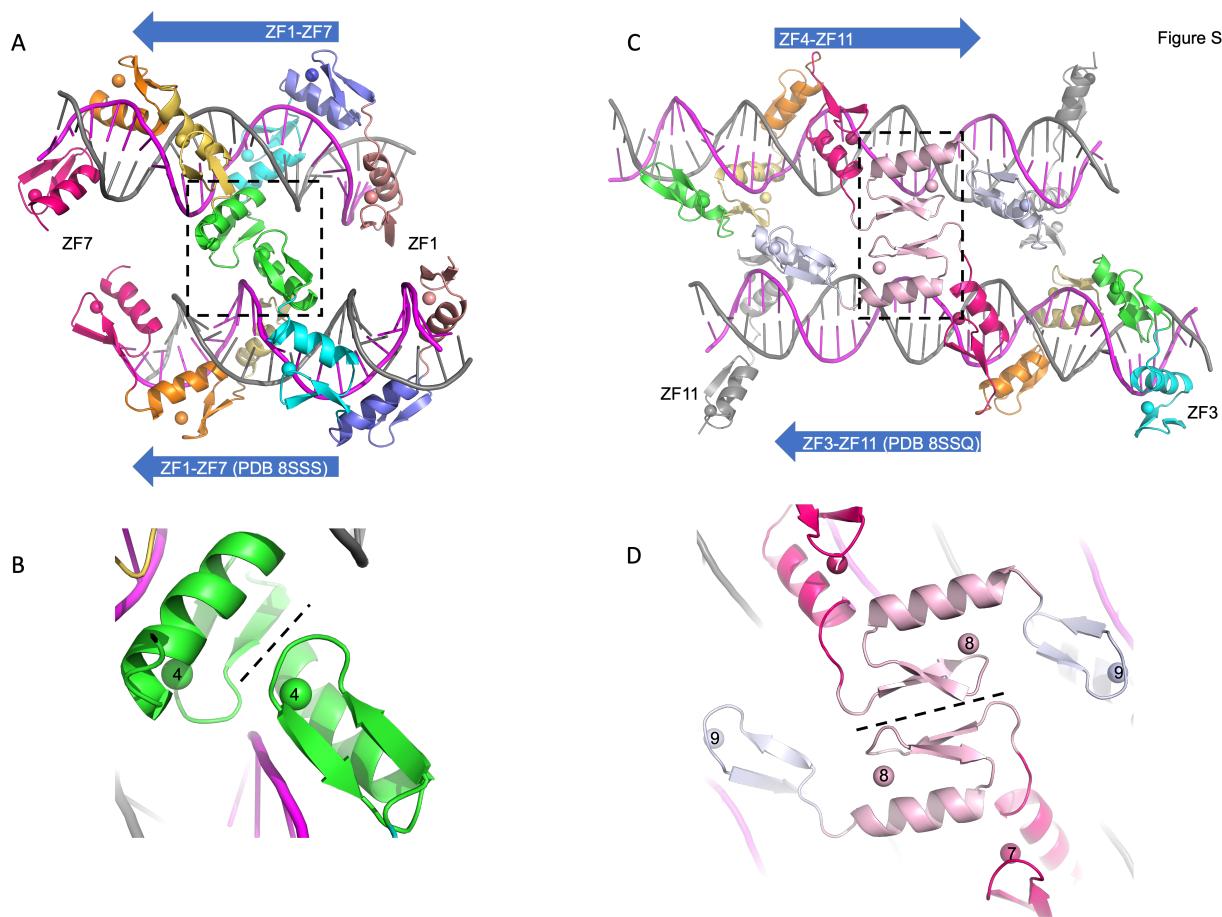


Figure S3. **(A)** Crystal of ZF1-7 in complex with DNA, with two protein-DNA complexes per crystallographic asymmetric unit. The two complexes are arranged in parallel. **(B)** The interface of two molecule is mainly mediated by ZF4 involving the hair loop between the antiparallel strands. **(C)** Crystal of ZF3-11 in complex with DNA, with two protein-DNA complexes per crystallographic asymmetric unit. One of the complexes has a missing ZF3 (top). The two complexes are arranged in antiparallel. **(D)** The interface of the two complexes is mainly mediated by hairpin β -strands of ZF8 (related to Figure 8A).

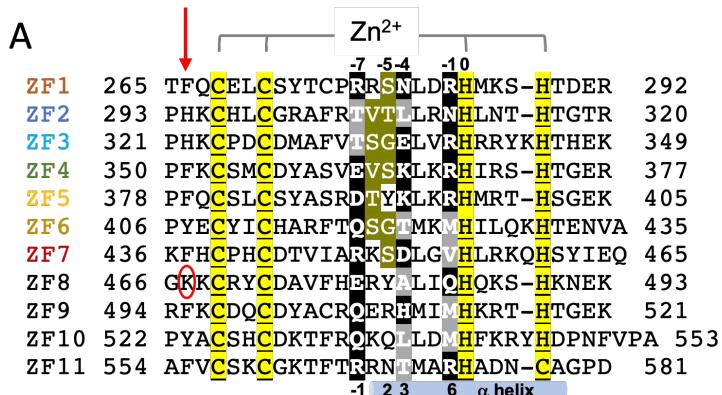


Figure S4

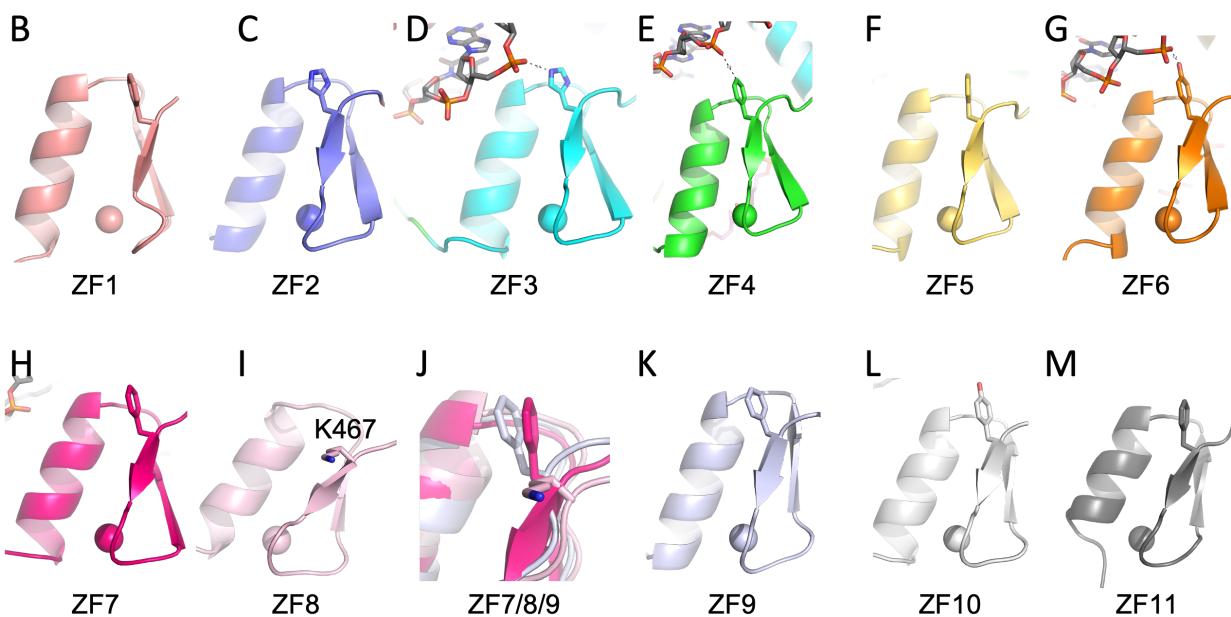


Figure S4. (A) In the corresponding position to Lys467 of ZF8 (indicated by a red arrow), all the other ten fingers in CTCF have a Phe, Tyr or His as the first residue of the first β strand. (B-H and K-M) This side chain packs between the β strand and the helix and provides stability for the finger. (D-E and G) In three fingers (ZF3, ZF4 and ZF6), when the finger gets deep into the DNA major groove, the corresponding His, Phe or Tyr provides an H-bond or van der Waals contact to the phosphate group of the non-recognition strand. (I) Lys467 of ZF8 points to DNA. (J) Superimposition of three figures, ZF7, ZF8 and ZF9.

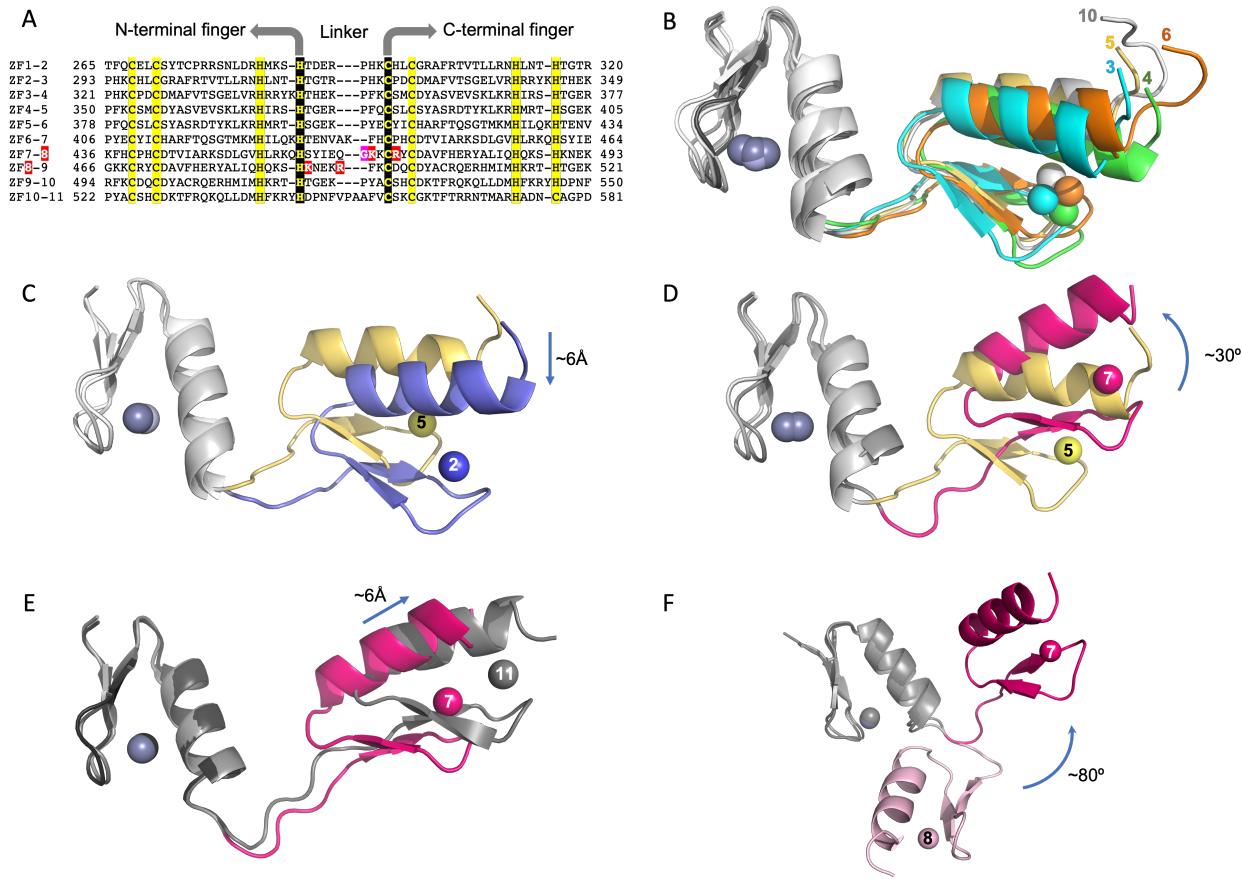


Figure S5. Effect of inter-finger linker length on the relative conformation of associated fingers. (A) we examined the effect of inter-finger linker on the relative positions of two neighboring fingers. We generated ten pairs of two-finger element. Seven pairs have the inter-finger length of seven residues between the last Zn-ligand His of the N-terminal finger and the first Zn-ligand Cys of the C-terminal finger (ZF1-2, ZF2-3, ZF3-4, ZF4-5, ZF5-6, ZF8-9 and ZF9-10), two pairs have the length of eight residues (ZF6-7 and ZF7-8), and one pair has ten residues between ZF10 and ZF11. (B) We superimposed the corresponding N-terminal fingers and resulting C-terminal fingers adopted at least four varied conformations. First, five pairs of seven-residue linker have similar conformations of the linker as well as the C-terminal finger, including ZF2-3, ZF3-4, ZF4-5, ZF5-6 and ZF9-10. Both fingers of these five pairs are involved in intimate DNA base contacts, as discussed in text (Figures 2 and 7). (C) Using ZF4-5 as a

reference, superimposition of ZF1-2 onto ZF4-5 resulted in a movement of ZF2 as well as the linker, away from the DNA base interface. Though this movement (~6Å) was sufficient to distance the base-interacting residues but till placed ZF2 in the DNA major groove. (D) Superimposing ZF6-7, which has eight-residue linker, onto ZF4-5, seen the rotation of ZF7 helix of ~30° in reference to ZF5 helix. (E) Further increasing the linker distance to ten residues between ZF10 and ZF11 placed ZF11 additional ~6Å away, which might correlate to the observation that the base-interacting residues at -1 and -4 positions of ZF11 are farthest away from their corresponding DNA bases. (F) Superimposition of ZF7-8 and ZF6-7, where both pairs include an eight-residue inter-finger linker, positioned ZF7 and ZF8 in opposite directions. The two conformations are approximately ~80° of rotation apart. Apparently, the inter-finger length alone is not the primary reason that that ZF8 spans the minor groove. As noted in the text, the linkers prior to and after ZF8 harbor unique basic residues (white letters against red background in panel A) which form interactions with DNA phosphate groups.

References

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Table S1. Summary of X-ray data collection and refinement statistics, beamline 22-ID (APS) and 1.0000 Å wavelength

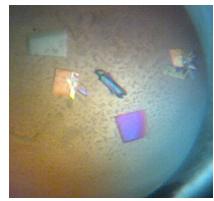
Protein Construct	ZF3-ZF11	ZF3-ZF11	ZF1-ZF7	ZF1-ZF7 (K365T)	ZF3-ZF11
DNA (see Table S2)	35-4	35-20	23-bp	23-bp	19-bp
PDB code	8SSQ	8SSR	8SSS	8SST	8SSU
Date Collected	07-17-20, 08-07-20, 09-26-20	07/10/22, 11/7/2022	09-26-2020	03-24-2021	02/06/2020
Space group	C2	C2	C2	C2	P3 ₂ 1
Cell dimensions (Å)	352.1, 67.8, 61.2	362.2, 68.1, 62.0	161.0, 41.2, 135.7	160.6, 41.6, 135.2	80.5, 80.5, 187.6
α, β, γ (°)	90, 92.6, 90	90, 94.9, 90	90, 105.2, 90	90, 105.6, 90	90, 90, 120
Resolution (Å)	41.86-3.12 (3.23-3.12)	37.59-3.14 (3.26-3.14)	43.18-2.30 (2.37-2.30)	40.17-2.19 (2.27-2.19)	38.34-2.89 (2.99-2.89)
^a R _{merge}	0.398 (2.173)	0.254 (1.068)	0.090 (0.989)	0.121 (1.315)	0.203 (2.56)
R _{pim}	0.076 (0.703)	0.061 (0.338)	0.038 (0.649)	0.053 (0.753)	0.065 (0.860)
CC _{1/2}	0.985 (0.397)	0.940 (0.883)	1.000 (0.513)	0.992 (0.387)	0.974 (0.706)
^b <I/σI>	11.0 (1.2)	10.9 (1.1)	17.0 (1.1)	13.3 (1.0)	12.8 (1.5)
Completeness (%)	100.0 (99.8)	95.7 (73.6)	94.0 (74.4)	97.2 (87.4)	99.9 (100.0)
Redundancy	27.1 (11.5)	15.8 (7.1)	6.0 (2.5)	5.8 (3.2)	10.3 (9.7)
Observed reflections	701,822	391,247	220,378	250,177	168,662
Unique reflections	25,877 (2,583)	24,779 (1,894)	36,617 (2,757)	43,468 (3,842)	16,333 (1,605)
Refinement					
Resolution (Å)	3.12	3.14	2.30	2.19	2.89
No. reflections	25,644	24,639	35,515	43,313	16,037
^c R _{work} / ^d R _{free}	0.238 / 0.268	0.253 / 0.285	0.205 / 0.231	0.220 / 0.243	0.205 / 0.240
No. Atoms					
Protein	4051	4051	3150	3140	1,888
DNA	2856	2856	1874	1874	802
Zn	17	17	14	14	8
Solvent	15	2	111	121	14
B Factors (Å ²)					
Protein	177.9	190.6	68.7	85.9	97.0
DNA	187.5	195.1	70.5	88.7	83.1
Zn	185.7	238.5	69.4	86.7	102.2
Solvent	100.7	144.4	49.2	54.8	73.2
R.m.s. deviations					
Bond lengths (Å)	0.003	0.003	0.003	0.003	0.003
Bond angles (°)	0.5	0.5	0.5	0.5	0.5

* Values in parenthesis correspond to highest resolution shell.

^a R_{merge} = $\sum |I - \langle I \rangle| / \sum I$, where I is the observed intensity and $\langle I \rangle$ is the averaged intensity from multiple observations.^b <I/σI> = averaged ratio of the intensity (I) to the error of the intensity (σI).^c R_{work} = $\sum |F_{\text{obs}} - F_{\text{cal}}| / \sum |F_{\text{obs}}|$, where F_{obs} and F_{cal} are the observed and calculated structure factors, respectively.^d R_{free} was calculated using a randomly chosen subset (5%) of the reflections not used in refinement.

Table S2. Oligonucleotides used in the study

Residues	name	pXC#	DNA oligos for crystallization	Crystallization conditions
263-465	ZF1-ZF7	1564	23-bp: 5'-G CCA GCA GGG GGC GCT AGT GAG G-3' 3'-C GGT CGT CCC CCG CGA TCA CTC C-5'	260 mM DL-Malic acid 7.0 26.5% PEG 3350
263-465	K365T	2332	23-bp: 5'-G CCA GCA GGG GGC GCT AGT GAG G-3' 3'-C GGT CGT CCC CCG CGA TCA CTC C-5'	200 mM ammonium acetate 100 mM BIS-TRIS pH 5.5 25% (w/v) PEG 3350
321-581	ZF3-ZF11	1566	35-4: 5'-GTG CAG TACC ACATTTAA CCA GCA GGG GGC GCT AA-3' 3'-CAC GTC ATGG TGTAAATT GGT CGT CCC CCG CGA TT-5' 35-20: 5'-GTG CAG TACC ACATTTAA CCA GCA GGT GGC GCT AA-3' 3'-CAC GAC TTGG TGTAAATT GGC CGT CCA CCG CGA TT-5' 19-bp: 5'-TGC GCC CCC TGC TGG TCC T-3' 3'-ACG CGG GGG ACG ACC AGG A-5'	200 mM sodium malonate pH 5.0 20% (w/v) PEG 3350 20% (w/v) PEG 3350 100 mM BIS-TRIS pH 5.0 50 mM NaCl 40 mM citric acid, 60 mM BIS-TRIS propane pH 6.4 20% (w/v) PEG 3350
			Primers for mutagenesis:	
	K365T		FP: 5'-GAA GTC AGC ACA TTA AAA CGT CAC ATT CG-3' RP: 5'-CGT TTT AAT GTG CTG ACT TCT ACA CTG GC-3'	
			For FP binding:	
			5'-A GGA CCA GCA GGG GXC GCA-3' 3'-T CCT GGT CGT CCC CYG CGT-5'-FAM X:Y=G:C, A:T, T:A, C:G	



ZF3-ZF11 + oligo 35-4



ZF3-ZF11 + 19 bp