

Supplemental information

**HIV-1 neutralizing antibodies elicited in humans
by a prefusion-stabilized envelope trimer form
a reproducible class targeting fusion peptide**

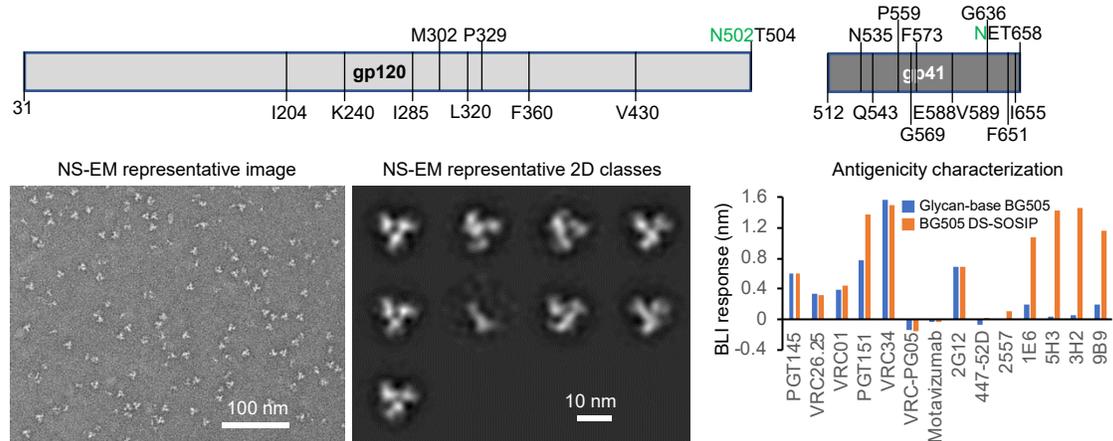
Shuishu Wang, Flavio Matassoli, Baoshan Zhang, Tracy Liu, Chen-Hsiang Shen, Tatsiana Bylund, Timothy Johnston, Amy R. Henry, I-Ting Teng, Prabhanshu Tripathi, Jordan E. Becker, Anita Changela, Ridhi Chaudhary, Cheng Cheng, Martin Gaudinski, Jason Gorman, Darcy R. Harris, Myungjin Lee, Nicholas C. Morano, Laura Novik, Sijy O'Dell, Adam S. Olin, Danealle K. Parchment, Reda Rawi, Jesmine Roberts-Torres, Tyler Stephens, Yaroslav Tsybovsky, Danyi Wang, David J. Van Wazer, Tongqing Zhou, Nicole A. Doria-Rose, Richard A. Koup, Lawrence Shapiro, Daniel C. Douek, Adrian B. McDermott, and Peter D. Kwong

A Env trimers used for B cell sorting and antibody-binding analyses

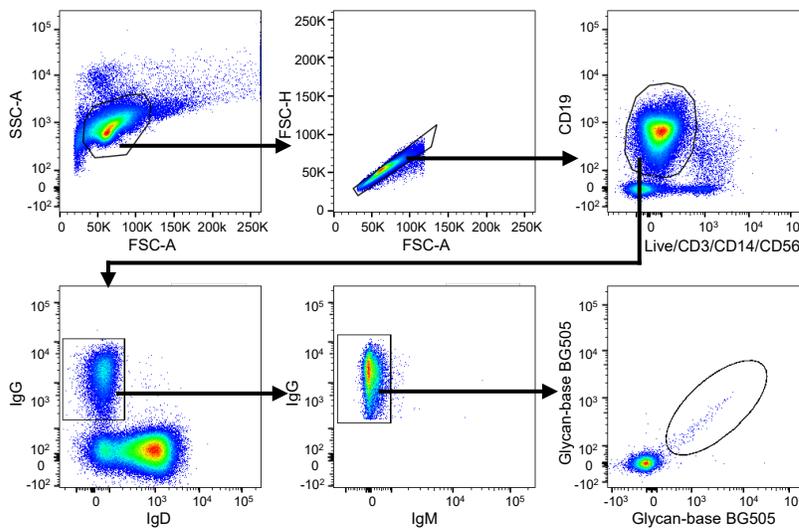


B Glycan-base BG505 design and characterization

Glycan-base BG505 sequence - relative to BG505 DS-SOSIP, additional stabilizing mutations and two *N*-glycans at trimer base



C Sorting gates



D Staining of IgG+ cells

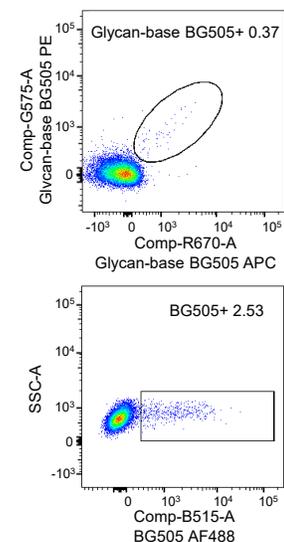


Figure S1. Env trimers and B cell sorting details, related to Figure 1.

(A) Env trimers used for B cell sorting and binding analyses with AlphaLISA, BLI, and SPR.

(B) The probe glycan-base BG505 was designed based on BG505 DS-SOSIP with additional mutations to stabilize the trimer in prefusion-closed conformation and two additional *N*-glycans to cover the trimer base. Negative stain (NS) EM revealed well-formed trimers, but with some monomeric shape present. Glycan-base BG505 bound most antibodies similarly to BG505 DS-SOSIP, except for the base-directed antibodies 1E6, 5H3, 3H2, and 9B9, to which it had negligible affinity.

(C) Details of B cell sorting gate strategy.

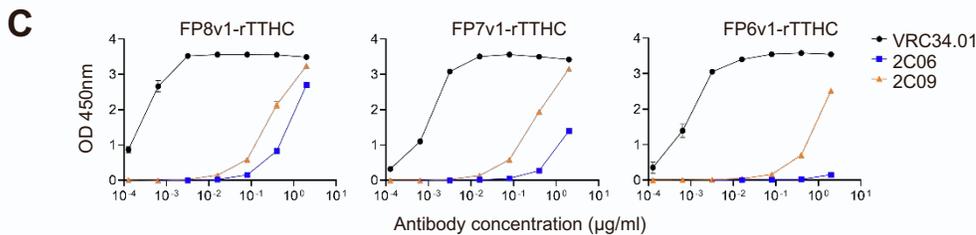
(D) Cytometry of IgG+ B cells from N751 serum. Cells were stained first with glycan-base BG505 to allow all non-base directed B cells to engage first in the trimer, washed the cells, and then stained with BG505 to allow the base-specific B cells to engage. BG505+ cell population is much larger than glycan-base BG505+ cell population. The glycan-base BG505-positive cells account for 0.37% of IgG+ memory B cells (Singlet, Live, CD19+, CD3-, CD14-, CD56-, IgG+) (upper panel). After absorption with glycan-base BG505, 2.53% of IgG+ memory B cells bind BG505 DS-SOSIP trimer (lower panel). Glycan-base BG505+ cells are 12.89% of all antigen-specific memory B cells, and the other 87.11% are likely targeting the trimer base.

A Pseudovirus neutralization assessments for 2D02 and 1B06

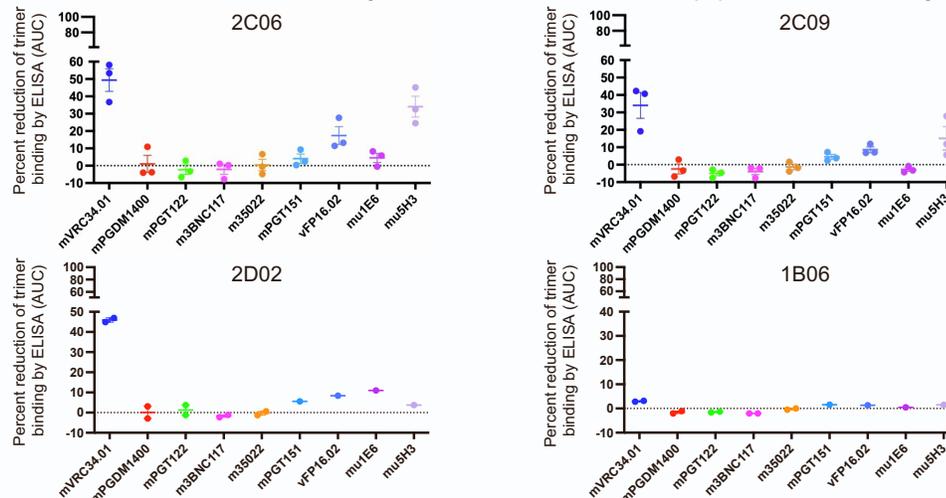
Viral strains	IC50 $\mu\text{g/ml}$			IC80 $\mu\text{g/ml}$		
	MW965.26	BG505.W6M.C2	BG505.W6M.C2.N611Q	MW965.26	BG505.W6M.C2	BG505.W6M.C2.N611Q
Clade	C	A		C	A	
2D02 IgG	>50	>50	>50	>50	>50	>50
1B06 IgG	>50	>50	>50	>50	>50	>50

B Neutralization assessments against a panel of viruses neutralized by most fusion peptide-directed antibodies

Viral strains	Clade	BG505.W6M.C2.N611Q	BG505.W6M.C2	HxB2.D.G	KER200.8.12	MB201.A1	MI369.A5	96ZM6.51.02	CNE40	Q23.17	CH117.4	3468.V.1.C12	3365.v2.c20	2M135.10a	BI369.9A	0815.v.3.c3	CNE19
		A	A	B	A	A	A	C	BC	A	BC	AD	A	C	A	ACD	BC
IC50 $\mu\text{g/ml}$	2C06 IgG	2.92	33.6	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100
	2C09 IgG	2.07	44.2	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100
	N123-VRC34.01	0.017	0.116	0.254	0.082	0.072	0.155	0.121	0.214	0.173	0.062	1.11	0.050	>100	0.134	0.039	0.035
IC80 $\mu\text{g/ml}$	2C06 IgG	10.4	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100
	2C09 IgG	10.9	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100
	N123-VRC34.01	0.084	0.641	1.05	0.317	0.323	1.03	0.414	0.832	1.21	0.604	>100	0.194	>100	1.25	0.14	>100



D Competition ELISA of 2C06 and 2C09 binding to BG505 DS-SOSIP with fusion peptide and base-binding antibodies



E Neutralization assessments against BG505.S241N as well as against all strains in 208-strain panel lacking a glycan at residue 241

Viral strains	BG505.S241N	CNE56	929-28	T257-31	0077.v1.c16	3168.v4.c10	6471.v1.c16	25710-2.43
	Tier 2	2	2/3	2	C	C	C	1B/2
Clade	A	AE	AG	AG	C	C	C	C
IC50 $\mu\text{g/ml}$	2C06	>100	>100	>100	>100	>100	>100	>100
	2C09	>100	>100	>100	>100	>100	>100	>100
	VRC34.01	0.758	>100	>100	>100	1.48	11.5	>100
IC80 $\mu\text{g/ml}$	2C06	>100	>100	>100	>100	>100	>100	>100
	2C09	>100	>100	>100	>100	>100	>100	>100
	VRC34.01	4.610	>100	>100	>100	>100	65.6	>100

Figure S2. Binding and neutralization data, related to Figures 1, 5, and 6.

(A) Neutralization assessments for 2D02 and 1B06 revealed no detectable neutralization activity.

(B) Neutralization assessments of VRC 018-elicited antibodies from donor N751 against a panel of viruses neutralized by most fusion peptide-directed antibodies.

(C) ELISA analysis of 2C06 and 2C09 IgG binding to fusion peptide. Measurements were carried out in triplicates, and mean and SE are reported for each data point.

(D) Competition ELISA revealed 2C06, 2C09, and 2D02 competed for binding to BG505 DS-SOSIP trimer with fusion peptide-directed antibodies and base binding antibody 1E6 or 5H3, whereas 1B06 did not compete with any antibody tested.

(E) Neutralization assessments focusing on strains in 208-strain panel lacking a glycan at residue 241.

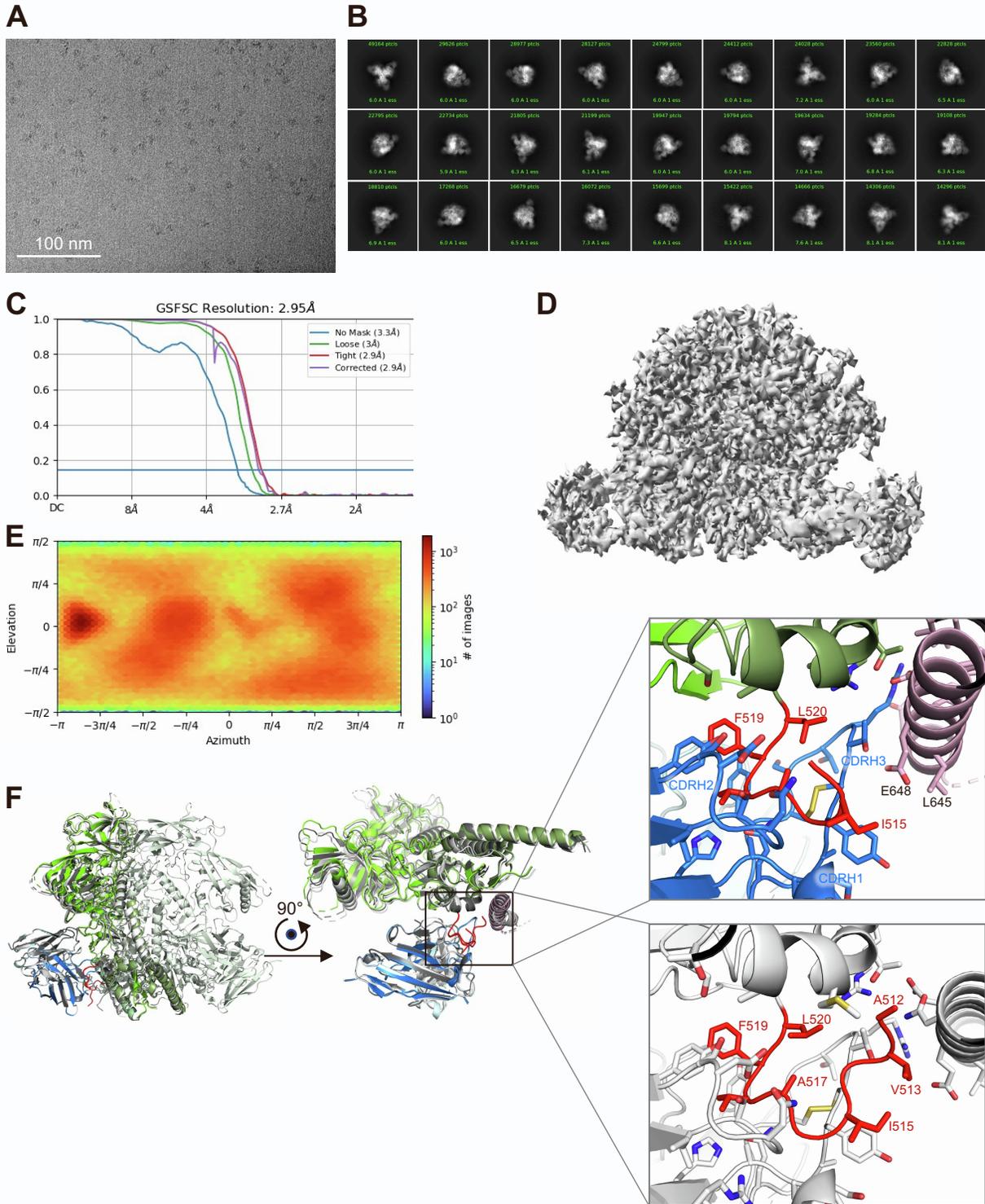


Figure S3. Cryo-EM validation for 2C06 in complex with BG505 DS-SOSIP, related to Figure 2.

(A) Representative micrograph. (B) 2D classes. (C) Fourier shell correlation (FSC) curves showing gold-standard FSC resolution at 2.95 Å. (D) 3D reconstruction density map at 2.95 Å from non-uniform refinement with C1 symmetry, showing antibody Fab binding near the trimer base at the fusion peptide-site of vulnerability. (E) Orientations of all particles used in the final refinement shown as a heatmap. (F) Superposition of three protomers revealing the trimer asymmetry induced by 2C06 binding. (Left) Protomers in right and back are superimposed on the one in left. (Middle) A 90° rotation of the left image to put the superimposed antibody chains at the bottom. Only the superimposed protomers are shown. One protomer is shown in colors, and the other two are in light or dark gray. (Right) Zoom-in views showing two slightly different modes of binding. The helix on the right is from a neighboring protomer.

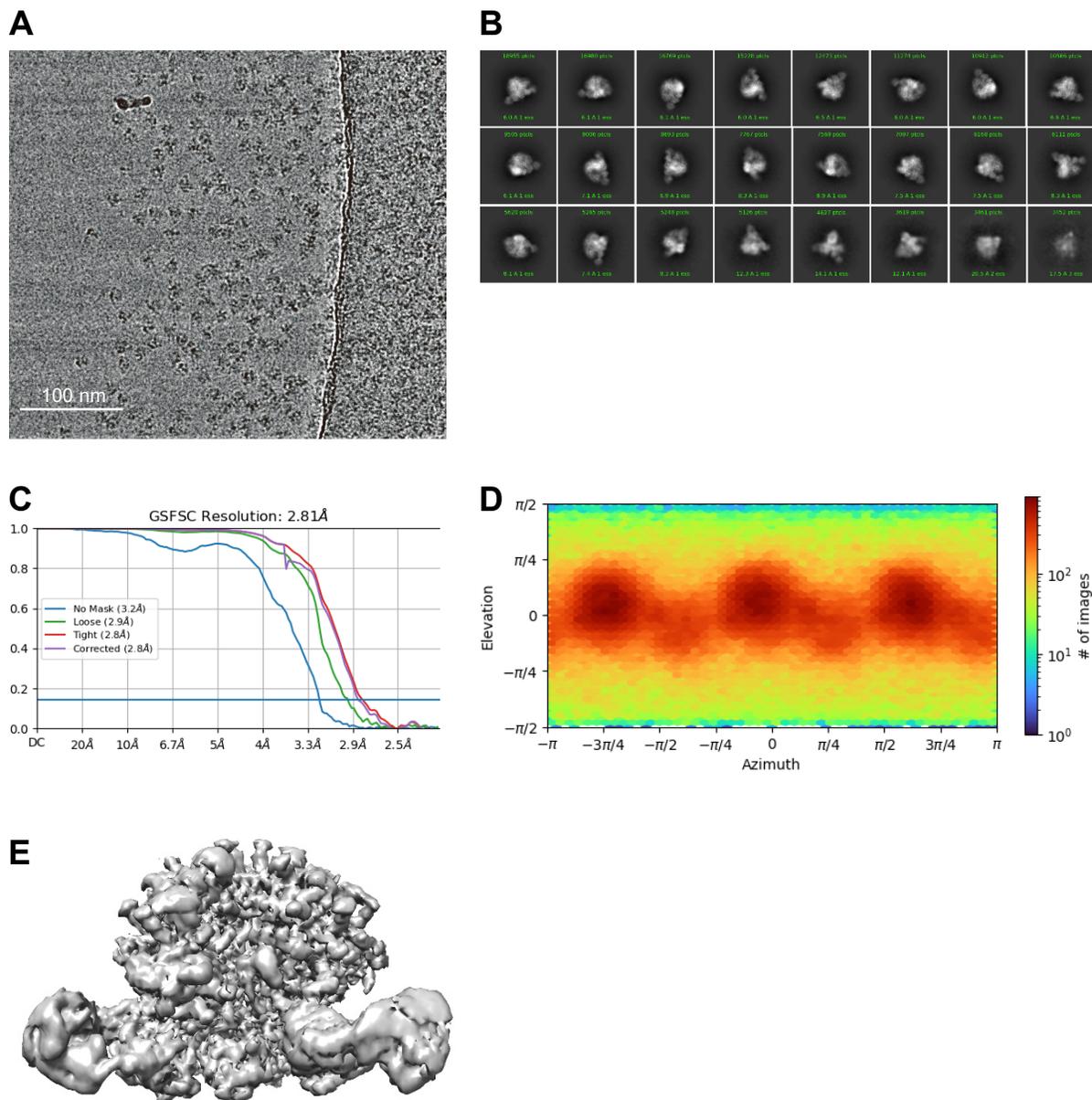


Figure S4. Cryo-EM validation for 2C09 in complex with BG505 DS-SOSIP, related to Figure 3.

(A) Representative micrograph. (B) 2D classes. (C) Fourier shell correlation (FSC) curves showing gold-standard FSC resolution at 2.81 Å. (D) Orientations of all particles used in the final refinement shown as a heatmap. (E) 3D reconstruction density map at 2.81 Å from non-uniform refinement with C3 symmetry, showing antibody Fab binding near the trimer base at the fusion peptide-site of vulnerability.

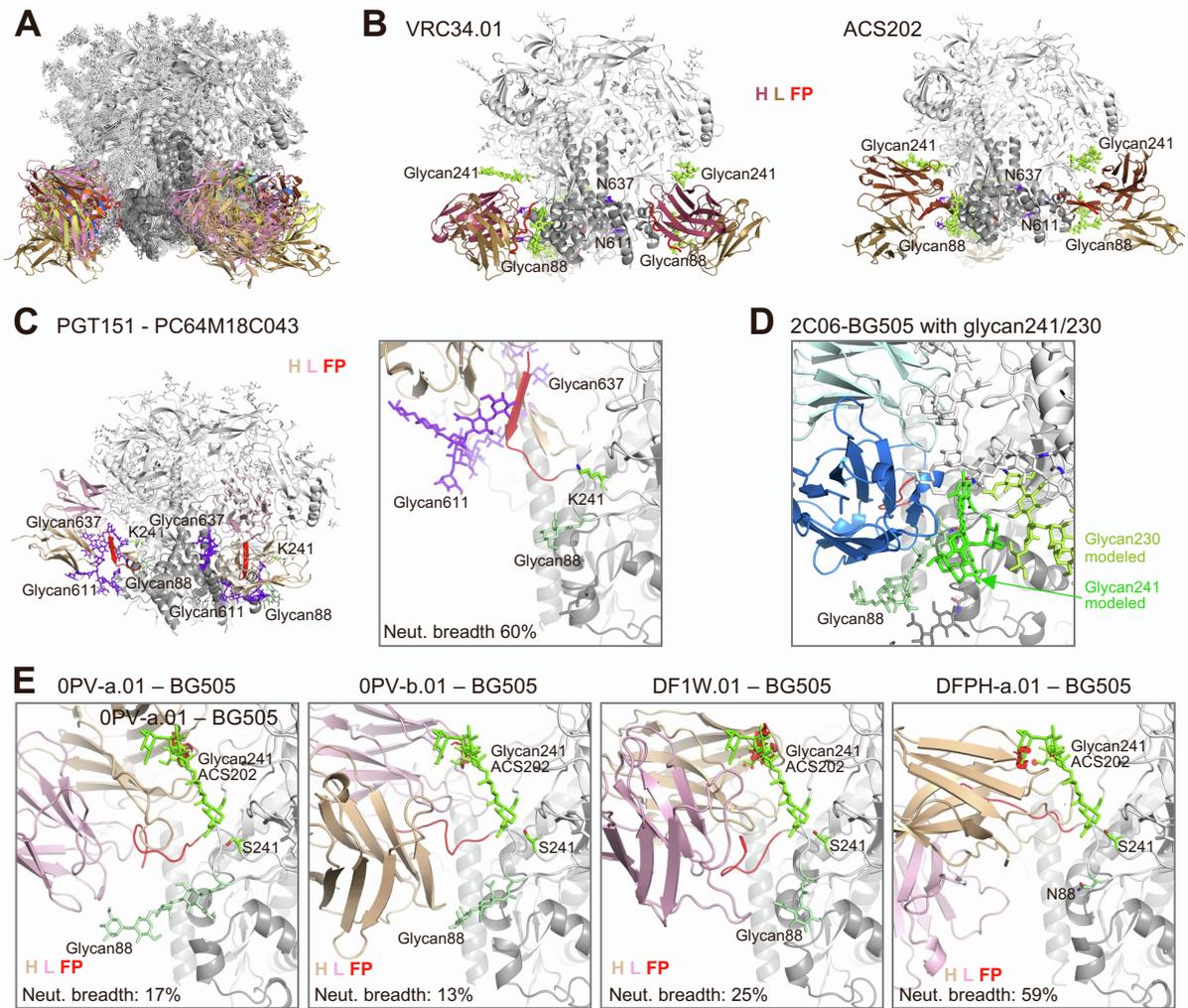


Figure S5. Fusion peptide-directed antibodies (except for PGT151) bind at a similar location and interact favorably with glycan88 but variably with glycan241, related to Figure 5.

(A) Structural superposition of fusion peptide-directed antibodies in complex with Env trimer. The structures, 2C06, 2C09, ACS202 (PDB: 6nc2), VRC34.01 (PDB: 6nc3), DFPH-a.01 (modeled based on DFPH-a.15 structure, PDB: 6n1w), DF1W-a.01 (PDB: 6mph), A12V163-a.01 (PDB: 6n1v), A12V163-b.01 (PDB: 6mpg), 0PV-a.01 (PDB: 6osy), 0PV-b.01 (PDB: 6ot1), 0PV-c.01 (PDB: 6nf2), and vFP16.02 (PDB: 6cdi), were aligned by one of the gp120 subunits (right).

(B) Binding of VRC34.01 and ACS202 involves both glycan88 and glycan241 (green sticks), but not glycan611 or glycan637, both not modeled in the structure, likely disordered. VRC34.01 and ACS202 bind in a similar location but with different orientations.

(C) PGT151 (PDB: 6dcq) binds to glycan611 and glycan637 (blue-purple sticks), away from glycan88 (green sticks) and the side chain of K241 (green sticks). Neutralization breadths were for IC₅₀ <50 µg/ml in the 208-strain panel.

(D) Structure of 2C06-BG505 complex with glycans at 241 and 230, as well as all other potential glycans modeled as mannose 5. With adjustment of glycan conformations, both glycans at 241 and 230 could be modeled without clashing with each other or with the antibody.

(E) Vaccine-elicited NHP or murine antibodies all bind at a similar location to that of VRC34.01 and ACS202. The aligned glycan241 from the ACS202 complex (green sticks) has some minor clashes with antibodies 0PV-a.01, 0PV-b.01, DF1W.01, and DFPH-a.01, shown as red disks in Pymol (<https://pymol.org/2/>). These clashes are from the terminal mannose residues and can be avoided by minor shifts of the glycan, unlike those observed in 2C06 and 2C09 and other vaccine-elicited antibodies shown in Figure 5.

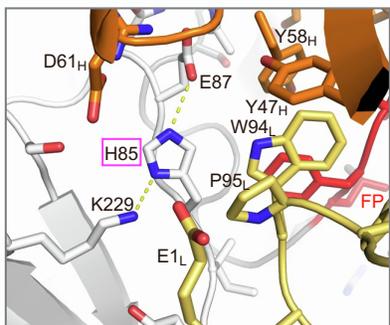
A

Position	Entropy	Most freq aa	2C06		2C09		A12V163-a.01		A12V163-b.01		VRC34.01 _a		vFP16.02		0PV-c.01		
			Contact atoms	BSA	Contact atoms	BSA	Contact atoms	BSA	Contact atoms	BSA	Contact atoms	BSA	Contact atoms	BSA	Contact atoms	BSA	
512	0.082	Ala (96.5)	Ala C, O	4.78	C, O	11.9	N, CA, C, O, CB	158.6	CA, C, O	20.1	N(H), CA, C, O, CB	127.3	N, CA, C, O(H), CB	161.6	N(H, S), CA, C, O, CB	83.1	
513	0.275	Val (80.6)	Val N, CA, C, O, CB, CG2	26.9	CA, C, O, CB, CG1, CG2	87.6	N, CA, C, O, CB, CG1, CG2	86.2	N, CA, C, O, CB	22.9	N(H), CA, C, O(H), CB, CG1, CG2	114.5	N(H), CA, C, O(H), CB, CG1, CG2	106.9	N(H), CA, C, O, CB, CG1, CG2	97.4	
514	0.226	Gly (84.9)	Gly N, CA, C, O	19.9	N, CA	11.9	N(H), CA, C, O(H)	73.7	N, CA, C, O	46.4	N, CA, C, O	22.6	N(H), CA, C, O	42.6	N, CA, C, O(H)	90.7	
515	0.411	Ile (50.0)	Ile N, CA, C, O, CB, CG1, CG2, CD1	75.6	N(H), C, O(H), CB, CG1, CG2, CD1	30.7	N, CA, C, O, CB, CG1, CG2, CD1	154.6	N, CA, C, O(H), CB, CG1, CG2, CD1	125.4	N(H), CA, C, O(H), CB, CG1, CG2, CD1	153.4	N, CA, C, O, CB, CG1, CG2, CD1	140.6	N, CA, C, O, CB, CG1, CG2, CD1	134.5	
516	0.023	Gly (99.0)	Gly N, CA, C, O(H)	81.5	N(H), CA, C, O(H)	55.9	N, CA, C, O	25.1	N, CA, C, O	41.3	N, CA, C, O(H), CB, CG1, CG2	38.7	N, CA, C, O	38.2	N(H), CA, C, O(H)	67.1	
517	0.043	Ala (98.1)	Ala N, CA, C, O, CB	31.1	N, CA, C, O, CB	36.3	N, CA, C, O, CB	44.8	N(H), CA, C, O, CB	108.9	N(H), CA, C, O(H), CB	46.9	N, CA, C, O, CB	40.3	N(H), CA, C, O(H), CB	70.3	
518	0.429	Val (45.1)	Val N, CA, C, O(H), CB, CG1, CG2	146.2	N, CA, O, CB, CG1, CG2	151.4	CA, C, CB, CG1, CG2	74.6	N, CA, C, O, CB, CG1, CG2	122.9	N, CA, C, O, CB, CG1, CG2	91	N, CA, CB, CG1, CG2	34	N(H), CA, C, O, CB, CG1, CG2	145.4	
519	0.183	Phe (85.9)	Phe N, CA, C, O, CB, CG, CD1, CD2, CE1, CE2, CZ	106.4	N, CA, O, CB, CG, CD1, CD2, CE1, CE2, CZ	108.6	N, O, CG, CD1, CE1, CE2, CZ	67.9	N, CA, C, O, CB, CG, CD1, CD2, CE1, CE2, CZ	132.3	N(H), C, O(H), CB, CG, CD1, CD2, CE2	72.1	CB, CG, CD1, CD2, CE1, CE2, CZ	46.2	N(H), CA, C, O(H), CB, CD1, CD2, CE1, CE2, CZ	83.2	
520	0.144	Leu (90.8)	Leu N, CA, O, CB, CG, CD1, CD2	45.5	N, O, CB, CD1	65	CA, C, CB, CG, CD1, CD2	63.6	N	6.02	N, CA, O(H)	47.7	CD1	17.2	N, CA, O, CB, CG, CD1, CD2	62.4	
535	0.472	Ile (47.6)	Met (17.6)	CG	5.52	SD, CE	17.7					CG	1.67	SD, CE	26.7		
85	0.733	Val (49.8)	His (2.5)	N, CA, O, CB, CG, CD2, ND1, CE1, NE2	67.9	N, CA, CB, CG, CD2, ND1, CE1, NE2	62.1	N, CA, C, O, CB, CG, CD2, ND1, CE1, NE2	94.6	N, CA, C, O, CB, CG, CD2, ND1, CE1, NE2	74.4	N, CB, CG1, CG2	40.9	N, CB, CG, CD2, ND1, CE1, NE2	43.2	N(H), O, CB, CG, CD2, ND1, CE1, NE2	69
87	0.573	Glu (52.7)	Glu	CA, CB, CG, CD, OE1, OE2	77.3	CB, CG, CD, OE1, OE2(H, S)	85.6	N, CA, C, O, CB, CG, CD	46.2	N, CA, C, O, CB, CG, CD, OE1, OE2	77.3	CB, CG, CD, OE1, OE2	39.2	CG, OE1, OE2	46.7		
640	0.743	Gly (24.8)	Gln (6.8)	CA, C, O	19.2												
644	0.729	Thr (26.5)	Gly (1.7)														
648	0.547	Glu (54.9)	Glu	N, CA, CB, CG, CD, OE1	23.2												
651	0.417	Asn (69.2)	Asn	CB, CG, ND2	28.8												

a – Numbers in parentheses are the percentage frequency of the amino acid in the 2020 HIV-1 group M sequences.
 b – Hydrogen bonds or salt bridges involved with the atom are designated as H or S, respectively.
 c – BSA, buried surface area in Å² per residue calculated with PISA. 0  160 Å²
 d – VRC34.01 in complex with AMC011 Env, which has a Val at position 85 instead of His.

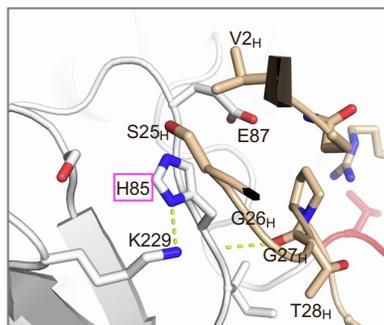
B

His85 interactions with 2C09



C

His85 interactions with 0PV-c.01



D

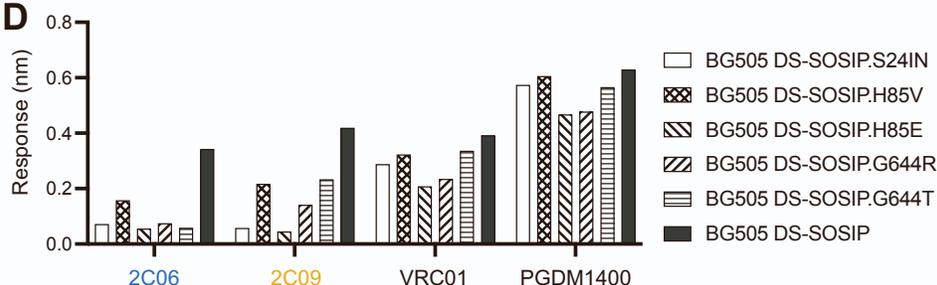


Figure S6. Analysis of entropy and binding interactions of the epitope residues for 2C06 and 2C09 in comparison with other fusion peptide-directed antibodies, related to Figure 6.

(A) Shannon entropy and contact surface areas of epitope residues for strain-specific and broadly neutralizing fusion peptide-directed antibodies. Residues analyzed include fusion peptide residues 512 to 520 and other residues involved in binding of 2C06 and 2C09 with normalized entropy above 0.4. Entropy values were calculated with Shannon Entropy-One (https://www.hiv.lanl.gov/content/sequence/ENTROPY/entropy_one.html) on the curated alignment of year 2020 HIV-1 M group Env of 5255 sequences (<https://www.hiv.lanl.gov/content/sequence/NEWALIGN/align.html>), and were normalized to have values between 0 and 1. Contact atoms and BSA were calculated with PISA (https://www.ebi.ac.uk/msd-srv/prot_int/cgi-bin/piserver).

(B) His85 has tight interactions with 2C09 heavy and light chain residues, similar to that in the 2C06 complex. (C) In 0PV-c.01, most interactions with H85 side chain are from heavy chain residues S25, G26, and G27. There is space and flexibility to accommodate other amino acids at position 85, despite large BSA with H85. (D) BLI assays for antibody binding to mutants of BG505 DS-SOSIP, S241N, H85V, H85E, G644R, and G644T.

Table S1. Cryo-EM data and refinement statistics, related to Figures 2 and 3.

	2C06	2C09
EMDB ID	EMD-29725	EMD-29731
PDB ID	8G4M	8G4T
<u>Data Collection</u>		
Microscope	FEI Titan Krios	FEI Titan Krios
Voltage (kV)	300	300
Electron dose (e ⁻ /Å ²)	58.06	58
Detector	Gatan K3 BioQuantum	Gatan K2 Summit
Pixel Size (Å)	0.83	1.11
Defocus Range (µm)	-0.8/-2.0	-1.0/-2.5
Magnification	105,000	29,000
 <u>Reconstruction</u>		
Software	cryoSPARC v3.3.1	cryoSPARC v3.3.1
Particles	645,442	445,733
Symmetry	C1	C3
Resolution (Å) (FSC _{0.143})	2.95	2.81
 <u>Refinement</u>		
Software	Phenix 1.20	Phenix 1.20
Protein residues	2432	2437
Ligands	BMA: 7; NAG: 98; MAN: 14	BMA: 9; NAG: 105; MAN: 15
CC (box)	0.80	0.83
CC (mask)	0.88	0.88
R.m.s. deviations		
Bond lengths (Å)	0.003	0.004
Bond angles (°)	0.459	0.453
 <u>Validation</u>		
Molprobit score	1.55	1.51
Clash score	4.25	4.00
Rotamer outliers (%)	2.43	1.38
Ramachandran		
Favored regions (%)	97.78	96.62
Allowed regions (%)	2.22	3.38
Disallowed regions (%)	0	0

Table S2. Epitope analysis for fusion peptide-directed antibodies in comparison to non-fusion peptide antibodies, related to Figure 6.

Epitope entropy was calculated as BSA weighted average of normalized Shannon's entropy based on 208-virus panel for the epitope residues of each antibody. Percentage of epitope overlap was calculated as number of overlapping epitope residues divided by the number of epitope residues for the antibody with the least number of epitope residues of the two. Epitope residue was defined as residue with BSA>0. Fusion peptide-directed antibodies have substantial epitope overlap, and they have similar epitope entropies, except PGT151, which binds at a slightly different location and has a lower epitope entropy. Non-fusion peptide antibodies have variable epitope entropies that correlates with neutralization breadth.

Category	PDB ID	Antibody	Epitope entropy	Neut breadth	Epitope BSA (Å ²)	Epitope overlap (%) with	
						2C06	2C09
Fusion peptide		2C06	0.29	<1%	1268	100	90
		2C09	0.26	<1%	1044	90	100
	6mpg	A12V163-b.01	0.28	0%	1083	84	87
	6n1v	A12V163-a.01	0.28	3%	1055	84	92
	6osy	0PV-a.01	0.26	17%	1108	71	71
	6ot1	0PV-b.01	0.26	13%	1017	67	57
	6nf2	0PV-c.01	0.27	23%	1320	62	71
	6mph	DF1W-a.01	0.26	25%	1068	75	65
	6nc2	ACS202	0.30	26%	902	62	74
	6cdi	vFP16.02	0.24	31%	1107	69	81
	6nc3	VRC34.01	0.27	50%	774	71	75
6n1w	DFPH-a.1	0.27	59%	1179	88	69	
5fuu	PGT151	0.15	60%	1465	54	55	
V1V2	3u2s	PG9	0.38	81%			
	5v8l	PGT145	0.31	75%			
	5vgj	VRC38.01	0.46	29%			
Glycan-V3	5aco	PGT128	0.38	69%			
	5cez	PGT121	0.34	65%			
	4jm2	PGT135	0.47	34%			
	6ozc	2G12	0.40	25%			
CD4-binding site	5fyj	VRC01	0.25	90%			
	4yjd	VRC13.01	0.16	83%			
	4ydk	VRC16.01	0.18	55%			
	5f9o	8ANC131	0.20	53%			
	4jan	CH103	0.23	53%			
	5t3z	IOMA	0.23	50%			
	5vn8	b12	0.29	43%			
4ye4	HJ16	0.51	31%				
Silent face	6bf4	VRC-PG05	0.60	28%			
Subunit	5cez	35O22	0.29	45%			
interface	5cix	8ANC195	0.21	67%			

Table S3. Analysis of antibody-trimer interfaces, by CDRs, related to Figures 2 and 3. Subunit interface buried surface areas (\AA^2) were analyzed using PISA. Blank entries are 0, i.e. no binding interactions.

	Chain	Protomer 1		Protomer 2		Protomer 3		Subtotal
		A gp41	C gp120	B gp41	G gp120	F gp41	I gp120	
2C06	CDR H1	54.67						54.67
Antibody 1	CDR H2	164.67	129.49					294.16
(chains DE)	CDR H3	316.52	14.32			126.34		457.18
	L N-term		27.07					27.07
	CDR L1		87.45					87.45
	CDR L3	58.34	176.47					234.81
	FR L3		8.97					8.97
	DE total	594.2	443.77			126.34		1164.31
Antibody 2	CDR H1			72.27				72.27
(chains JK)	CDR H2			184.65	118.97			303.62
	CDR H3	204.07		280.25	13.11			497.43
	L N-term				7.71			7.71
	CDR L1				95.57			95.57
	CDR L3			59.48	176.94			236.42
	FR L3				7.66			7.66
	JK total	204.07		596.65	419.96			1220.68
Antibody 3	CDR H1					65.28		65.28
(chains HL)	CDR H2					198.22	122.92	321.14
	CDR H3			202		255.54	15.73	473.27
	L N-term						11.5	11.5
	CDR L1						92.69	92.69
	CDR L3					58.82	178.01	236.83
	FR L3						6.7	6.7
	HL total			202		577.86	427.55	1207.41
2C09	CDR H1	93.24						93.24
Antibody 1	CDR H2	164.88	98.03					262.91
(chains DE)	CDR H3	256.08	14.32					270.4
	L N-term		27.07					27.07
	CDR L1		83.25					83.25
	CDR L3	92.08	170.84					262.92
	FR L3		8.97					8.97
	DE total	606.28	402.48					1008.76
antibody	CDR H1			92.27				92.27
(chains JK)	CDR H2			164.61	100.5			265.11
	CDR H3			258.57	13.11			271.68
	L N-term				7.71			7.71
	CDR L1				83.33			83.33
	CDR L3			88.73	170.05			258.78
	FR L3				7.66			7.66
	JK total			604.18	382.36			986.54
Antibody	CDR H1					91.49		91.49
(chains HL)	CDR H2					166.05	98.03	264.08
	CDR H3					259.94	15.73	275.67
	L N-term						11.5	11.5
	CDR L1						83.74	83.74
	CDR L3					86.97	170.6	257.57
	FR L3						6.7	6.7
	HL total					604.45	386.3	990.75

Table S4. Analysis of antibody-trimer interfaces, by trimer sequence segments, related to Figures 2 and 3. Subunit interface buried surface areas (\AA^2) were analyzed using PISA. Blank entries are 0, i.e. no binding interactions.

			Antibody 1 DE		Antibody 2 JK		Antibody 3 HL	
Epitope subregions			Heavy	Light	Heavy	Light	Heavy	Light
2C06								
Protomer 1	A gp41	FP 512-520	469.77	64.41	149.14			
		524-542	109.63					
		592-648						
	C gp120	79-88	125.14	276.3				
		glycan88	249.6					
		227+229		45.91				
Protomer 2	B gp41	FP 512-520	99.28		489.37	67.96	142.4	
		524-542			154.52			
		588-648						
	G gp120	79-88			119.21	292.67		
		glycan88			247.2			
		227+229				19.07		
Protomer 3	F gp41	FP 512-520	99.28		1180.1	379.7	455.95	67.28
		524-542					146.62	
		644-651						
	I gp120	79-88					125.64	299.58
		glycan88					246	
		227+229						20.52
	241	22.27						
Subtotal			1073.05	386.62	1180.1	379.7	1138.9	387.38
2C09								
Protomer 1	A gp41	FP 512-520	463.99	91.9				
		524-542	70.96					
		592-648						
	C gp120	85-88	125.14	279.91				
		glycan88	285					
		229		10				
Protomer 2	B gp41	FP 512-520	99.28		470.71	90.22		
		524-542			71.48			
		588-648						
	G gp120	85-88			101.4	279.55		
		glycan88			285			
		229				7.5		
Protomer 3	F gp41	FP 512-520	99.28		944.58	377.27	469.98	89.36
		524-542					71.83	
		644-651						
	I gp120	85-88					102.66	276.51
		glycan88					285	
		229					0.17	8.55
	241	14.1						
Subtotal			964.72	381.81	944.58	377.27	943.74	374.42

Table S5. Frequency of N-glycosylation sites in HIV-1 Env sequences, related to Figure 5. HIV-1 Env protein sequence alignment was from HIV sequence database (<https://www.hiv.lanl.gov/content/sequence/NEWALIGN/align.html>) M group without recombinants, year 2020, a total of 5255 sequences. The aligned sequences were input to N-GlycoSite (<https://www.hiv.lanl.gov/content/sequence/GLYCOSITE/glycosite.html>), excluding NP[ST] and first position N in NN[ST][ST], with HXB2 as reference sequence.

Position in sequence alignment	HXB2 sequence	HXB2 residue number	Number of sequences with N-glycan sequon	Frequency of N-glycosite	Position in sequence alignment	HXB2 sequence	HXB2 residue number	Number of sequences with N-glycan sequon	Frequency of N-glycosite	Position in sequence alignment	HXB2 sequence	HXB2 residue number	Number of sequences with N-glycan sequon	Frequency of N-glycosite	Position in sequence alignment	HXB2 sequence	HXB2 residue number	Number of sequences with N-glycan sequon	Frequency of N-glycosite
10	h	9	19	0.36	268	N	156	5030	95.72	541	N	301	4969	94.56	739	n	425	1	0.02
13	w	11	1	0.02	272	N	160	4741	90.22	542	n	302	74	1.41	756	s	440	1	0.02
14	r	12	1	0.02	273	l	161	3	0.06	544	r	304	1	0.02	758	q	442	1524	29.00
41	i	27	19	0.36	274	s	162	1	0.02	568	g	321	2	0.04	759	i	443	1	0.02
43	s	29	185	3.52	276	s	164	10	0.19	569	k	322	1	0.02	760	r	444	462	8.79
44	a	30	1	0.02	277	i	165	1	0.02	588	h	330	8	0.15	762	s	446	370	7.04
45	t	31	7	0.13	281	-	166	1	0.02	591	N	332	4207	80.06	764	N	448	4519	85.99
46	-	31	1	0.02	288	g	167	1	0.02	593	s	334	670	12.75	776	g	459	3	0.06
47	-	31	1	0.02	289	k	168	1	0.02	594	r	335	11	0.21	777	n	460	694	13.21
48	-	31	1	0.02	291	q	170	1	0.02	597	k	337	303	5.77	778	s	461	1101	20.95
52	-	31	3	0.06	292	k	171	10	0.19	598	w	338	1	0.02	779	n	462	1552	29.53
54	-	31	2	0.04	293	e	172	1	0.02	600	N	339	3615	68.79	780	N	463	1044	19.87
55	-	31	1	0.02	295	y	173	1	0.02	601	n	340	10	0.19	781	-	463	189	3.60
56	-	31	1	0.02	299	f	176	1	0.02	602	t	341	4	0.08	782	-	463	102	1.94
58	e	32	16	0.30	300	y	177	1	0.02	604	k	343	47	0.89	783	-	463	47	0.89
74	e	47	18	0.34	303	l	179	1	0.02	605	q	344	138	2.63	784	-	463	2	0.04
76	t	49	619	11.78	307	p	183	1	0.02	606	i	345	1	0.02	788	-	463	7	0.13
82	c	54	1	0.02	308	l	184	1	0.02	607	a	346	7	0.13	804	-	463	1	0.02
87	k	59	1	0.02	309	d	185	293	5.58	609	k	348	1	0.02	805	-	463	1	0.02
96	n	67	1	0.02	310	N	186	723	13.76	611	r	350	6	0.11	806	-	463	5	0.10
126	v	85	15	0.29	311	-	186	199	3.79	612	-	350	2	0.04	808	-	463	11	0.21
128	v	87	1	0.02	312	-	186	287	5.46	617	e	351	6	0.11	809	-	463	8	0.15
131	-	87	1	0.02	313	-	186	184	3.50	618	q	352	1	0.02	810	-	463	13	0.25
132	N	88	5186	98.69	314	-	186	95	1.81	620	g	354	378	7.19	811	-	463	16	0.30
136	n	92	23	0.44	315	-	186	34	0.65	621	-	354	14	0.27	812	-	463	13	0.25
155	k	97	20	0.38	316	-	186	12	0.23	622	-	354	5	0.10	813	-	463	23	0.44
156	n	98	2	0.04	317	-	186	8	0.15	623	-	354	6	0.11	814	-	463	43	0.82
173	d	113	5	0.10	318	-	186	12	0.23	624	-	354	1	0.02	815	-	463	51	0.97
180	c	119	1	0.02	319	-	186	10	0.19	628	-	354	3	0.06	816	-	463	50	0.95
184	-	122	1	0.02	320	-	186	9	0.17	630	-	354	4	0.08	817	-	463	127	2.42
188	-	125	1	0.02	322	-	186	3	0.06	631	-	354	7	0.13	818	-	463	1617	30.77
191	s	128	7	0.13	323	-	186	6	0.11	632	-	354	8	0.15	819	e	464	5	0.10
192	l	129	1	0.02	324	-	186	1	0.02	633	n	355	211	4.02	820	s	465	1101	20.95
193	k	130	3353	63.81	325	-	186	3	0.06	634	N	356	3992	75.97	833	n	478	4	0.08
195	t	132	55	1.05	326	-	186	1	0.02	635	k	357	25	0.48	843	v	488	1	0.02
196	d	133	872	16.59	328	-	186	1	0.02	636	t	358	232	4.41	855	t	499	1	0.02
197	l	134	167	3.18	329	-	186	1	0.02	637	i	359	3	0.06	952	q	567	1	0.02
198	k	135	1243	23.65	330	-	186	3	0.06	638	l	360	166	3.16	984	g	597	7	0.13
199	N	136	881	16.76	331	-	186	1	0.02	639	f	361	1	0.02	994	a	607	7	0.13
200	d	137	1268	24.13	332	-	186	2	0.04	640	k	362	2177	41.43	998	N	611	5179	98.55
201	t	138	824	15.68	334	-	186	3	0.06	642	-	362	3	0.06	999	a	612	1	0.02
202	n	139	760	14.46	335	-	186	3	0.06	643	q	363	304	5.78	1000	s	613	36	0.69
203	t	140	812	15.45	336	-	186	4	0.08	653	t	373	2	0.04	1003	-	615	11	0.21
204	N	141	778	14.80	337	-	186	8	0.15	655	h	374	1	0.02	1004	-	615	15	0.29
205	s	142	743	14.14	338	-	186	9	0.17	656	s	375	1	0.02	1005	-	615	3	0.06
206	-	142	466	8.87	339	-	186	16	0.30	662	e	381	1	0.02	1006	-	615	1	0.02
207	-	142	312	5.94	340	-	186	26	0.49	668	N	386	4523	86.07	1007	-	615	3	0.06
208	-	142	250	4.76	341	-	186	44	0.84	670	t	388	1	0.02	1009	-	615	1	0.02
209	-	142	160	3.04	342	-	186	104	1.98	672	q	389	2	0.04	1012	-	615	1	0.02
210	-	142	135	2.57	343	-	186	218	4.15	674	l	390	1	0.02	1013	-	615	2	0.04
211	-	142	111	2.11	344	-	186	301	5.73	675	f	391	1	0.02	1014	-	615	1	0.02
212	-	142	71	1.35	345	-	186	453	8.62	676	N	392	4100	78.02	1015	-	615	19	0.36
213	-	142	63	1.20	346	-	186	511	9.72	677	s	393	89	1.69	1016	N	616	4780	90.96
215	-	142	12	0.23	347	d	187	1505	28.64	678	t	394	216	4.11	1017	k	617	3	0.06
217	-	142	6	0.11	348	t	188	1490	28.35	679	w	395	94	1.79	1018	s	618	287	5.46
218	-	142	6	0.11	349	t	189	8	0.15	680	f	396	1387	26.39	1024	-	623	1	0.02
219	-	142	13	0.25	350	s	190	16	0.30	681	N	397	642	12.22	1025	-	623	1	0.02
220	-	142	6	0.11	353	l	193	1	0.02	682	s	398	1722	32.77	1026	N	624	36	0.69
221	-	142	15	0.29	355	s	195	7	0.13	683	t	399	381	7.25	1027	h	625	5077	96.61
222	-	142	2	0.04	356	c	196	4	0.08	684	w	400	382	7.27	1038	i	635	1	0.02
223	-	142	3	0.06	357	N	197	5145	97.91	685	s	401	518	9.86	1040	N	637	5082	96.71
224	-	142	2	0.04	358	t	198	1	0.02	686	t	402	1112	21.16	1043	s	640	1	0.02
225	-	142	4	0.08	374	k	207	2	0.04	687	e	403	582	11.08	1048	s	644	1	0.02
226	-	142	3	0.06	395	k	227	1	0.02	688	g	404	598	11.38	1052	e	648	1	0.02
227	-	142	4	0.08	397	n	229	59	1.12	689	s	405	451	8.58	1083	n	656	1	0.02
228	-	142	4	0.08	401	-	229	1	0.02	690	N	406	301	5.73	1094	k	665	5	0.10
231	-	142	2	0.04	402	N	230	2153	40.97	691	n	407	262	4.99	1101	n	671	1	0.02
232	-	142	1	0.02	404	t	232	86	1.64	692	t	408	237	4.51	1104	N	674	687	13.07
233	-	142	1	0.02	407	N	234	4098	77.98	693	-	408	102	1.94	1130	v	698	1	0.02
236	-	142	1	0.02	411	t	236	10	0.19	694	-	408	41	0.78	1142	n	706	5	0.10
238	-	142	2	0.04	413	p	238	7	0.13	696	-	408	4	0.08	1157	-	718	5	0.10
239	-	142	5	0.10	416	N	241	5072	96.52	700	-	408	1	0.02	1188	r	740	1	0.02
240	-	142	9	0.17	440	s	256	1	0.02	703	-	408	1	0.02	1189	d	741	6	0.11
241	-	142	21	0.40	441	t	257	1	0.02	706	-	408	1	0.02	1190	r	742	4	0.08
242	-	142	38	0.72	447	N	262	5220	99.33	709	-	408	3	0.06	1191	d	743	245	4.66
243	-	142	31	0.59	449	s	264	4	0.08	710	-	408	2	0.04	1192	r	744	3	0.06
244	-	142	54	1.03	452	e	267	1	0.02	712	-	408	2	0.04	1195	i	746	6	0.11
245	-	142	52	0.99	453	e	268	41											