

Supplementary Information

Tumor Suppressor p53 Mediates Interleukin-6 Expression to Enable Cancer Cell Evasion of Genotoxic Stress

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Fig. S1 IL-6 is induced by genotoxic stress and drives cancer cell tolerance to genotoxic treatments. **A-C** IL-6 gene expression in A549 or H1299 cells treated with increasing concentrations of (A) sodium arsenite (SA), (B) doxorubicin (Dox), or (C) cisplatin (CisPt) for 24 h was measured by quantitative real-time PCR (qRT-PCR). **D** IL-6 mRNA levels in primary human liposarcoma cell cultures were analyzed using the GSE12972 dataset and their \log_2 -transformed values were compared between control (n = 19) and paired Dox-treated (n = 19) cells. **E** The concentration of IL-6 present in the culture supernatants of A549 or H1299 cells treated with the indicated concentrations of CisPt for 24 h was determined by ELISA. **F, G** Flow cytometric analysis of the cell surface expression of IL-6R α in A549 cells treated with increasing concentrations of (F) SA or (G) CisPt for 24 h. Cells were stained with PE-conjugated antibody against IL-6R α . **H** IL-6 gene expression in control (scramble) and IL-6-silenced (shIL-6-1 and shIL-6-2) A549 cells untreated or treated with 20 μ M SA for 24 h was measured by qRT-PCR. **I** IL-6 protein concentration in the culture supernatants of scramble, shIL-6-1, and shIL-6-2 A549 cells untreated or treated with 20 μ M SA for 24 h was determined by ELISA. **J** IL-6 gene expression in control (siCtrl) and IL-6-silenced (siIL-6) A549 cells untreated or treated with 20 μ M SA, 0.5 μ M Dox, or 20 μ M CisPt for 24 h was measured by qRT-PCR. **K** IL-6 protein concentration in the culture supernatants of siCtrl and siIL-6 A549 cells untreated or treated with 20 μ M SA, 0.5 μ M Dox, or 20 μ M CisPt for 24 h was determined by ELISA. **L** Dose-response curves showing the survival of siCtrl and siIL-6 A549 cells in response to increasing concentrations of CisPt treatment for 24 h. **M** The IC₅₀ values of CisPt against siCtrl and siIL-6 A549 cells were calculated from the nonlinear regression curves in Fig. S1L. Cell viability was measured by MTT assays. Error bars represent mean \pm SD, n = 3. Statistical analysis was performed using unpaired two-tailed Student's *t*-test (A-C, E-G, M), paired two-tailed Student's *t*-test (D), or two-way ANOVA with Tukey's multiple comparisons test (H-K). **p* \leq 0.05; ***p* \leq 0.01; ****p* \leq 0.001; ns, not significant.

Fig. S2 IL-6 silencing heightens genotoxic stress-induced cell death in human breast cancer MCF-7 and cervical cancer HeLa cells. **A, H** IL-6 gene expression in control (siCtrl) and IL-6-silenced (siIL-6) (A) MCF-7 or (H) HeLa cells untreated or treated with 20 μ M SA, 0.5 μ M Dox, or 20 μ M CisPt for 24 h was measured by qRT-PCR. **B, D, F, I, K, M** Dose-response curves showing the survival of siCtrl and siIL-6 (B, D, F) MCF-7 or (I, K, M) HeLa cells in response to increasing concentrations of (B, I) SA, (D, K) Dox, or (F, M) CisPt treatment for 24 h. **C, E, G, J, L, N** The IC₅₀ values of (C, J) SA, (E, L) Dox, or (G, N) CisPt against siCtrl and siIL-6 (C, E, G) MCF-7 or (J, L, N) HeLa cells were calculated from the nonlinear regression curves in Fig. S2B, D, F, I, K, M, respectively. Cell viability was measured by MTT assays. Error bars represent mean \pm SD, n = 3. Statistical analysis was performed using two-way ANOVA with Tukey's multiple comparisons test (A, H) or unpaired two-tailed Student's *t*-test (C, E, G, J, L, N). **p* \leq 0.05; ***p* \leq 0.01; ****p* \leq 0.001.

Fig. S3 IL-6 mitigates genotoxic stress-induced cell death. **A, C, E** Dose-response curves showing the survival of control and 50 or 20 ng/ml IL-6-pretreated (A) A549 or (C, E) H1299 cells, respectively, in response to increasing concentrations of (A, C) SA or (E) CisPt treatment for 24 h. **B, D, F** The IC₅₀ values of (B, D) SA or (F) CisPt against control and 50 or 20 ng/ml IL-6-pretreated (B) A549 or (D, F) H1299 cells were calculated from the nonlinear regression curves in Fig. S3A, C, E, respectively. **G, I** Dose-response curves showing the survival of siCtrl, siIL-6, and siIL-6+IL-6 A549 cells in response to increasing concentrations of (G) SA or (I) Dox treatment for 24 h. **H, J** The IC₅₀ values of (H) SA or (J) Dox against siCtrl, siIL-6, and siIL-6+IL-6 A549 cells were calculated from the nonlinear regression curves in Fig. S3G and S3I, respectively. Cell viability was measured by MTT assays. **K-M** The mRNA levels of the anti-apoptotic genes (K)

Bcl-xL, (L) Mcl-1, and (M) Bcl-2 in A549 cells treated with increasing concentrations of SA for 24 h were measured by qRT-PCR. **N** The proliferation of scramble, shIL-6-1, and shIL-6-2 A549 cells untreated or treated with 20 μ M SA for 24 h was assessed by manual cell counting with a hemocytometer. Error bars represent mean \pm SD, n = 3. Statistical analysis was performed using unpaired two-tailed Student's *t*-test (B, D, F, K-M) or one-way ANOVA with Tukey's multiple comparisons test (H, J). **p* \leq 0.05; ***p* \leq 0.01; ****p* \leq 0.001; ns, not significant.

Fig. S4 p53 silencing attenuates IL-6 expression. qRT-PCR analysis of (A, C) p53 and (B, D) IL-6 gene expressions in control (siCtrl) and p53-silenced (sip53) (A, B) MCF-7 or (C, D) HeLa cells untreated or treated with 0.5 μ M Dox for 24 h. Error bars represent mean \pm SD, n = 3. Statistical analysis was performed using two-way ANOVA with Tukey's multiple comparisons test. **p* \leq 0.05; ***p* \leq 0.01; ****p* \leq 0.001; ns, not significant.

Fig. S5 Genotoxic-treated cancer cells secrete IL-6 to enable alternative (M2) macrophage polarization. **A** THP-1 monocytes were differentiated into M0 macrophages by treatment with 10 ng/ml PMA for 24 h followed by culture in serum-free RPMI medium for an additional 24 h and M0 macrophages were then stimulated with conditioned media from untreated or SA/Dox-treated A549 cells to induce macrophage polarization as depicted. **B** Phase-contrast imaging of THP-1 monocytes and 10 ng/ml PMA-primed macrophages before and after being cultured in serum-free RPMI medium for 24 h. Scale bar: 100 μ m. **C-F** The mRNA levels of the macrophage markers (C) CD14, (D) CD68, (E) IL-1 β , and (F) TNF- α were analyzed in THP-1 monocytes and THP-1-derived M0 macrophages using qRT-PCR. **G** Flow cytometric analysis of the cell surface expression of the macrophage marker CD14 in THP-1 monocytes and THP-1-derived M0 macrophages. Cells were stained with PE-conjugated antibody against CD14. **H-J** The mRNA

levels of (H, I) M2- (CD206 and CD163) or (J) M1-associated (CD80) macrophage markers were analyzed in THP-1-derived macrophages co-cultured with RPMI, control (scramble), or IL-6-silenced (shIL-6-1 and shIL-6-2) A549 cells with and without 20 μ M SA or 0.1 μ M Dox treatment for 24 h using qRT-PCR. Error bars represent mean \pm SD, n = 3. Statistical analysis was performed using unpaired two-tailed Student's *t*-test (C-G) or two-way ANOVA with Tukey's multiple comparisons test (H-J). **p* \leq 0.05; ***p* \leq 0.01; ****p* \leq 0.001; ns, not significant.

Fig. S6 IL-6 expression in SA-treated A549 cells after SA removal and IL-6 concentration in macrophage-stimulating conditioned media. **A** qRT-PCR analysis of IL-6 mRNA levels in A549 cells after being treated with 20 μ M SA for 24 h and cultured in SA-free RPMI medium for the indicated time periods (1, 3, 6, 12, and 24 h). **B** The concentration of IL-6 present in the culture supernatants of A549 cells after being treated with SA (20 or 40 μ M) or Dox (0.1 or 0.5 μ M) for 24 h and cultured in SA/Dox-free RPMI medium for an additional 24 h. **C** The concentration of IL-6 present in the conditioned medium collected from the bottom wells of a 24-well cell culture plate was determined by ELISA. The upper chamber was plated with control (scramble) or IL-6-silenced (shIL-6-1 and shIL-6-2) A549 cells with and without 20 μ M SA or 0.1 μ M Dox treatment for 24 h. Error bars represent mean \pm SD, n = 3. Statistical analysis was performed using one-way ANOVA with Tukey's multiple comparisons test (A, B) or two-way ANOVA with Tukey's multiple comparisons test (C). **p* \leq 0.05; ***p* \leq 0.01; ****p* \leq 0.001; ns, not significant.

Supplementary Tables

Table S1. IC₅₀ of SA, Dox, or CisPt under various treatment conditions

Figure	Cell line	Treatment	IC ₅₀ of SA (μM)	IC ₅₀ of Dox (μM)	IC ₅₀ of CisPt (μM)
1H, I	A549	Scramble	256.90 ± 11.75		
		shIL-6-1	183.03 ± 2.72		
		shIL-6-2	192.80 ± 14.64		
1J ,K	A549	Scramble		112.05 ± 25.18	
		shIL-6 -1		11.60 ± 2.71	
		shIL-6 -2		28.09 ± 0.93	
S1L, M	A549	siCtrl			113.09 ± 12.73
		siIL-6			19.27 ± 0.39
S2B, C	MCF-7	siCtrl	47.45 ± 2.12		
		siIL-6	23.45 ± 5.06		
S2D, E	MCF-7	siCtrl		0.81 ± 0.12	
		siIL-6		0.44 ± 0.08	
S2F, G	MCF-7	siCtrl			214.97 ± 11.37
		siIL-6			107.30 ± 3.03
S2I, J	Hela	siCtrl	70.72 ± 8.53		
		siIL-6	33.69 ± 3.46		
S2K, L	Hela	siCtrl		24.80 ± 7.29	
		siIL-6		1.89 ± 0.93	
S2M, N	Hela	siCtrl			62.63 ± 2.60
		siIL-6			32.72 ± 2.82
S3A, B	A549	Vehicle	254.20 ± 11.37		
		IL-6	392.87 ± 84.06		

S3C, D	H1299	Vehicle	17.02 ± 0.76		
		IL-6	22.27 ± 2.85		
S3E, F	H1299	Vehicle			48.67 ± 8.22
		IL-6			76.80 ± 2.73
S3G, H	A549	siCtrl	258.87 ± 12.25		
		siIL-6	158.33 ± 11.40		
		siIL-6+IL-6	195.57 ± 14.62		
S3I, J	A549	siCtrl		93.91 ± 3.66	
		siIL-6		2.41 ± 0.29	
		siIL-6+IL-6		13.07 ± 0.80	
3G, H	A549	Vehicle	249.40 ± 1.51		
		BAPTA-AM	138.00 ± 11.15		
3N, O	A549	Vehicle	264.27 ± 9.24		
		LY294002	124.73 ± 18.16		
5M, N	A549	siCtrl	249.80 ± 21.85		
		si53	148.30 ± 6.12		
6H, I	A549	RPMI	252.27 ± 11.25		
		Ctrl CM	38.30 ± 0.80		
		SA CM	60.73 ± 9.76		
6J, K	H1299	RPMI	16.82 ± 1.10		
		Ctrl CM	11.83 ± 1.19		
		SA CM	17.06 ± 2.85		

Table S2. List of primers used for qRT-PCR

Target protein	Sequence (5'-3')
IL-6	Forward: AGACAGCCACTCACCTCTTCAG
	Reverse: TTCTGCCAGTGCCTCTTTGCTG
Panx1	Forward: CCACGGAGTACGTGTTCTCG
	Reverse: CCGCCCAGCAATATGAATCC
Bcl-2	Forward: ATGTGTGTGGAGAGCGTCAA
	Reverse: ACAGTTCCACAAAGGCATCC
Bcl-xL	Forward: GAGCTGGTGGTTGACTTTCTC
	Reverse: TCCATCTCCGATTCAGTCCCT
Mcl-1	Forward: TAAGGACAAAACGGGACTGG
	Reverse: ACCAGCTCCTACTCCAGCAA
p53	Forward: AAGGAAATTTGCGTGTGGAGT
	Reverse: AAAGCTGTTCCGTCCCAGTA
CD206	Forward: GGGAAAGGTTACCCTGGTGG
	Reverse: GTCAAGGAAGGGTCGGATCG
CD163	Forward: TTTGTCAACTTGAGTCCCTTCAC
	Reverse: TCCCGCTACACTTGTTTTAC
CCL18	Forward: TGGCAG ATTCCACAAAAGTTCA
	Reverse: GGATGACACCTGGCTTGGG
CD80	Forward: GGCCCGAGTACAAGAACCG
	Reverse: TCGTATGTGCCCTCGTCAGAT
CXCL10	Forward: TGTACGCTGTACCTGCATCA
	Reverse: GGACAAAATTGGCTTGCAGGA

IL-1 β	Forward: CTCGCCAGTGAAATGATGGCT
	Reverse: GTCGGAGATTCGTAGCTGGAT
CD14	Forward: CTGCAACTTCTCCGAACCTC
	Reverse: CCAGTAGCTGAGCAGGAACC
CD68	Forward: CTTCTCTCATTCCCCTATGGACA
	Reverse: GAAGGACACATTGTACTIONCCACC
TNF- α	Forward: TTCTGCCTGCTGCACTTTGGA
	Reverse: TTGATGGCAGAGAGGAGGTTG
UBC	Forward: CTGGAAGATGGTCGTACCCTG
	Reverse: GGTCTTGCCAGTGAGTGTCT
GAPDH	Forward: TGCACCACCAACTGCTTAGC
	Reverse: GGCATGGACTGTGGTCATGAG

Table S3. shRNA sequences

shRNA name	Clone ID	Oligo sequence (5'-3')
Scramble	ASN0000000004	CCGGCCTAAGGTTAAGTCGCCCTCGCTCGAGCGA GGGCGACTTAACCTTAGGTTTTT
shIL-6-1	TRCN0000059207	CCGGCAGAACGAATTGACAAACAAACTCGAGTTT GTTTGTCAATTCGTTCTGTTTTTG
shIL-6-2	TRCN0000059206	CCGGGACATGTAACAAGAGTAACATCTCGAGATG TACTCTTGTTACATGTCTTTTTG

Table S4. siRNA sequences

Target protein	siRNA ID	Target sequence (5'-3')
p53	VHS40366	Sense: CCAUCCACUACAACUACAUGUGUAA
		Antisense: UUACACAUGUAGUUGUAGUGGAUGG
Caspase-2	HSS141454	Sense: UGCACGUGGCCGACAUGCUGGUUAA
		Antisense: UUAACCAGCAUGUCGGCCACGUGCA
IL-6	s7311	Sense: GAACGAAUUGACAAACAAAtt
		Antisense: UUUGUUUGUCAAUUCGUUCtg

Table S5. List of primers used for ChIP-qPCR

Gene locus	Target region	Sequence (5'-3')
PANX1	ChIP 1 (-1911 to -1768)	Forward: GGGAGAATTTACAAAGGGACC
		Reverse: CTTCCACCCAAAACACTCC
	ChIP 2 (-1441 to -1329)	Forward: TGGTTATGGCAGCTGTGAGA
		Reverse: CCTGAACTCCATCTTCCCA
	ChIP 3 (-1277 to -1169)	Forward: TATGAGCCATCTACGCGGAG
		Reverse: GGGTCCACAGATACAAGCCT
	ChIP 4 (-817 to -652)	Forward: CCTGTGTACCATACCCTCCG
		Reverse: GGAAGCTAAATGTGAGGATCCA
	ChIP 5 (-622 to -488)	Forward: CCTTAATTTCCGTGGACACTGG
		Reverse: GAGCCAGCAGAGAGAGGG
	ChIP 6 (-218 to -119)	Forward: GCCATCCCATCCACAGAAAG
		Reverse: GAAATTCTCCCGTCCCTCCC
3'-UTR (+2675 to +2791)	Forward: TGCTTCATATGGCGTGCTTG	
	Reverse: CCAAATGGCCTCAATTCGGA	
CDKN1A	-2292 to -2169	Forward: GGCTCTGATTGGCTTTCTGG
		Reverse: TCCTACCATCCCCTTCCTCA