

Photoinduced Nitroarenes as Versatile Anaerobic Oxidants for Accessing Carbonyl and Imine Derivatives

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Supporting Information

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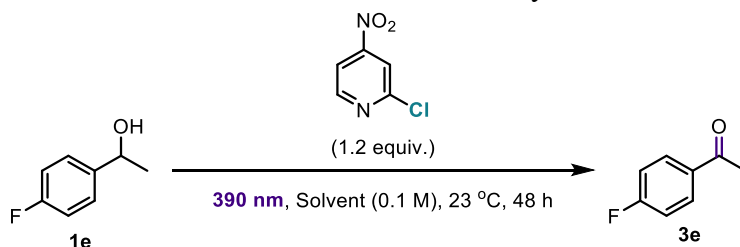
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General Information

All requisite chemicals were purchased from Fisher Scientific, Sigma Aldrich (Merck), Oakwood Chemical (Oakwood Products), Ambeed, TCI, and used without further purification unless otherwise stated. ^1H NMR spectra were recorded at 400 and 500 MHz, ^{13}C NMR spectra were obtained at 126 MHz, and ^{19}F NMR spectra at 377 MHz. All NMR were obtained on a Bruker 400 and 500 MHz Advance spectrometer and are referenced to the deuterated solvent resonance. Chemical shifts (δ) are reported in parts per million (ppm), multiplicity (s = singlet, br = broad, d = doublet, t = triplet, q = quartet, qt = quintet, sext = sextet, m = multiplet), and coupling constants (J) are in Hertz (Hz). All reactions were carried out under an inert nitrogen atmosphere in glassware that had been either flame-dried under vacuum or oven-dried unless otherwise noted. Thin-layer chromatography (TLC) was performed on 250- μm glass-backed silica gel plates and column chromatography were performed using 200–300 mesh silica gel. Solvents were dried and deoxygenated by passing through alumina in a solvent purification system. Deuterated chloroform (CDCl_3), dimethyl sulfoxide (DMSO-d_6), and acetonitrile (CD_3CN) were purchased from Cambridge Isotopes. GC chromatograms were taken on an Agilent 8890 GC with 5977B MSD, and helium as the carrier gas. High-resolution mass spectra (HRMS) were obtained on an Agilent 6224 TOF LC/MS which was acquired through the support of New York University. We utilized 34 W Kessil Lamps with varying wavelengths for our photochemical set ups.

Optimization of the reaction parameters for the oxidation of benzylic alcohols.

Table S1. Solvent screen for the anaerobic oxidation of benzylic alcohols to ketones.

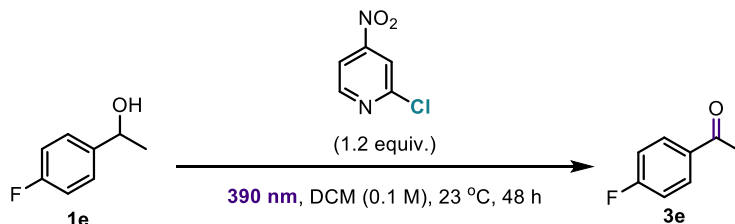


Entry	Conditions	Conversion ^a	3e ^b
1	DCM	95%	88%
2	CHCl ₃	79%	65%
3	CCl ₄	>99%	54%
4	PhCF₃	90%	89%
5	Acetone	75%	74%
6	MeNO ₂	72%	58%
7	MeCN	75%	70%
8	Benzene	50%	50%
9	HFB	72%	50%
10	Cyrene TM	0%	0%

Reactions were performed on a 0.1 mmol scale. ^aConversion of **1e**. ^bDenotes ¹H NMR yields of **3e** using CH₂Br₂ as an internal standard.

*Solvents found to react with the photoexcited nitroarene: DMSO, DCE, 1,4, dioxane, MTBE, THF, DMA, Toluene, IPA, EtOAc, and EtOH.

Table S2. Control experiments for the anaerobic oxidation of alcohols to ketones.



Entry	Conditions	Conversion ^a	3e ^b
1	-20°C	0%	0%
2	0°C	21%	16%
3	dark	0%	0%
4	degassed	>99%	>99% ^c
5	Air	76%	73%
6	N ₂	95%	88%
7	N ₂	>99%	>99% ^c

Reactions were performed on a 0.1 mmol scale. ^aConversion of **1e**. ^bDenotes ¹H NMR yields of **3e** using CH₂Br₂ as an internal standard. ^cUsing 4-methoxyphenyl methyl carbinol.

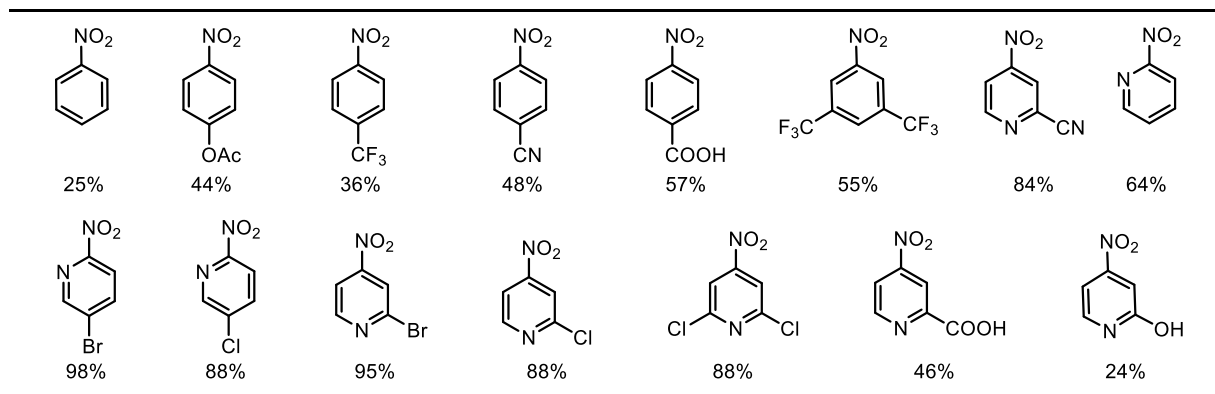
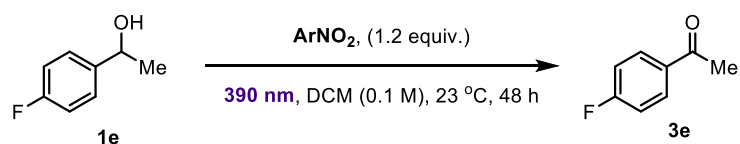
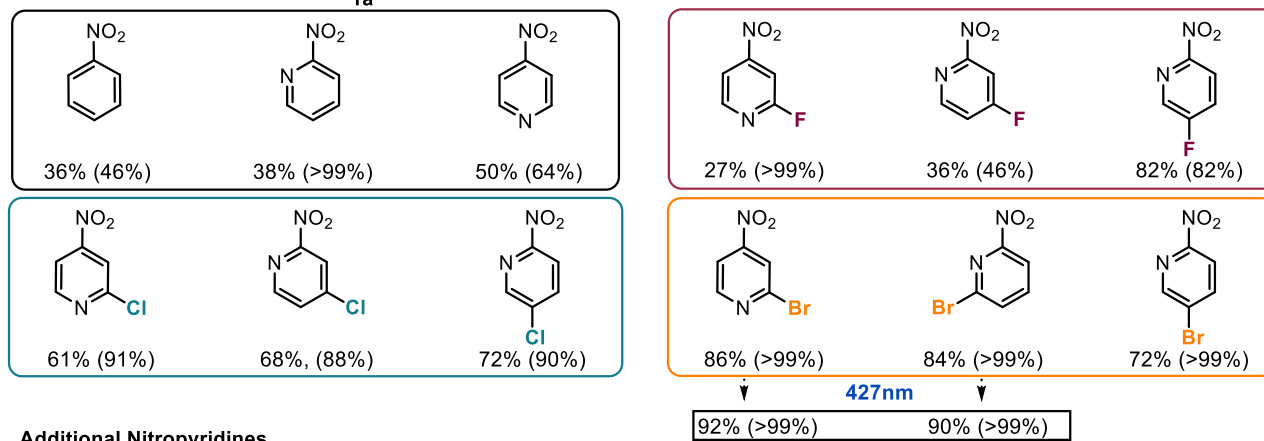
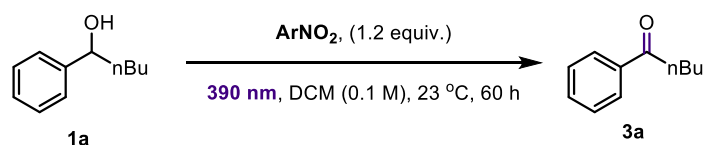


Figure S1. Nitroarene screen for oxidation of benzylic alcohols. Reactions were performed on a 0.1 mmol scale. Yields of **3e** were determined by ¹H NMR using CH₂Br₂ as an internal standard.



Additional Nitropyridines

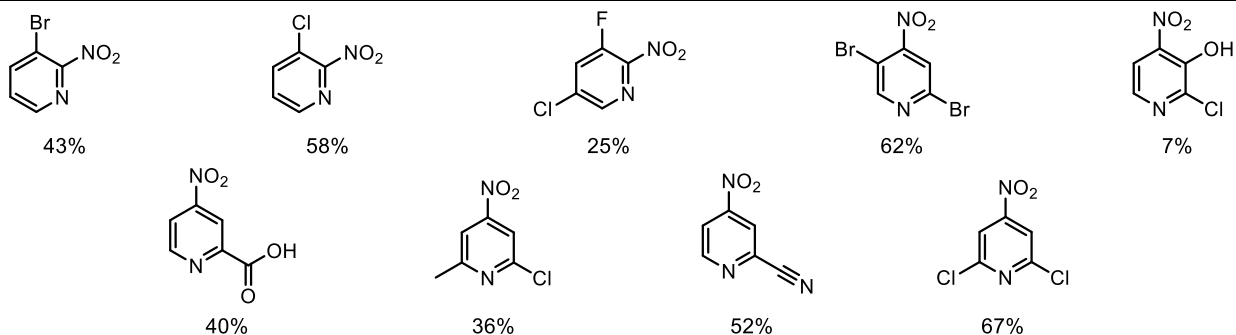
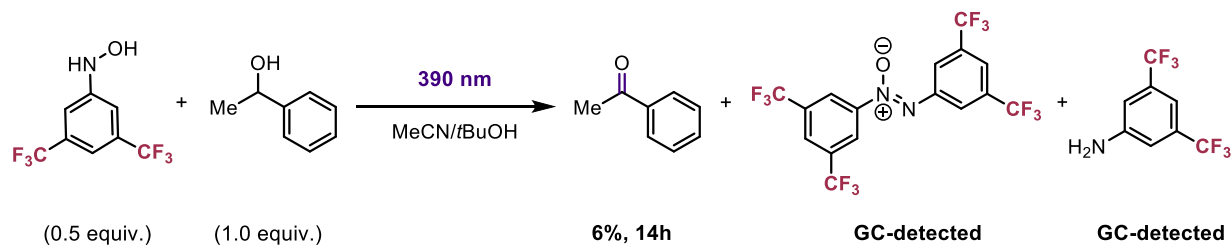
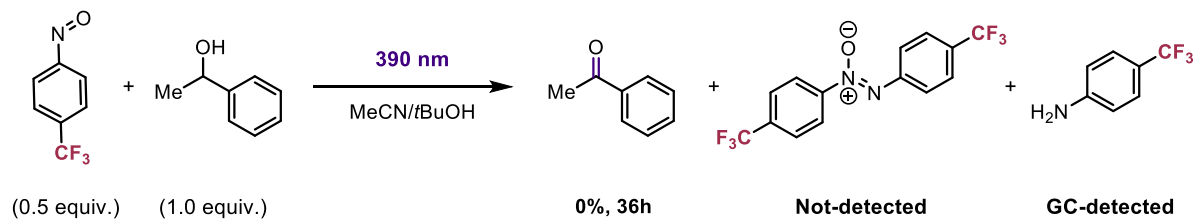


Figure S2. Nitropyridine screen for the oxidation of benzylic alcohols. Reactions performed on a 0.1 mmol scale. Yields of **3d** were determined by ¹H NMR using CH₂Br₂ as an internal standard.

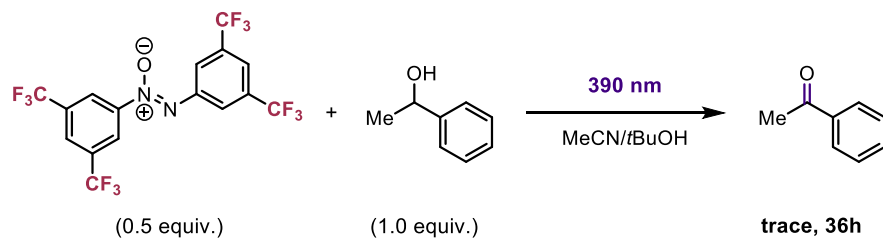
A) Anaerobic Oxidations with hydroxylaniline



B) Anaerobic Oxidations with nitrosoarene



C) Anaerobic Oxidations with azoxyarene



D) Anaerobic Oxidations with aniline

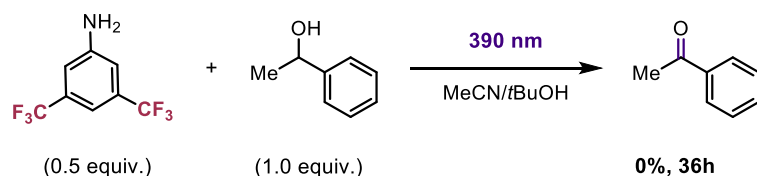
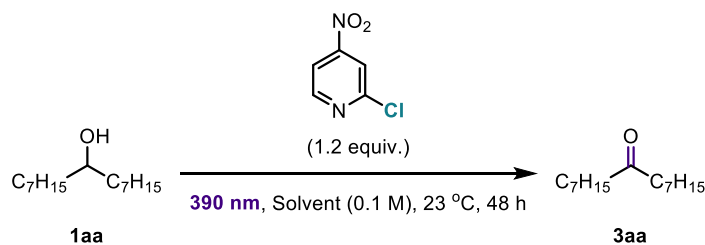


Figure S3. Byproduct Experiments: anaerobic oxidations using synthesized *N*-(3,5-bis(trifluoromethyl)phenyl)hydroxylamine¹ (A), synthesized 1-nitroso-4-(trifluoromethyl)benzene² (B), 1,2-bis(3,5-bis(trifluoromethyl)phenyl)diazene 1-oxide (C), and 3,5-bis(trifluoromethyl)aniline (D) as oxidants. Reactions were performed on a 0.5 mmol scale and the yield of acetophenone was determined by ¹H NMR using CH₂Br₂ as an internal standard.

Optimization of the reaction parameters for the oxidation of unactivated alcohols.

Table S3. Solvent screen for the oxidation of unactivated alcohols to ketones.

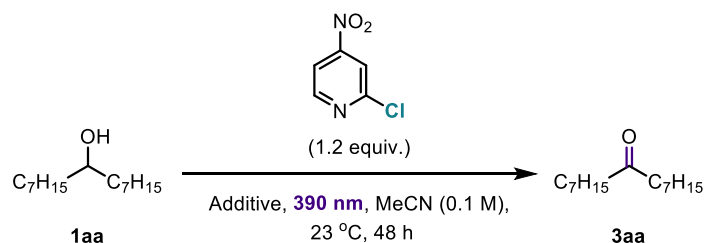


Entry	Solvent	3aa ^a
1	DCM	27%
2	PhCF ₃	30%
3	Benzene	ND
4	Cyrene TM	ND
5	Acetone	39%
6	EtOAc	ND
7	HFIP	ND
8	MeCN	41%

Reactions were performed on a 0.1 mmol scale. ^aDenotes ¹H NMR Yield using CH₂Br₂ as an internal standard.

*Solvents found to react with the photoexcited nitroarene: DMSO, DCE, 1,4, dioxane, MTBE, THF, DMA, Toluene, IPA, EtOAc, and EtOH.

Table S4. Additives screen for the oxidation of unactivated alcohols to ketones.



Entry	Additive (1 equiv.)	3aa ^a
1	-	41%
2	LiOAc	58%
3	KH ₂ PO ₄	40%
4	Quinuclidine	3%
5	Aceclidine	27%
6	Quinuclidinol	3%
7	LiCl	42%
8	LiBF ₄	33%
9	KBr	41%
10	KCl	34%
11	CsF	12%

Reactions were performed at 0.1 mmol scale. ^aDenotes ¹H NMR yield using CH₂Br₂ as an internal standard.

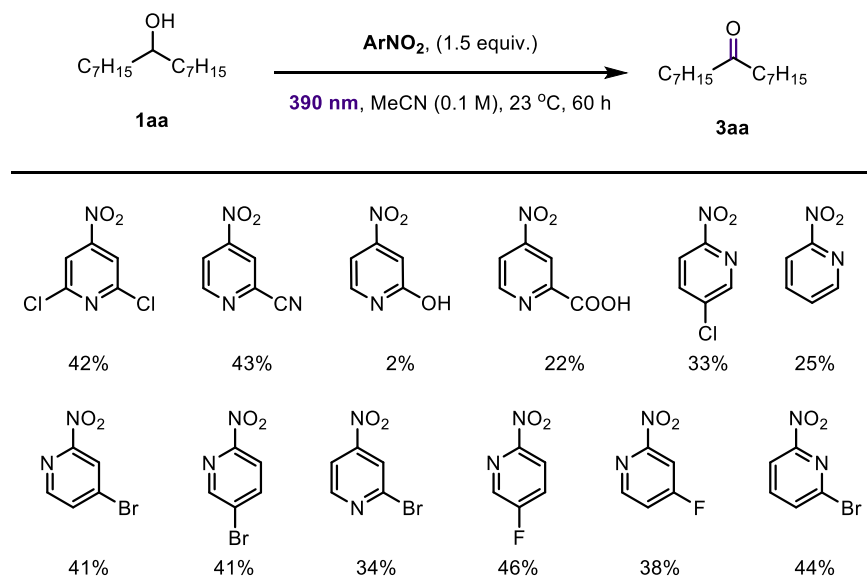
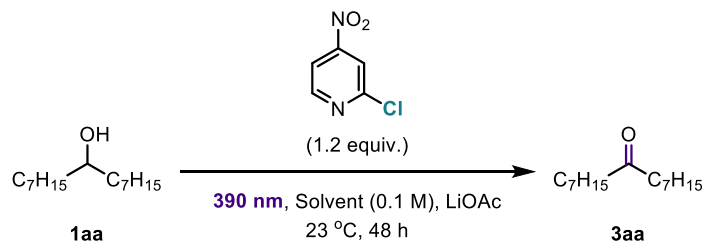


Figure S4. Nitroarene screen for oxidation of unactivated alcohols to ketones. Reactions performed on a 0.1 mmol scale. Yields were calculated using CH_2Br_2 as an internal standard.

Table S5. Solvent Screen for the oxidation of unactivated alcohols to ketones.

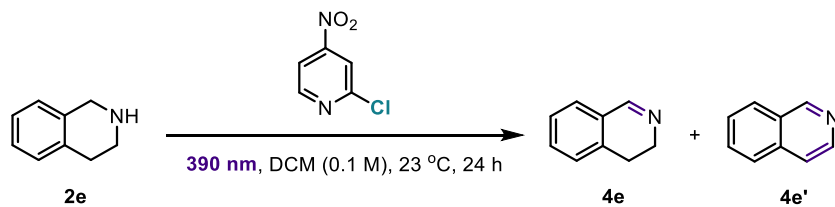


Entry	Time	Solvent	LiOAc equiv.	3aa ^a
1	48 h	MeCN	1	54%
2	48 h	MeCN	0.25	59%
3	60 h	MeCN	0.25	67%
4	60 h	MeCN/ <i>t</i> BuOH (5:1)	-	65%
5	60 h	MeCN/ <i>t</i> BuOH (5:1)	-	69%
6	60 h	MeCN/ <i>t</i> BuOH (5:1)	0.25	77%
7	72 h	MeCN/ <i>t</i> BuOH (5:1)	0.25	84%
8	72 h	MeCN/<i>t</i>BuOH (5:1)	0.25	85% ^b

Reactions were performed at 0.1 mmol scale. ^aDenotes ¹H NMR yield using CH_2Br_2 as an internal standard. ^bUsing 3,5-bis(trifluoromethyl)nitrobenzene.

Optimization of the reaction parameters for the oxidation of amines.

Table S6. Nitropyridine equivalence screen for the oxidation of amines to imines.^a



<i>Equiv.</i>	4e	4e'
0.25	47%	5%
0.5	49%	12%
1.0	41%	18%
1.5	32%	21%
2.0	28%	22%

Reactions were performed on a 0.1 mmol scale. ¹H NMR Yields calculated using CH₂Br₂ as an internal standard.

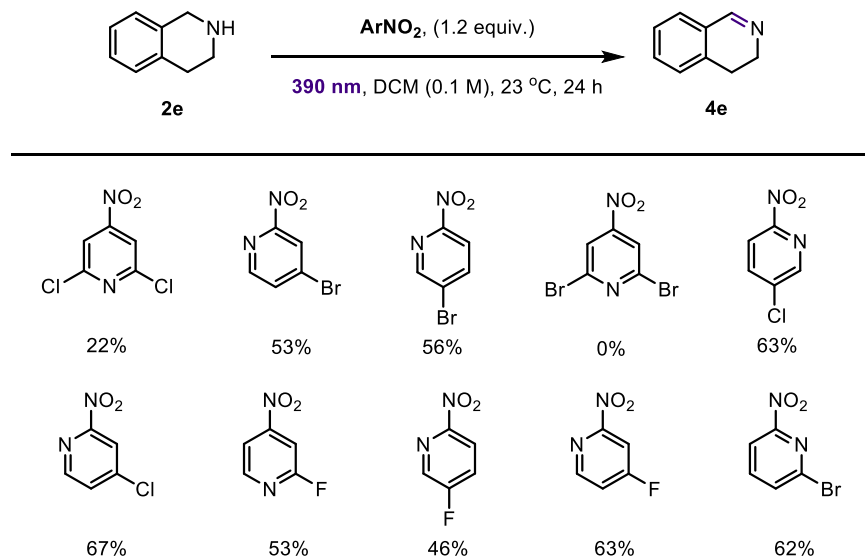
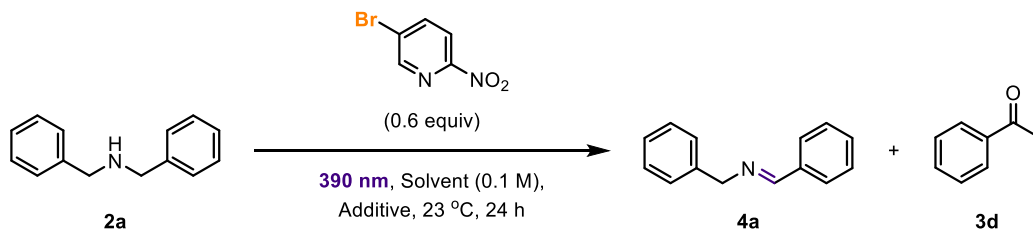


Figure S5. Nitropyridine screen for the oxidation of amines to imines. Reactions were performed on a 0.1 mmol scale. ¹H NMR Yields calculated using CH₂Br₂ as an internal standard.

Table S7. Further wavelength, solvent, time, and additive screen for the oxidation of amines to imines.

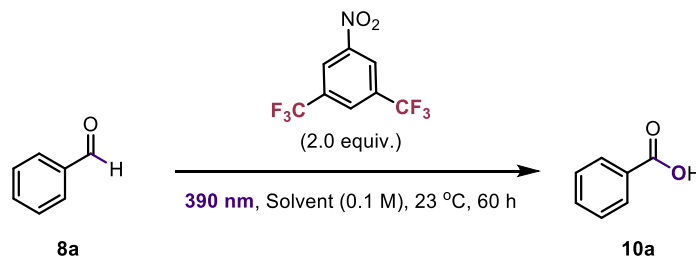


Entry	Solvent	Wavelength th	Time	Additive (0.1 equiv.)	4a ^a	3d ^a
1	DCM	440 nm	72 h	-	84% ^b	16%
2	DCM	440 nm	72 h	-	57%	36%
3	DCM	440 nm	72 h	Pyr	80% ^b	16%
4	DCM	440 nm	72 h	Pyr	71%	22%
5	DCM/ <i>t</i> BuOH (5:1)	440 nm	72 h	-	73%	24%
6	DCM/ <i>t</i> BuOH (5:1)	440 nm	72 h	Pyr	58%	2%
7	DCM/ <i>t</i> BuOH (5:1)	440 nm	72 h	<i>t</i> BuNH ₂	88%	12%
8	PhCF ₃	390 nm	24 h	<i>t</i> BuNH ₂	90%	16%
9	PhCF₃	390 nm	24 h	<i>t</i>BuNH₂	96%^c	<1%

Reactions were performed on a 0.1 mmol scale. ^a¹H NMR yields calculated using CH₂Br₂ as an internal standard. ^bUsing 2-bromo-6-nitropyridine. ^cUsing 1.0 equiv. of nitro.

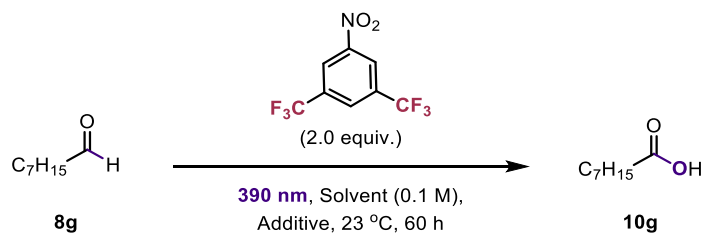
Optimization of the reaction parameters for the oxidation of aldehydes.

Table S8. Optimization for the oxidation of benzylic aldehydes to carboxylic acids.



Entry	Solvent	Conc.	Time	Conversion ^a	Yield ^b
1	DCM	0.1 M	60 h	75%	70%
2	MeCN	0.1 M	60 h	70%	64%
3	MeCN	0.9 M	60 h	90%	80%
4	MeCN	1.0 M	60 h	90%	80%
5	Neat	-	12 h	70%	65%
6	Neat	-	24 h	>99%	93%

Reactions were performed on a 0.5 mmol scale. ^aAmount of **8a** converted. ^b¹H NMR Yields calculated using CH₂Br₂ as an internal standard.

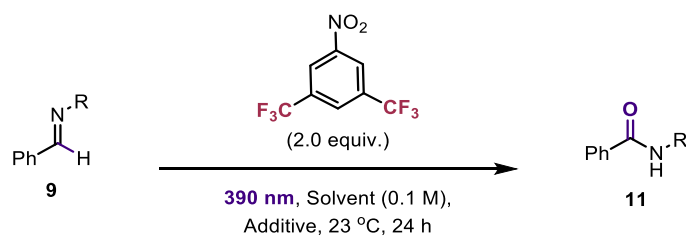
Table S9. Optimization for the oxidation of aliphatic aldehydes to carboxylic acids.

Entry	Nitro (equiv.)	Solvent	Conc.	Additive	Time	Conversion ^a	10g ^b
1	2	Neat	-	-	4 h	-	22%
2	2	Neat	-	-	12 h	-	27%
3	2	DCM	0.1 M	-	60 h	-	10%
4	2	MeCN	0.1 M	-	60 h	-	15%
5	2	MeCN	1.0 M	-	60 h	-	22%
6	2	MeCN	1.0 M	-	24 h	-	32%
7	1.5	MeCN	1.0 M	-	24 h	-	34%
8	1	MeCN	1.0 M	-	24 h	6%	40%
9	0.5	MeCN	1.0 M	-	24 h	30%	22%
10	1	MeCN	1.0 M	<i>t</i> BuOH (0.1 equiv.)	24 h	-	19%
11	1	MeCN	1.0 M	CH ₃ COOH (0.1 equiv.)	24 h	-	37%
12	1	MeCN	1.0 M	H ₂ O (0.1 equiv.)	24 h	-	40%
13	1	MeCN	1.0 M	H ₂ O (1.0 equiv.)	24 h	-	55%

Reactions were performed on a 0.5 mmol scale. ^aAmount of **8g** converted. ^b¹H NMR Yields calculated using CH₂Br₂ as an internal standard.

Optimization of the reaction parameters for the oxidation of imines.

Table S10. Optimization for the oxidation of imines to amides.

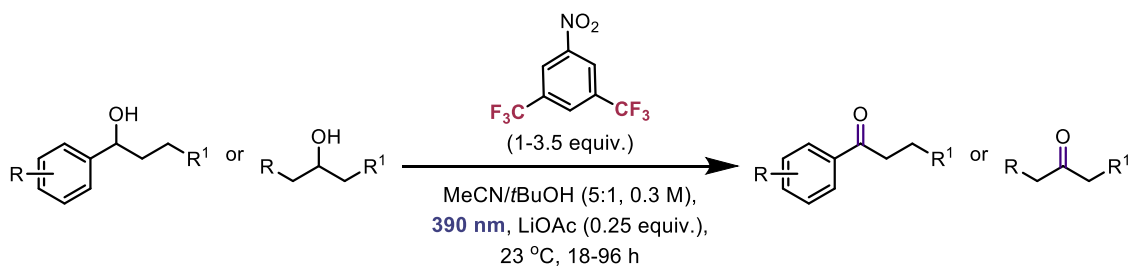


Entry	R	Nitro (equiv.)	Solvent	Conc.	Time	Yield ^a
1	<i>t</i> Bu	2	<i>t</i> BuOH	2 M	24 h	56%
2	<i>t</i> Bu	2	MeCN	2 M	24 h	64%
3	<i>t</i> Bu	2	MeCN	2 M	24 h	81%
4	<i>t</i> Bu	2	DCM/ <i>t</i> BuOH (5:1)	0.4 M	24 h	56%
5	<i>t</i>Bu	2	Neat	-	24 h	91%
6	<i>t</i> Bu	3	Neat	-	24 h	75%
7	Cy	2	DCM/ <i>t</i> BuOH (5:1)	0.4 M	24 h	96%
8	Cy	1.5	DCM/ <i>t</i> BuOH (5:1)	0.4 M	24 h	46%
9	Cy	1	DCM/ <i>t</i> BuOH (5:1)	0.4 M	24 h	50%
10	Cy	0.5	DCM/ <i>t</i> BuOH (5:1)	0.4 M	24 h	18%

Reactions were performed on a 0.5 mmol scale. ¹H NMR Yields calculated using CH₂Br₂ as an internal standard.

General Procedures

General Procedure A. Standard Anaerobic Oxidations for Benzylic and Aliphatic Alcohols.



Oxidation for benzylic/activated alcohols to ketones:

Except when otherwise stated, in an oven-dried 0.5-dram vial equipped with a stir bar was added 3,5-bis(trifluoromethyl)nitrobenzene (**5**) (130 mg, 0.50 mmol, 1.0 equiv.), benzylic alcohol (0.50 mmol, 1.0 equiv.), lithium acetate (0.13 mmol, 0.25 equiv.) and 5:1 MeCN/*t*BuOH (2.3 mL, 0.30 M) under N₂ gas. The reaction vessel was left to stir at 1000 rpm and irradiated under 390 nm Kessil lamps with a cooling fan, for 24-96 h. Completion of the reaction was determined by GCMS analysis. After the reaction was complete, the solvent was removed under a stream of nitrogen. The crude product was purified by column chromatography (0 – 5% EtOAc/Hexane) to afford the ketone products.

Oxidation for unactivated alcohols to ketones:

Except when otherwise stated, in a glove box an oven dried 1.5-dram vial equipped with a stir bar was charged 3,5-bis(trifluoromethyl)nitrobenzene (**5**) (1 – 3 equiv.), alcohol (1.0 mmol, 1 equiv.), and where applicable, lithium acetate (0.40 mmol, 0.25 equiv.). Anhydrous, degassed 5:1 MeCN/*t*BuOH (1.0 mL, 0.36 M) was added. The reaction vessel was sealed and left to stir at 900 rpm, and the reaction was irradiated under 390 nm Kessil lamps with a cooling fan, for 24-96 h. Completion of the reaction was determined by GCMS analysis. The seal vessel was opened and K₂HPO₄ (2.5 mmol, 5.0 equiv.), urea (2.5 mmol, 5.0 equiv.) and formaldehyde (2.5 mmol, 5.0 equiv.) were added in the flask and left to stir for another 5-10 h or until the appearance of white solid. The contents of vial were diluted with EtOAc (20 mL) and washed with deionized H₂O (100 mL) and extracted using EtOAc (3 × 30 mL). Combined organic phases were dried with NaHSO₄ and concentrated in vacuo. The crude product was then purified by column chromatography (2.5 – 5% Acetone/ Hexane) to afford the ketone products.

Analysis of volatile ketone products:

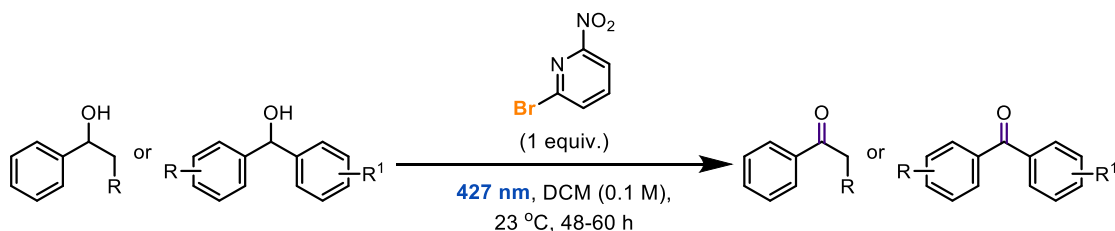
Owing to the volatility of products **3ac**, **3ad**, **3ae**, and **3af**, these products were not isolated. The reaction efficiency was determined by crude ¹H NMR yield using CH₂Br₂ as an internal standard in CD₃CN. The corresponding ¹H NMR yield spectra for **3ac**, **3ad**, **3ae**, and **3af**, are provided in the NMR spectra section. To validate that the measured ¹H NMR yields were accurate, the crude mixtures of **3ac**, **3ad**, **3ae**, and **3af** were subjected to hydrazone derivatization (see the procedure below). It was found that the isolated yields of the hydrazone-derivatized products (**3ac'**, **3ad'**,

3ae', and **3af'**) were within the error of the crude ^1H NMR yield measurements. The ^1H NMR and ^{13}C NMR spectra of the hydrazone-derivatized products are provided in the characterization and NMR spectra sections.

Hydrazone derivatization for the isolation of volatile ketones:

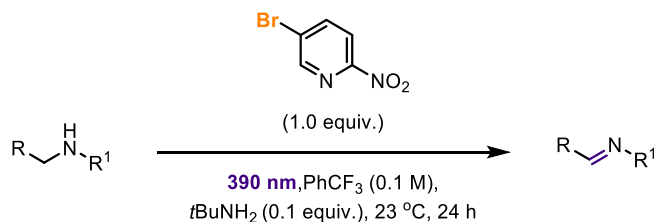
Upon completion, the reaction mixture featuring 1.16 mmol of the volatile ketone (**3ac**, **3ad**, **3ae**, and **3af**) was mixed with H_2O (5 mL), HCl (600 μL , 6M aq.), then 2,4-dinitrophenylhydrazine (237 mg, 1.19 mmol, 1.03 equiv.) and stirred. The formation of the hydrazone was indicated by the appearance of reddish precipitates. Completion of the reaction was monitored by GCMS in 3 hours. The slurry was diluted with H_2O (50 mL), filtered, and stirred in 6 M HCl (20 ml) for additional 30 minutes. The filtered precipitate was washed with water, redissolved in dichloromethane, and dried with NaHSO_4 . The crude product was then purified by column chromatography to afford the hydrazone products.

General Procedure B. Standard Anaerobic Oxidations for α -Substituted and Dibenzyllic Alcohols.



Except when otherwise stated, in an oven-dried 2-dram vial equipped with a stir bar was added 2-bromo-6-nitropyridine (**6**) (102 mg, 0.500 mmol, 1.0 equiv.), target alcohol (0.50 mmol, 1.0 equiv.), and DCM (5.0 mL, 0.10 M) under N_2 gas. The reaction vessel was left to stir at 1000 rpm and irradiated under 427 nm Kessil lamps with a cooling fan, for 48-60 h. Completion of the reaction was determined by GCMS analysis. After the reaction was completed, the solvent was removed under a stream of nitrogen. The crude product was purified by column chromatography (0 – 5% EtOAc/Hexane) to afford the ketone product.

General Procedure C. Standard Anaerobic Oxidations for amines to imines.



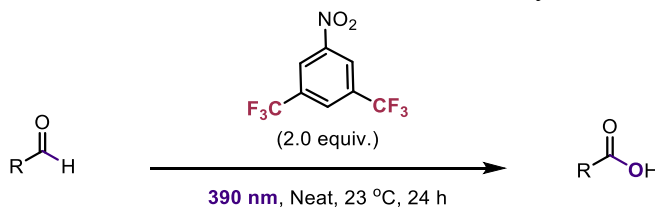
Except when otherwise stated, in an oven-dried 2-dram vial equipped with a stir bar was added 5-bromo-2-nitropyridine (**7**) (102 mg, 0.5 mmol, 1.0 equiv.), target amine (0.5 mmol, 1 equiv.), $t\text{BuNH}_2$ (0.05 mmol, 0.1 equiv.), and the vial was purged with N_2 flow for 15 min, followed by the addition of dry PhCF_3 (5 mL, 0.1 M). The reaction vessel was left to stir at 1000 rpm and

irradiated under 390 nm Kessil lamps with a cooling fan, for 24 h. Completion of the reaction was determined by GCMS analysis. After the reaction was completed, the solvent was removed under a stream of nitrogen. The crude product was purified by column chromatography (0 – 5% NEt₃/Hexane) to afford the imine product.

Analysis of hydrolytically unstable imine products:

Imine products **4c**, **4d**, **4e**, **4g**, **4h**, and **4i** were found to be hydrolytically unstable. Numerous attempts to isolate the above compounds using NEt₃-treated silica gel, neutral and basic alumina, and Devisil failed to provide any appreciable products. The reaction efficiency for **4c**, **4d**, **4e**, and **4g** was determined by crude ¹H NMR yield using CH₂Br₂ as an internal standard in CD₃CN. The reaction efficiency for **4h** and **4i** was determined by crude ¹H NMR yield using CH₂Cl₂ as an internal standard in CD₃CN. The corresponding ¹H NMR yield spectra for **4c**, **4d**, **4e**, **4g**, **4h**, and **4i** are provided in the NMR spectra section. Unfortunately, attempts to derivatize the crude products were unsuccessful.

General Procedure D. Standard Anaerobic Oxidation for Aldehydes to Acids.



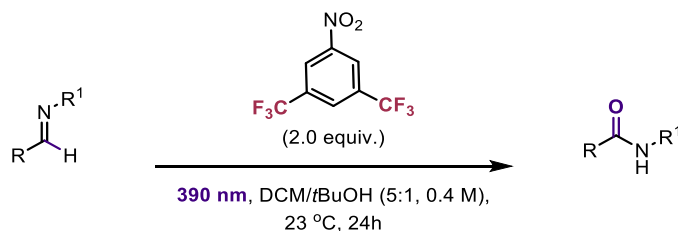
Oxidation for aromatic aldehydes to acids:

Except when otherwise stated, in an oven-dried 0.5-dram vial equipped with a stir bar was purged with N₂ flow for 15 min, 3,5-bis(trifluoromethyl)nitrobenzene (**5**) (170 μL, 1.00 mmol, 2.00 equiv.) and aldehyde (0.50 mmol, 1.0 equiv.) were added. The reaction vessel was left to stir at 1000 rpm and irradiated under 390 nm Kessil lamps with a cooling fan, for 24-36 h. Completion of the reaction was determined by GCMS analysis. After the reaction was completed, the mixture was concentrated down, and the crude product was then purified by column chromatography (0 – 50% EtOAc/Hexane) to afford the carboxylic acid product.

Oxidation for aliphatic aldehydes to acids:

Except when otherwise stated, in a flame- or oven-dried 0.5-dram vial equipped with a stir bar, 3,5-bis(trifluoromethyl)nitrobenzene (**5**) (84 μL, 0.50 mmol, 1.0 equiv.) and aldehyde (0.50 mmol, 1.0 equiv.) were added. The reaction vial was purged with N₂ flow for 15 min followed by the addition of H₂O (0.5 mmol, 1 equiv.), and CH₃CN (400 μL, 1.0 M). The vial was left to stir at 1000 rpm and irradiated under 390 nm Kessil lamps with a cooling fan, for 24-36 h. Completion of the reaction was determined by GCMS analysis. After the reaction was completed, the mixture was concentrated down, and the crude product was then purified by column chromatography (0 – 50% EtOAc/Hexane) to afford the carboxylic acid product.

General Procedure E. Standard Anaerobic Oxidation for imines to amide.



Except when otherwise stated, in a flame- or oven-dried 0.5-dram vial equipped with a stir bar was purged with N₂ flow for 15 min, 3,5-bis(trifluoromethyl)nitrobenzene (**5**) (170 μL, 0.50 mmol, 2.0 equiv.), aldehyde (0.5 mmol, 1.0 equiv.), and amine (0.50 mmol, 1.0 equiv.) were added, followed by the addition of 5:1 DCM/*t*BuOH (1.2 mL, 0.40 M). The reaction mixture was allowed to stir until imine formation was complete as determined by GCMS analysis. Then, the vessel was irradiated under a 390 nm Kessil lamp with a cooling fan, the stir rate was set to 1000 rpm, and the reaction was irradiated for 24-36 h. Completion of the reaction was determined by GCMS analysis. The crude product was then purified by column chromatography (0 – 50% EtOAc/Hexane) to afford the amide product.

General Procedure F. Continuous-Flow Conditions for the Photoinduced Anaerobic Oxidation Reactions.

The setup for the flow reactor used is as follows: 10 ft. of 0.03-inch diameter Fluorinated Ethylene Propylene (FEP) tubing was coiled around a 100 mL glass bottle (**Figure S6**). Two 390 nm Kessil lamps (34 W each) were placed approximately 3.0 cm from the coil of tubing. An electric fan was placed behind to prevent the tubing from overheating. A syringe pump was connected to control the flow rate of the reaction.

To a dry 1-dram vial was added nitroarene (0.75 - 2.0 equiv.) and substrate (1.0 equiv.). The vial was sealed with a septum cap and purged with nitrogen for 5 minutes. Then solvent (2.0 mL) was added, and the mixture was taken up in a 5.0 mL syringe which was attached to the FEP tubing. The flow rate was set using the syringe pump, the blue LEDs were switched on and the reaction flowed through (residence time of 3-5 h). A 2-dram vial was used to collect the reaction solution. After the given reaction time, the mixture was concentrated down and CDCl₃ (1.0 mL) and CH₂Br₂ (1.0 equiv.) was added as an internal ¹H NMR standard. The NMR yield of the oxidation product was recorded.

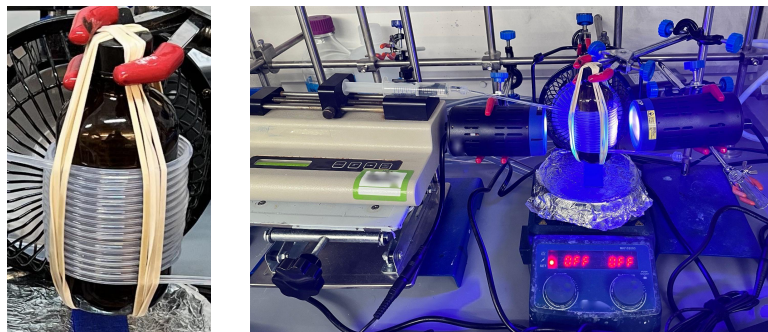
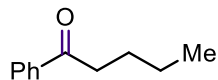


Figure S6. Images of the photochemical continuous-flow setup

Characterization of oxidized products

Characterization data of ketones

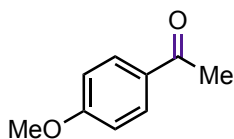


1-phenylpentan-1-one (**3a**)

Following General Procedure A, prepared from 0.50 mmol of 1-phenylpentan-1-ol (**1a**) and 3.0 equiv. of **5** in 24 h, the title compound was isolated via flash column chromatography (0 – 20% EtOAc/Hexane) as a clear oil (73 mg, 91% yield). All analytical data for **3a** was in accordance with literature data.³

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 7.99 – 7.93 (m, 2H), 7.58 – 7.52 (m, 1H), 7.49 – 7.43 (m, 2H), 3.00 – 2.94 (m, 2H), 1.77 – 1.67 (m, 2H), 1.46 – 1.37 (m, 2H), 0.96 (t, *J* = 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 200.8, 137.3, 133.0, 128.7, 128.2, 38.5, 26.6, 22.6, 14.1.

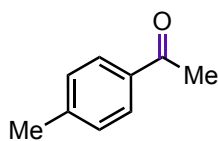


1-(4-methoxyphenyl)ethan-1-one (**3b**)

Following General Procedure A, prepared from 1.00 mmol of 1-(4-methoxyphenyl)ethan-1-ol (**1b**) in 24 h, the title compound was isolated via flash column chromatography (0 – 5% EtOAc/Hexane) as a yellow solid (141 mg, 98% yield). All analytical data for **3b** was in accordance with literature data.⁴

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 7.94 (d, *J* = 8.9 Hz, 2H), 6.93 (d, *J* = 9.0 Hz, 2H), 3.87 (s, 3H), 2.56 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 196.9, 163.6, 130.7, 130.5, 113.8, 55.6, 26.5.

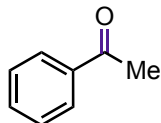


1-(*p*-tolyl)ethan-1-one (**3c**)

Following General Procedure A, prepared from 0.50 mmol of 1-(*p*-tolyl)ethan-1-ol (**1c**) and 3.0 equiv. of **5** in 36 h. The title compound was isolated via flash column chromatography (0 – 5% EtOAc/Hexane) as a yellow oil (56 mg, 82% yield). All analytical data for **3c** was in accordance with literature data.⁵

¹H NMR (500 MHz, CD₃CN) (δ, ppm): 7.86 (d, *J* = 8.1 Hz, 2H), 7.31 (d, *J* = 7.9 Hz, 2H), 2.53 (s, 3H), 2.40 (s, 3H).

¹³C NMR (126 MHz, CD₃CN) (δ, ppm): 198.6, 144.9, 135.8, 130.2, 129.3, 26.9, 21.6.

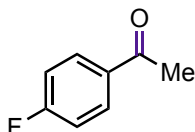


Acetophenone (**3d**)

Following General Procedure A, prepared from 1.0 mmol of 1-phenylethan-1-ol (**1d**) in 36 h. The title compound was isolated via flash column chromatography (0 – 5% EtOAc/Hexane) as a yellow oil (96 mg, 80% yield). All analytical data for **3d** was in accordance with literature data.⁴

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 7.96 (dd, *J* = 8.4, 1.3 Hz, 2H), 7.59 – 7.54 (m, 1H), 7.47 (dd, *J* = 8.4, 7.0 Hz, 2H), 2.61 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 198.3, 137.3, 133.3, 128.72, 128.71, 128.5, 128.5, 128.5, 128.4, 26.7.



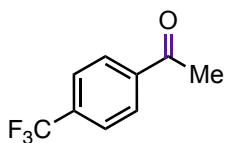
1-(4-Fluorophenyl)ethan-1-one (**3e**)

Following General Procedure A, prepared from 1.00 mmol of 1-(4-fluorophenyl)ethan-1-ol (**1e**) and 3.00 equiv. of **5** in 48 h. The title compound was isolated via flash column chromatography (0 – 5% EtOAc/Hexane) as a yellow oil (134 mg, 97% yield). All analytical data for **3e** was in accordance with literature data.⁶

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 8.02 – 7.94 (m, 2H), 7.17 – 7.09 (m, 2H), 2.59 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 196.5, 165.8 (d, *J* = 255.5 Hz), 133.6 (d, *J* = 3.0 Hz), 131.0 (d, *J* = 9.0 Hz), 115.7 (d, *J* = 22.2 Hz), 26.6.

¹⁹F NMR (377 MHz, CDCl₃) (δ, ppm): -63.13.

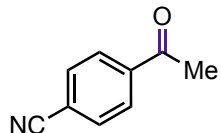


1-(4-(Trifluoromethyl)phenyl)ethan-1-one (**3f**)

Following General Procedure A, prepared from 0.50 mmol of 1-(4-(trifluoromethyl)phenyl)ethan-1-ol (**1f**) and 2.0 equiv. of **5** in 36 h. The title compound was isolated via flash column chromatography (0 – 5% EtOAc/Hexane) as a yellow oil (65 mg, 70% yield). All analytical data for **3f** was in accordance with literature data.⁴

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 8.06 (d, *J* = 8.2 Hz, 2H), 7.74 (d, *J* = 8.1 Hz, 2H), 2.65 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 197.1, 139.8, 134.5 (q, *J* = 32.8 Hz), 128.8, 125.8 (q, *J* = 3.8 Hz), 123.7 (q, *J* = 273.4 Hz), 27.0.

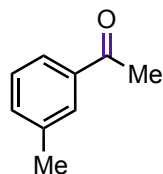


4-Acetylbenzotrile (**3g**)

Following General Procedure A, prepared from 0.50 mmol of synthesized 4-(1-hydroxyethyl)benzotrile (**1g**)^{7a} in 36 h. The title compound was isolated via flash column chromatography (0 – 20% EtOAc/Hexane) as a white solid (70 mg, 98% yield). All analytical data for **3g** was in accordance with literature data.^{7b}

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 8.04 (d, *J* = 8.4 Hz, 2H), 7.78 (d, *J* = 8.4 Hz, 2H), 2.65 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 196.7, 140.1, 132.7, 128.8, 118.1, 116.6, 26.9.

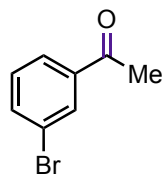


1-(*m*-Tolyl)ethan-1-one (**3h**)

Following General Procedure A, prepared from 0.50 mmol of 1-(*m*-tolyl)ethan-1-ol (**1h**) and 3.0 equiv. of **5** in 36 h, the title compound was isolated via flash column chromatography (0 – 5% EtOAc/Hexane) as a yellow oil (48 mg, 70% yield). All analytical data for **3h** was in accordance with literature data.⁸

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 7.79 – 7.73 (m, 2H), 7.40 – 7.33 (m, 2H), 2.59 (s, 3H), 2.41 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 198.6, 138.5, 137.3, 134.0, 128.9, 128.6, 125.7, 26.8, 21.5.

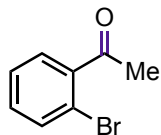


1-(3-Bromophenyl)ethan-1-one (**3i**)

Following General Procedure A, prepared from 0.50 mmol of 1-(3-bromophenyl)ethan-1-ol (**1i**) and 2.0 equiv. of **5** in 36 h. The title compound was isolated via flash column chromatography (0 – 5% EtOAc/Hexane) as a yellow oil (34 mg, 35% yield). All analytical data for **3i** was in accordance with literature data.⁹

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 8.08 (q, *J* = 1.7 Hz, 1H), 7.87 (dq, *J* = 7.7, 1.5 Hz, 1H), 7.71 – 7.66 (m, 1H), 7.34 (td, *J* = 7.8, 1.8 Hz, 1H), 2.59 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 196.8, 138.9, 136.1, 131.5, 130.3, 127.0, 123.1, 26.8.

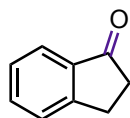


1-(2-Bromophenyl)ethan-1-one (**3k**)

Following General Procedure A, prepared from 0.50 mmol of 1-(2-bromophenyl)ethan-1-ol (**1k**) and 3.0 equiv. of **5** in 24 h, the title compound was isolated via flash chromatography (0 – 5 % EtOAc/Hexane) as a yellow oil (46 mg, 46% yield). All analytical data for **3k** was in accordance with the literature data.¹⁰

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 7.62 (dd, *J* = 8.1, 1.1 Hz, 1H), 7.47 (dd, *J* = 7.6, 1.8 Hz, 1H), 7.37 (td, *J* = 7.5, 1.2 Hz, 1H), 7.30 (td, *J* = 7.7, 1.8 Hz, 1H), 2.64 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 201.6, 141.6, 134.0, 132.0, 129.1, 127.6, 119.1, 30.5.

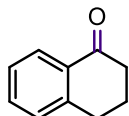


2,3-Dihydro-1H-inden-1-one (**3l**)

Following General Procedure A, prepared from 0.50 mmol of 1,2,3,4-tetrahydronaphthalen-1-ol (**1l**) in 36 h, the title compound was isolated via flash chromatography (0 – 5% EtOAc/Hexane) as a brown oil (67 mg, 51% yield). All analytical data for **3l** was in accordance with the literature data.¹¹

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 7.75 (dt, *J* = 7.7, 1.0 Hz, 1H), 7.58 (td, *J* = 7.4, 1.3 Hz, 1H), 7.47 (dp, *J* = 7.7, 1.0 Hz, 1H), 7.36 (ddt, *J* = 8.0, 7.2, 0.9 Hz, 1H), 3.18 – 3.10 (m, 2H), 2.72 – 2.64 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 207.2, 155.3, 137.2, 134.7, 127.4, 126.8, 123.8, 36.3, 25.9.

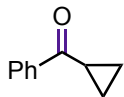


3,4-Dihydronaphthalen-1(2H)-one (**3m**)

Following General Procedure A, prepared from 0.50 mmol of 1,2,3,4-tetrahydronaphthalen-1-ol (**1m**) in 36 h, the title compound was isolated via flash chromatography (0 – 5% EtOAc/Hexane) as a colorless oil (70 mg, 48% yield). All analytical data for **3m** was in accordance with the literature data.¹¹

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 8.03 (dd, *J* = 7.9, 1.5 Hz, 1H), 7.47 (td, *J* = 7.5, 1.5 Hz, 1H), 7.31 (t, *J* = 7.5 Hz, 1H), 7.25 (d, *J* = 7.9 Hz, 1H), 2.97 (t, *J* = 6.1 Hz, 2H), 2.66 (dd, *J* = 7.3, 5.7 Hz, 2H), 2.15 (q, *J* = 6.3 Hz, 2H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 198.5, 144.6, 133.5, 132.8, 128.9, 127.3, 126.8, 39.3, 29.9, 23.4.

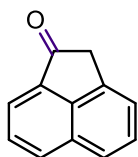


Cyclopropyl(phenyl)methanone (**3n**)

Following General Procedure A, prepared from 1.00 mmol of cyclopropyl(phenyl)methanol (**1n**) in 36 h, the title compound was isolated via flash column chromatography (0 – 5% EtOAc/Hexane) as a yellow oil (142 mg, 98% yield). All analytical data for **3n** was in accordance with literature data.¹²

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 8.05 – 7.96 (m, 2H), 7.61 – 7.53 (m, 1H), 7.52 – 7.42 (m, 2H), 2.68 (tt, *J* = 7.8, 4.6 Hz, 1H), 1.28 – 1.22 (m, 2H), 1.08 – 1.01 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 200.8, 138.2, 132.9, 128.6, 128.2, 17.3, 11.8.

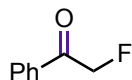


Acenaphthylen-1(2H)-one (**3o**)

Following General Procedure B, prepared from 0.50 mmol of 1,2-dihydroacenaphthylen-1-ol (**1o**) in 36 h, the title compound was isolated via flash column chromatography (0 – 5% EtOAc/Hexane) as a yellow solid (67 mg, 86% yield). All analytical data for **3o** was in accordance with literature data.¹³

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 8.10 (dd, *J* = 8.1, 0.7 Hz, 1H), 7.97 (dd, *J* = 7.1, 0.8 Hz, 1H), 7.83 (dd, *J* = 8.5, 0.8 Hz, 1H), 7.72 (dd, *J* = 8.1, 7.0 Hz, 1H), 7.60 (dd, *J* = 8.4, 6.8 Hz, 1H), 7.47 (dd, *J* = 6.9, 1.0 Hz, 1H), 3.83 (s, 2H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 203.0, 143.0, 135.1, 134.7, 131.5, 131.0, 128.5, 128.1, 124.0, 121.5, 121.1, 42.1.

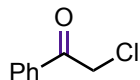


2-Fluoro-1-phenylethan-1-one (**3p**)

Following General Procedure B, prepared from 0.20 mmol of synthesized 2-fluoro-1-phenylethan-1-ol (**1p**)¹⁴ at 390 nm in 96 h, the title compound was isolated via flash column chromatography (0 – 25% DCM/Hexane) as a colorless liquid (27 mg, 93% yield). All analytical data for **3p** was in accordance with literature data.¹⁵

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 7.95 – 7.87 (m, 2H), 7.63 (ddt, *J* = 7.9, 6.9, 1.3 Hz, 1H), 7.57 – 7.45 (m, 2H), 5.53 (d, *J* = 46.9 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 193.6 (d, *J* = 15.2 Hz), 134.29, 133.89, 129.09, 128.05, 128.02, 83.7 (d, *J* = 183.8 Hz).

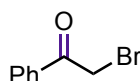


2-Chloro-1-phenylethan-1-one (3q)

Following General Procedure B, prepared from 0.50 mmol of 2-chloro-1-phenylethan-1-ol (**1q**) and 2-bromo-4-nitropyridine in 96 h, the title compound was isolated via flash column chromatography (0 – 25% DCM/Hexane) as a white solid (65 mg, 84% yield). All analytical data for **3q** was in accordance with literature data.⁴

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 8.11 – 7.90 (m, 2H), 7.75 – 7.57 (m, 1H), 7.55 – 7.38 (m, 2H), 4.72 (s, 2H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 191.1, 134.3, 134.0, 128.9, 128.6, 46.0.

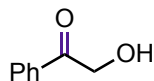


2-Bromo-1-phenylethan-1-one (3r)

Following General Procedure B, prepared from 0.50 mmol of 2-bromo-1-phenylethan-1-ol (**1r**) and 2-bromo-4-nitropyridine in 48 h, the title compound was isolated via flash column chromatography (0 – 25% DCM/Hexane) as a colorless liquid (78 mg, 78% yield). All analytical data for **3r** was in accordance with literature data.⁴

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 8.02 – 7.96 (m, 2H), 7.67 – 7.57 (m, 1H), 7.53 – 7.45 (m, 2H), 4.46 (s, 2H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 191.4, 134.11, 134.09, 129.1, 129.0, 31.1.

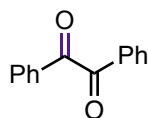


2-Hydroxy-1-phenylethan-1-one (3s)

Following General Procedure A, prepared from 0.50 mmol of 1-phenylethane-1,2-dioland (**1s**) and 1 equiv. of **5** in MeCN/H₂O (1:1, 0.30 M), in 48 h, the title compound was isolated via flash column chromatography (0 – 25% DCM/Hexane) as a white solid (32 mg, 47% yield). All analytical data for **3s** was in accordance with literature data.⁴

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 7.96 – 7.89 (m, 2H), 7.70 – 7.59 (m, 1H), 7.54 – 7.46 (m, 2H), 4.89 (s, 2H), 3.50 (br, 1H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 198.5, 134.5, 133.5, 129.1, 127.9, 65.6.



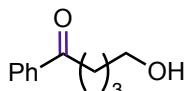
Benzil (3t)

Following General Procedure A, prepared from 0.50 mmol of 2-hydroxy-1,2-diphenylethan-1-one (**1t**) and 1.0 equiv. of **5** in 24h, the title compound was isolated via flash chromatography (0 – 5 %

EtOAc/Hexane) as a yellow solid (67 mg, 64% yield). All analytical data for **3t** was in accordance with the literature data.¹⁶

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 8.02 – 7.94 (m, 4H), 7.74 – 7.60 (m, 2H), 7.57 – 7.45 (m, 4H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 194.7, 135.0, 133.2, 130.1, 129.2.

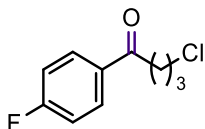


5-Hydroxy-1-phenylpentan-1-one (**3u**)

Following General Procedure A, prepared from 0.50 mmol of 1-phenylpentane-1,5-diol (**1u**) and 1 equiv. of **5** in MeCN/H₂O (1:1, 0.30 M), in 36 h, the title compound was isolated via flash column chromatography (0 – 5% Acetone/DCM) as a colorless oil (37 mg, 42% yield). All analytical data for **3u** was in accordance with literature data.¹⁷

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 8.05 – 7.91 (m, 2H), 7.59 – 7.53 (m, 1H), 7.50 – 7.41 (m, 2H), 3.68 (t, *J* = 6.3 Hz, 2H), 3.03 (t, *J* = 7.1 Hz, 2H), 1.90 – 1.81 (m, 2H), 1.77 – 1.54 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 200.5, 137.1, 133.2, 128.8, 128.23, 128.20, 62.6, 38.3, 32.4, 20.3.

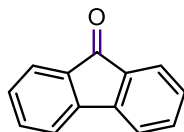


4-Chloro-1-(4-fluorophenyl)butan-1-one (**3v**)

Following General Procedure A, prepared from 0.500 mmol of synthesized 4-chloro-1-(4-fluorophenyl)butan-1-one (**1v**)^{18a} and 3.00 equiv. of **5** in 24 h, the title compound was isolated via flash chromatography (0 – 5% EtOAc/Hexane) as a yellow oil (120 mg, 61% yield). All analytical data for **3v** was in accordance with the literature data.^{18b}

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 8.05 – 7.96 (m, 2H), 7.14 (t, *J* = 8.6 Hz, 2H), 3.68 (t, *J* = 6.2 Hz, 2H), 3.15 (t, *J* = 6.9 Hz, 2H), 2.22 (p, *J* = 6.7 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 197.5, 166.0 (d, *J* = 254.9 Hz), 133.3 (d, *J* = 3.0 Hz), 130.8 (d, *J* = 9.3 Hz), 115.9 (d, *J* = 21.9 Hz), 44.8, 35.3, 26.8.

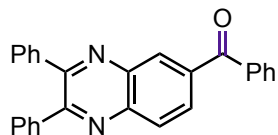


9H-Fluoren-9-one (**3w**)

Following General Procedure B, prepared from 0.50 mmol of 9H-fluoren-9-ol (**1w**) and in 36 h, the title compound was isolated via flash column chromatography (0 – 5% EtOAc/Hexane) as a yellow solid (67 mg, 75% yield). All analytical data for **3w** was in accordance with literature data.¹³

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 7.67 (dd, *J* = 7.4, 1.0 Hz, 1H), 7.53 (d, *J* = 7.4 Hz, 1H), 7.49 (td, *J* = 7.4, 1.2 Hz, 1H), 7.30 (td, *J* = 7.3, 1.2 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 194.1, 144.6, 134.8, 134.3, 129.2, 124.6, 120.5.

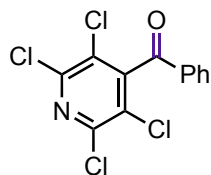


(2,3-Diphenylquinoxalin-6-yl)(phenyl)methanone (**3x**)

Following General Procedure B, prepared from 0.17 mmol of synthesized (2,3-diphenylquinoxalin-6-yl)(phenyl)methanol (**1x**)^{18a, 19a} in 36h, the title compound was isolated via flash column chromatography (0 – 5% EtOAc/Hexane) as a white solid (55 mg, 85% yield). All analytical data for **3x** was in accordance with literature data.^{19b}

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 8.54 (dd, *J* = 1.7, 0.8 Hz, 1H), 8.32 – 8.25 (m, 2H), 7.93 – 7.89 (m, 2H), 7.67 – 7.62 (m, 1H), 7.59 – 7.50 (m, 6H), 7.43 – 7.31 (m, 6H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 195.8, 155.2, 154.6, 143.0, 140.2, 138.7, 138.6, 138.3, 137.2, 132.8, 132.5, 130.2, 129.9, 129.8, 129.7, 129.7, 129.3, 129.2, 128.6, 128.4, 128.3.



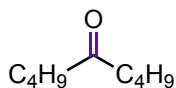
(Perchloropyridin-4-yl)(phenyl)methanone (**3y**)

Following General Procedure A, prepared from 0.14 mmol of perchloropyridin-4-yl)(phenyl)methanol (**1y**) and 3 eq of nitropyridine **6**, the title compound was isolated via flash chromatography (0 – 5% EtOAc/Hexane) as a white solid (5.0 mg, 11% yield). All analytical data for **3y** was in accordance with the literature data.²⁰

¹H NMR (600 MHz, CDCl₃) (δ, ppm): 7.80 (dd, *J* = 1.3 Hz, 2H), 7.74-7.68 (m, 1H), 7.55 (dd, *J* = 8.4, 7.4 Hz, 2H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 188.8, 149.4, 147.0, 135.6, 133.5, 129.7, 129.6, 126.5.

HRMS (ESI-TOF): *m/z* calculated for C₁₂H₅Cl₄NO [M]⁺ = 318.9125, found 318.9141.

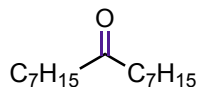


Nonan-5-one (**3z**)

Following General Procedure A, prepared from 0.500 mmol of nonan-5-ol (**1z**) and 1.10 equiv. of **5** in 96 h, the title compound was isolated via flash chromatography (1 – 5% Acetone/Hexane) as a clear oil (61 mg, 86% yield). All analytical data for **3z** was in accordance with literature data.²¹

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 2.37 (t, *J* = 7.5 Hz, 4H), 1.60 – 1.47 (m, 4H), 1.36 – 1.20 (m, 4H), 0.88 (t, *J* = 7.4 Hz, 6H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 212.0, 42.9, 26.4, 22.8, 14.2.

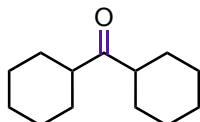


Pentadecan-8-one (**3aa**)

Following General Procedure A, prepared from 0.50 mmol of 8-pentadecanol (**1aa**) and 3.5 equiv. of **5** in 96 h, the title compound was isolated via flash chromatography (1 – 5% Acetone/Hexane) as a tan oil (45 mg, 78% yield). All analytical data for **3aa** was in accordance with literature data.^{22a, 22b}

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 2.38 (t, *J* = 7.5 Hz, 4H), 1.56 (dd, *J* = 13.1, 5.9 Hz, 4H), 1.27 (s, 16H), 0.87 (t, *J* = 6.9 Hz, 6H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 211.7, 42.7, 31.6, 29.1, 29.0, 23.8, 22.5, 14.0.

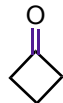


Dicyclohexylmethanone (**3ab**)

Following General Procedure A, prepared from 0.48 mmol of dicyclohexylmethanol (**1ab**) and 1.1 equiv. of **5**, the title compound was isolated via flash chromatography (1 – 5% Acetone/Hexane) as a pale oil (52 mg, 56% yield). All analytical data for **3ab** was in accordance with literature data.²³

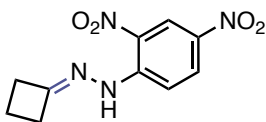
¹H NMR (500 MHz, CDCl₃) (δ, ppm): 2.48 (ddd, *J* = 11.0, 8.2, 2.8 Hz, 2H), 1.81 – 1.73 (m, 8H), 1.70 – 1.63 (m, 2H), 1.39 – 1.13 (m, 10H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 217.6, 49.6, 29.0, 26.3, 26.2.



Cyclobutanone (**3ac**)

Following General Procedure A, prepared from 0.49 mmol of cyclobutanol (**1ac**) and 3.5 equiv. of **5** in 96 h. ¹H NMR yield determined using CH₂Br₂ as an internal standard (>99% yield). All analytical data for **3ac** was in accordance with literature data.²⁴

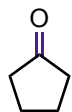


1-cyclobutylidene-2-(2,4-dinitrophenyl)hydrazine (**3ac'**)

Following General Procedure A, prepared from 1.16 mmol of cyclobutanol (**1ac**) and 2.5 equiv. of **5**, the title compound **3ac** was subjected to hydrazone derivatization and isolated as 2,4-dinitrophenylhydrazone of cyclobutanone **3ac'**. The title compound was purified by silica gel chromatography (Pentane:EtOAc 9:1) to give a reddish crystalline solid (270 mg) in 93% yield. All analytical data of **3ac'** was in accordance with the reported literature.²⁵

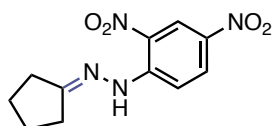
¹H NMR (500 MHz, CDCl₃) δ 10.72 (s, 1H), 9.12 (d, *J* = 2.6 Hz, 1H), 8.29 (dd, *J* = 9.6, 2.6 Hz, 1H), 7.87 (d, *J* = 9.6 Hz, 1H), 3.18 – 3.03 (m, 4H), 2.19 (p, *J* = 8.1 Hz, 2H).

^{13}C NMR (126 MHz, CDCl_3) δ 161.2, 144.7, 137.5, 129.9, 128.6, 123.5, 115.9, 33.8, 31.7, 13.7.



Cyclopentanone (**3ad**)

Following General Procedure A, prepared from 0.50 mmol of cyclopentanol (**1ad**) and 3.5 equiv. of **5** in 96 h. ^1H NMR yield determined using CH_2Br_2 as an internal standard (90% yield). All analytical data for **3ad** was in accordance with literature data.²⁶

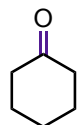


1-cyclopentylidene-2-(2,4-dinitrophenyl)hydrazine(**3ad'**)

Following General Procedure A, prepared from 1.16 mmol of cyclopentanol (**1ad**) and 2.5 equiv. of **5**, the title compound **3ad** was subjected to hydrazone derivatization and isolated as 2,4-dinitrophenylhydrazone of cyclopentanone **3ad'**. The title compound was purified by silica gel chromatography (Pentane:EtOAc 9:1.5) to give a yellow crystalline solid (265 mg) in 86% yield. All analytical data of **3ad'** was in accordance with the reported literature.²⁷

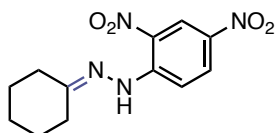
^1H NMR (500 MHz, CDCl_3) δ 10.81 (s, 1H), 9.12 (d, $J = 2.6$ Hz, 1H), 8.28 (dd, $J = 9.7, 2.7$ Hz, 1H), 7.92 (d, $J = 9.6$ Hz, 1H), 2.58 (t, $J = 7.2$ Hz, 2H), 2.47 (t, $J = 7.4$ Hz, 2H), 1.99 (p, $J = 7.0$ Hz, 2H), 1.88 (p, $J = 7.0$ Hz, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 168.2, 144.9, 137.4, 129.8, 128.6, 123.5, 116.1, 33.5, 28.0, 24.8, 24.7.



Cyclohexanone (**3ae**)

Following General Procedure A, prepared from 0.50 mmol of cyclohexanol (**1ae**) and 3.5 equiv. of **5** in 96 h. ^1H NMR yield determined using CH_2Br_2 as an internal standard (>99% yield). All analytical data for **3ae** was in accordance with literature data.²⁶



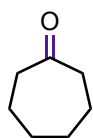
1-cyclohexylidene-2-(2,4-dinitrophenyl)hydrazine(**3ae'**)

Following General Procedure A, prepared from 1.16 mmol of cyclohexanol (**1ae**) and 2.5 equiv. of **5**, the title compound **3ae** was subjected to hydrazone derivatization and isolated as 2,4-

dinitrophenylhydrazone of cyclopentanone **3ae'**. The title compound was purified by silica gel chromatography (Pentane:EtOAc 9:1.5) to give a yellow crystalline solid (295 mg) in 91% yield. All analytical data for 2,4-dinitrophenylhydrazone of **3ae'** was in accordance with the reported literature.²⁸

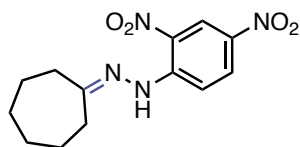
¹H NMR (500 MHz, CDCl₃) δ 11.20 (s, 1H), 9.13 (d, *J* = 2.6 Hz, 1H), 8.29 (dd, *J* = 9.6, 2.6 Hz, 1H), 7.97 (d, *J* = 9.6 Hz, 1H), 2.47 (q, *J* = 6.2 Hz, 4H), 1.79 (dt, *J* = 11.5, 6.2 Hz, 4H), 1.72 (td, *J* = 6.6, 3.3 Hz, 2H) of **5** in 96 h.

¹³C NMR (126 MHz, CDCl₃) δ 161.5, 145.5, 137.6, 130.1, 128.9, 123.8, 116.4, 35.7, 27.3, 27.1, 26.1, 25.6.



Cycloheptanone (**3af**)

Following General Procedure A, prepared from 0.50 mmol of cycloheptanol (**1af**) and 3.5 equiv. of **5** in 96 h. ¹H NMR yield determined using CH₂Br₂ as an internal standard (>99% yield). All analytical data for **3ae** was in accordance with literature data.²⁹

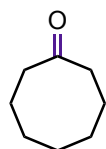


1-cycloheptylidene-2-(2,4-dinitrophenyl)hydrazine(**3af'**)

Following General Procedure A, prepared from 1.16 mmol of cycloheptanol (**1af**) and 2.5 equiv. of **5**, the title compound **3af** was subjected to hydrazone derivatization and isolated as 2,4-dinitrophenylhydrazone of cyclopentanone **3af'**. The title compound was recrystallized from pentane to give a bright yellow solid (310 mg) in 91% yield. All analytical data for **3af'** was in accordance with the reported literature.³⁰

¹H NMR (500 MHz, CDCl₃) δ 11.05 (s, 1H), 9.13 (q, *J* = 2.8, 2.0 Hz, 1H), 8.29 (dd, *J* = 9.6, 2.6 Hz, 1H), 7.99 (d, *J* = 9.7 Hz, 1H), 2.66 – 2.60 (m, 2H), 2.60 – 2.53 (m, 2H), 1.91 – 1.83 (m, 2H), 1.76 – 1.70 (m, 2H), 1.69 – 1.62 (m, 4H).

¹³C NMR (126 MHz, CDCl₃) δ 163.96, 145.0, 137.5, 129.8, 128.9, 123.4, 116.3, 37.1, 30.7, 30.1, 30.0, 27.5, 24.2.

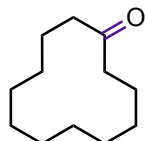


Cyclooctanone (**3ag**)

Following General Procedure A, prepared from 0.60 mmol of cyclooctanol(**1ag**) and 3.5 equiv. of **5** in 96 h, the title compound was isolated via flash chromatography (1 – 5% Acetone/Hexane) as a clear oil (50 mg, 64% yield). All analytical data for **3ag** was in accordance with literature data.^{23, 29}

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 2.44 – 2.33 (m, 4H), 1.92 – 1.79 (m, 4H), 1.52 (qt, *J* = 5.9, 2.9 Hz, 4H), 1.36 (tt, *J* = 5.6, 2.8 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 218.1, 41.7, 27.0, 25.5, 24.5.

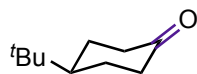


Cyclododecanone (**3ah**)

Following General Procedure A, prepared from 0.45 mmol of cyclododecanol (**1ah**) and 3.5 equiv. of **5** in 96 h, the title compound was isolated via flash chromatography (1 – 5% Acetone/Hexane) as a pale oil (40 mg, 48% yield). All analytical data for **3ah** was in accordance with literature data.²¹

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 2.49 – 2.43 (m, 4H), 1.71 (p, *J* = 6.4 Hz, 4H), 1.28 (dd, *J* = 13.2, 5.6 Hz, 14H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 213.1, 40.5, 24.9, 24.7, 24.3, 22.7, 22.5.

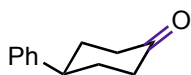


4-(*tert*-Butyl)cyclohexan-1-one (**3ai**)

Following General Procedure A, prepared from 0.57 mmol of 4-(*tert*-butyl)cyclohexan-1-ol (**1ai**) and 1.1 equiv. of **5** in 24 h, the title compound was isolated via flash chromatography (1 – 5% Acetone/Hexane) as a pale white solid (66 mg, 75% yield). All analytical data for **3ai** was in accordance with literature data.³¹

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 2.40 (d, *J* = 16.0 Hz, 2H), 2.39 – 2.25 (m, 2H), 2.13 – 2.04 (m, 2H), 1.54 – 1.38 (m, 3H), 0.92 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 213.1, 47.1, 41.7, 32.9, 28.0.

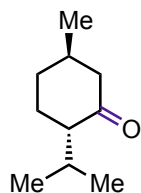


4-Phenylcyclohexan-1-one (**3aj**)

Following General Procedure A, prepared from 0.49 mmol of 4-phenylcyclohexan-1-ol (**1aj**) and 1.1 equiv. of **5** in 24 h, the title compound was isolated via flash chromatography (1 – 5% Acetone/Hexane) as pale white crystals (46 mg, 54% yield). All analytical data for **3aj** was in accordance with literature data.³²

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 7.37 – 7.29 (m, 2H), 7.27 – 7.18 (m, 3H), 3.03 (tt, *J* = 12.1, 3.4 Hz, 1H), 2.52 (dd, *J* = 11.7, 5.2 Hz, 4H), 2.28 – 2.19 (m, 2H), 2.03 – 1.88 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 211.6, 145.2, 129.1, 127.1, 127.0, 43.2, 41.8, 34.4.

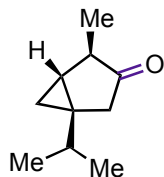


2-Isopropyl-5-methylcyclohexan-1-one (3ak)

Following General Procedure A, prepared from 0.60 mmol of 2-isopropyl-5-methylcyclohexan-1-ol (**1ak**) in 24 h. Title compound menthone was prepared using 1.0 equiv. of 2-chloro 4-nitropyridine. The crude reaction mixture was concentrated on *vacuo* and added a minimal amount of diethyl ether. The precipitates were filtered and added 2 N HCl in diethyl ether (0.5 ml). Precipitates were filtered again, and the desired compound was isolated via flash chromatography (0 – 2.5% Acetone/Hexane) as a white solid (49 mg, 50% yield). All analytical data for **3ak** was in accordance with literature data.³³

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 2.33 (ddd, *J* = 12.9, 3.8, 2.2 Hz, 1H), 2.17 – 2.08 (m, 1H), 2.08 – 1.98 (m, 2H), 1.95 (d, *J* = 12.8 Hz, 1H), 1.86 (ddd, *J* = 19.1, 8.0, 3.8 Hz, 2H), 1.43 – 1.26 (m, 2H), 0.99 (d, *J* = 6.4 Hz, 3H), 0.89 (d, *J* = 6.8 Hz, 3H), 0.83 (d, *J* = 6.8 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 212.6, 56.0, 51.0, 35.6, 34.0, 28.0, 26.0, 22.4, 21.3, 18.8.

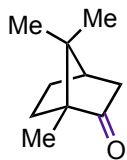


4-Phenylcyclohexan-1-one (3al)

Following General Procedure A, prepared from 0.50 mmol of synthesized 4-phenylcyclohexan-1-one (**1al**)³⁴ and 3.5 equiv. of **5** in 24 h. The title compound was isolated via flash chromatography (1 – 5% Acetone/Hexane) as a clear oil (41 mg, 54% yield). All analytical data for **3al** was in accordance with literature data.³⁵

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 2.54 (ddd, *J* = 18.8, 2.6, 1.3 Hz, 1H), 2.21 (qd, *J* = 7.5, 1.4 Hz, 1H), 2.06 (d, *J* = 18.8 Hz, 1H), 1.35 (dt, *J* = 13.7, 6.8 Hz, 1H), 1.15 (d, *J* = 7.5 Hz, 3H), 1.08 (dd, *J* = 8.1, 4.0 Hz, 1H), 1.00 (d, *J* = 6.7 Hz, 3H), 0.95 (d, *J* = 6.8 Hz, 3H), 0.75 (ddd, *J* = 8.1, 5.6, 2.4 Hz, 1H), 0.12 (dd, *J* = 5.7, 4.0 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 221.7, 47.5, 39.9, 33.1, 31.4, 29.8, 25.7, 20.2, 19.9, 18.9, 18.4.

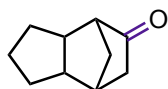


(1*S*,4*R*)-1,7,7-Trimethylbicyclo[2.2.1]heptan-2-one (Camphor) (3am)

Following General Procedure A, prepared from 0.50 mmol of (1*S*)-1,7,7-trimethylbicyclo[2.2.1]heptan-2-ol (borneol) (**1a**) and 3.5 equiv. of **5** in 90 h. The title compound was isolated via flash chromatography (1 – 5% Acetone/Hexane) as a white solid (67 mg, 81% yield). All analytical data for **3am** was in accordance with literature data.³⁶

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 2.34 (dt, *J* = 18.2, 3.9 Hz, 1H), 2.08 (t, *J* = 4.5 Hz, 1H), 1.94 (tq, *J* = 12.0, 4.1 Hz, 1H), 1.83 (d, *J* = 18.2 Hz, 1H), 1.67 (td, *J* = 13.3, 12.8, 3.7 Hz, 1H), 1.45 – 1.28 (m, 2H), 0.95 (s, 3H), 0.90 (s, 3H), 0.82 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 220.2, 58.1, 47.2, 43.7, 43.5, 30.3, 27.5, 20.2, 19.6, 9.7.

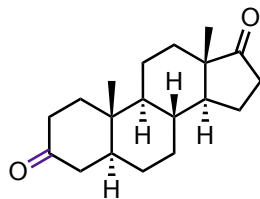


(3*aR*,4*S*,7*S*,7*aR*)-Octahydro-5*H*-4,7-methanoinden-5-one (3an)

Following General Procedure A, prepared from 0.49 mmol (3*aR*,4*S*,7*S*,7*aR*)-octahydro-5*H*-4,7-methanoinden-5-ol and 1.1 equiv. of **5** in 24 h. The title compound was isolated via flash chromatography (1 – 5% Acetone/Pentane) as a pale-yellow oil (59 mg, 81% yield). All analytical data for **3an** was in accordance with literature data.³⁷

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 2.44 – 2.34 (m, 2H), 2.12 (q, *J* = 7.0 Hz, 2H), 2.07 – 1.91 (m, 3H), 1.82 – 1.71 (m, 3H), 1.52 (dp, *J* = 10.8, 1.5 Hz, 1H), 1.42 – 1.29 (m, 1H), 1.15 – 1.02 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 218.0, 54.2, 46.7, 44.4, 41.8, 39.6, 32.2, 31.5, 31.3, 27.9.



(8*R*,9*S*,10*S*,13*S*,14*S*)-10,13-Dimethyltetradecahydro-3*H*-cyclopenta[*a*]phenanthrene-3,17(2*H*)-dione (3ao)

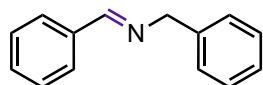
Following General Procedure A, prepared from 0.45 mmol of (8*R*,9*S*,10*S*,13*S*,14*S*)-3-hydroxy-10,13-dimethylhexadecahydro-17*H*-cyclopenta[*a*]phenanthren-17-one and 2.0 equiv. of **5** in 24 h with minimal amount of dry and degassed DCM, the title compound was purified via flash

chromatography (1 – 15% EtOAc/Hexane) as a white solid (120 mg, 86% yield). All analytical data for **3a** was in accordance with literature data.²⁹

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 2.49 – 2.24 (m, 4H), 2.14 – 1.99 (m, 3H), 1.98 – 1.90 (m, 1H), 1.83 (dt, *J* = 12.9, 3.5 Hz, 2H), 1.69 (dd, *J* = 13.6, 2.9 Hz, 1H), 1.65 – 1.48 (m, 3H), 1.47 – 1.22 (m, 7H), 1.03 (s, 3H), 0.88 (s, 3H), 0.79 (td, *J* = 12.0, 3.8 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 220.9, 211.5, 53.8, 51.1, 47.6, 46.5, 44.5, 38.3, 38.0, 35.7, 34.8, 31.3, 30.4, 28.5, 21.7, 20.6, 13.7, 11.4.

Characterization data of imines

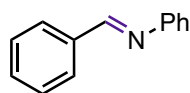


(*E*)-*N*-Benzyl-1-phenylmethanimine (**4a**)

Following General Procedure C, prepared from 0.50 mmol of dibenzylamine (**2a**) in 24 h, the title compound was purified by flash column chromatography (5% NEt₃/Hexane) as a colorless oil (69 mg, 70% yield). All analytical data for **4a** is in accordance with literature data.³⁸

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 8.41 (s, 1H), 7.82 – 7.75 (m, 2H), 7.45 – 7.39 (m, 3H), 7.35 (d, *J* = 4.6 Hz, 4H), 7.31 – 7.22 (m, 1H), 4.84 (s, 2H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 163.5, 139.4, 136.3, 130.9, 128.7, 128.6, 128.4, 128.1, 127.1, 65.2.

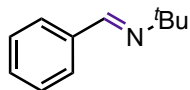


(*E*)-*N*-1-Diphenylmethanimine (**4b**)

Following General Procedure C, prepared from 0.50 mmol of *N*,1-diphenylmethanamine (**2b**) in 24 h, the title compound was purified by flash column chromatography (5% NEt₃/Hexane) as a yellow oil (40 mg, 44% yield). All analytical data for **4b** is in accordance with literature data.³⁸

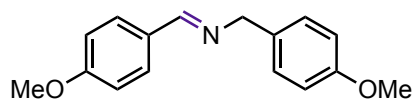
¹H NMR (500 MHz, CDCl₃) (δ, ppm): 8.47 (s, 1H), 7.95 – 7.87 (m, 2H), 7.53 – 7.44 (m, 3H), 7.44 – 7.37 (m, 2H), 7.28 – 7.21 (m, 3H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 160.6, 152.2, 136.4, 131.5, 129.3, 129.0, 128.9, 126.1, 121.0.



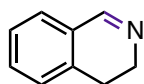
(*E*)-*N*-*tert*-Butyl-1-phenylmethanimine (**4c**)

Following General Procedure C, prepared from 0.50 mmol of *N*-*tert*-butyl-1-phenylmethanimine (**2c**). ¹H NMR yield determined using CH₂Br₂ as an internal standard (62% yield). All analytical data for **4c** is in accordance with literature data.³⁹



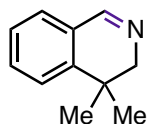
(E)-N-(4-Methoxybenzyl)-1-(4-methoxyphenyl)methanimine (4d)

Following General Procedure C, prepared from 0.50 mmol of bis(4-methoxybenzyl)amine (**2d**) in 24 h. ¹H NMR yield determined using CH₂Br₂ as an internal standard (43% yield). All analytical data for **4d** was in accordance with literature data.⁴⁰



3,4-Dihydroisoquinoline (4e)

Following General Procedure C, prepared from 0.50 mmol of 1,2,3,4-tetrahydroisoquinoline (**2e**), in 1.0 mL of PhCF₃ for 24h. ¹H NMR yield determined using CH₂Br₂ as an internal standard (>99% yield, 4.6:1 of 3,4-dihydroisoquinoline to isoquinoline). All analytical data for **4e** was in accordance with literature data.⁴¹

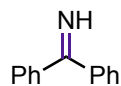


4,4-Dimethyl-3,4-dihydroisoquinoline (4f)

Following General Procedure C, prepared from 0.50 mmol of 4,4-dimethyl-1,2,3,4-tetrahydroisoquinoline (**2f**) in 24h, the title compound was purified by flash column chromatography (5% NEt₃/Hexane) as a pale-yellow solid (69 mg, 86% yield). All analytical data for **4f** is in accordance with the literature data.⁴²

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 8.37 (s, 1H), 7.42 (ddd, J = 7.7, 5.9, 2.8 Hz, 1H), 7.38 – 7.33 (m, 1H), 7.32 – 7.27 (m, 2H), 3.62 (s, 2H), 1.24 (s, 6H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 160.7, 145.6, 131.9, 127.8, 126.8, 123.6, 61.1, 31.6, 26.5.



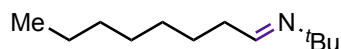
Diphenylmethanimine (4g)

Following General Procedure C, prepared from 0.50 mmol of diphenylmethanimine (**2g**) in 24 h. ¹H NMR yield determined using CH₂Br₂ as an internal standard (85% yield). All analytical data for **4g** was in accordance with literature data.⁴³



(E)-N-Octyloctan-1-imine (4h)

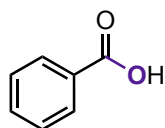
Following General Procedure C, prepared from 0.50 mmol of dioctylamine (**2h**) in 24 h. ¹H NMR yield determined using CH₂Cl₂ as an internal standard (40% yield). All analytical data for **4h** was in accordance with literature data.⁴⁰



(E)-N-(*tert*-Butyl)octan-1-imine (**4i**)

Following General Procedure C, prepared from 0.50 mmol of synthesized *N*-(*tert*-butyl)octan-1-amine (**2i**)^{44a} in 24 h. ¹H NMR yield determined using CH₂Cl₂ as an internal standard (22% yield). All analytical data for **4i** was in accordance with literature data.^{44b}

Characterization data of carboxylic acids

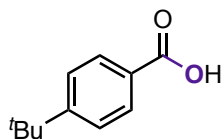


Benzoic acid (**10a**)

Following General Procedure D, prepared from 0.50 mmol of benzaldehyde (**8a**) in 24 h, the title compound was isolated via flash chromatography (0 – 50% EtOAc/Hexane) as a white solid (53 mg, 87% yield). All analytical data for **10a** was in accordance with literature data.⁴⁵

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 11.75 (s, 1H), 8.13 (d, *J* = 7.3 Hz, 2H), 7.62 (t, *J* = 7.4 Hz, 1H), 7.49 (t, *J* = 7.7 Hz, 2H).

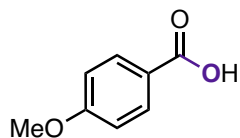
¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 171.8, 134.0, 130.4, 129.4, 128.7.



4-(*tert*-Butyl)benzoic acid (10b**)** Following General Procedure D, prepared from 0.50 mmol of 4-(*tert*-butyl)benzaldehyde (**8b**) in 36 h, the title compound was isolated via flash chromatography (0 – 50% EtOAc/Hexane) as a white solid (60 mg, 67% yield). All analytical data for **10b** was in accordance with literature data.⁴⁶

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 8.06 (d, *J* = 8.2 Hz, 2H), 7.50 (d, *J* = 7.9 Hz, 2H), 1.36 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 172.5, 157.8, 130.3, 126.7, 125.6, 35.4, 31.4, 31.3.

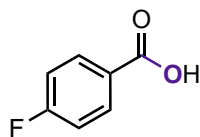


4-Methoxybenzoic acid (**10c**)

Following General Procedure D, prepared from 0.50 mmol of 4-methoxybenzaldehyde (**8c**) in 36 h, the title compound was isolated via flash chromatography (0 – 50% EtOAc/Hexane) as a white solid (52 mg, 69% yield). All analytical data for **10c** was in accordance with literature data.⁴⁵

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 8.07 (d, *J* = 8.7 Hz, 2H), 6.95 (d, *J* = 8.6 Hz, 2H), 3.88 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) (δ , ppm): 171.8, 164.2, 132.5, 121.8, 113.9, 55.6.

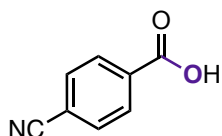


4-Fluorobenzoic acid (10d)

Following General Procedure D, prepared from 0.50 mmol of 4-fluorobenzaldehyde (**8d**) in 24 h, the title compound was isolated via flash chromatography (0 – 50% EtOAc/Hexane) as a white solid (60 mg, 85% yield). All analytical data for **10d** was in accordance with the literature data.⁴⁵

^1H NMR (500 MHz, CDCl_3) (δ , ppm): 8.18 – 8.10 (m, 2H), 7.15 (t, $J = 8.6$ Hz, 2H).

^{13}C NMR (126 MHz, CDCl_3) (δ , ppm): 171.1, 166.5 (d, $J = 255.4$ Hz), 133.1 (d, $J = 9.6$ Hz), 125.6 (d, $J = 3.1$ Hz), 115.9 (d, $J = 22.1$ Hz).

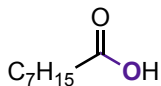


4-Cyanobenzoic acid (10e)

Following General Procedure D, prepared from 0.50 mmol of 4-cyanobenzaldehyde (**8e**) in 24 h, the title compound was isolated via flash chromatography (0 – 50% EtOAc/Hexane) as a white solid (64 mg, 87% yield). All analytical data for **10e** was in accordance with the literature data.⁴⁷

^1H NMR (500 MHz, CDCl_3) (δ , ppm): 8.21 (d, $J = 8.4$ Hz, 2H), 7.80 (d, $J = 8.4$ Hz, 2H).

^{13}C NMR (126 MHz, CDCl_3) (δ , ppm): 170.0, 133.0, 132.5, 130.9, 117.9, 117.5.

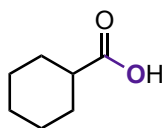


Octanoic acid (10f)

Following General Procedure D, prepared from 0.50 mmol of octanal (**8f**) with 1.0 equiv. of **5**, and 1.0 equiv. H_2O in MeCN (1.0M) in 12 h, the title compound was isolated via flash chromatography (0 – 50% EtOAc/Hexane) as a colorless oil (35 mg, 49% yield). All analytical data for **10f** was in accordance with literature data.⁴⁸

^1H NMR (400 MHz, CDCl_3) (δ , ppm): 2.35 (t, $J = 7.5$ Hz, 2H), 1.63 (p, $J = 7.4$ Hz, 2H), 1.40 – 1.21 (m, 8H), 0.92 – 0.83 (m, 3H).

^{13}C NMR (126 MHz, CDCl_3) (δ , ppm): 180.7, 34.3, 31.8, 29.2, 29.0, 24.8, 22.7, 14.2.

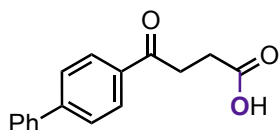


Cyclohexanecarboxylic acid (**10g**)

Following General Procedure D, prepared from 0.50 mmol of cyclohexanecarbaldehyde (**8g**) with 1.0 equiv. of **5**, and 1.0 equiv. H₂O in MeCN (1.0M) in 12 h, the title compound was isolated via flash chromatography (0 – 50% EtOAc/Hexane) as a yellow oil (42 mg, 65% yield). All analytical data for **10g** was in accordance with literature data.⁴⁸

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 11.71 (s, 1H), 2.33 (tt, *J* = 11.3, 3.7 Hz, 1H), 1.93 (dt, *J* = 12.9, 3.9 Hz, 2H), 1.76 (dp, *J* = 11.1, 3.8 Hz, 2H), 1.64 (dd, *J* = 11.0, 4.7 Hz, 1H), 1.45 (qd, *J* = 11.6, 3.4 Hz, 2H), 1.35 – 1.17 (m, 3H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 182.8, 43.1, 28.9, 25.8, 25.5.



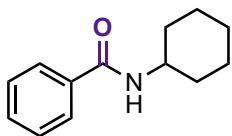
4-([1,1'-Biphenyl]-4-yl)-4-oxobutanoic acid (Fenbufen) (**10h**)

Following General Procedure D, prepared from 0.50 mmol of 4-([1,1'-biphenyl]-4-yl)-4-oxobutanal (**8h**) with 1.0 equiv. of **5**, and 1.0 equiv. H₂O in MeCN (1.0M) in 12 h, the title compound was isolated via flash chromatography (0 – 50% EtOAc/Hexane) as a white solid (70 mg, 56% yield). All analytical data for **10h** was in accordance with literature data.⁴⁹

¹H NMR (400 MHz, DMSO-*d*₆) (δ, ppm): 12.21 (s, 1H), 8.07 (d, *J* = 8.1 Hz, 2H), 7.83 (d, *J* = 8.2 Hz, 2H), 7.75 (d, *J* = 7.3 Hz, 2H), 7.51 (t, *J* = 7.5 Hz, 2H), 7.43 (dd, *J* = 8.4, 6.1 Hz, 1H), 3.29 (t, *J* = 6.2 Hz, 2H), 2.61 (t, *J* = 6.2 Hz, 2H).

¹³C NMR (101 MHz, DMSO-*d*₆) (δ, ppm): 198.1, 173.8, 144.5, 138.9, 135.3, 129.1, 128.6, 128.4, 127.0, 126.9, 33.2, 27.9.

Characterization data of amides

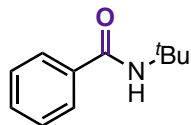


N-Cyclohexylbenzamide (**11a**)

Following General Procedure E, prepared from cyclohexanamine (0.50 mmol, 1.00 equiv.) and benzaldehyde (0.50 mmol, 1.00 equiv.) in 24 h, the title compound was isolated via flash chromatography (0 – 50% EtOAc/Hexane) as a yellow solid (86 mg, 85% yield). All analytical data for **11a** was in accordance with literature data.⁵⁰

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 7.75 (d, *J* = 7.2 Hz, 2H), 7.48 (t, *J* = 7.3 Hz, 1H), 7.42 (t, *J* = 7.3 Hz, 2H), 5.97 (s, 1H, NH), 3.98 (tdt, *J* = 11.4, 7.9, 3.9 Hz, 1H), 2.03 (dq, *J* = 12.3, 3.9 Hz, 2H), 1.75 (dp, *J* = 11.8, 3.9 Hz, 2H), 1.66 (dp, *J* = 11.5, 4.0, 3.3 Hz, 1H), 1.43 (qt, *J* = 12.5, 3.5 Hz, 2H), 1.34 – 1.13 (m, 3H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 166.8, 135.3, 131.4, 128.7, 127.0, 48.8, 33.4, 25.7, 25.0.

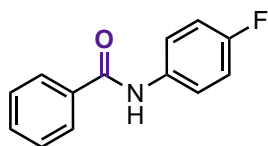


***N*-(*tert*-Butyl)benzamide (11b)**

Following General Procedure E, prepared from *tert*-butylamine (0.50 mmol, 1.00 equiv.) and benzaldehyde (0.50 mmol, 1.00 equiv.) in 24 h, the title compound was isolated via flash chromatography (0 – 50% EtOAc/Hexane) as a white solid (72 mg, 81% yield). All analytical data for **11b** was in accordance with literature data.⁵¹

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 7.75 – 7.68 (m, 2H), 7.53 – 7.36 (m, 3H), 5.93 (s, 1H, NH), 1.48 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 167.1, 136.1, 131.2, 128.6, 126.8, 51.8, 29.0.

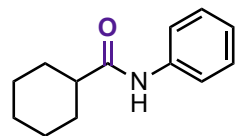


***N*-(4-Fluorophenyl)benzamide (11c)**

Following General Procedure E, prepared from 4-fluoroaniline (0.50 mmol, 1.00 equiv.) and benzaldehyde (0.50 mmol, 1.00 equiv.) in 24 h, the title compound was isolated via flash chromatography (0 – 50% EtOAc/Hexane) as a yellow solid (73 mg, 68% yield). All analytical data for **11c** was in accordance with literature data.⁵²

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 7.86 (d, *J* = 7.6 Hz, 2H), 7.81 (s, 1H, NH), 7.60 (dd, *J* = 8.7, 4.9 Hz, 2H), 7.55 (d, *J* = 7.3 Hz, 1H), 7.49 (t, *J* = 7.6 Hz, 2H), 7.06 (t, *J* = 8.4 Hz, 2H).

¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 165.9, 159.7 (d, *J* = 244.1 Hz), 134.9, 134.0 (d, *J* = 2.9 Hz), 132.1, 129.0, 127.1, 122.2 (d, *J* = 7.9 Hz), 115.9 (d, *J* = 22.4 Hz).

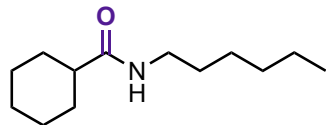


***N*-Phenylcyclohexanamide (11d)**

Following General Procedure E, prepared from aniline (0.50 mmol, 1.00 equiv.) and cyclohexanecarbaldehyde (0.50 mmol, 1.00 equiv.) in 24 h, the title compound was isolated via flash chromatography (0 – 50% EtOAc/Hexane) as a white solid (72 mg, 71% yield). All analytical data for **11d** was in accordance with literature data.⁵²

¹H NMR (400 MHz, CDCl₃) (δ, ppm): 7.52 (d, *J* = 7.4 Hz, 2H), 7.31 (dd, *J* = 8.6, 7.3 Hz, 2H), 7.09 (t, *J* = 7.4 Hz, 1H), 2.23 (tt, *J* = 11.7, 3.5 Hz, 1H), 1.96 (d, *J* = 13.0 Hz, 2H), 1.85 (dd, *J* = 13.0, 3.4 Hz, 2H), 1.75 – 1.65 (m, 1H), 1.62 – 1.48 (m, 2H), 1.39 – 1.18 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) (δ, ppm): 174.4, 138.2, 129.1, 124.2, 119.8, 46.8, 29.8, 25.8.



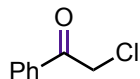
N-Hexylcyclohexanecarboxamide (11e)

Following General Procedure E, prepared from hexylamine (0.50 mmol, 1.00 equiv.) and cyclohexanecarbaldehyde (0.50 mmol, 1.00 equiv.) in 24 h, the title compound was isolated via flash chromatography (0 – 50% EtOAc/Hexane) as a white solid (65 mg, 62% yield). All analytical data for **11e** was in accordance with literature data.⁵¹

¹H NMR (500 MHz, CDCl₃) (δ, ppm): 5.51 (s, 1H, NH), 3.21 (q, *J* = 6.7 Hz, 2H), 2.04 (tt, *J* = 11.8, 3.5 Hz, 1H), 1.83 (d, *J* = 13.1 Hz, 2H), 1.77 (d, *J* = 10.6 Hz, 2H), 1.68 – 1.62 (m, 1H), 1.51 – 1.35 (m, 4H), 1.36 – 1.09 (m, 9H), 0.86 (t, *J* = 6.6 Hz, 3H).

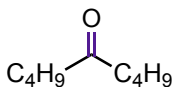
¹³C NMR (126 MHz, CDCl₃) (δ, ppm): 176.2, 45.8, 39.5, 31.6, 29.9, 29.8, 26.7, 25.9, 22.7, 14.1.

Photo-Flow reaction data



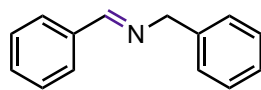
2-Chloro-1-phenylethan-1-one (3q)

Following General Procedure F, using 2-chloro-1-phenylethan-1-ol (0.20 mmol, 1.00 equiv.) and 2-bromo-6-nitropyridine (0.15 mmol, 0.75 equiv.) in trifluorotoluene (2.0 mL). The residence time was 5 h. The ¹H NMR yield was determined using CH₂Br₂, yield = 82%.



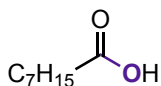
5-Nonanone (3z)

Following General Procedure F, using nonan-5-ol (0.20 mmol, 1.00 equiv.) and 3,5-bis(trifluoromethyl)nitrobenzene (0.40 mmol, 2.00 equiv.) in MeCN/*t*BuOH (5:1, 2.0 mL). The residence time was 5 h. The ¹H NMR yield was determined using CH₂Br₂, yield = 97%.



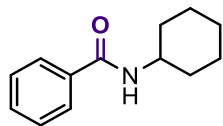
(E)-N-Benzyl-1-phenylmethanimine (4a)

Following General Procedure F, using dibenzylamine (0.20 mmol, 1.00 equiv.) and 5-bromo-2-nitropyridine (0.40 mmol, 2.00 equiv.) in DCM (2.0 mL). The residence time was 3 h. The ¹H NMR yield was determined using CH₂Br₂, yield = 75%.



Octanoic acid (10f)

Following General Procedure F, using octanal (0.20 mmol, 1.00 equiv.) and 3,5-bis(trifluoromethyl)nitrobenzene (0.20 mmol, 1.00 equiv.) in MeCN/H₂O (10:1, 2.0 mL). The residence time was 4 h. The ¹H NMR yield was determined using CH₂Br₂, yield = 58%.



***N*-Cyclohexylbenzamide (11a)**

Following General Procedure F, using cyclohexanamine (0.20 mmol, 1.00 equiv.), benzaldehyde (0.20 mmol, 1.00 equiv.) and 3,5-bis(trifluoromethyl)nitrobenzene (0.20 mmol, 1.00 equiv.) in DCM/*t*BuOH (5:1, 2.0 mL). The mixture was first pre-stirred for 4 h at room temperature to form *N*-cyclohexyl-1-phenylmethanimine in situ which was then subject to flow photochemical conditions. The residence time was 5 h. The ¹H NMR yield was determined using CH₂Br₂, yield = 82%.

Mechanism studies

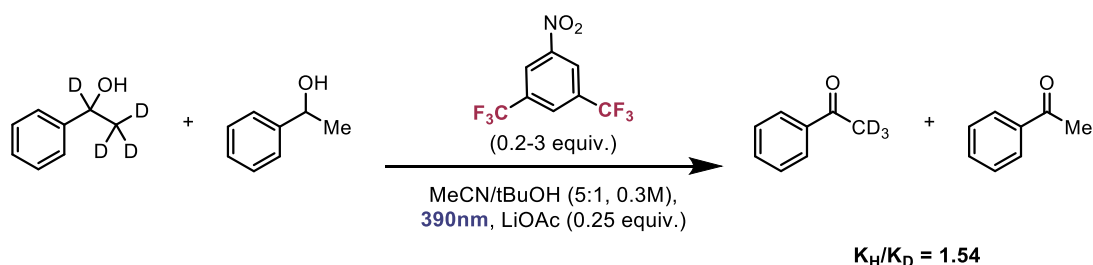
Kinetic isotope effect studies

Benzylic C(sp³) Anaerobic oxidation.

Independent Rate Measurement. To an oven-dried, 0.5-dram vial, equipped with a stir bar was added of 3,5-Bis(trifluoromethyl)nitrobenzene (16 mg, 1.0 equiv., 0.060 mmol), LiOAc (1 mg, 0.20 equiv., 0.12 mmol), and synthesized 1-phenylethan-1,2,2,2-d₄-1-ol⁵³ (38 mg, 5.0 equiv., 0.30 mmol). In a separate oven-dried, 0.5-dram vial, a second vial equipped with a stir bar was added of 3,5-Bis(trifluoromethyl)nitrobenzene (16 mg, 1.0 equiv., 0.060 mmol), LiOAc (1 mg, 0.20 equiv., 0.12 mmol), and 1-phenylethan-ol (37 mg, 5.0 equiv., 0.30 mmol). Each reaction vessel was subjected to 390 nm light for 1 h. A crude ¹H NMR was taken for each vessel and the ratio of the relative abundance was observed ((δ , ppm): 7.51). Each KIE measurement is the average of two independent experiments.

Intermolecular Competition Rate Measurement. To an oven-dried, 0.5-dram vial, equipped with a stir bar was added of 3,5-Bis(trifluoromethyl)nitrobenzene (16 mg, 1.0 equiv., 0.060 mmol), LiOAc (1 mg, 0.20 equiv., 0.12 mmol), and prepared 1-phenylethan-1,2,2,2-d₄-1-ol² (38 mg, 5.0 equiv., 0.30 mmol) and 1-phenylethan-ol (37 mg, 5.0 equiv., 0.30 mmol). The reaction vessel was subjected to 390 nm light for 1 h. A crude ¹H NMR was taken using CH₂Br₂ as an internal standard ((δ , ppm): 2.57). The KIE measurement is the average of two independent experiments.

Intermolecular Competition



Independent Rates

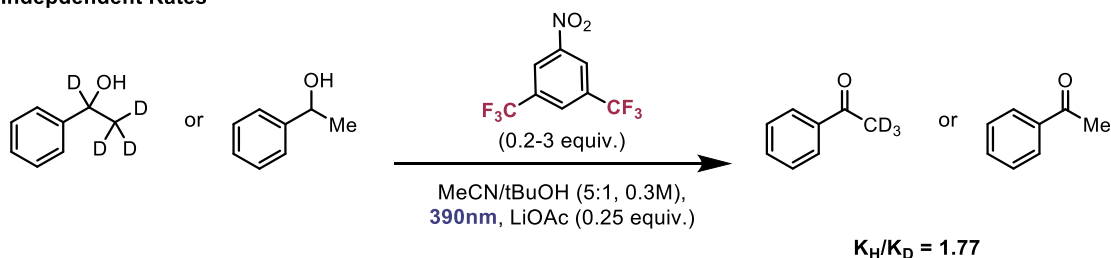


Figure S7. Intermolecular competition and independent rate measurements to determine KIE.

Table S11. Experimental KIE data for the reaction with **6** and **1d** and **1d-d₄**.

Independent	Standard	3d (7.51)	3d-d₃ (7.51)	3d / Standard	3d-d₃ / Standard	KIE
Run 1	2.00	0.38	-	0.19	-	
Run 2	2.00	0.45	-	0.23	-	
Run 1	2.00	-	0.26	--	0.13	1.46
Run 2	2.00	-	0.22	--	0.11	2.09
Average						1.77
Competition	(3d + 3d-d₃) _{obs} (2.57)	3d (2.57)		(3d-d₃) _{cal}		KIE
Run 1	2.00	1.82		1.18		1.54
Run 2	2.00	1.82		1.18		1.54
Average						1.54

 α -C(Sp³)-O–D/H Anaerobic oxidation.

Independent Rate Measurement. To an oven-dried, 0.5-dram vial, equipped with a stir bar was added of 3,5-Bis(trifluoromethyl)nitrobenzene (65 mg, 1.0 equiv., 0.25 mmol), and 1-phenylethan-1-ol (62 mg, 2.0 equiv., 0.20 mmol) in 0.5 mL of Acetone-d₆ and 0.5 mL of D₂O and left stirring until the OH peak of 1-phenylethan-1-ol was not detected by ¹H NMR, indicating formation of O–D. In a separate oven-dried, 0.5-dram vial, a second vial equipped with a stir bar was added of 3,5-Bis(trifluoromethyl)nitrobenzene (65 mg, 1.0 equiv., 0.25 mmol), and 1-phenylethan-1-ol (61.6 mg, 2.00 equiv., 0.20 mmol) in 0.5 mL of Acetone-d₆ and 0.5 mL of H₂O and left stirring for the same amount of time as the prior was used to confirm retention of the O–H peak. The reaction vessels were then subjected to 390 nm light for 1 h. A crude ¹H NMR was taken using CH₂Br₂ as an internal standard ((δ , ppm): 7.51). The KIE measurement is the average of two independent experiments.

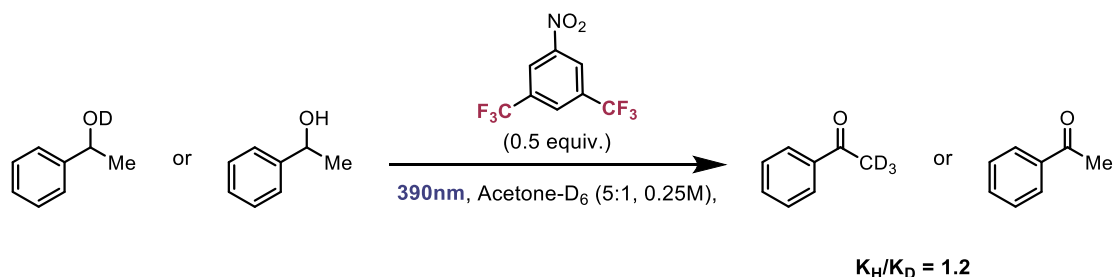
**Figure S8.** Independent rate measurements to determine KIE for O–D vs O–H.

Table S12. Experimental KIE data for the reaction of with **6** and **1d** and **1d-d₁**.

Independent	Standard	3d (7.51)	3d-d₃ (7.51)	3d / Standard	3d-d₃ / Standard	KIE
Run 1	1.00	0.50	-	0.50	-	
Run 2	2.00	0.59	-	0.59	-	
Run 1	2.00	-	0.59	--	0.59	1.18
Run 2	2.00	-	0.72	--	0.72	1.22
Average						1.20

Overall Results. To shed light on whether abstraction of a proton at either carbon or oxygen was occurring, first Kinetic Isotope Effects (KIEs) of PhCH(OH)Me were determined using independent rate measurements and intermolecular competition experiments (Table S11). Independent anaerobic oxidations of PhCD(OH)CD₃ revealed a KIE for the C(sp³)–H abstraction step, $k_H/k_D = 1.77$. Measurement of the competition experiments also showed an isotope effect for the benzylic ($k_H/k_D = 1.54$). These results indicate that for benzylic secondary alcohols, HAT at the α -C–H bond is possibly rate-limiting. Intermolecular competition experiments with PhCH(OD)Me and PhCH(OH)Me were not assessed due to the possible rapid exchange between alcohols of the H/D, which would lead to inconclusive results. However, Independent rate measurements were obtained, showing $k_H/k_D = 1.2$. This KIE is within the range for a secondary KIE of C(sp³) to C(sp²) transformations. This indicates that O–H/D bond breakage does occur after C–H breakage.

Radical clock study

Anaerobic Oxidation of a Radical Clock.

Anaerobic oxidation with radical clock, **18**⁵⁴ (112 mg, 1.00 equiv., 0.500 mmol). To a 0.5-dram oven-dried vial equipped with a stir bar was added 3,5-Bis(trifluoromethyl)nitrobenzene (389 mg, 3.00 equiv., 1.50 mmol) and LiOAc (8 mg, 0.25 equiv., 0.125 mmol) in a 5:1 MeCN/*t*BuOH mixture under N₂. The mixture was stirred at 1000 rpm under 390 nm light for 36 h. ¹H NMR spectrum was recorded using CH₂Br₂ as an internal standard. Products **20** and **21** were detected and their NMRs are in accordance with literature values.^{55a, 55b}

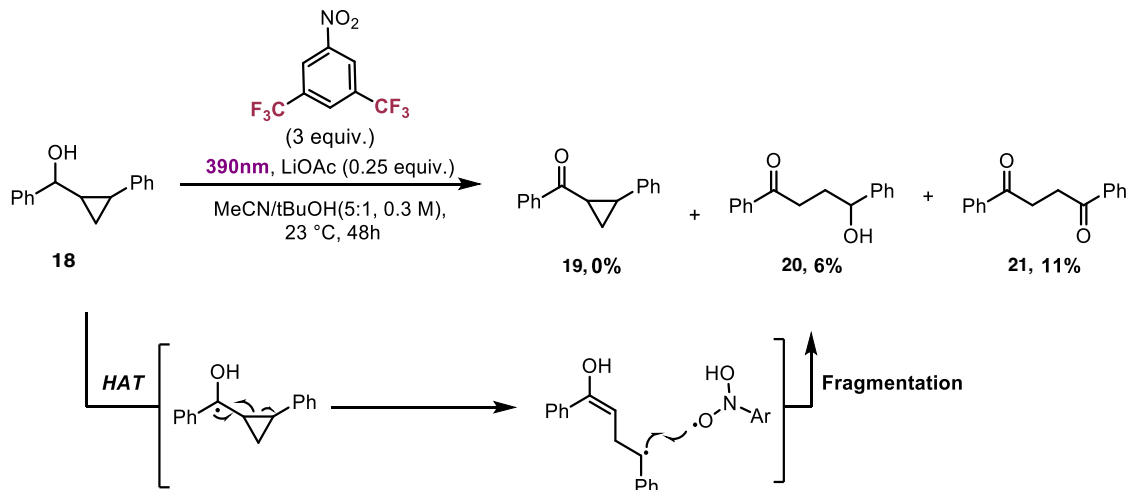
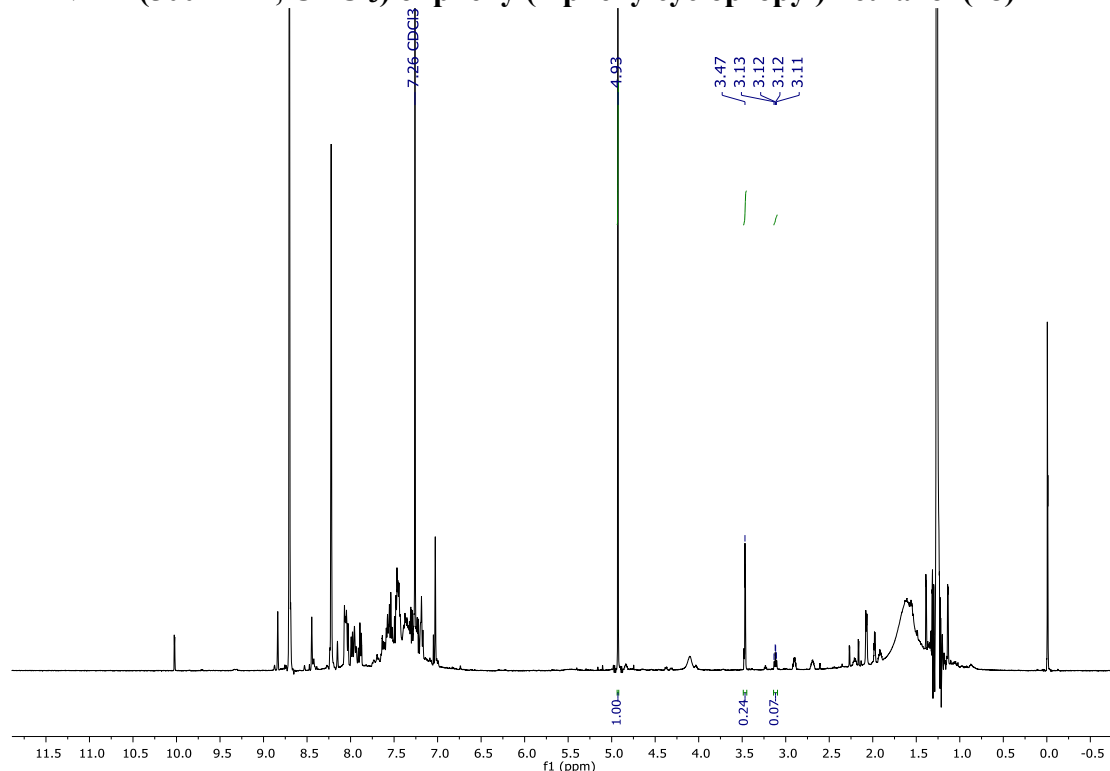


Figure S9. Radical clock subjected to the general reaction conditions.

Results. Support for a radically initiated process, at the α -C(sp³) center for these oxidations was provided from a radical clock study using phenyl(2-phenylcyclopropyl)methanol (Figure S9). The anaerobic oxidation revealed a mixture of 4-hydroxy-1,4-diphenylbutan-1-one (**20**) (δ , ppm): 3.13-3.11) and 1,4-diphenylbutane-1,4-dione (**21**) (δ , ppm): 3.47) as major products. These products likely occur after an HAT event, which ring opens to form a benzylic carbon-centered-radical that can be captured by the photoexcited-nitroarene, resulting in hydroxylation, a recombination step similar to previously reported methods. No ketone product (**19**) was observed.

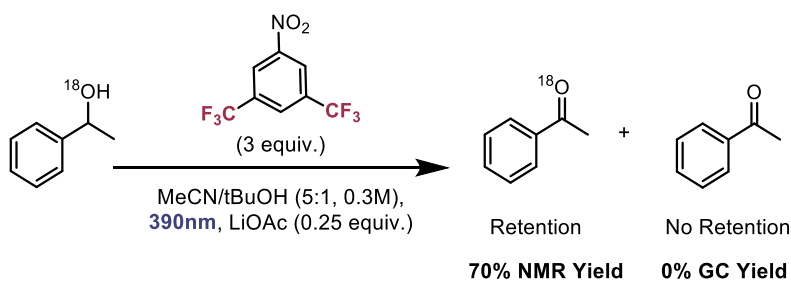
¹H NMR (500 MHz, CDCl₃) of phenyl(2-phenylcyclopropyl)methanol (18**)**



Labeling study

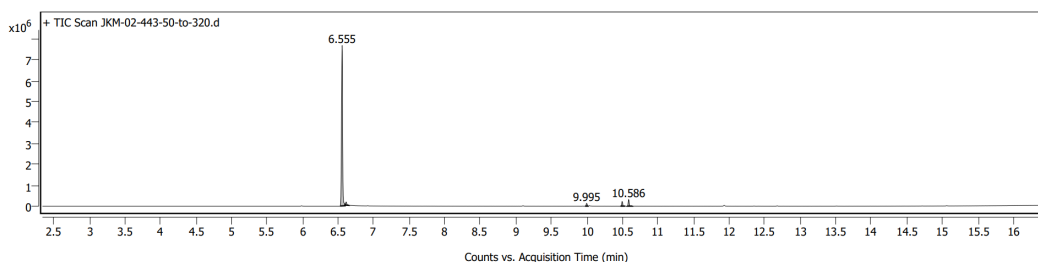
Anaerobic Oxidation of an ^{18}O -labelled Alcohol.

The anaerobic oxidation of ^{18}O -enriched phenylethane-1-ol (**1d- ^{18}O**)⁵⁶ was carried out with 3.0 equiv. of 3,5-Bis(trifluoromethyl)nitrobenzene under General Procedure A for 24 h. Based on GCMS analysis of the reaction mixtures, product **3d- ^{18}O** was solely found as opposed to **3d**, indicating ^{18}O was retained. ^1H NMR yield obtained using CH_2Br_2 as an internal standard showed 70% of product formation.

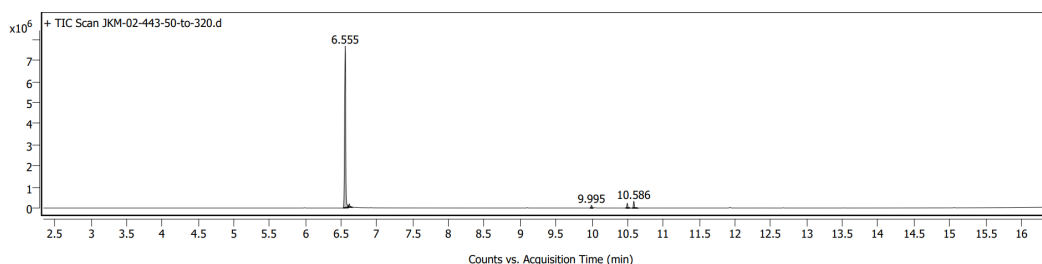


GC chromatograms of reactions with **1d- ^{18}O** .

Commercial Phenylethan-1-ol, **1d**



Synthesized Phenylethan-1-ol- d_1 , **1d- ^{18}O**



Reaction run with 1d-¹⁸O

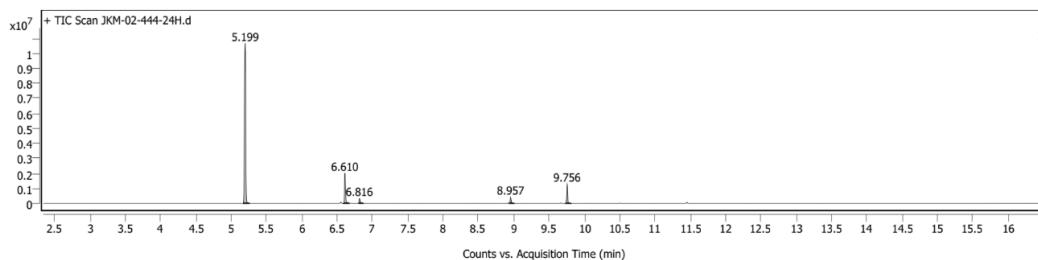
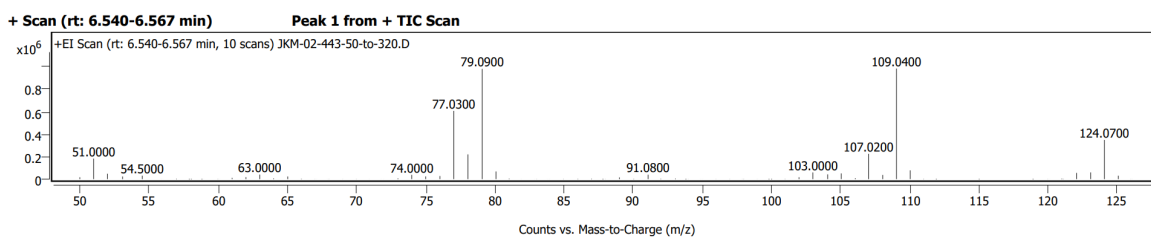


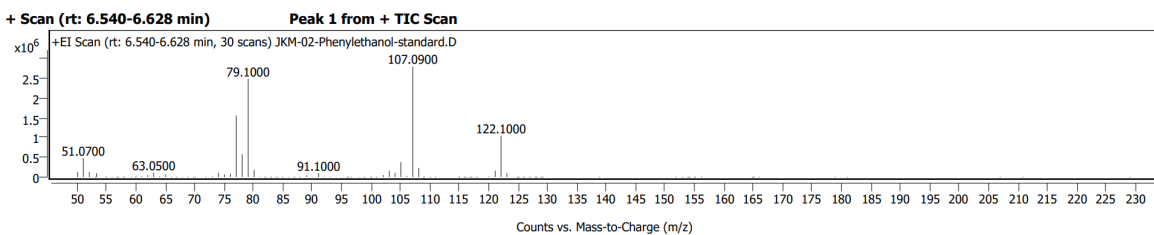
Figure S10. GCMS spectra of labeling study reactions with commercial Phenylethan-1-ol for comparison.

GCMS spectra of 1d, prepared 1d-¹⁸O, and reaction run with 1d-¹⁸O.

Commercial Phenylethan-1-ol, 1d



Synthesized Phenylethan-1-ol-d₁, 1d-¹⁸O



Reaction run with 1d-¹⁸O

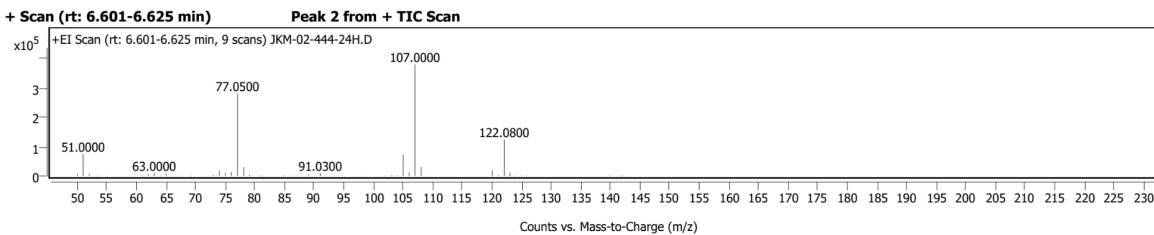


Figure S11. GCMS spectra of labeling studies reactions with commercial phenylethan-1-ol for comparison.

Hammett plot study

Independent experiments. The anaerobic oxidation of *para*-substituted phenylethane-1-ol (**1c-g**) were carried out independently. In an oven-dried vial equipped with a stir bar was charged alcohol (0.2 mmol, 1.0 equiv.), 3,5-Bis(trifluoromethyl)nitrobenzene (155 mg, 3 equiv., 0.6 mmol), and lithium acetate (3 mg, 0.2 equiv., 0.04 mmol) with MeCN/*t*BuOH (5:1, 0.3 M) under N₂. The reaction mixture was then irradiated under 390 nm light for 2 h. ¹H NMR yield obtained using CH₂Br₂ as an internal standard. Reactions were taken in duplicates.

Competition experiments. The anaerobic oxidation of *para*-substituted phenylethan-1-ol (**1c-g**) with phenylethane-1-ol were carried out independently. In an oven-dried vial equipped with a stir bar was charged *para*-substituted alcohol (0.2 mmol, 1.0 equiv.), phenylethane-1-ol (24 mg, 1.0 equiv., 0.2 mmol), 3,5-Bis(trifluoromethyl)nitrobenzene (155 mg, 3.00 equiv., 0.600 mmol), and lithium acetate (3 mg, 0.2 equiv., 0.04 mmol) with MeCN/*t*BuOH (5:1, 0.3 M) under N₂. The reaction mixture was then irradiated under 390 nm light for 2 h. ¹H NMR yield obtained using CH₂Br₂ as an internal standard. Reactions were taken in duplicates.

Results. For several *para*-substituted α -methyl benzyl alcohols, Hammett studies were conducted via independent rate measurements (Figure S12, Left) and competition experiments (Figure S12, Right). Of the varying Hammett Parameter (σ_p), independent rate measurements did not show significant electronic dependence among some of the substituents, but competition experiments did. The Hammett plots reveal preferential oxidation of electron-rich alcohols ($\rho = -0.45$). This observed Hammett is possible from loss of electron density and build-up of positive charge at the C(sp³)-H adjacent to the hydroxyl in the transition state, which has been observed in previous studies.

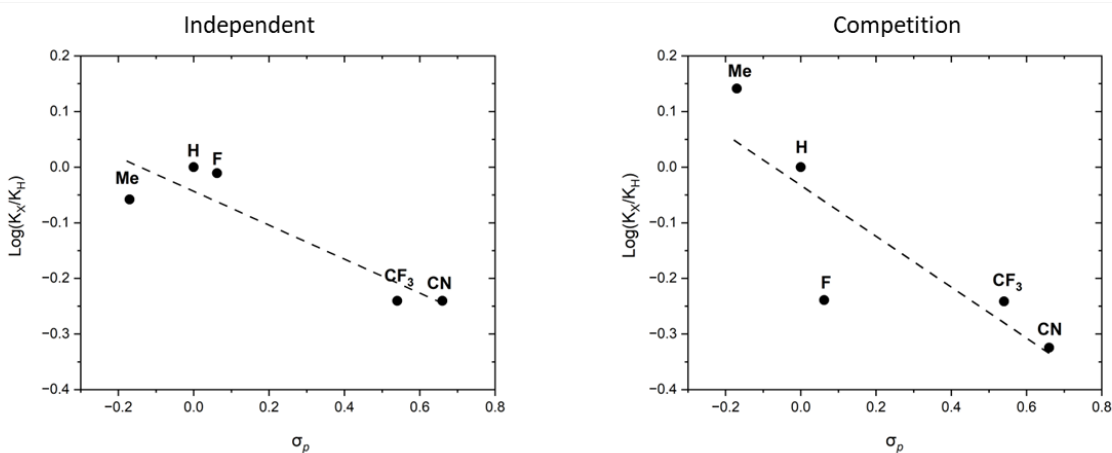


Figure S12. Hammett plots showing averaged $\log(K_H/K_X)$ vs. Hammett parameter for both independent (left) and competition (right) experiments.

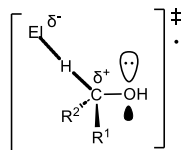


Figure S13. Representative loss of electron density as an electrophilic radical approaches the C(sp³)-H adjacent to the hydroxyl group in the transition state.

Pinacol probe study

Anaerobic Oxidation of a Pinacol Probe.

A carbocation formation was tested for the anaerobic oxidation of alcohols by subjecting a synthesized pinacol probe, 2-phenylbutane-2,3-diol⁵⁷ to General Procedure A. In an oven-dried vial equipped with a stir bar was charged 2-phenylbutane-2,3-diol (133 mg, 1.00 equiv., 0.800 mmol), 3,5-Bis(trifluoromethyl)nitrobenzene (207 mg, 1.00 equiv., 0.800 mmol), and lithium acetate (13 mg, 0.25 equiv., 0.20 mmol) with MeCN/*t*BuOH (5:1, 0.7 M) under N₂. The reaction mixture was then irradiated under 390 nm light for 18 h. The reaction was monitored by GCMS, and after completion, the crude reaction mixture was concentrated down and the product isolated by column chromatography.

Results. Methyl shift of by the pinacol probe was not observed, indicating the reaction likely does not proceed through a carbocation intermediate.

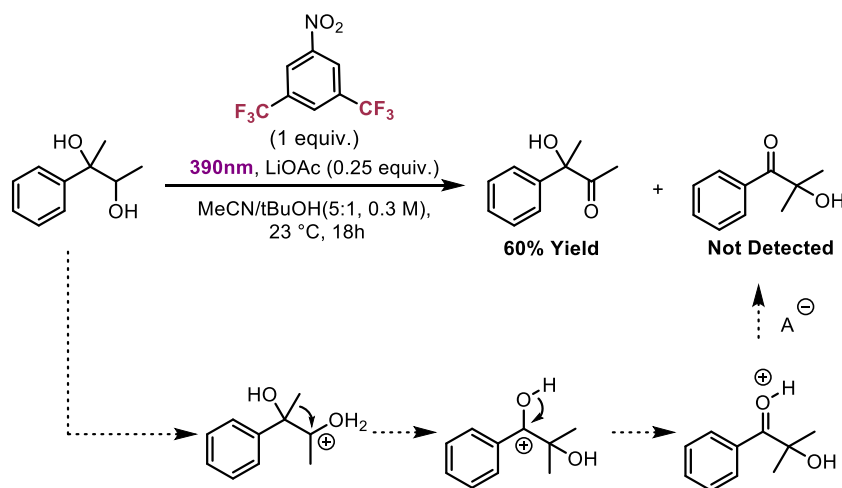
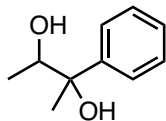


Figure S14. Pinacol Probe results after being subjected to Conditions A.

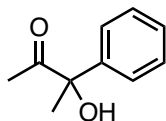


2-Phenylbutane-2,3-diol

To a solution of Acetoin (617 mg, 1.0 equiv., 7.00 mmol) in dry THF (5 mL) under argon atmosphere at $-78\text{ }^{\circ}\text{C}$, a solution of phenyl lithium dibutyl ether (706 mg, 4.42 mL, 1.9 molar, 1.2 equiv., 8.40 mmol) was added dropwise. The resulting mixture was then stirred at room temperature for 40 minutes, quenched with water (10 mL), and extracted with EtOAc ($3 \times 10\text{ mL}$). The combined organic layers were dried over Na_2SO_4 , filtered, and concentrated under reduced pressure. The residue was purified by flash chromatography (25 % EtOAc/Hexane) to give a colorless oil, 531 mg (yield: 86%). All analytical data for the title compound was in accordance with literature data.⁵⁷

$^1\text{H NMR}$ (500 MHz, CDCl_3) (δ , ppm): 7.39 – 7.33 (m, 2H), 7.30 – 7.22 (m, 2H), 7.21 – 7.14 (m, 1H), 3.87 (q, $J = 6.4\text{ Hz}$, 1H), 2.61 (s, 1H), 2.10 (s, 1H), 1.39 (s, 3H), 1.03 (d, $J = 6.5\text{ Hz}$, 3H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) (δ , ppm): 145.9, 128.4, 127.3, 125.6, 76.9, 74.3, 23.3, 16.6.



3-Hydroxy-3-phenylbutan-2-one

Following General Procedure A, prepared from 0.50 mmol of synthesized 2-phenylbutane-2,3-diol in 24 h. The title compound was isolated via flash chromatography (1 – 5% Acetone/Hexane). All analytical data for the title compound was in accordance with literature data.⁵⁸

$^1\text{H NMR}$ (400 MHz, CD_3CN) (δ , ppm): 7.50 – 7.42 (m, 2H), 7.42 – 7.33 (m, 2H), 7.35 – 7.26 (m, 1H), 4.34 (s, 1H), 2.04 (s, 3H), 1.65 (d, $J = 1.1\text{ Hz}$, 3H).

$^{13}\text{C NMR}$ (101 MHz, CD_3CN) (δ , ppm): 210.9, 143.6, 129.4, 128.5, 126.5, 81.1, 25.5, 24.2.

Light on/off study

3,5-Bis-trifluoromethylnitrobenzene (69 mg, 0.25 mmol, 1.0 equiv.), 1-(4-methoxyphenyl)ethan-1-ol (76 mg, 0.50 mmol, 2.0 equiv.), and deuterated-MeCN (0.40 mL, 250 mM) were added under nitrogen to an NMR tube. Under a PhotoNMR setup, a fiber optic cable connected to a 395 nm lamp was added to the reaction, and ^1H NMR experiments were taken every 90 seconds. Product growth was monitored by the integration of the peak from 6.94-7.02 ppm; starting material conversion was measured from the integration of the peak from 6.80-6.92 ppm. An internal standard of CH_2Cl_2 was used. The lack of product formation in the dark during these studies is evidence against a radical chain mechanism.

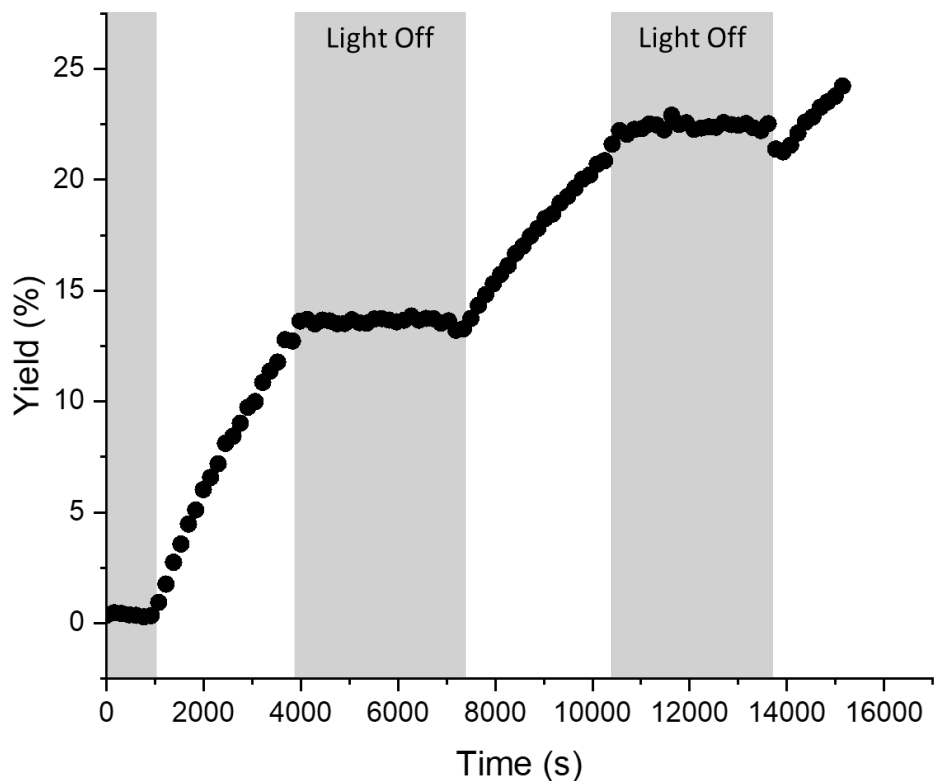
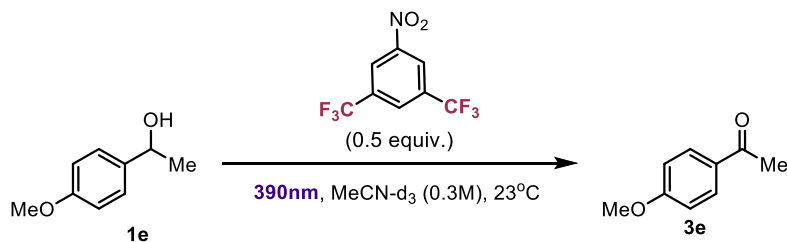


Figure S15. A Photo-NMR, light on/off study for the production of **3b** at 395 nm.

Discussion of mechanistic studies

We proposed that the oxidation reaction is initiated by hydrogen atom transfer (HAT) of the α -C(sp³)-H bond with the photoexcited state of the nitroarene. To verify this, kinetic isotope effect (KIE) studies of **1d/1d-d₉** were conducted (Scheme S16A). KIE values of 1.77 and 1.54 were obtained for parallel and intermolecular studies, respectively. Next, a parallel study of the second HAT step was investigated with **1d/1d-d₁** and resulted in a secondary KIE of 1.2 (Scheme S16B). These studies suggest that the first HAT participates in the rate-limiting step of the transformation. To support the radical nature of the transformation, radical clock **18** was subjected to the reaction conditions (Scheme S16C). Although low conversion of 23% was observed, the oxidation product with the cyclopropane moiety preserved was not observed **19**. Only the ring opening over-oxidation products **19** and **21** were detected. The presence of **19** indicates that radical recombination of the formed alkyl radical and the formed *N*-hydroxy-*N*-phenylhydroxylamine radical via oxygen atom transfer (OAT) from the nitroarene could be possible in the oxidation reaction. To distinguish if a recombination event is operative, ¹⁸O-labeled substrate **1d-¹⁸O** was tested (Scheme S16D). Upon exposure to the reaction conditions, the ¹⁸O-incorporated retention product **3d-¹⁸O** was obtained with no scrambling product detected. Hence, supporting that the oxidation of C(sp³)-heteroatom systems do not occur through radical recombination but rather a double HAT event.

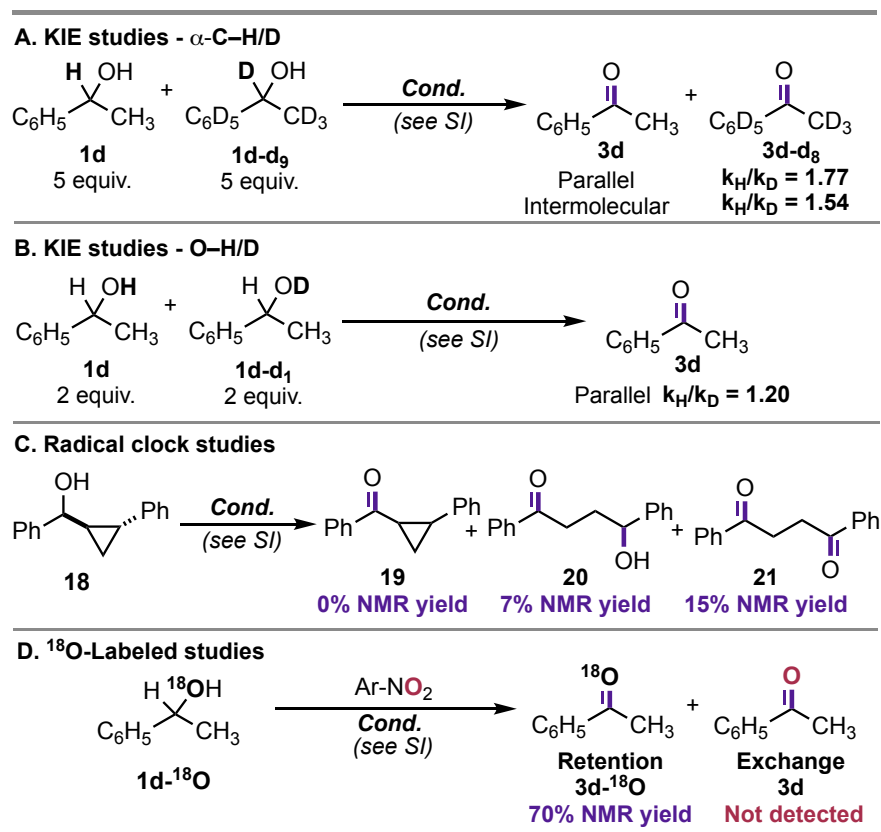
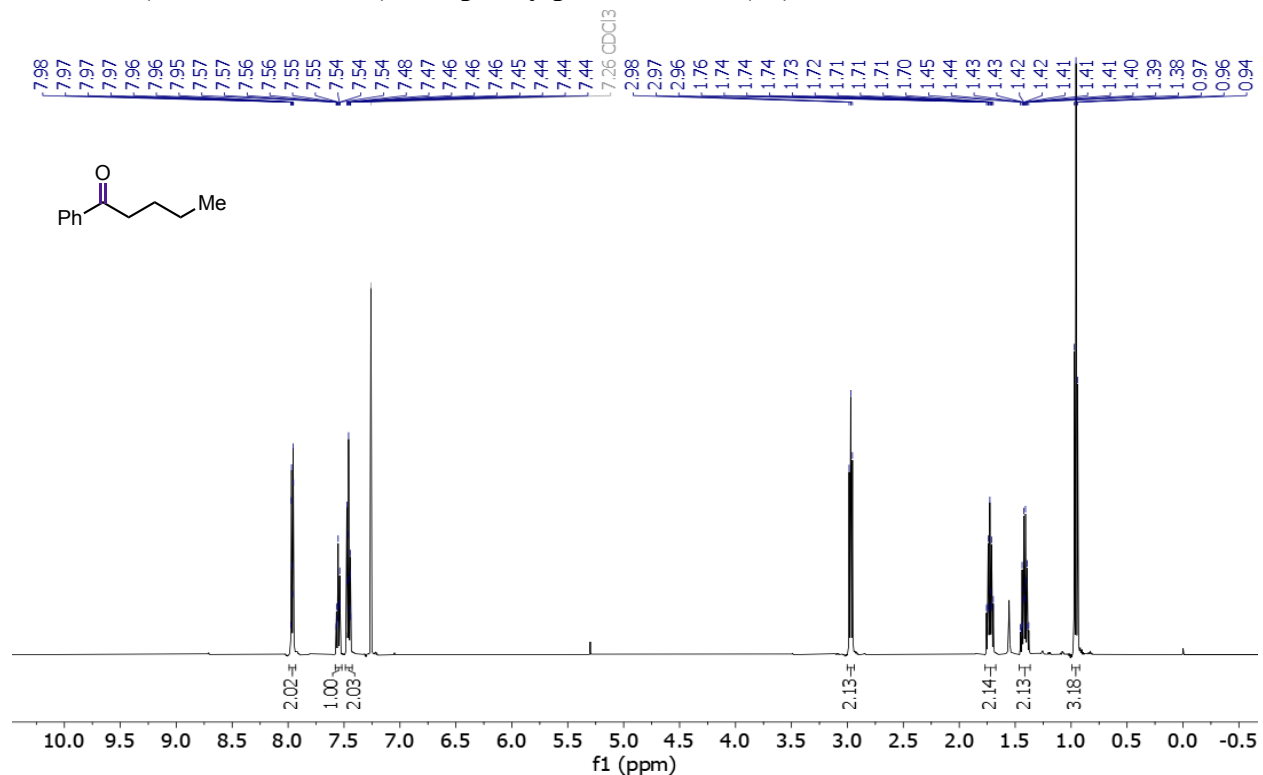


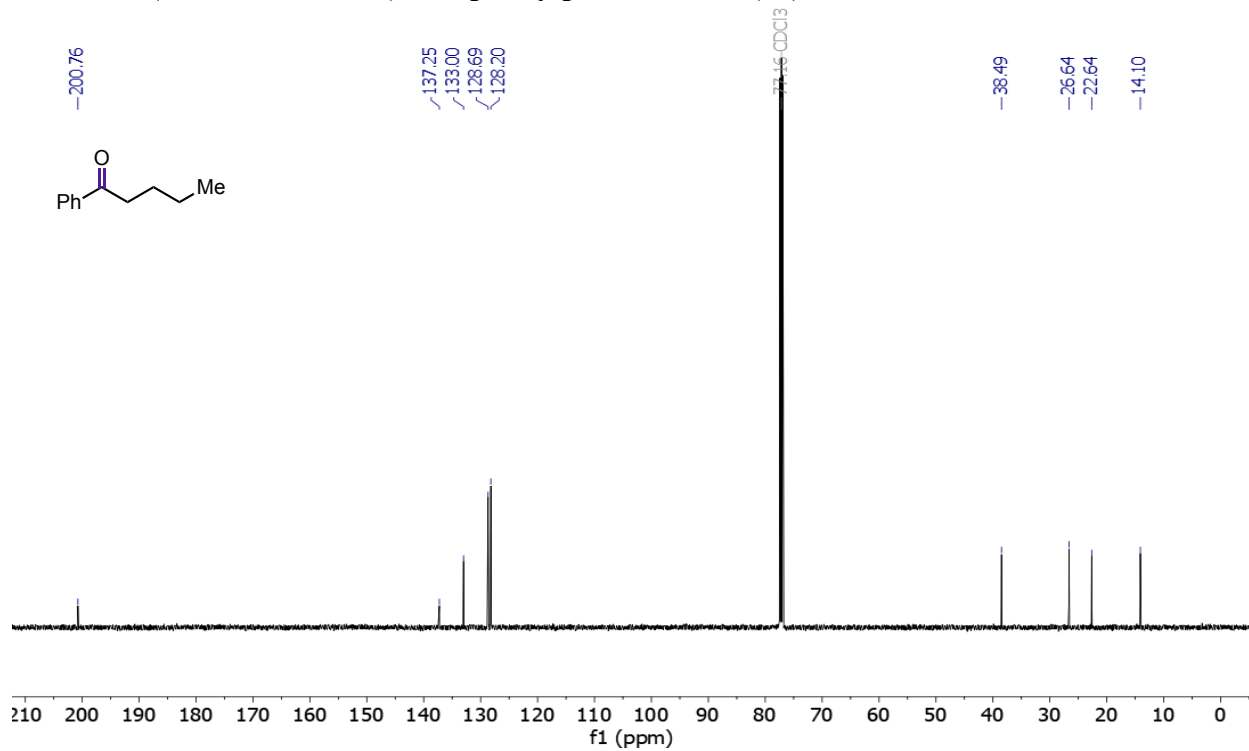
Figure S16. Summary of mechanistic studies

NMR spectra

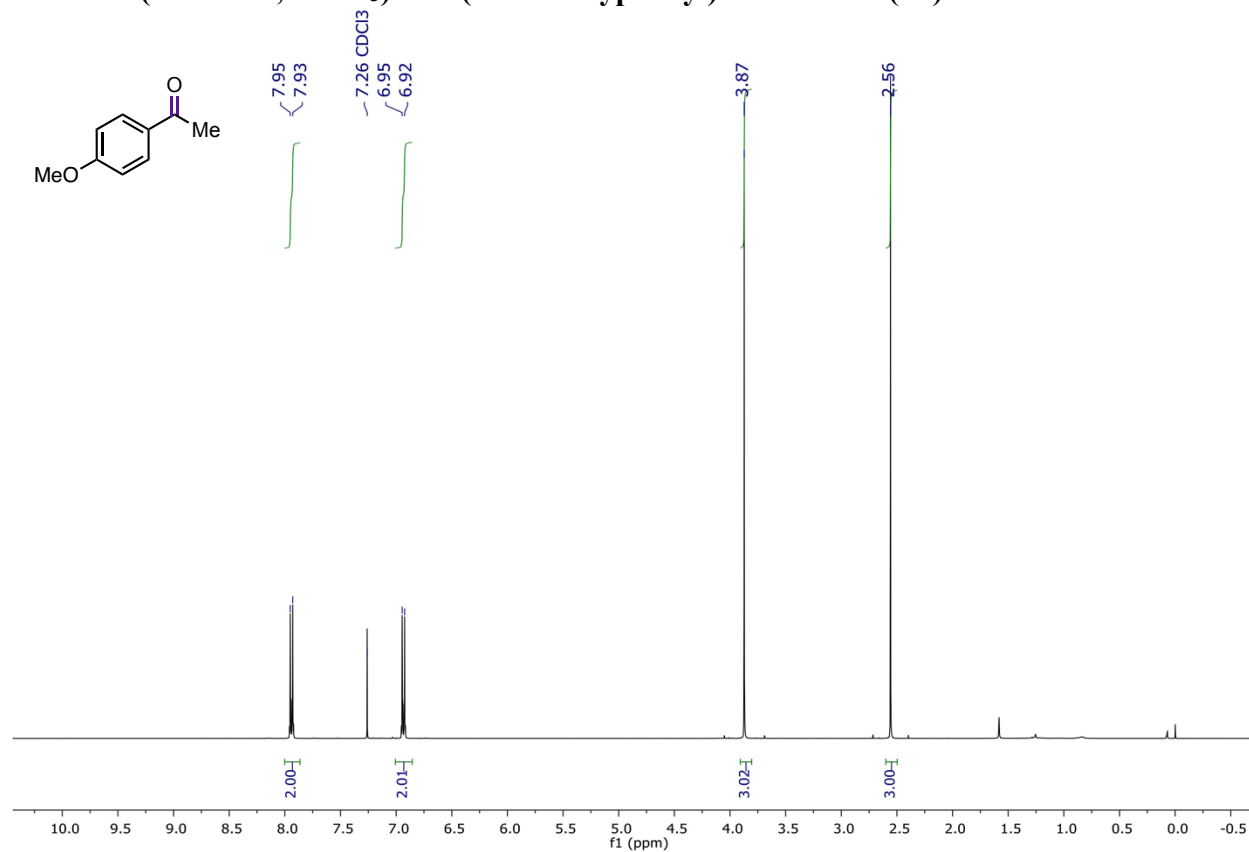
^1H NMR (500 MHz, CDCl_3) of 1-phenylpentan-1-one (3a)



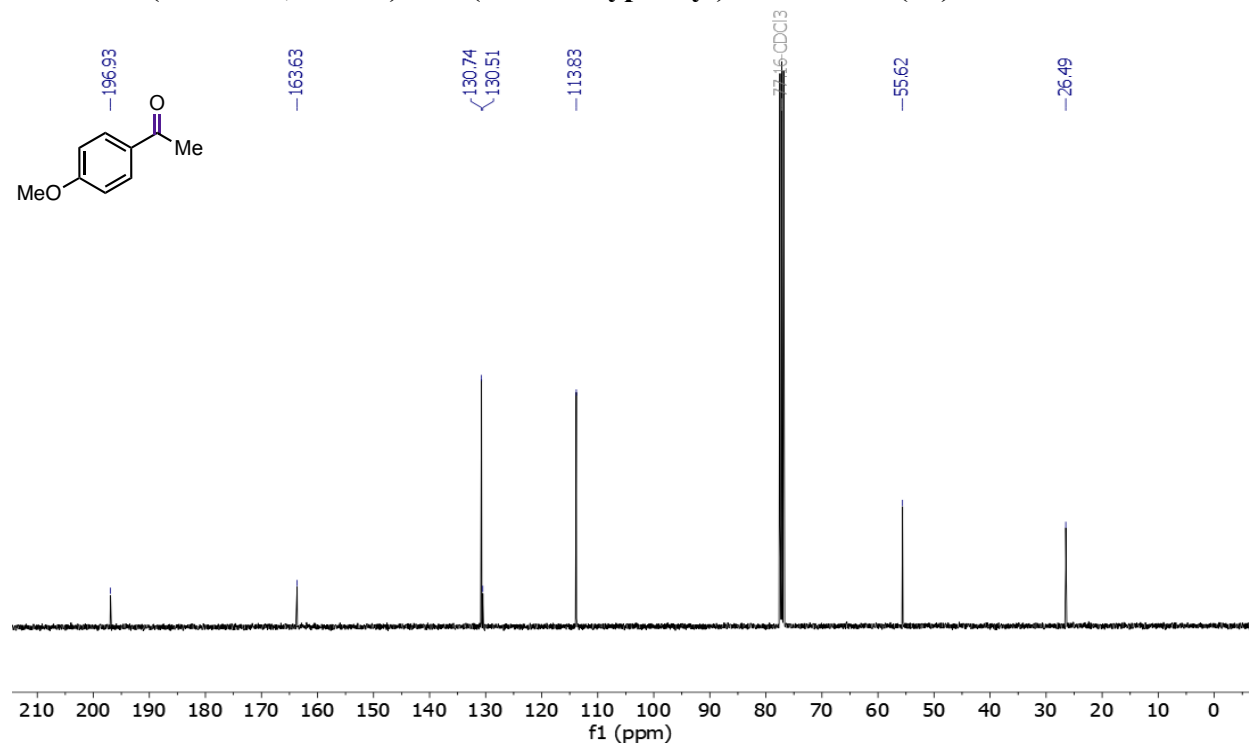
^{13}C NMR (126 MHz, CDCl_3) of 1-phenylpentan-1-one (3a)



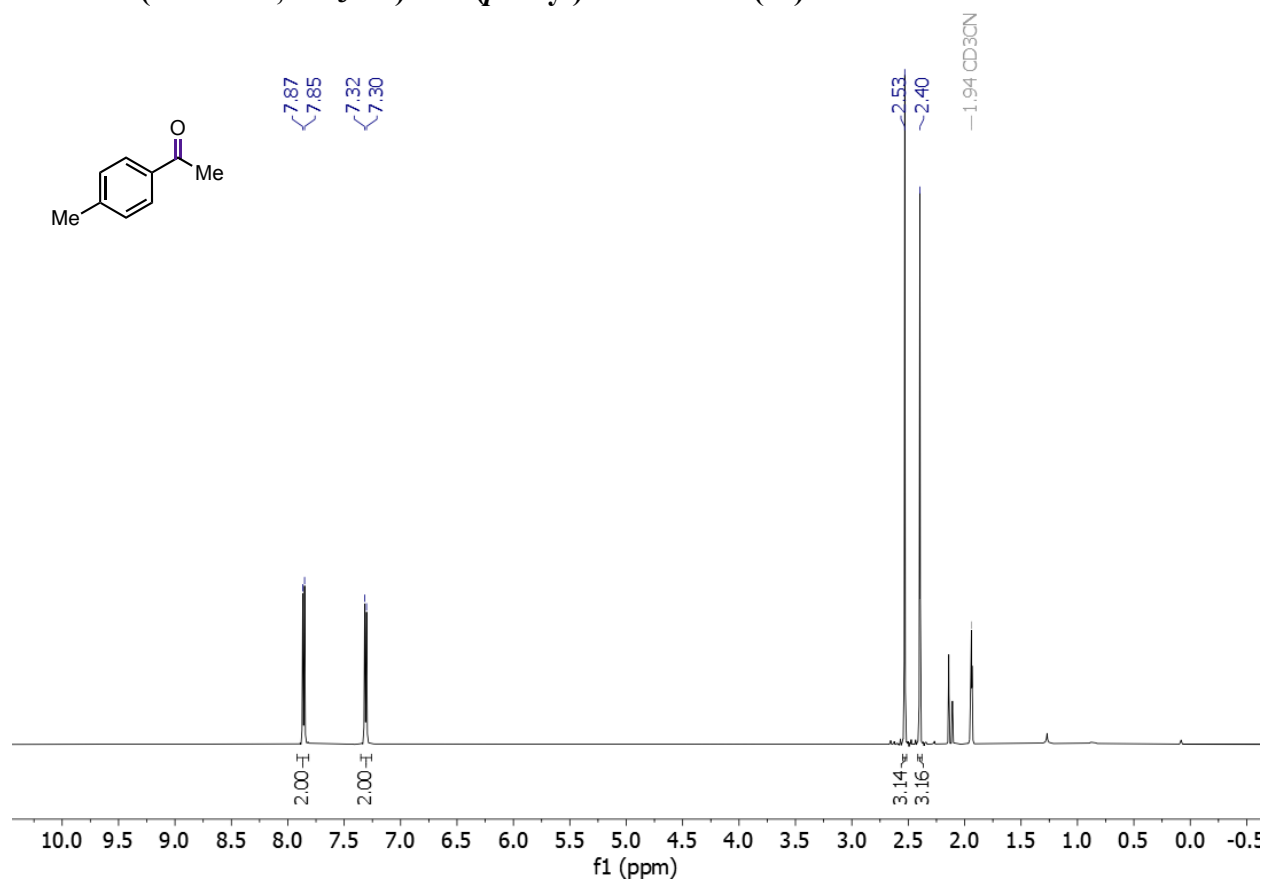
¹H NMR (400 MHz, CDCl₃) of 1-(4-methoxyphenyl)ethan-1-one (3b)



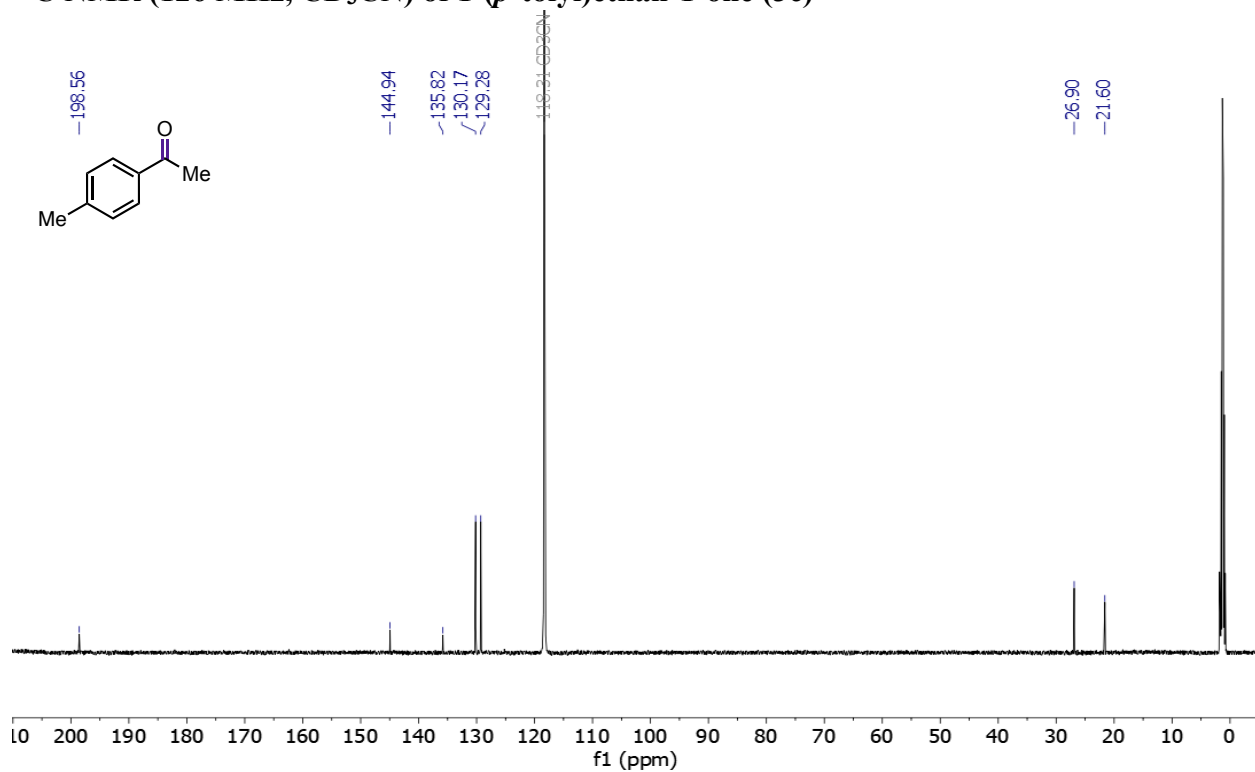
¹³C NMR (101 MHz, CDCl₃) of 1-(4-methoxyphenyl)ethan-1-one (3b)



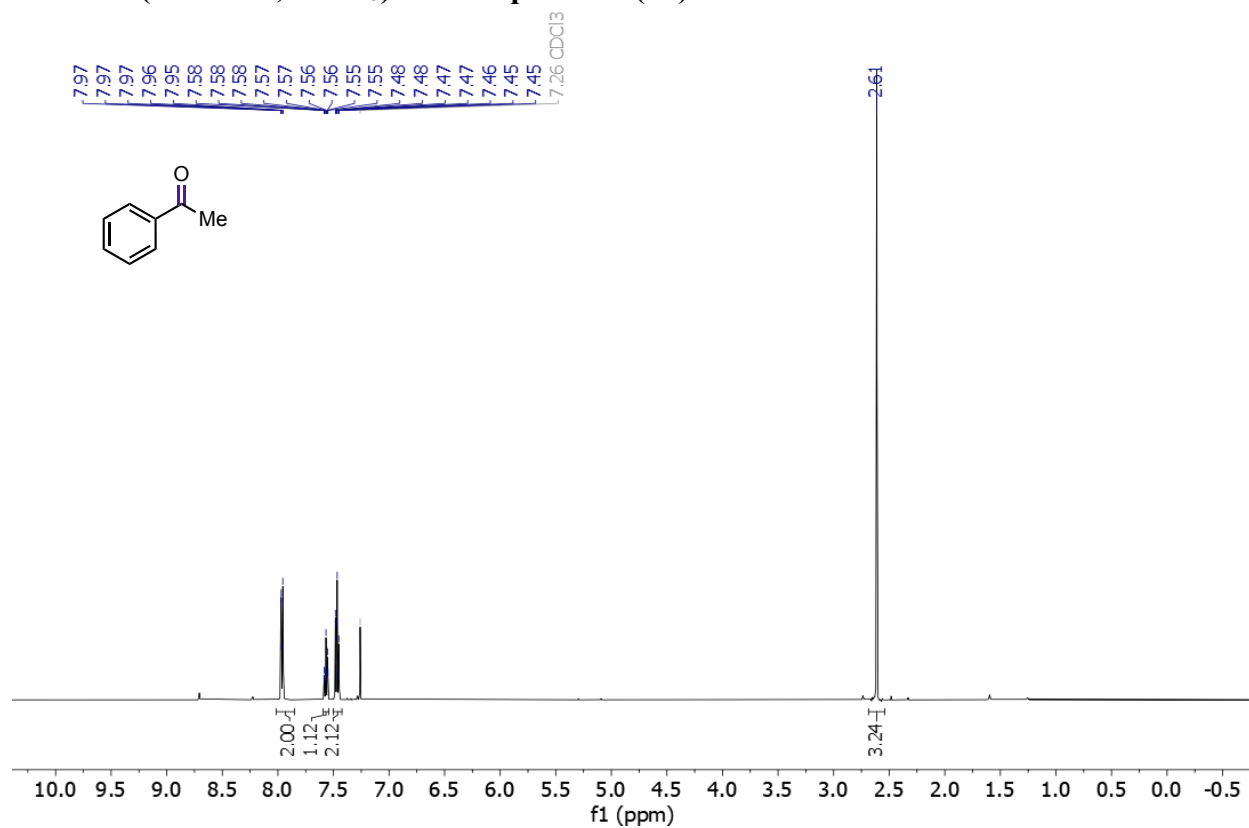
¹H NMR (500 MHz, CD₃CN) of 1-(*p*-tolyl)ethan-1-one (3c)



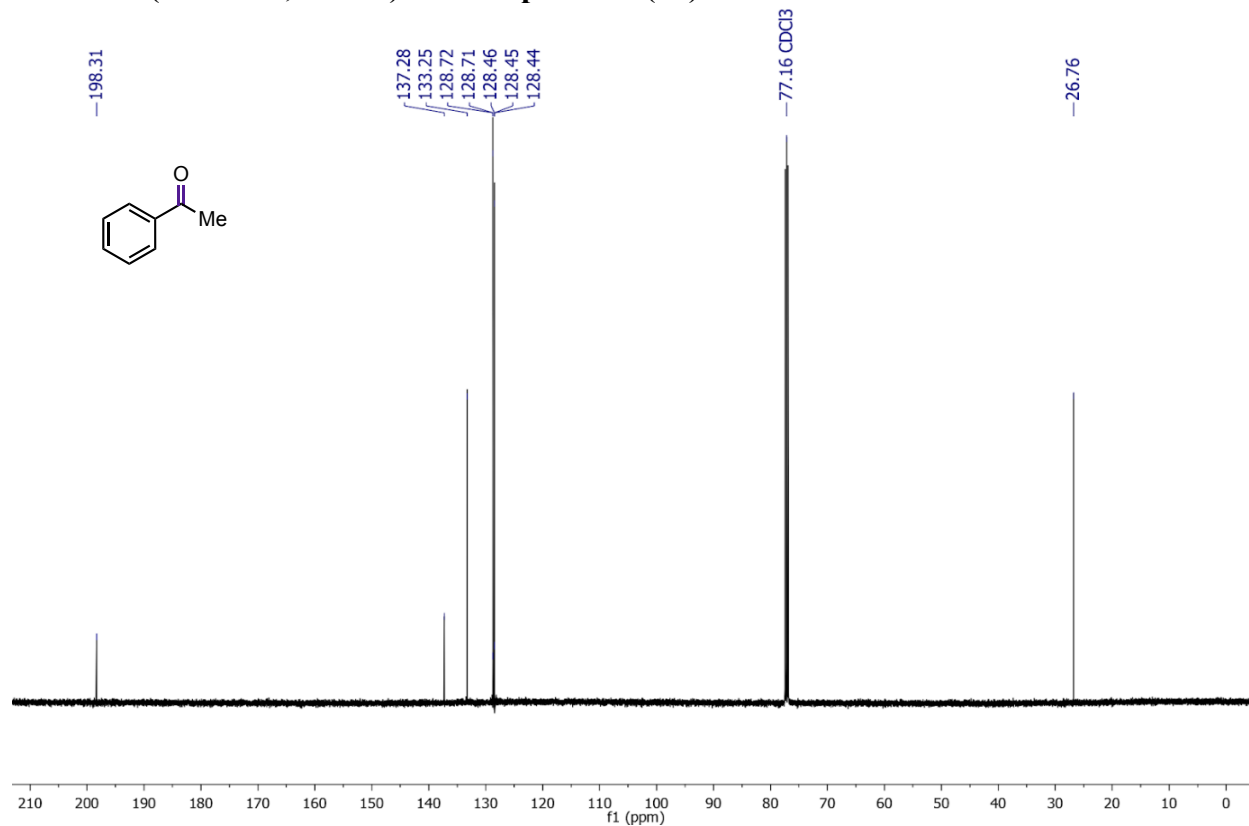
¹³C NMR (126 MHz, CD₃CN) of 1-(*p*-tolyl)ethan-1-one (3c)



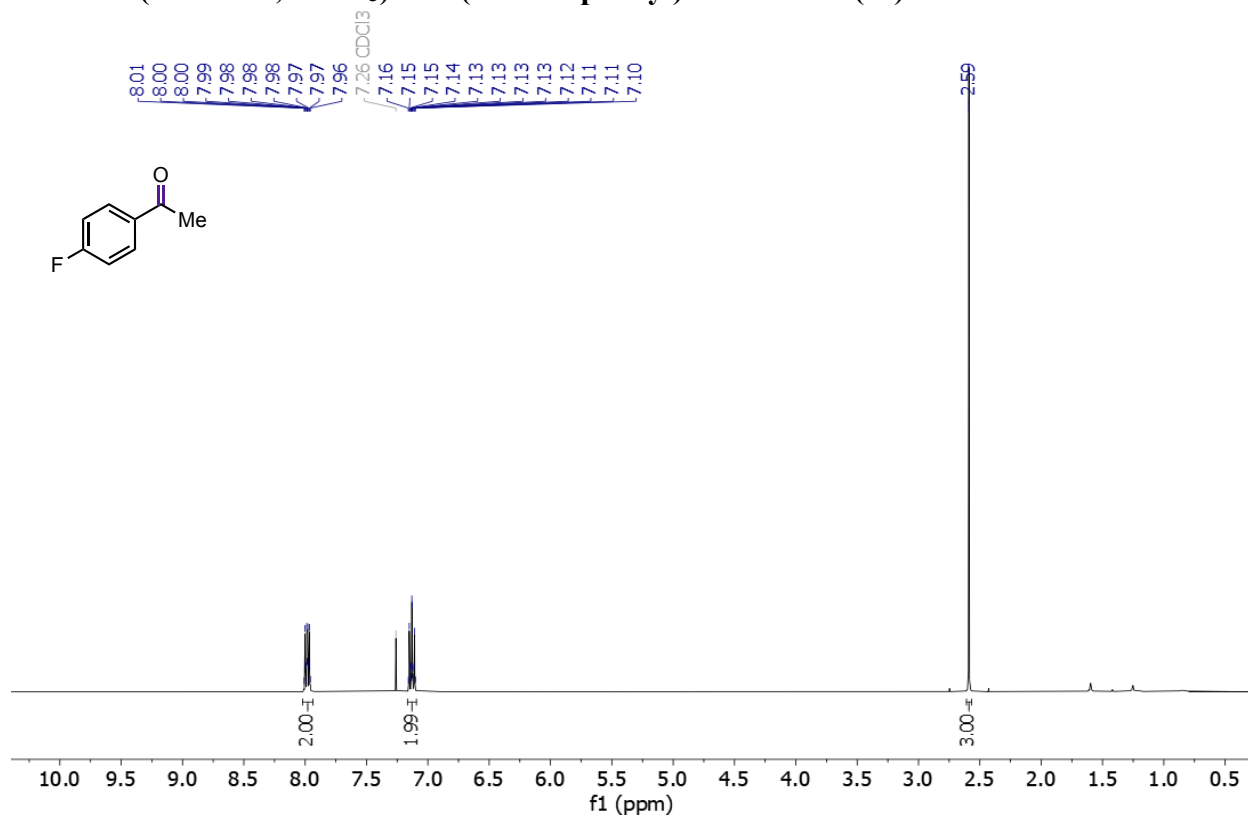
¹H NMR (500 MHz, CDCl₃) of Acetophenone (3d)



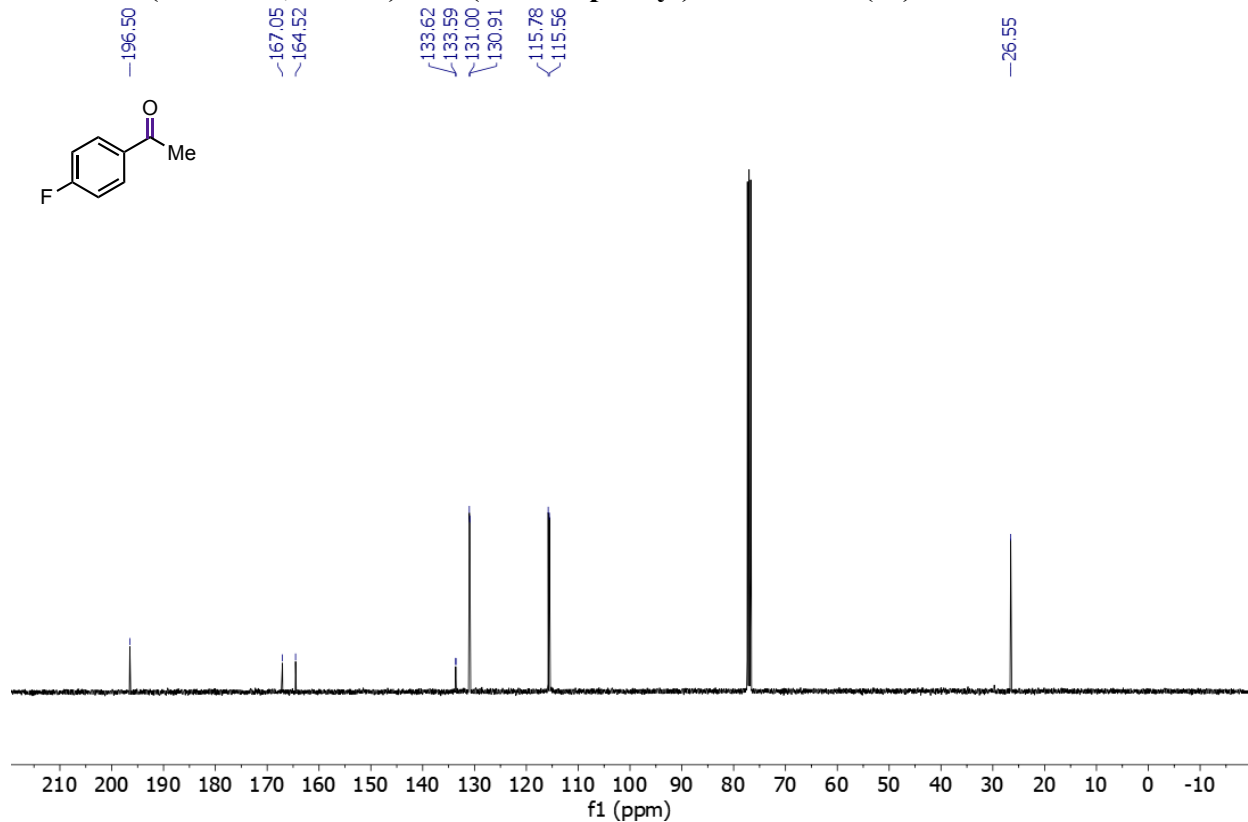
¹³C NMR (126 MHz, CDCl₃) of Acetophenone (3d)



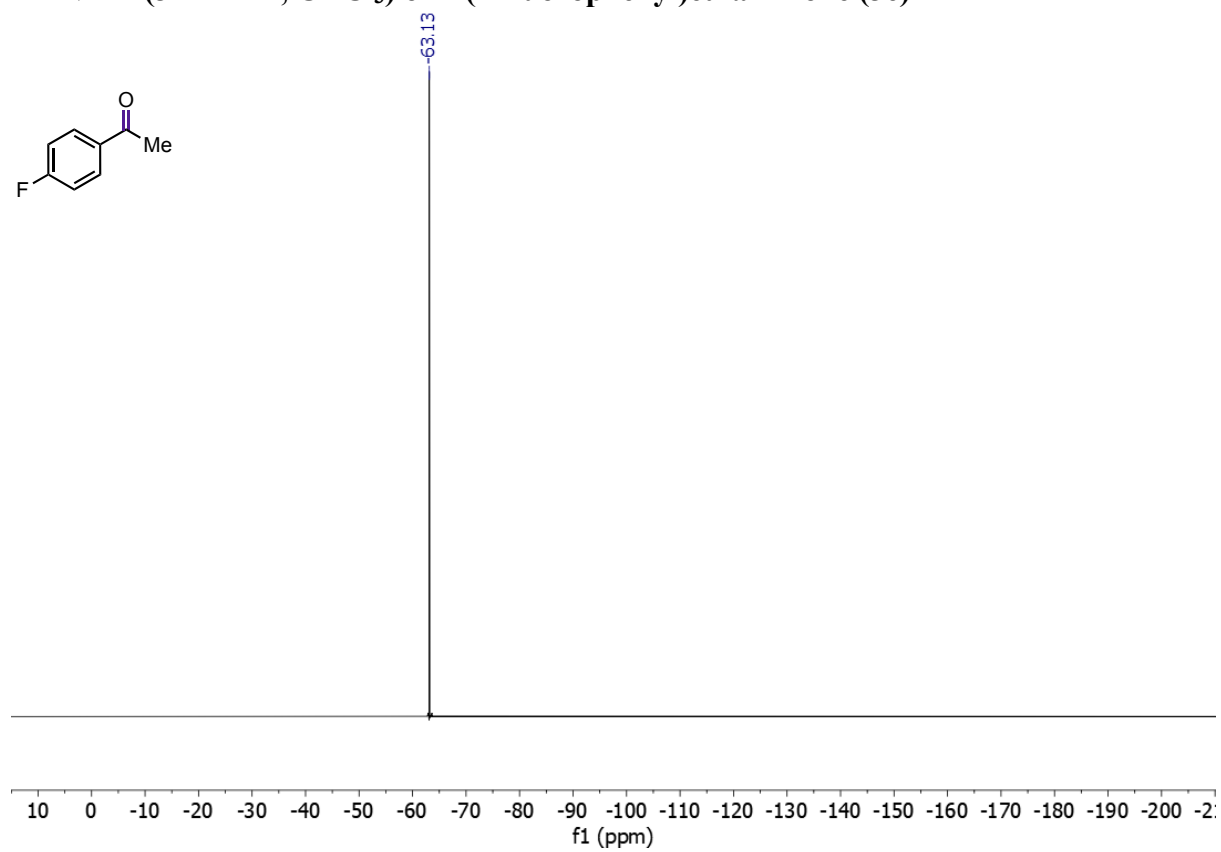
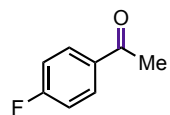
¹H NMR (400 MHz, CDCl₃) of 1-(4-fluorophenyl)ethan-1-one (3e)



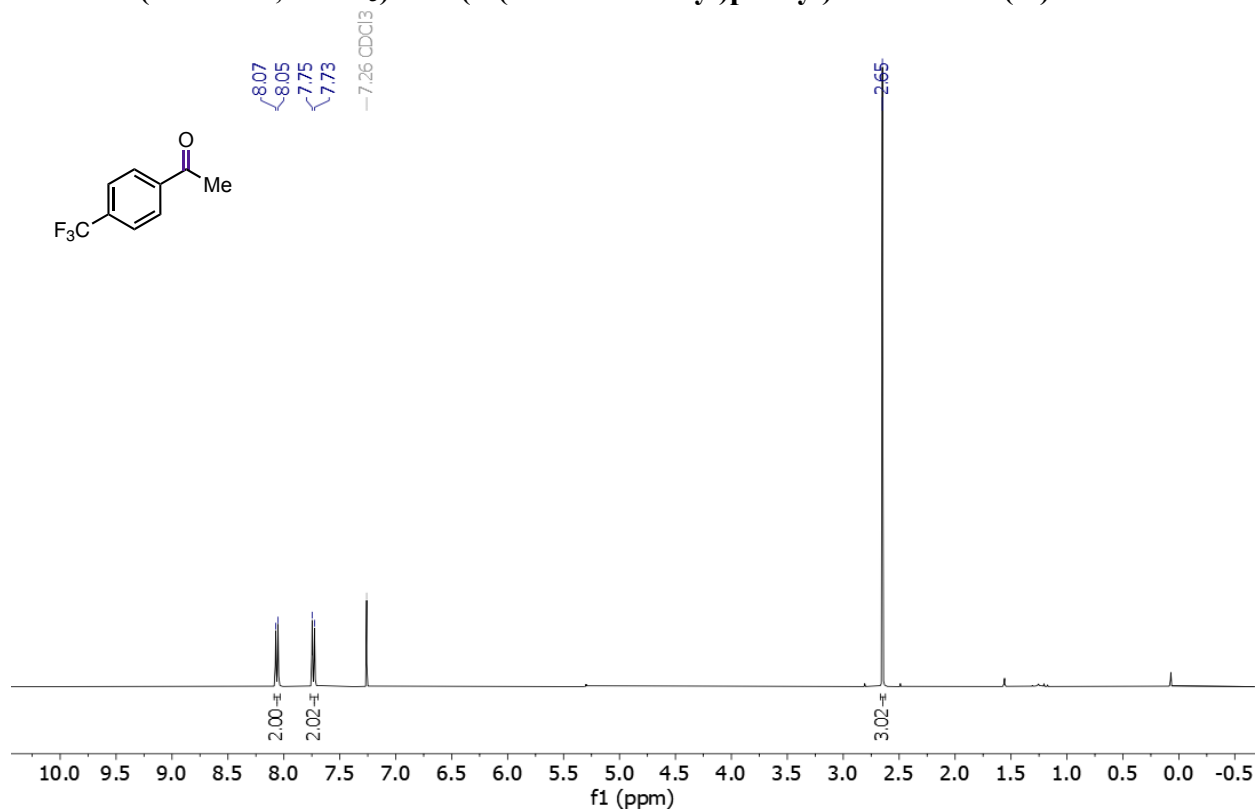
¹³C NMR (101 MHz, CDCl₃) of 1-(4-fluorophenyl)ethan-1-one (3e)



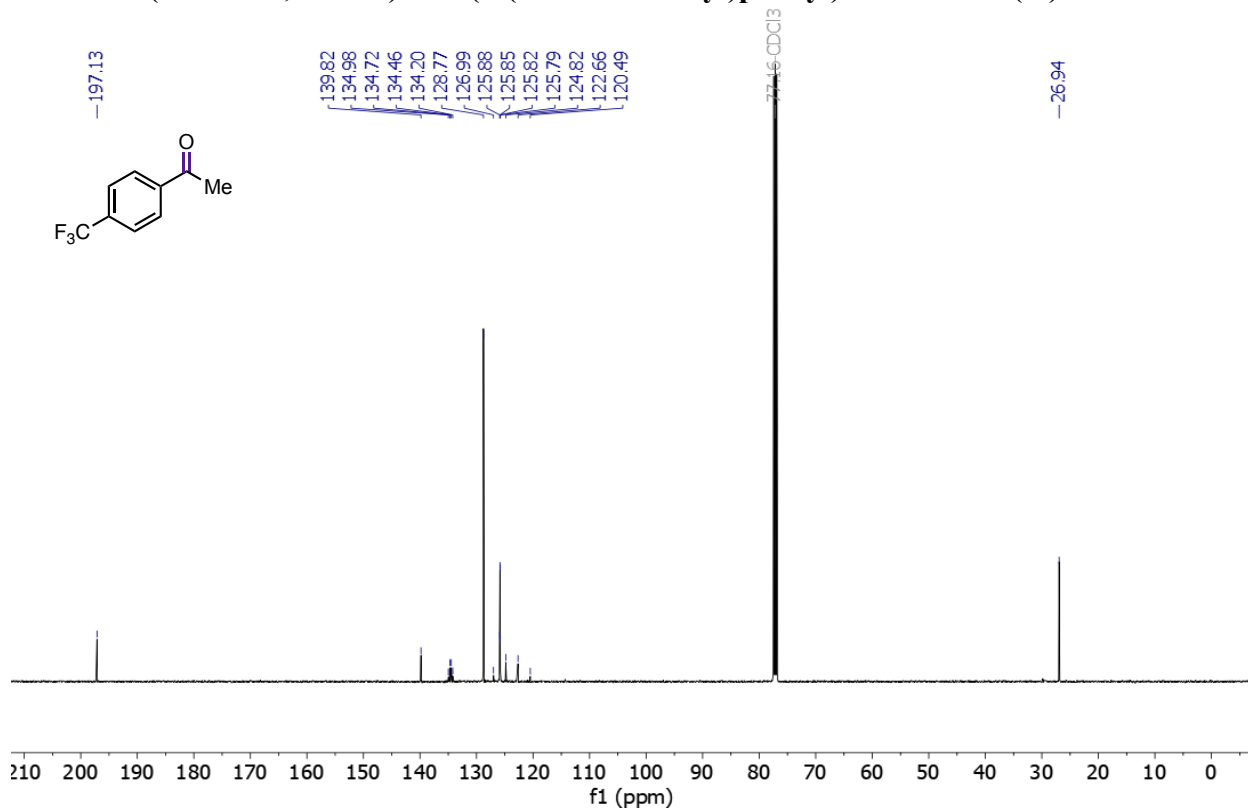
^{19}F NMR (377 MHz, CDCl_3) of 1-(4-fluorophenyl)ethan-1-one (3e)



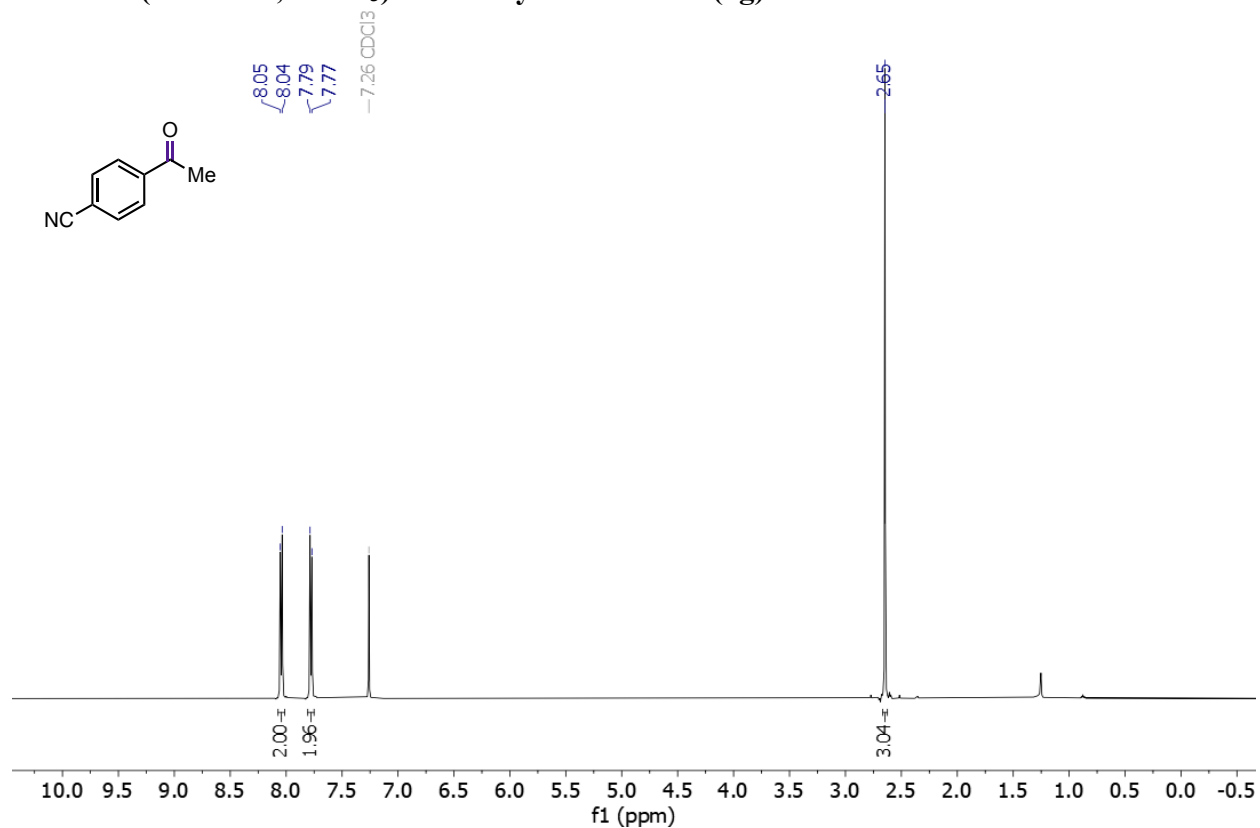
¹H NMR (400 MHz, CDCl₃) of 1-(4-(trifluoromethyl)phenyl)ethan-1-one (3f)



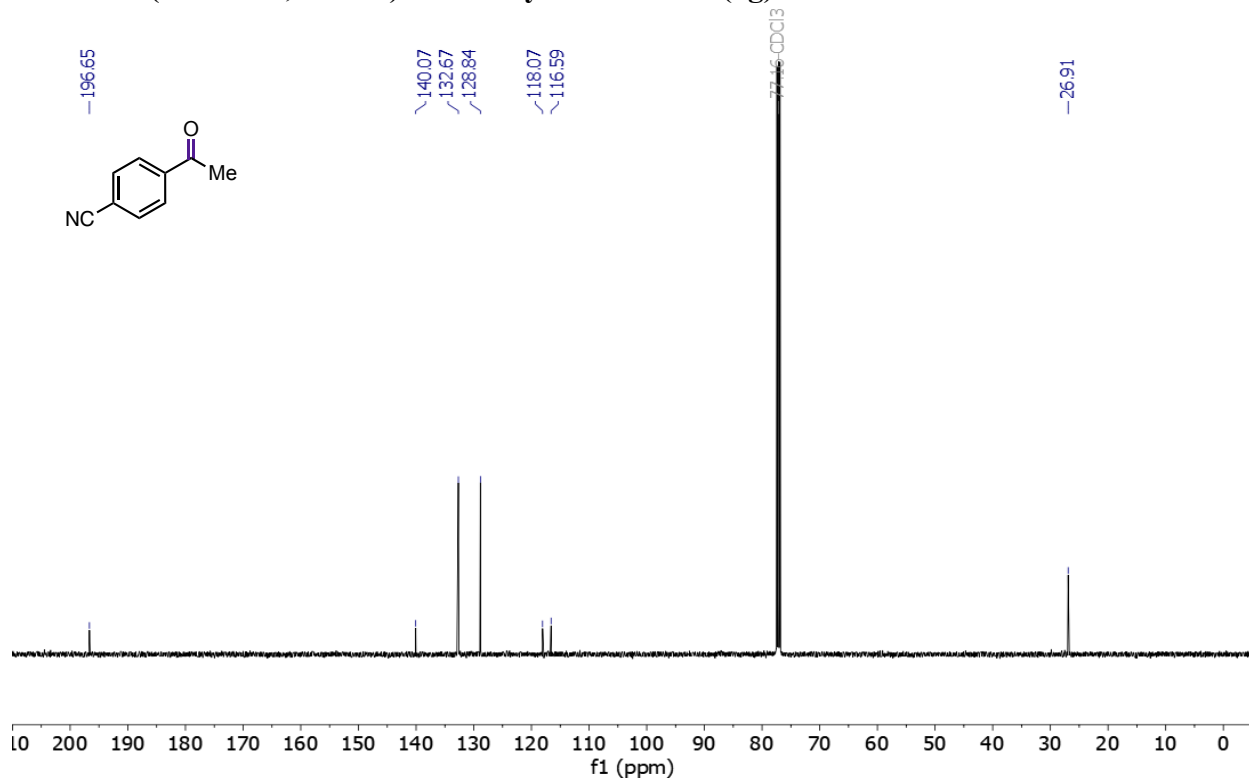
¹³C NMR (126 MHz, CDCl₃) of 1-(4-(trifluoromethyl)phenyl)ethan-1-one (3f)



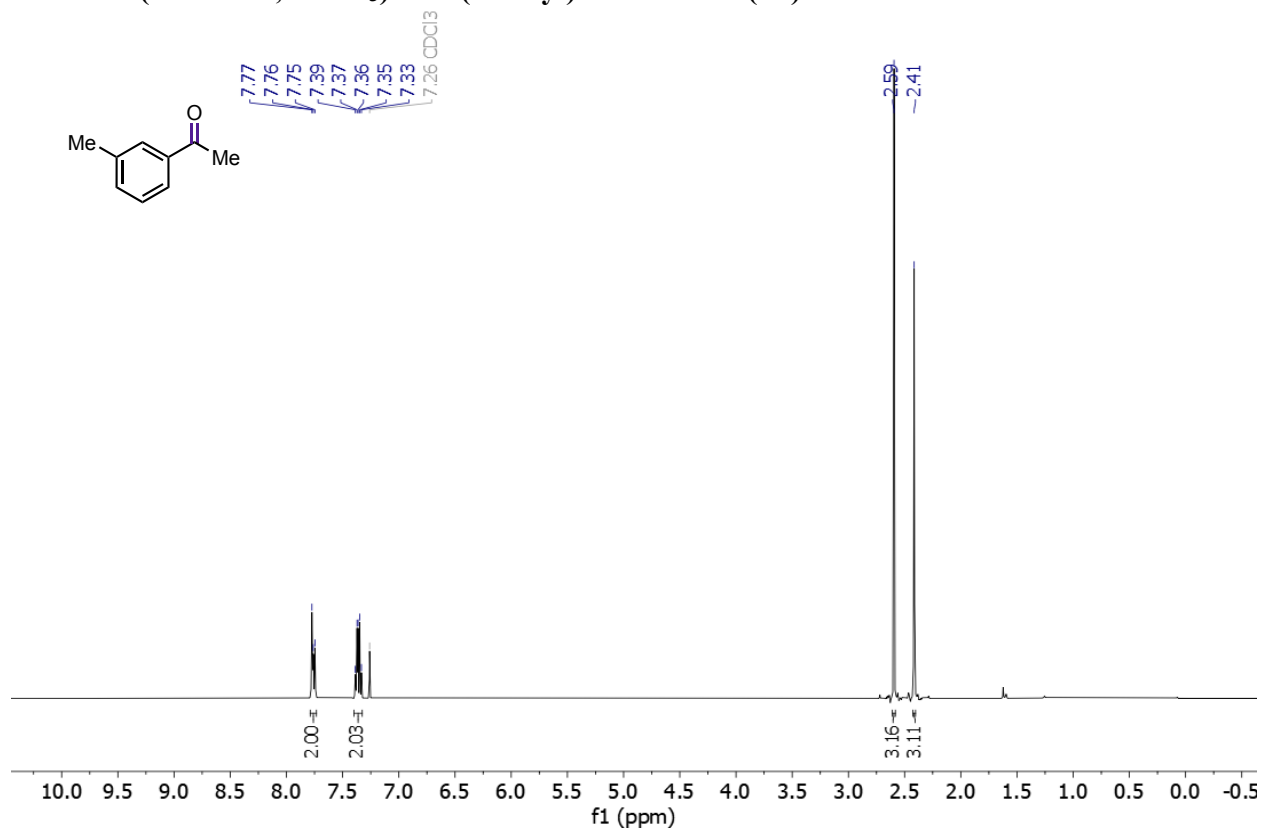
¹H NMR (500 MHz, CDCl₃) of 4-acetylbenzotrile (3g)



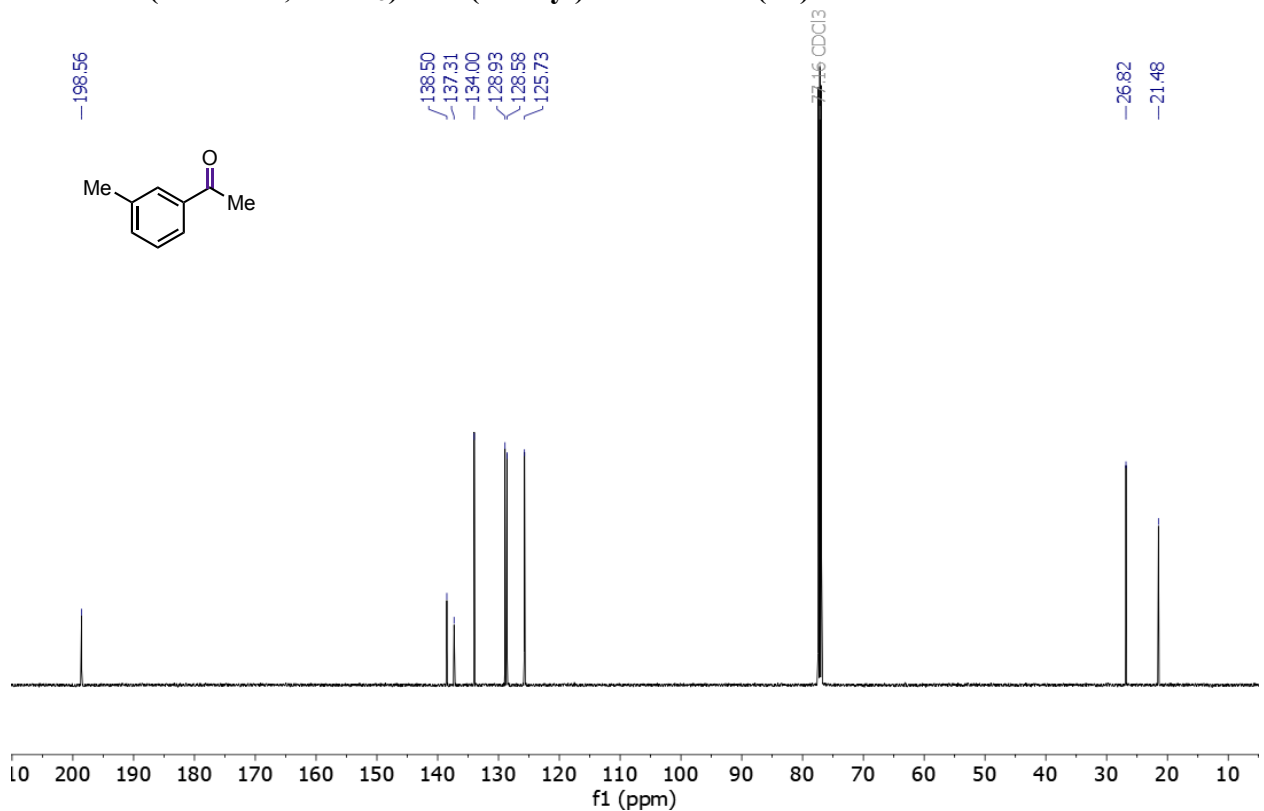
¹³C NMR (126 MHz, CDCl₃) of 4-acetylbenzotrile (3g)



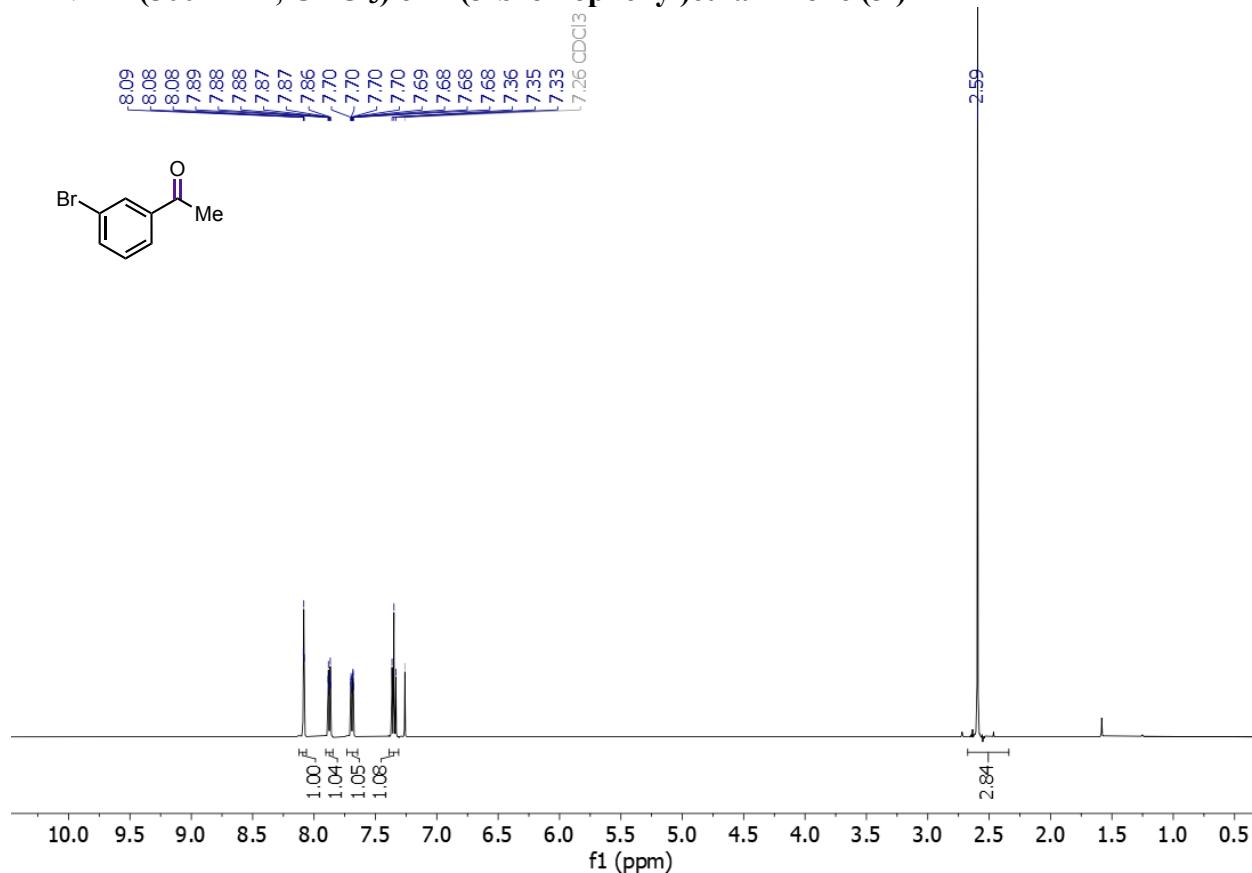
¹H NMR (500 MHz, CDCl₃) of 1-(*m*-tolyl)ethan-1-one (3h)



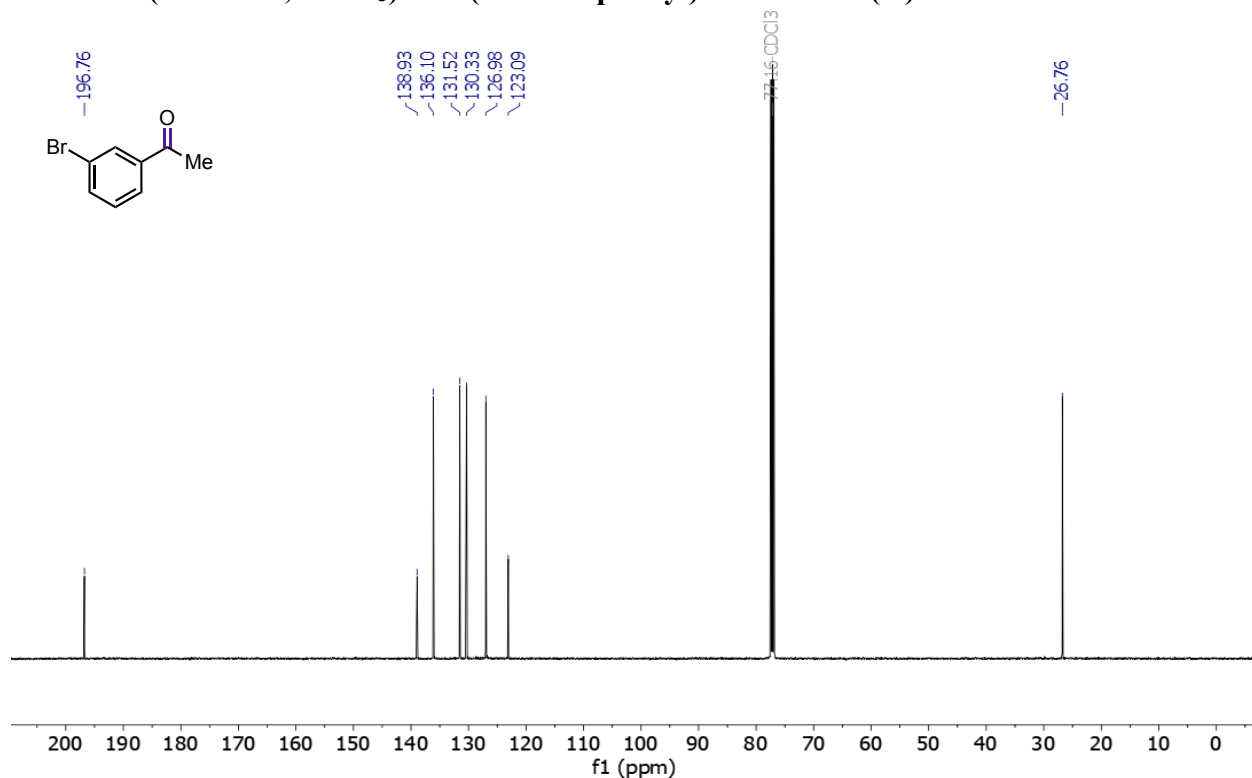
¹³C NMR (126 MHz, CDCl₃) of 1-(*m*-tolyl)ethan-1-one (3h)



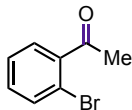
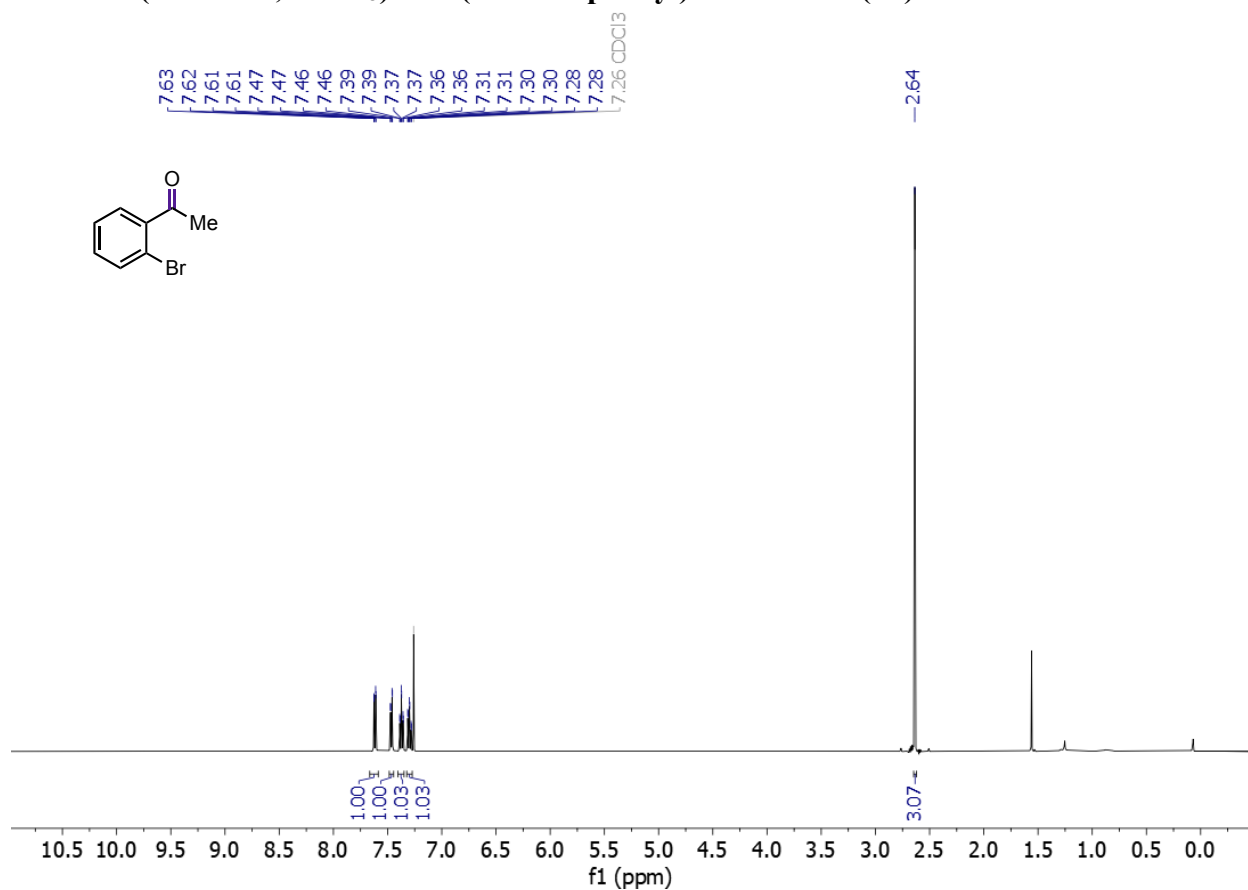
¹H NMR (500 MHz, CDCl₃) of 1-(3-bromophenyl)ethan-1-one (3i)



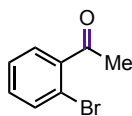
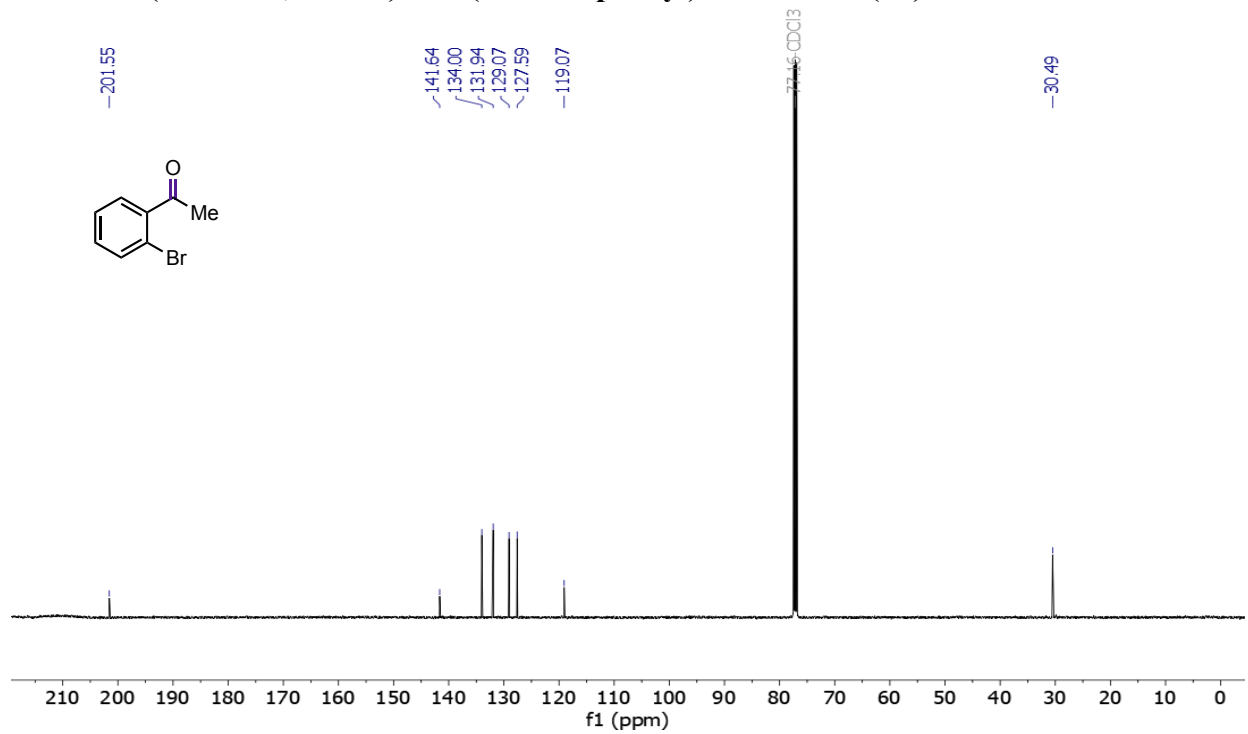
¹³C NMR (126 MHz, CDCl₃) of 1-(3-bromophenyl)ethan-1-one (3i)



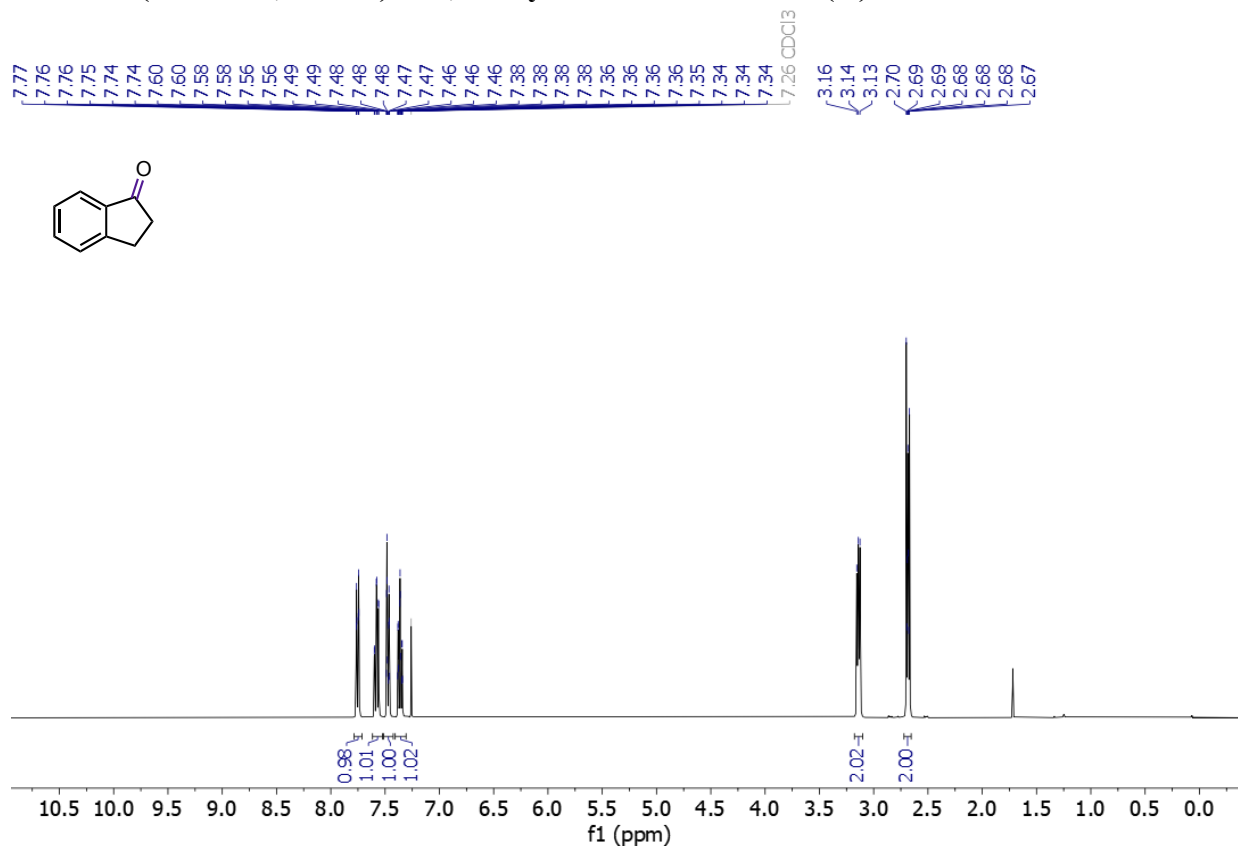
¹H NMR (500 MHz, CDCl₃) of 1-(2-bromophenyl)ethan-1-one (3k)



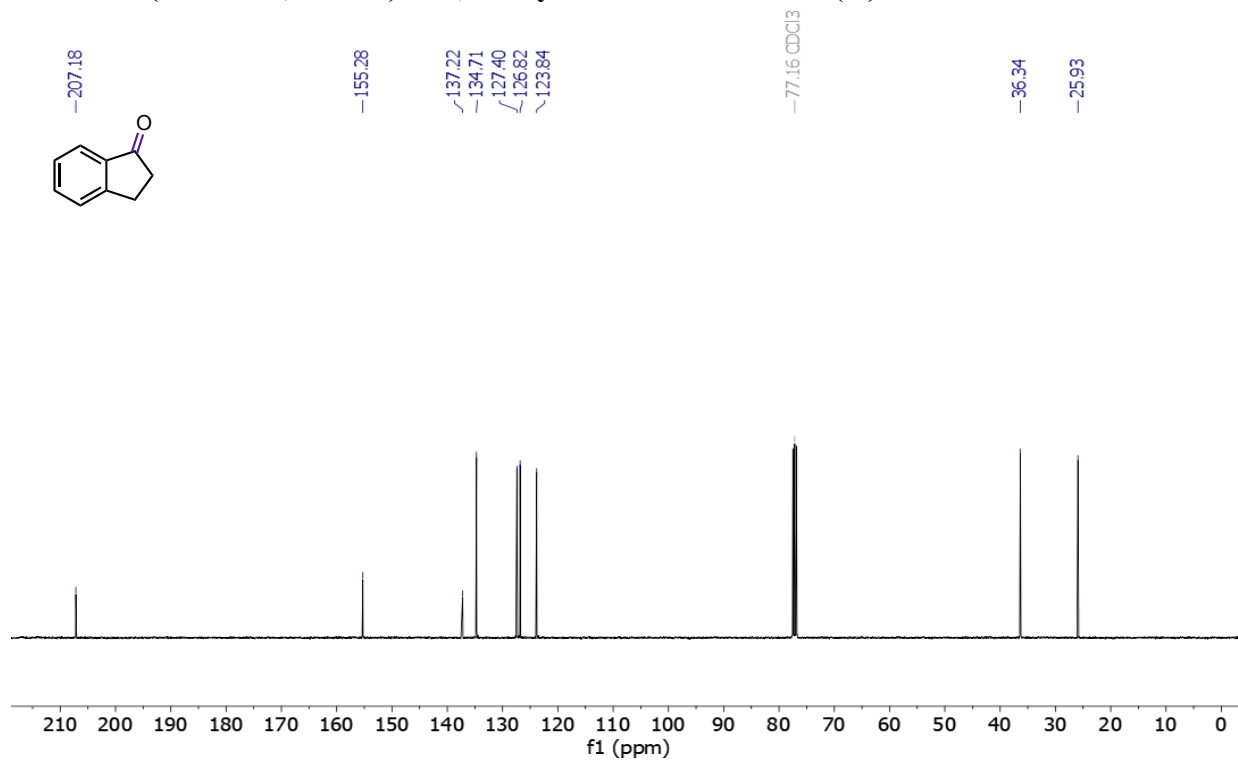
¹³C NMR (126 MHz, CDCl₃) of 1-(2-bromophenyl)ethan-1-one (3k)



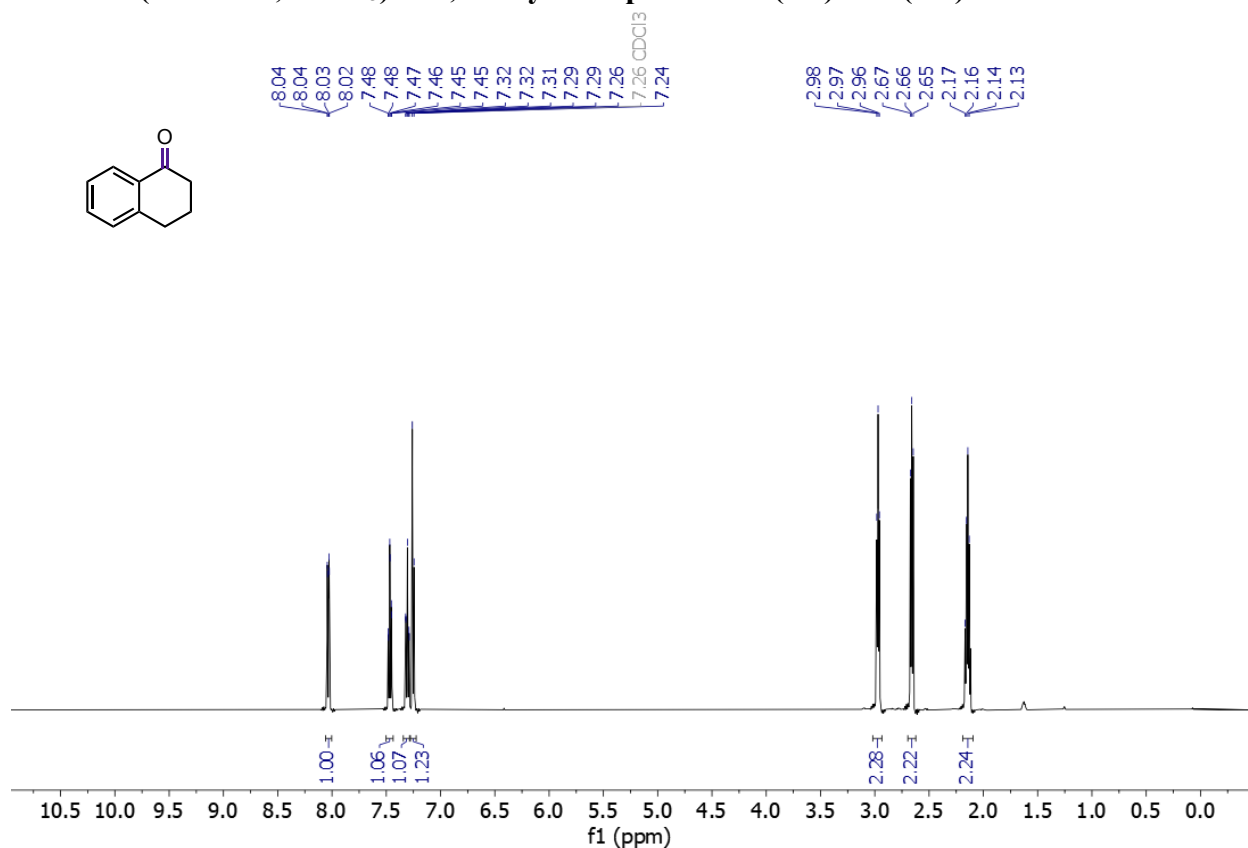
¹H NMR (400 MHz, CDCl₃) of 2,3-dihydro-1*H*-inden-1-one (3l)



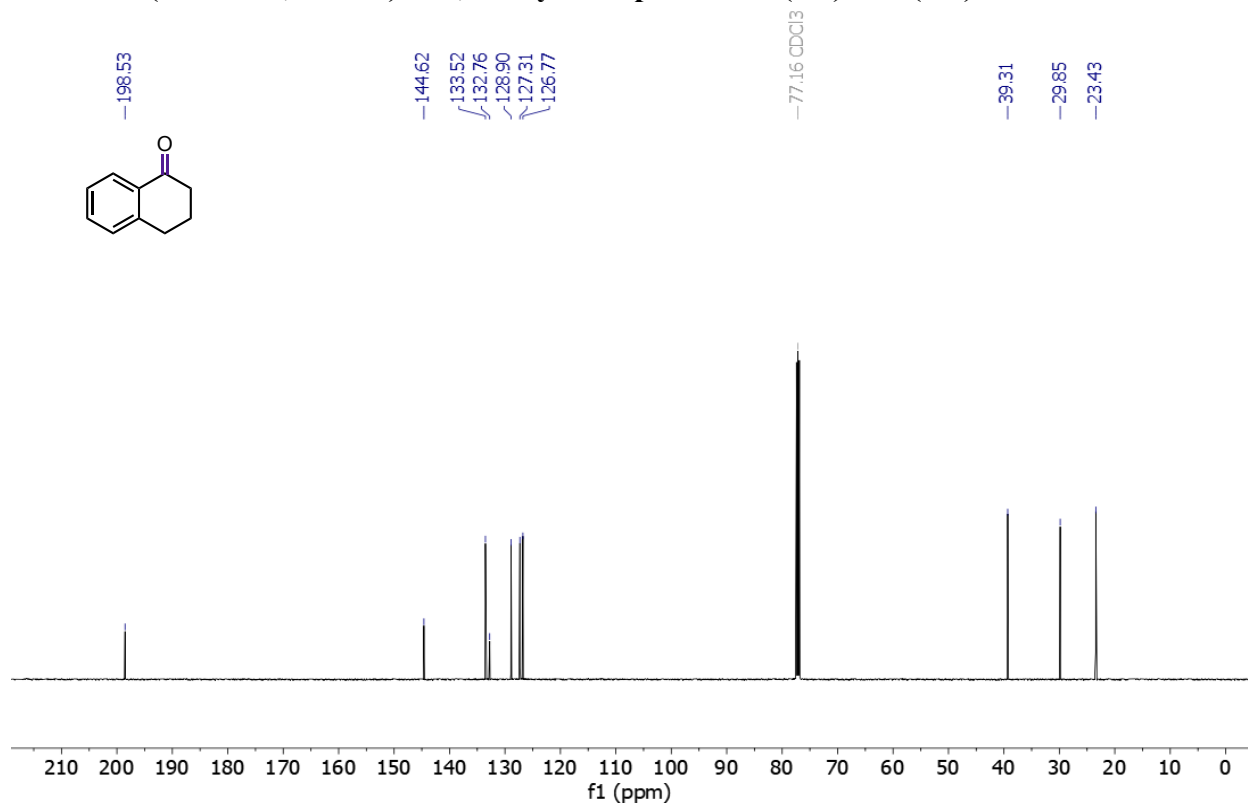
¹³C NMR (101 MHz, CDCl₃) of 2,3-dihydro-1*H*-inden-1-one (3l)



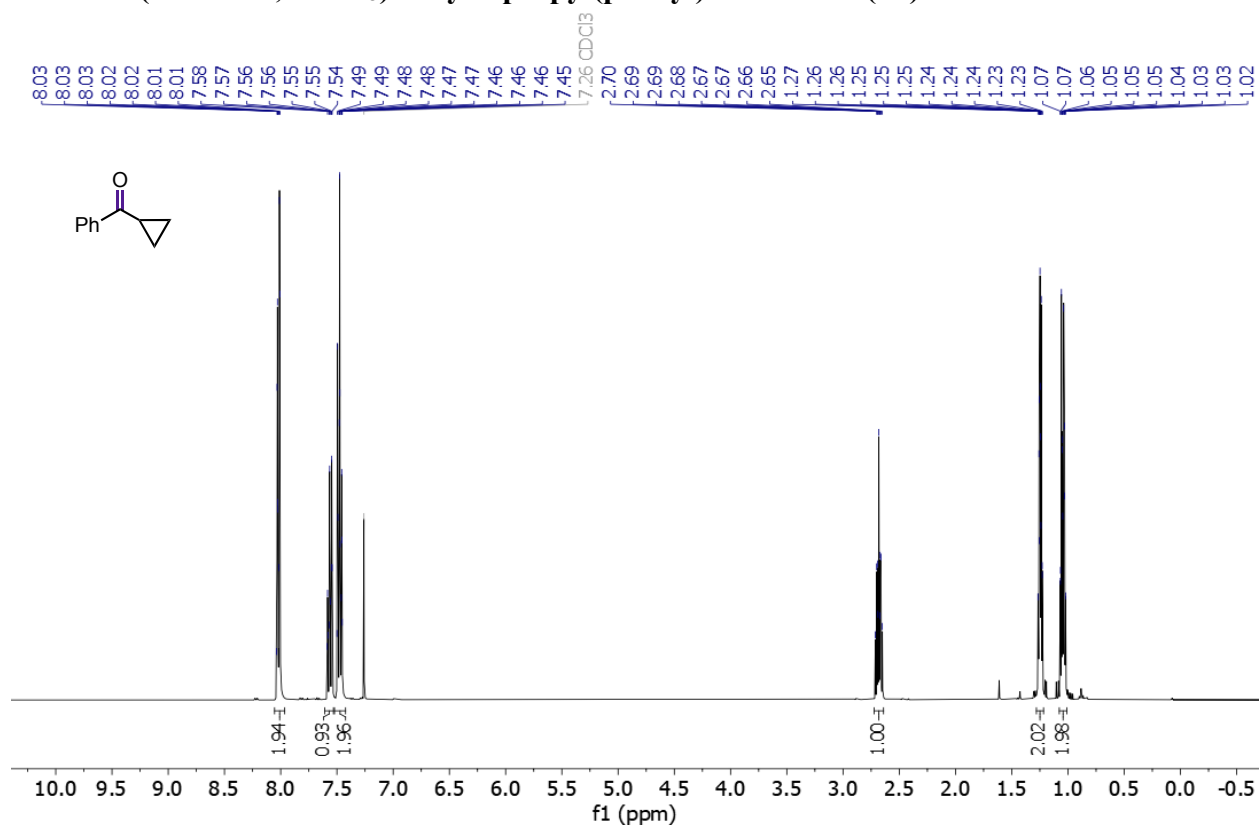
¹H NMR (500 MHz, CDCl₃) of 3,4-dihydronaphthalen-1(2H)-one (3m)



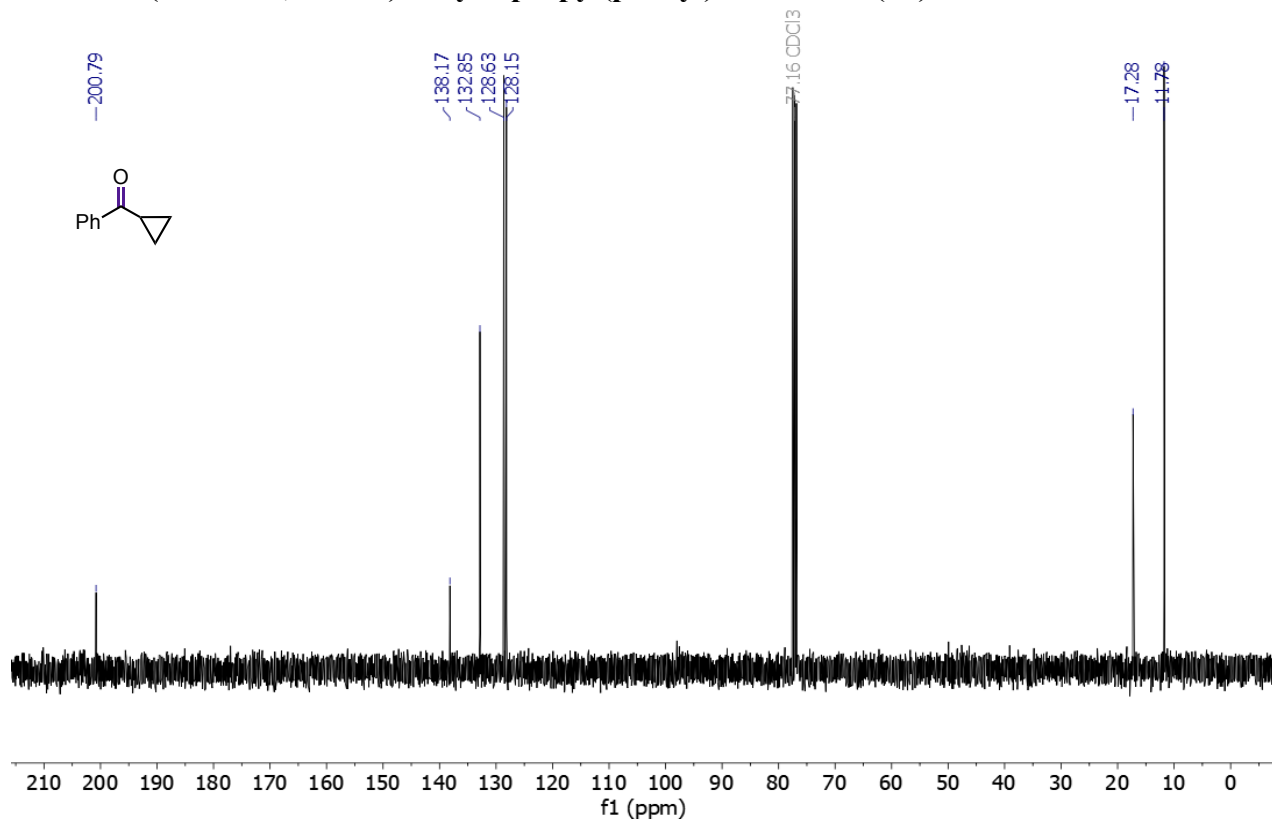
¹³C NMR (126 MHz, CDCl₃) of 3,4-dihydronaphthalen-1(2H)-one (3m)



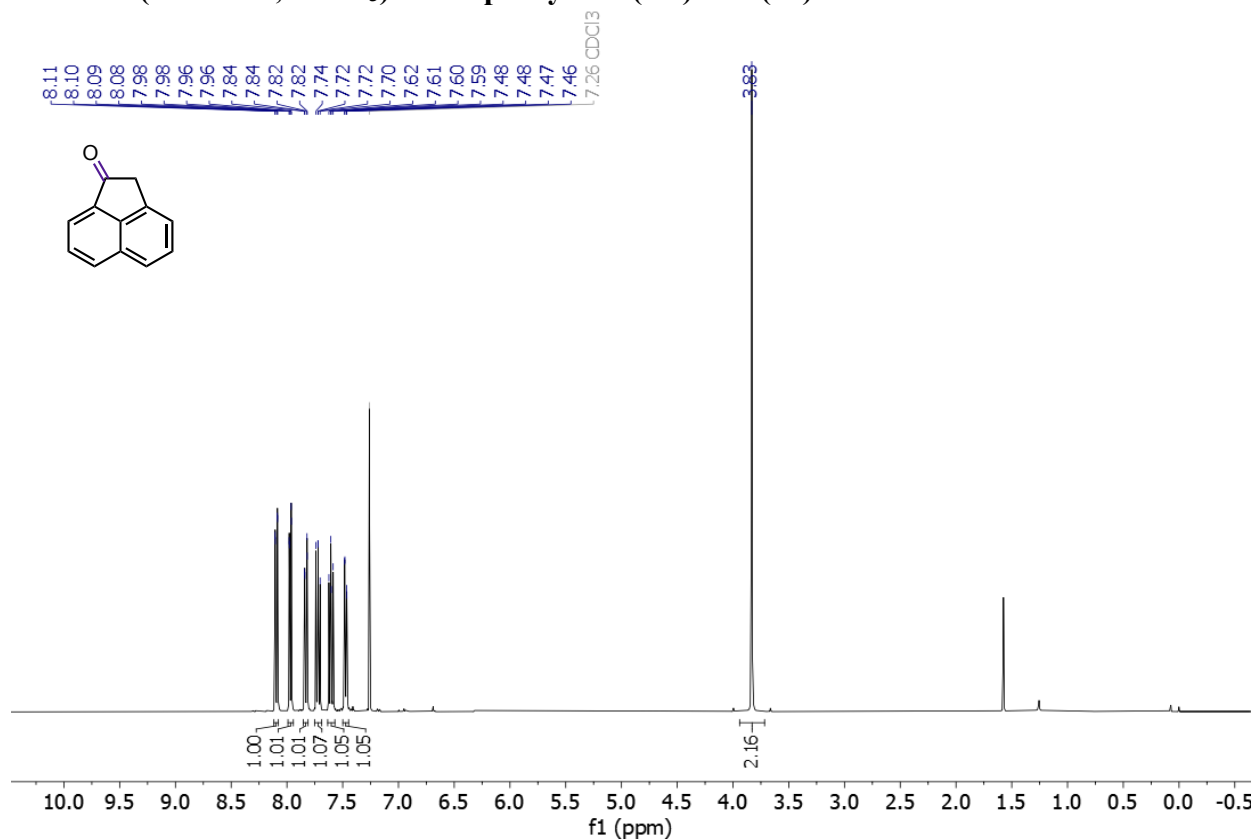
¹H NMR (400 MHz, CDCl₃) of cyclopropyl(phenyl)methanone (3n)



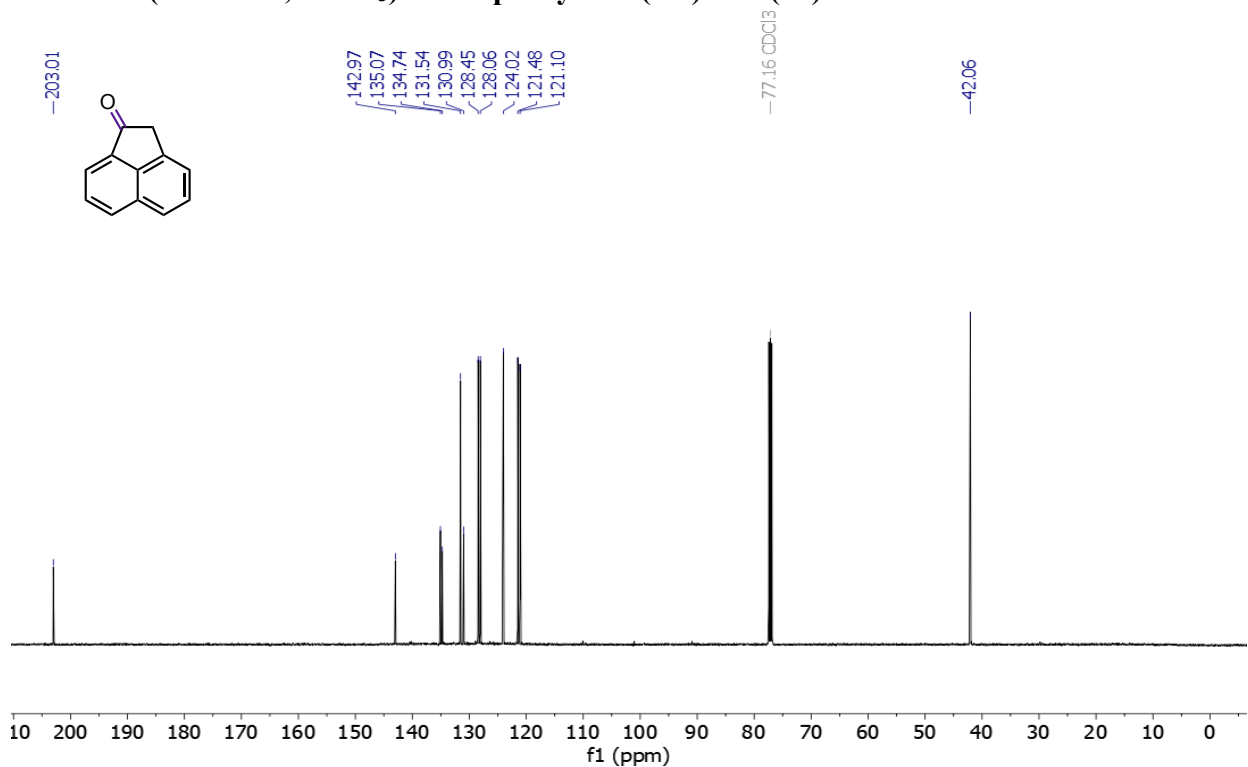
¹³C NMR (101 MHz, CDCl₃) of cyclopropyl(phenyl)methanone (3n)



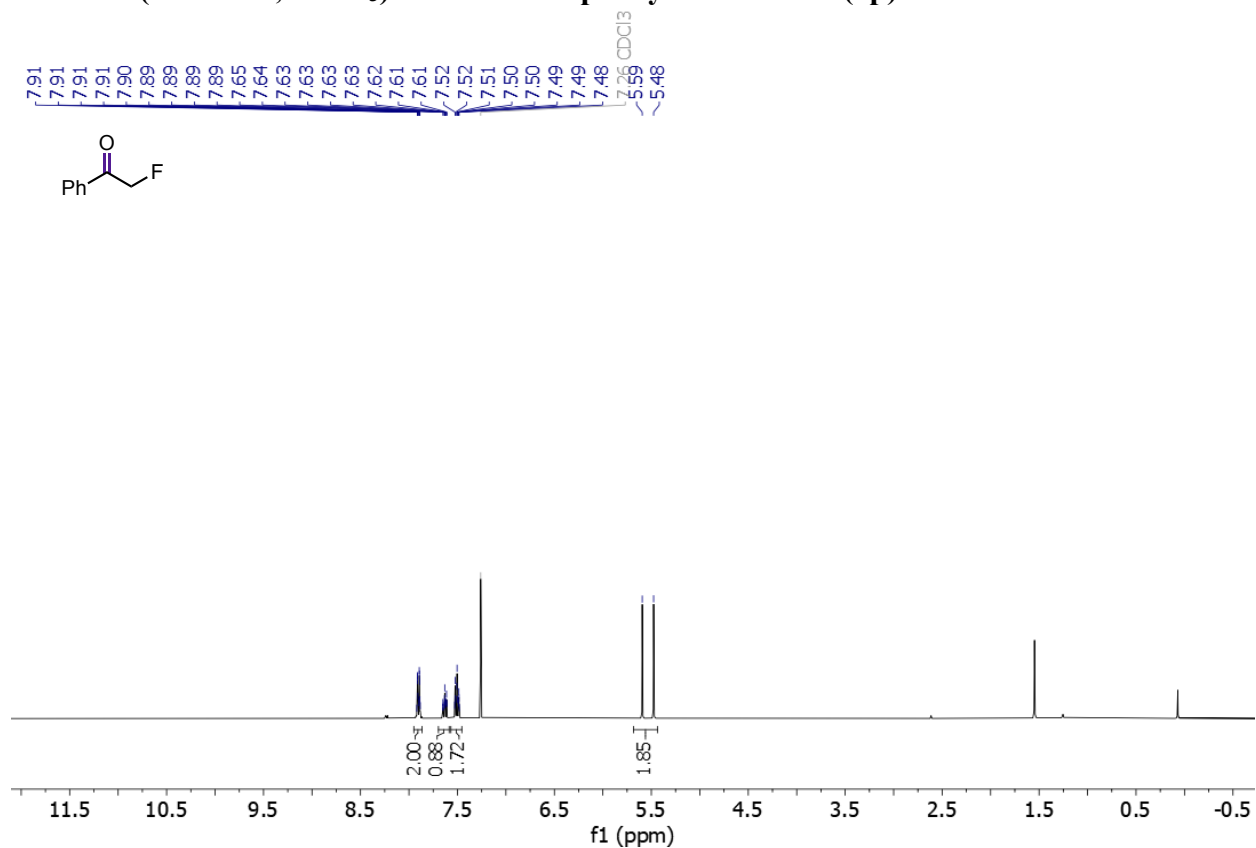
¹H NMR (400 MHz, CDCl₃) Acenaphthylen-1(2H)-one (3o)



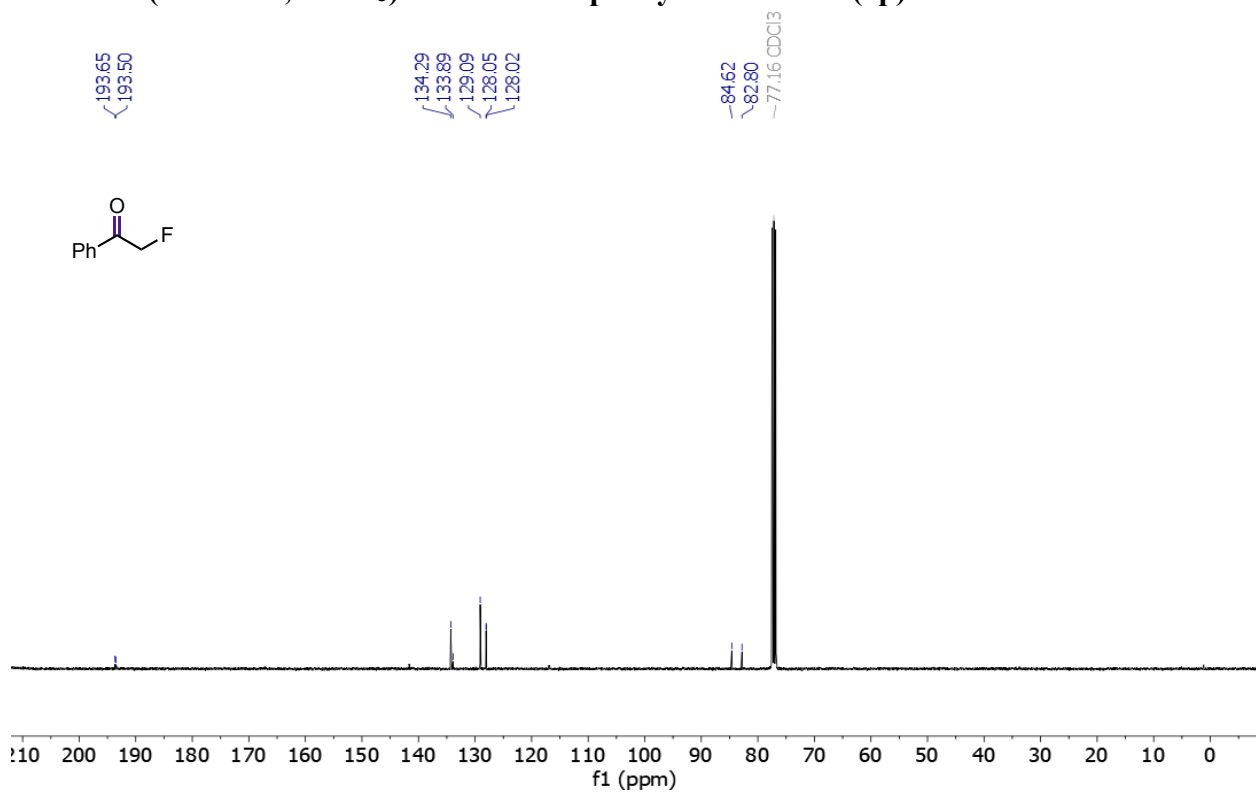
¹³C NMR (126 MHz, CDCl₃) Acenaphthylen-1(2H)-one (3o)



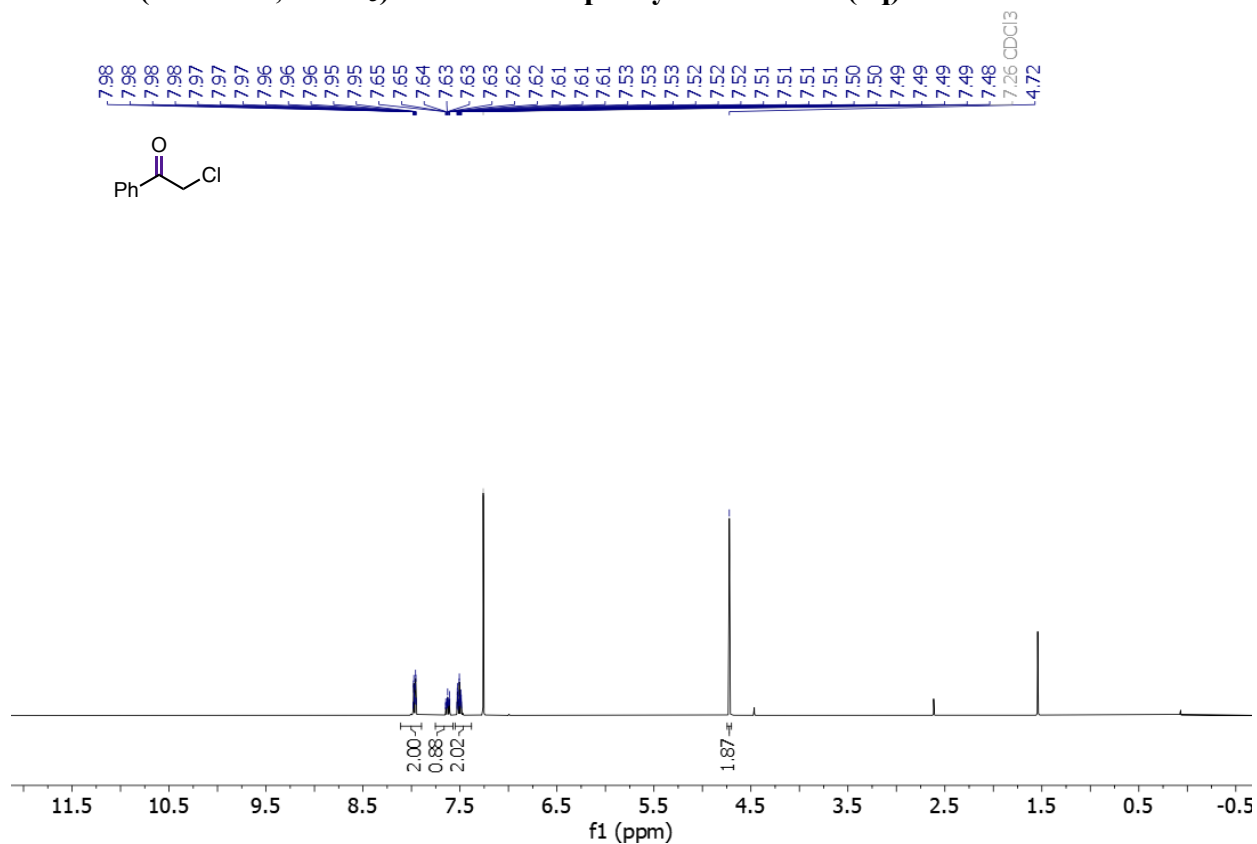
¹H NMR (400 MHz, CDCl₃) of 2-fluoro-1-phenylethan-1-one (3p)



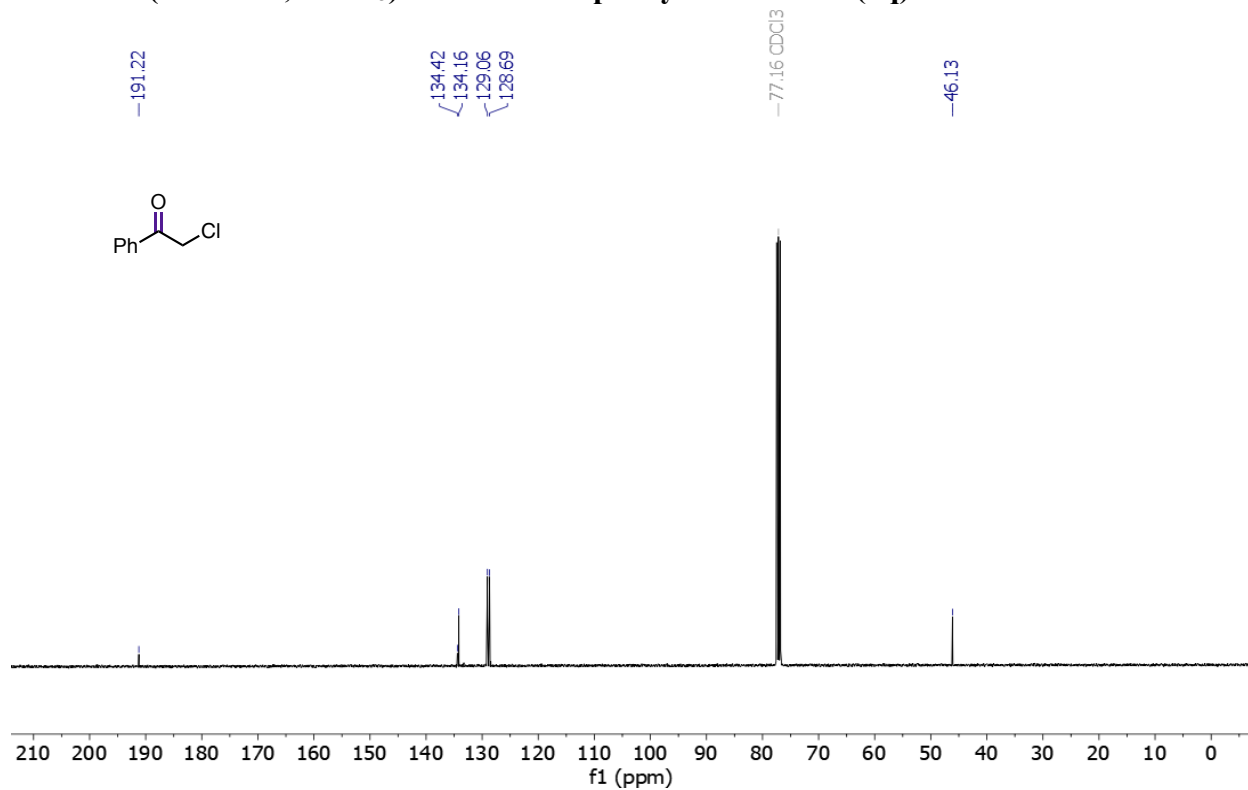
¹³C NMR (101 MHz, CDCl₃) of 2-fluoro-1-phenylethan-1-one (3p)



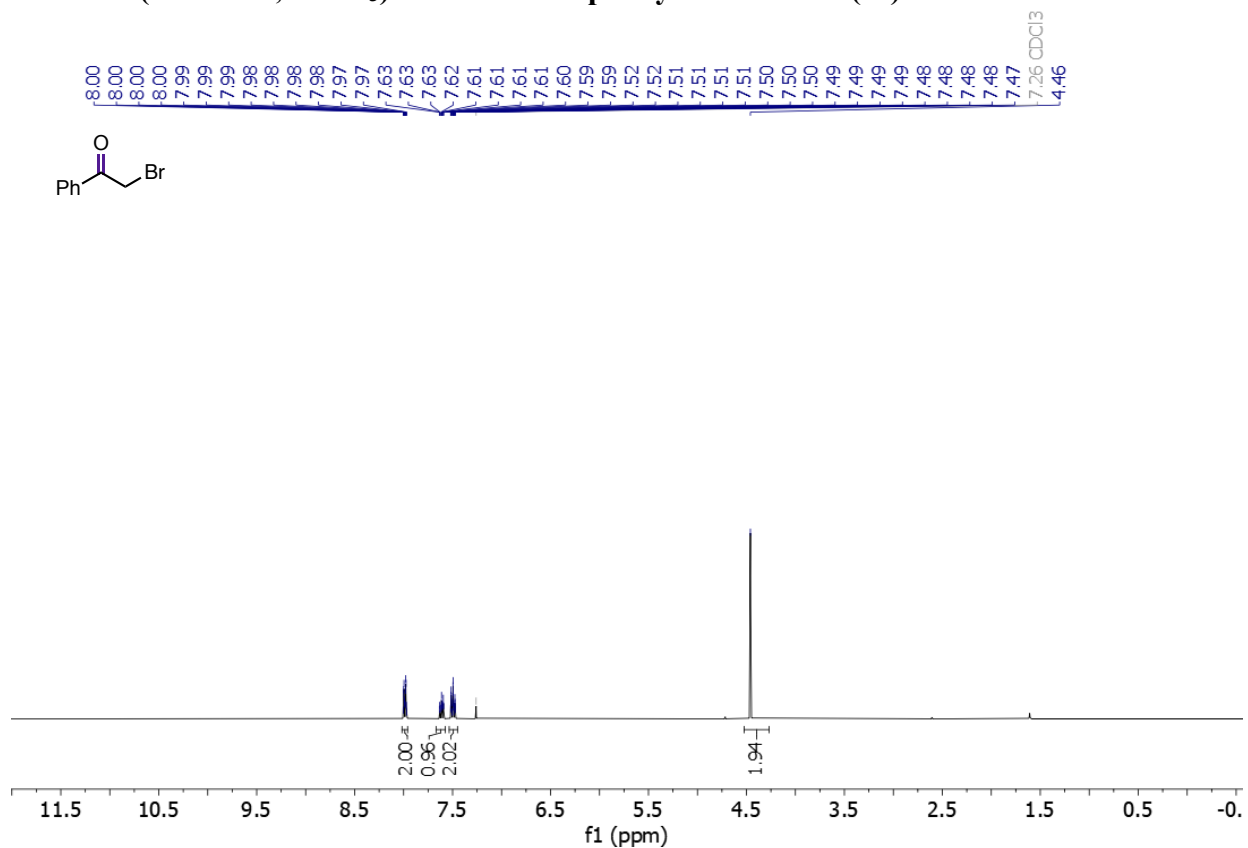
¹H NMR (400 MHz, CDCl₃) of 2-chloro-1-phenylethan-1-one (3q)



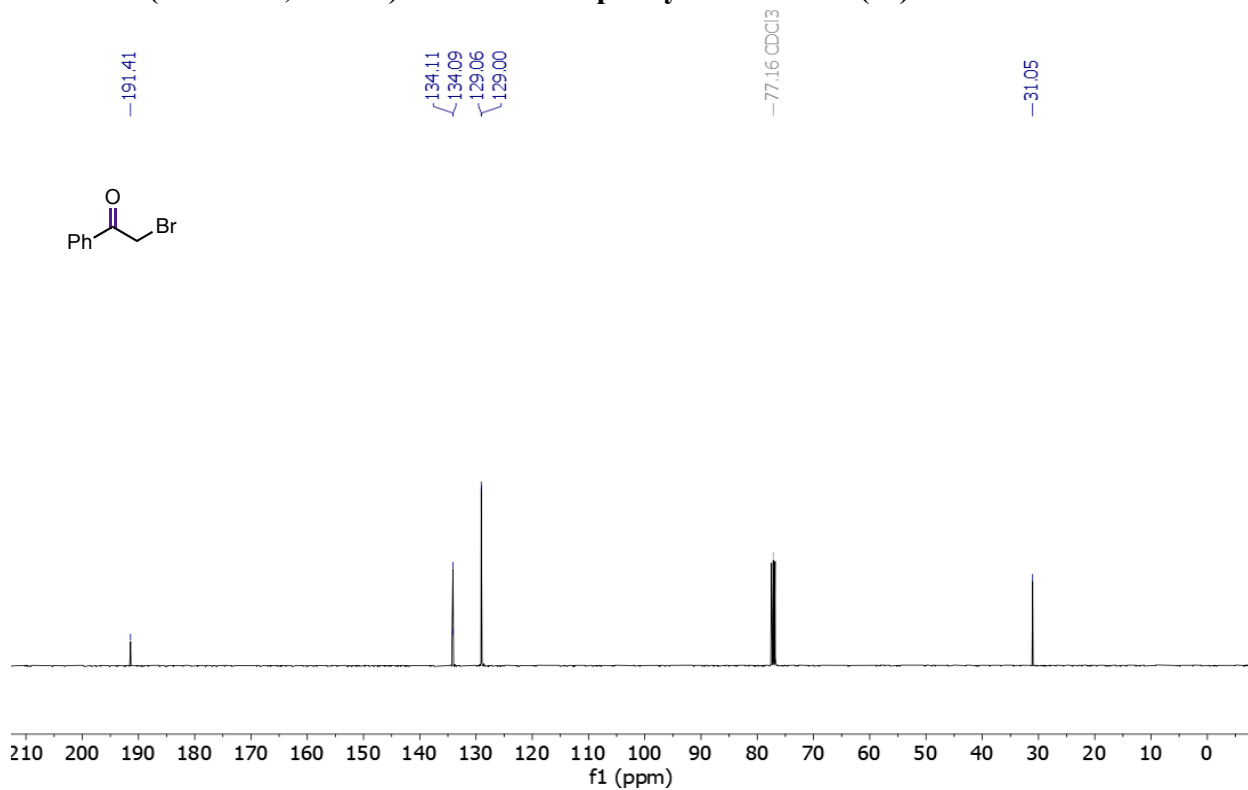
¹³C NMR (101 MHz, CDCl₃) of 2-chloro-1-phenylethan-1-one (3q)



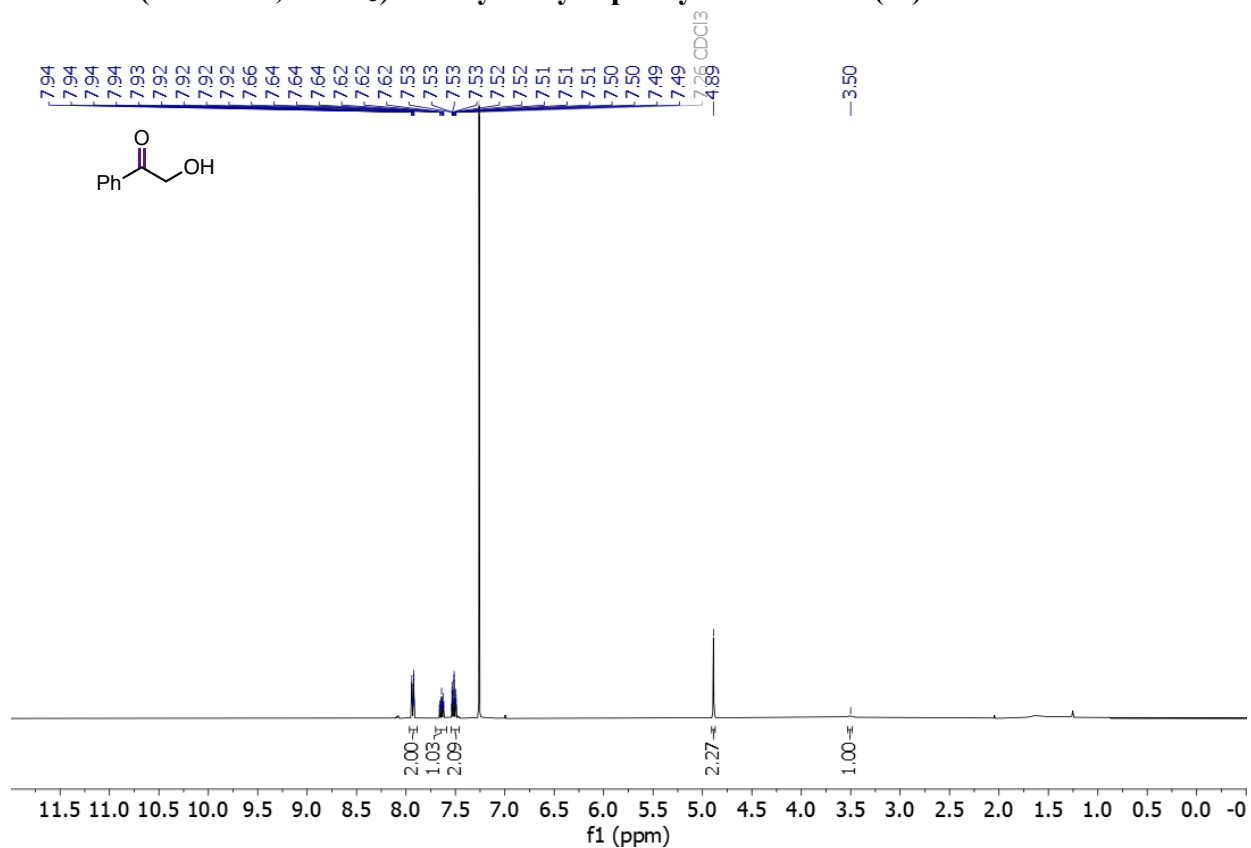
¹H NMR (400 MHz, CDCl₃) of 2-bromo-1-phenylethan-1-one (3r)



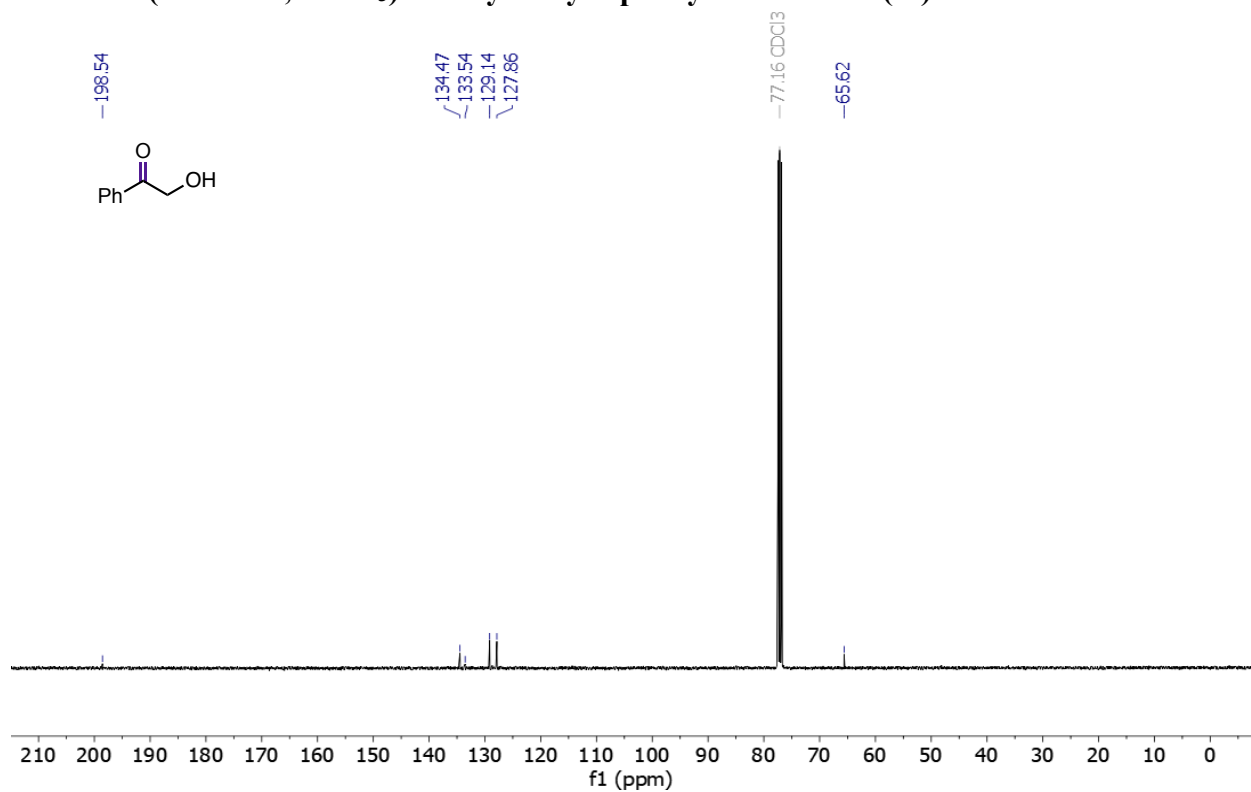
¹³C NMR (101 MHz, CDCl₃) of 2-bromo-1-phenylethan-1-one (3r)



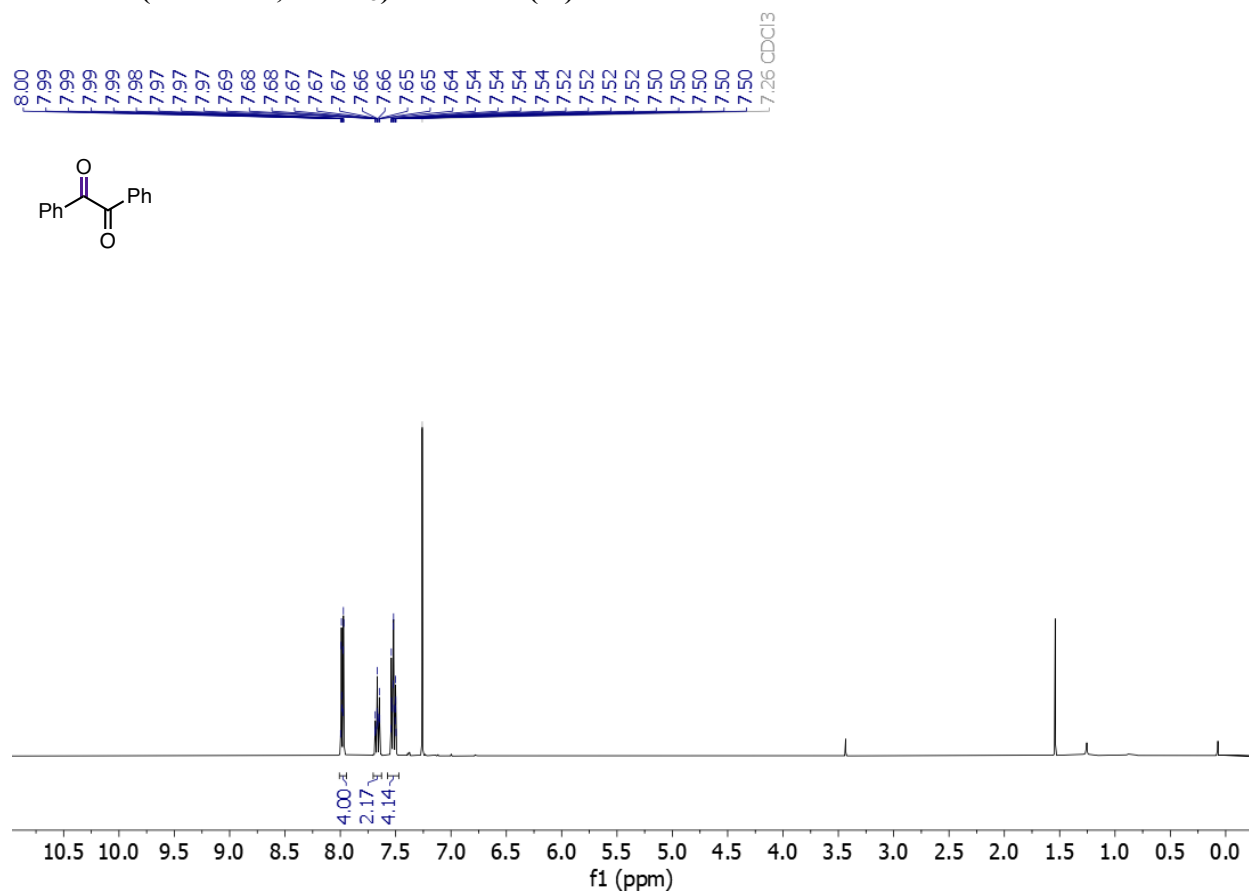
¹H NMR (400 MHz, CDCl₃) of 2-hydroxy-1-phenylethan-1-one (3s)



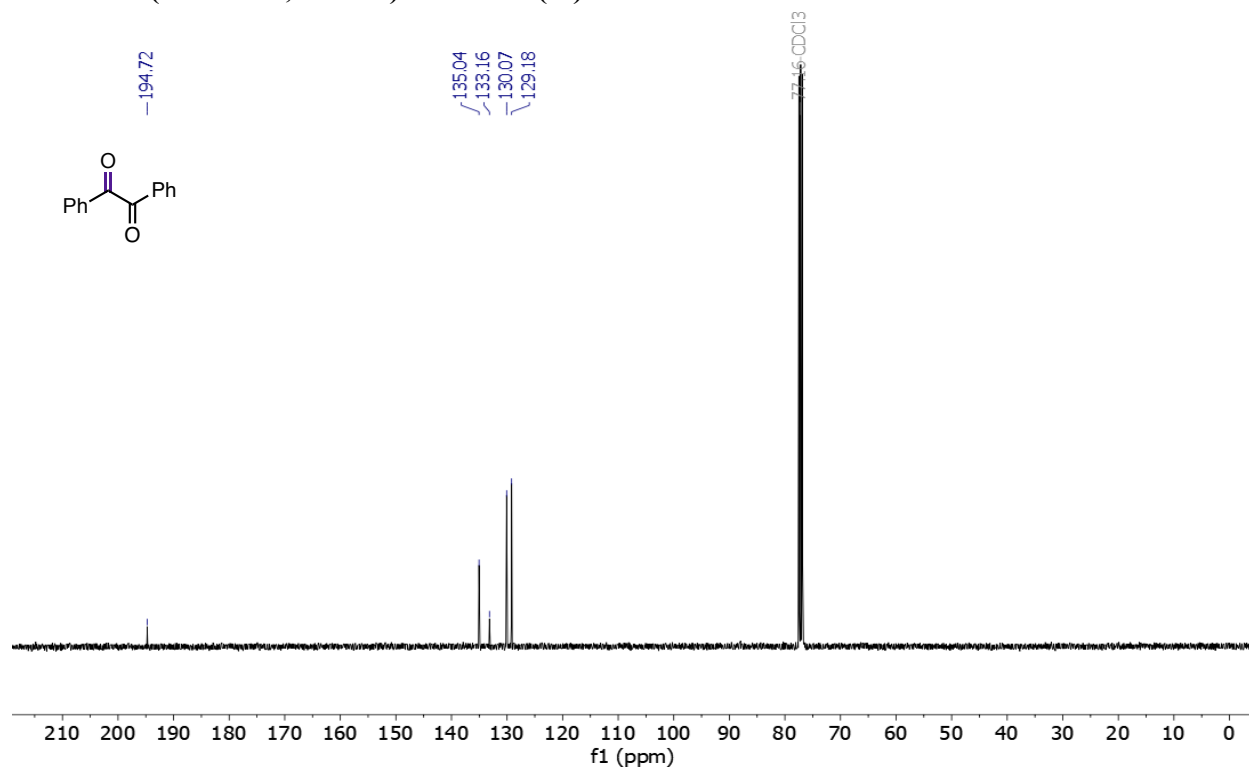
¹³C NMR (101 MHz, CDCl₃) of 2-hydroxy-1-phenylethan-1-one (3s)



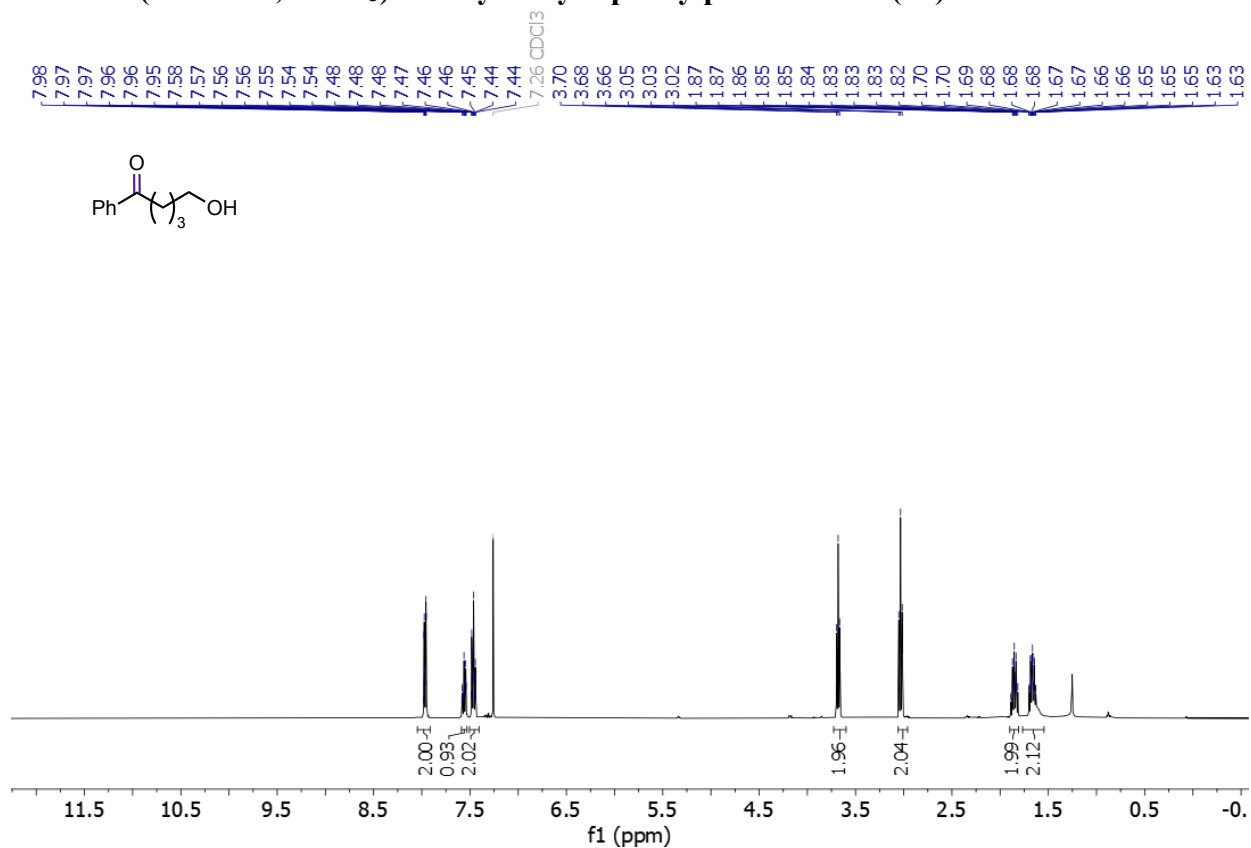
¹H NMR (400 MHz, CDCl₃) of Benzil (3t)



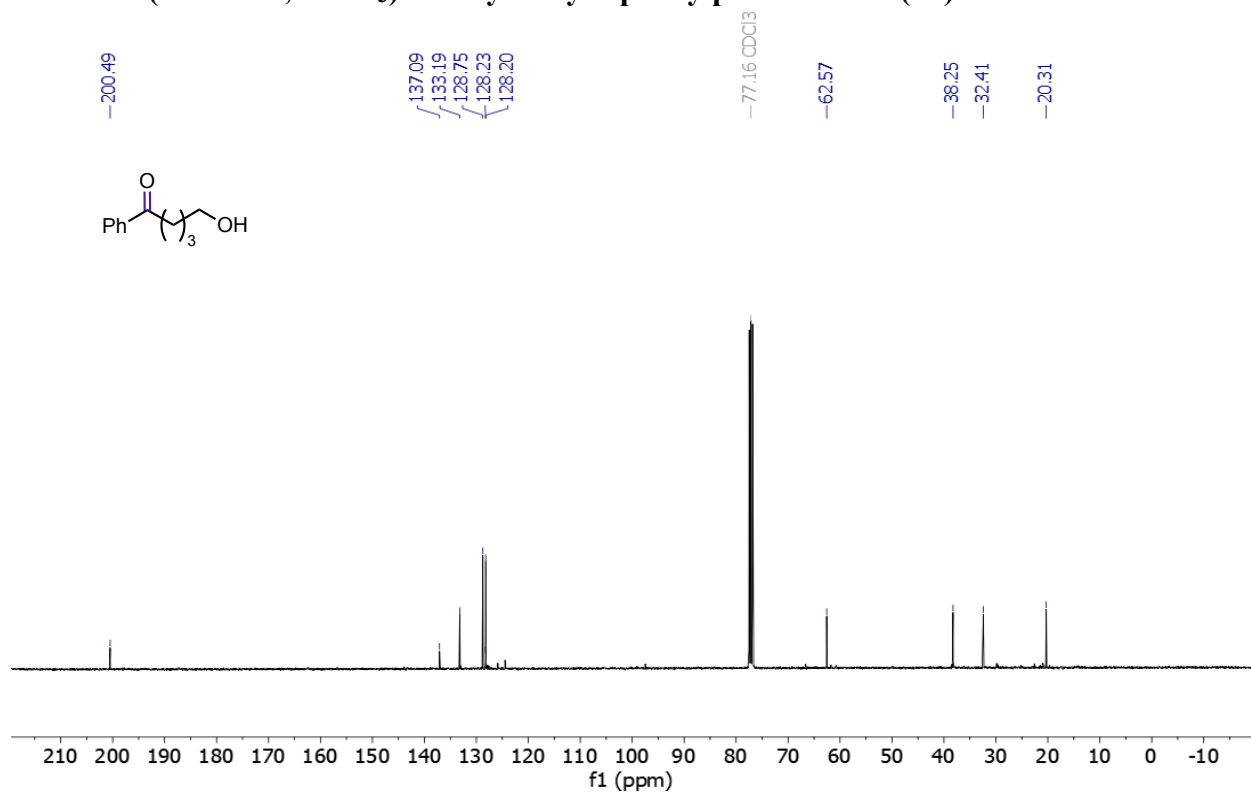
¹³C NMR (101 MHz, CDCl₃) of Benzil (3t)



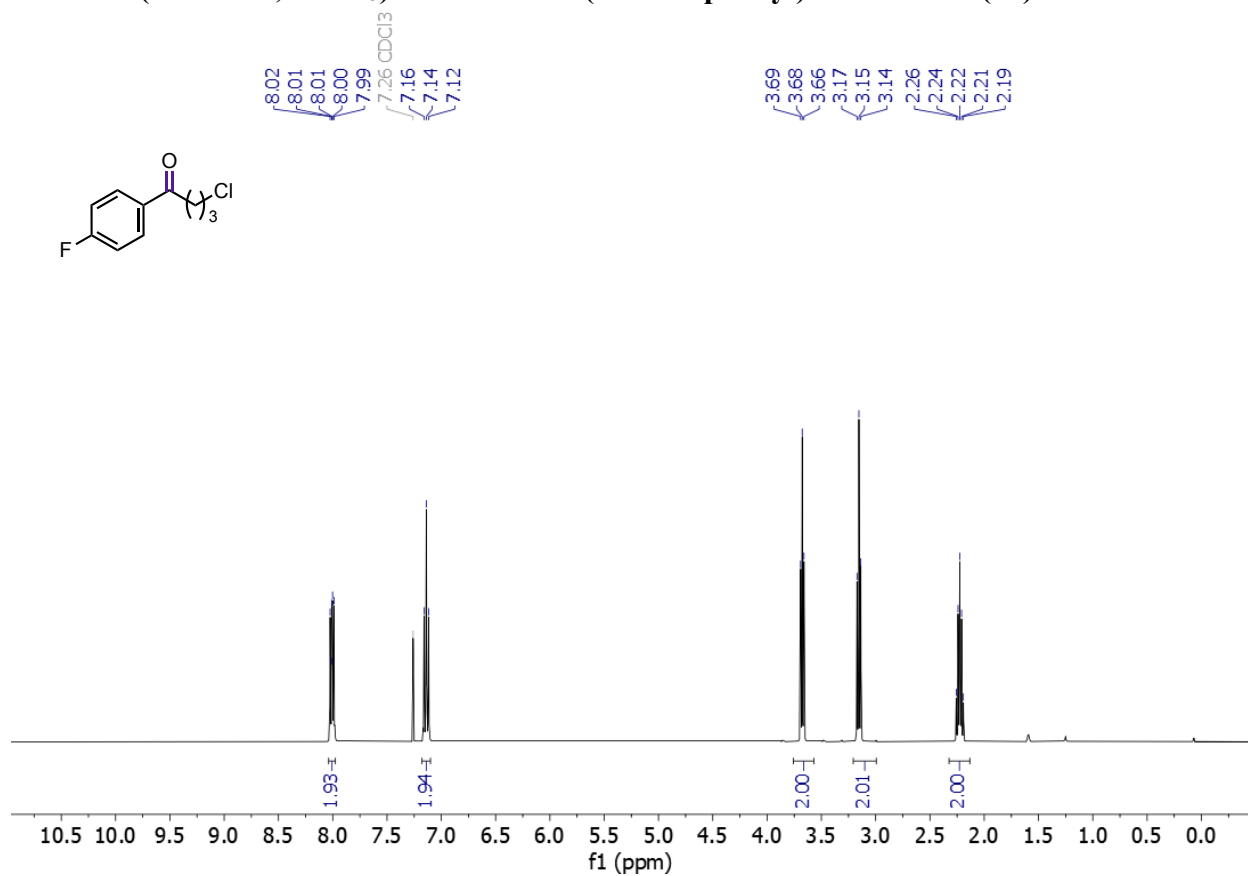
¹H NMR (400 MHz, CDCl₃) of 5-hydroxy-1-phenylpentan-1-one (3u)



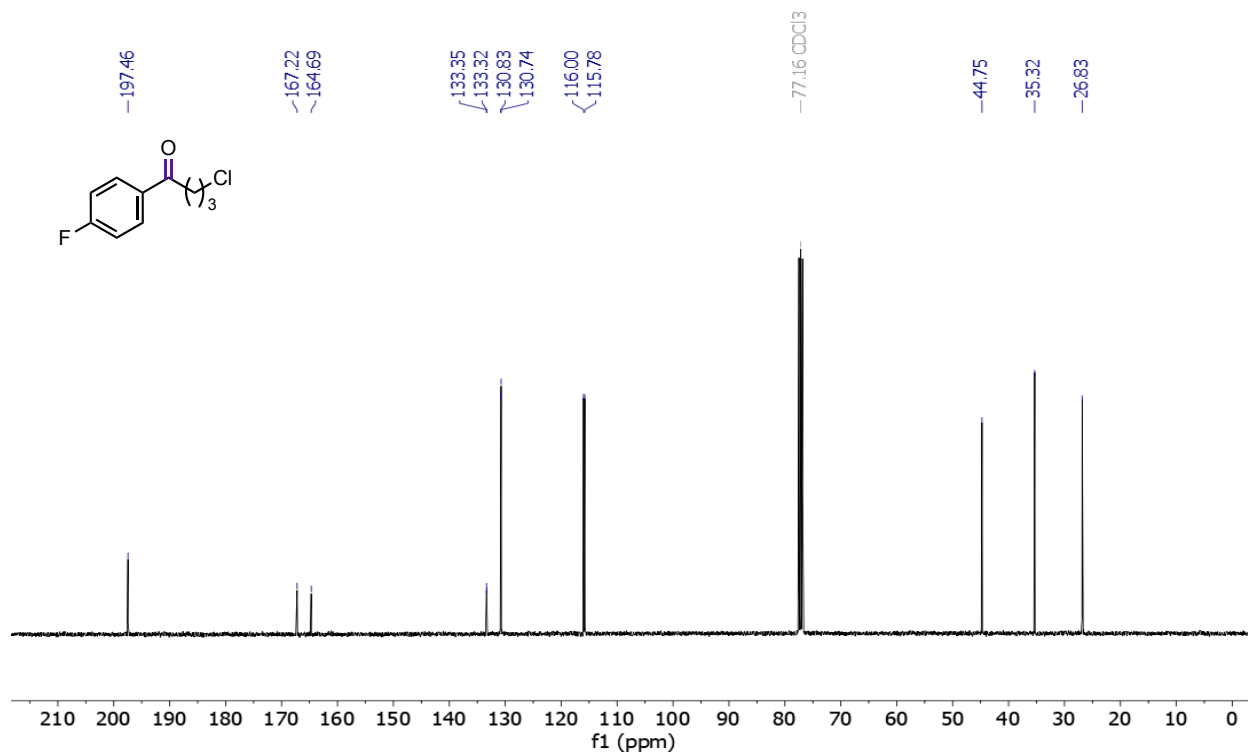
¹³C NMR (101 MHz, CDCl₃) of 5-hydroxy-1-phenylpentan-1-one (3u)



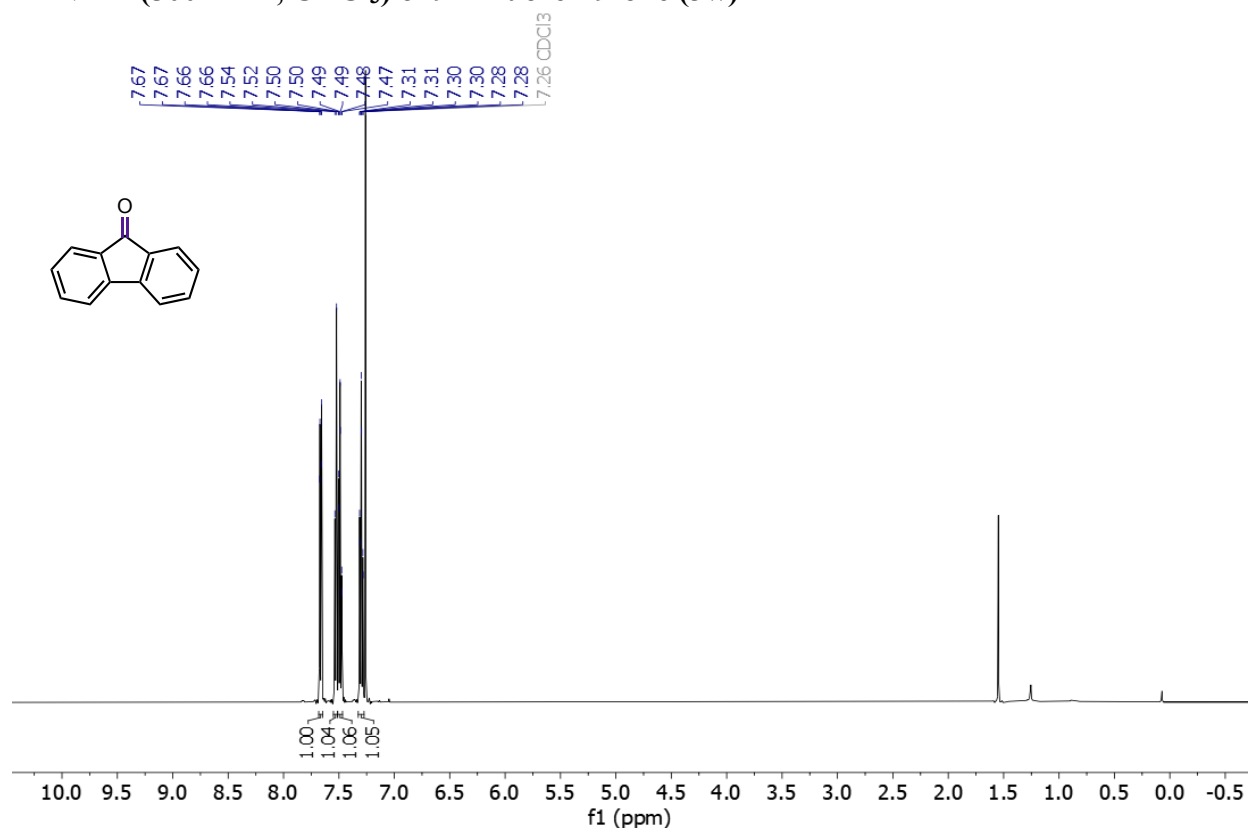
¹H NMR (400 MHz, CDCl₃) of 4-chloro-1-(4-fluorophenyl)butan-1-one (3v)



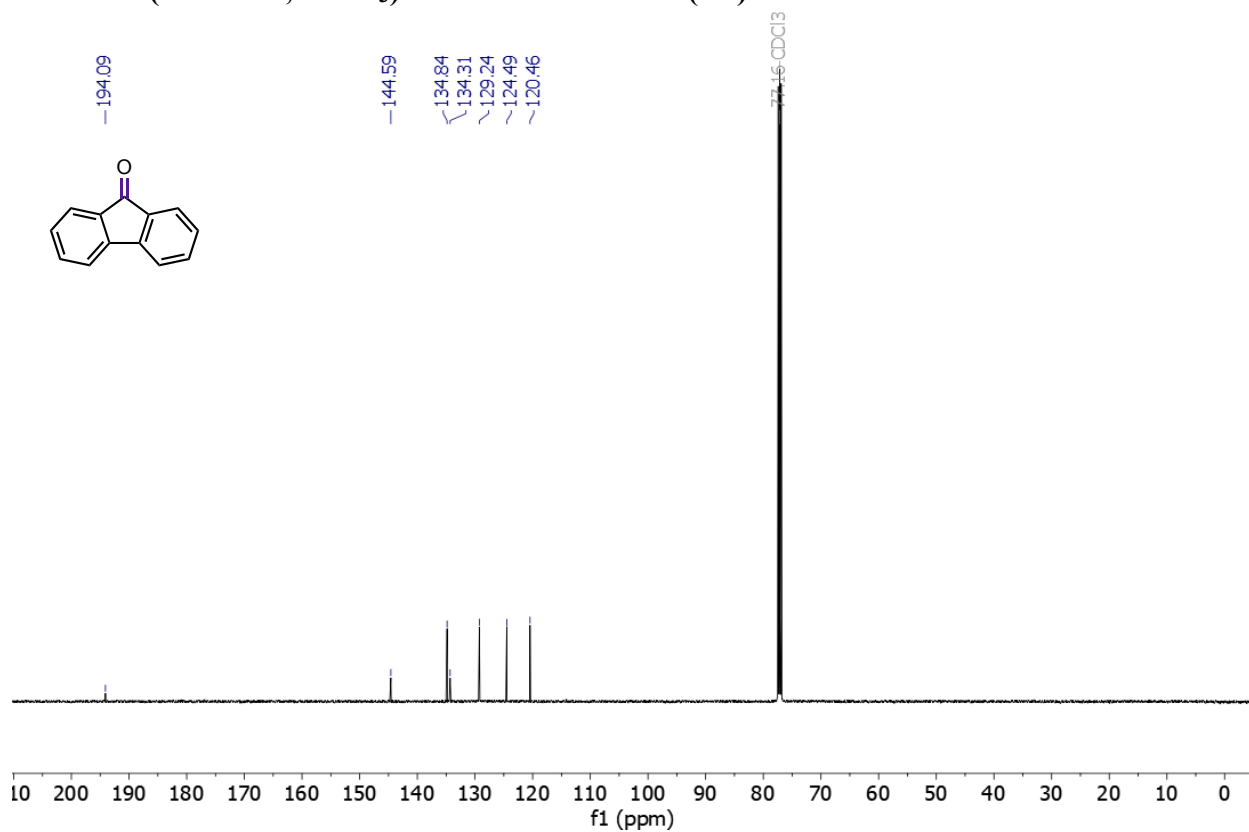
¹³C NMR (101 MHz, CDCl₃) of 4-Chloro-1-(4-fluorophenyl)butan-1-one (3v)



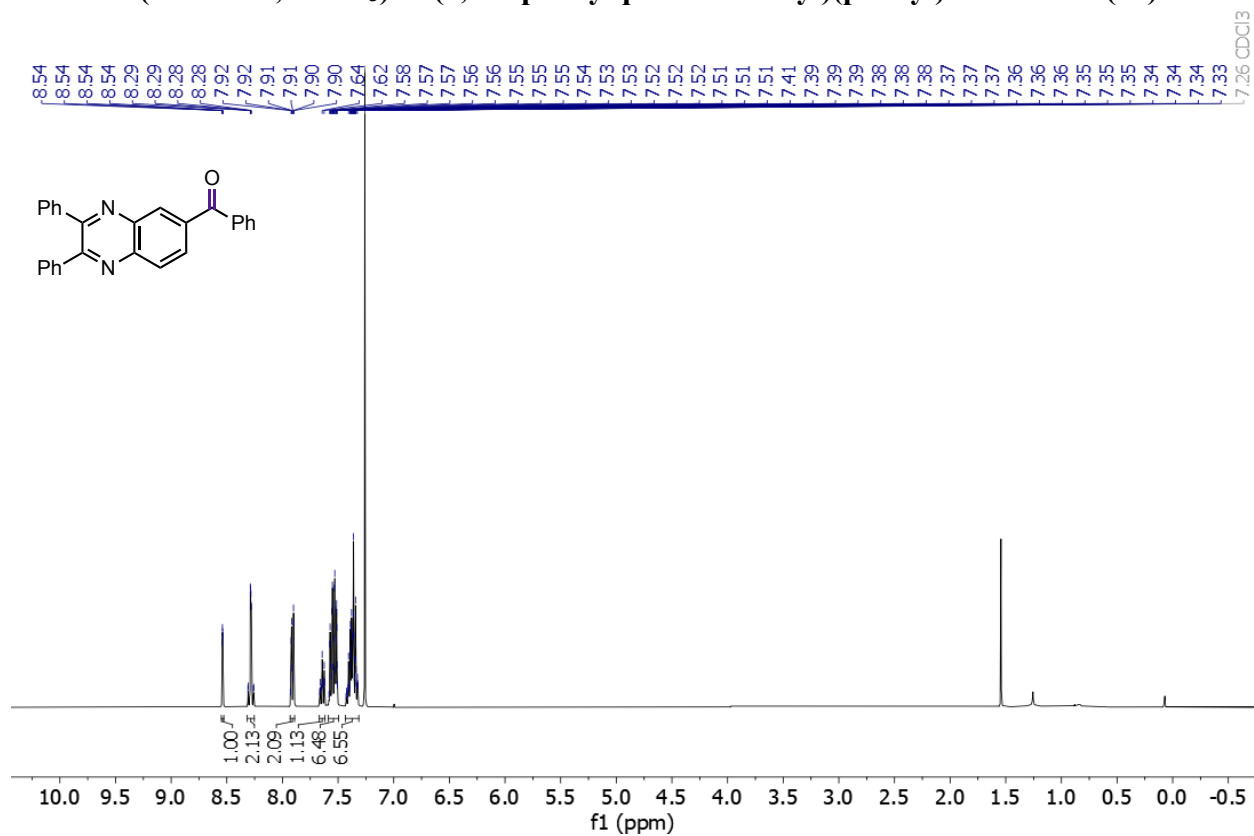
¹H NMR (500 MHz, CDCl₃) of 9H-fluoren-9-one (3w)



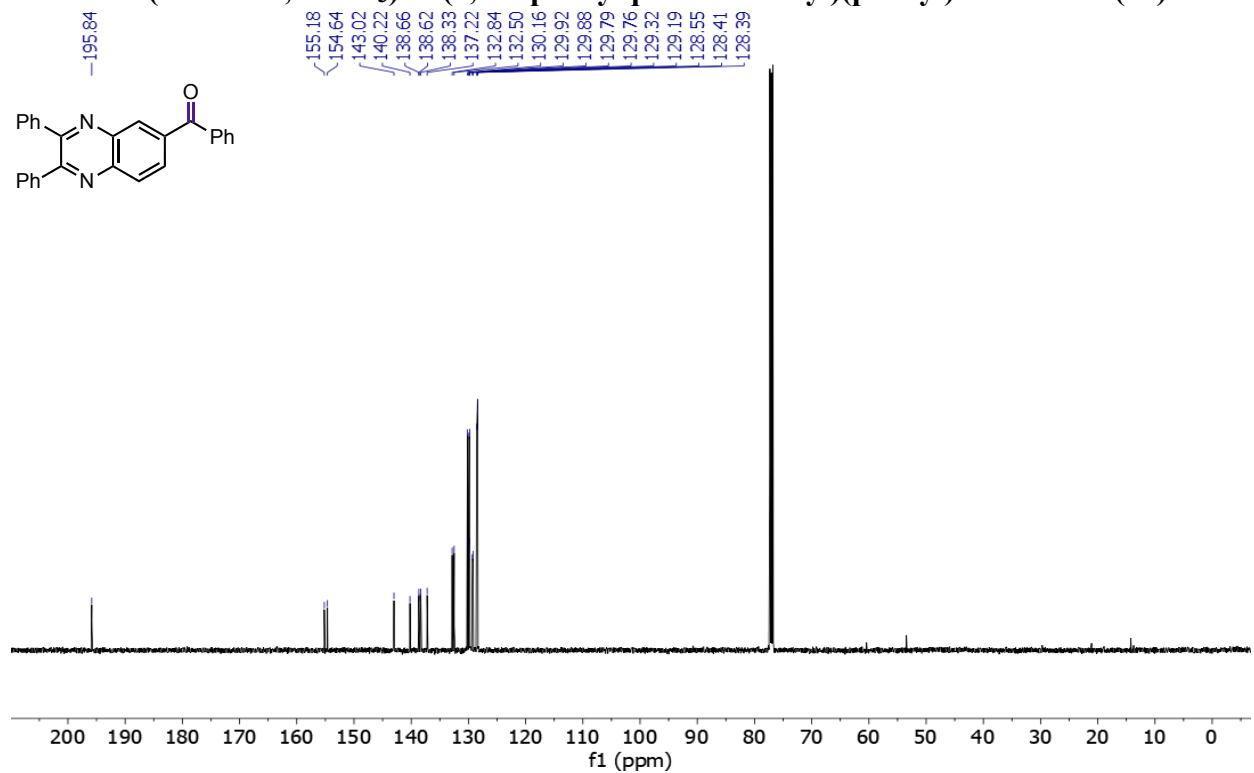
¹³C NMR (126 MHz, CDCl₃) of 9H-fluoren-9-one (3w)



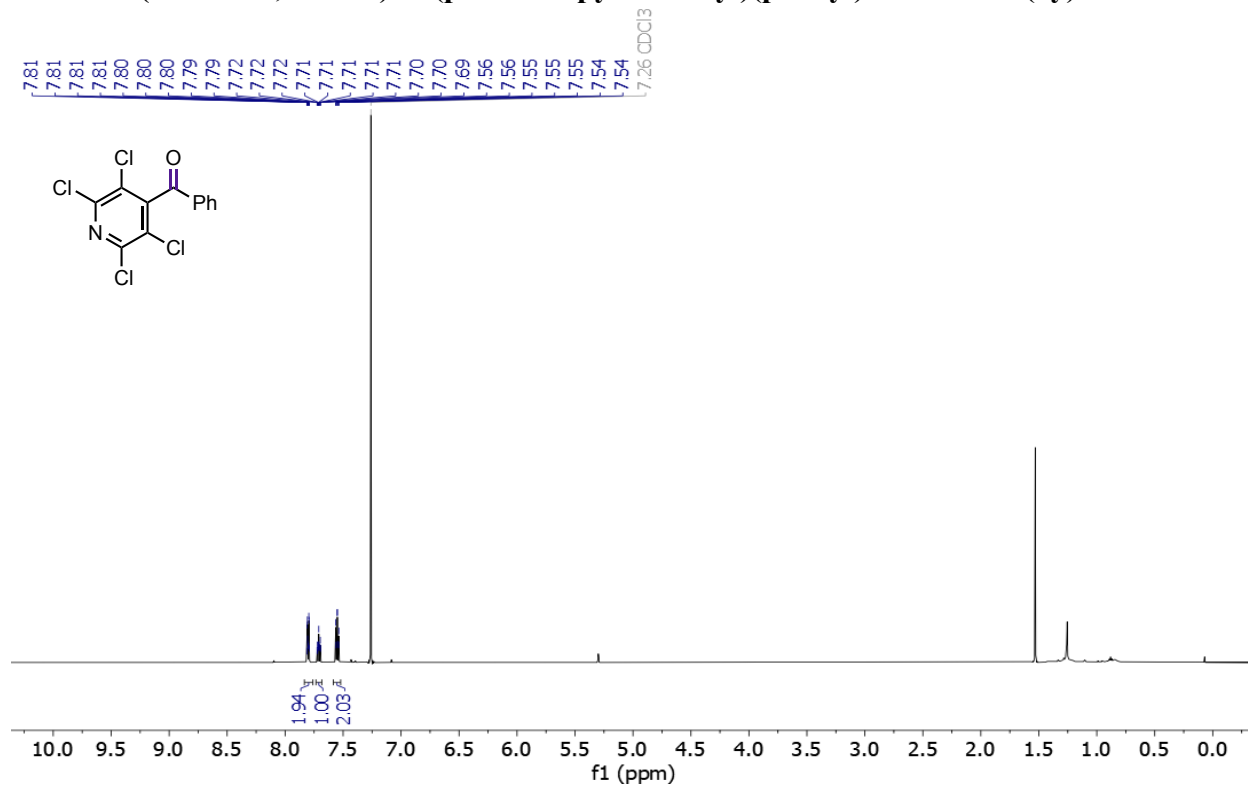
¹H NMR (400 MHz, CDCl₃) of (2,3-diphenylquinoxalin-6-yl)(phenyl)methanone (3x)



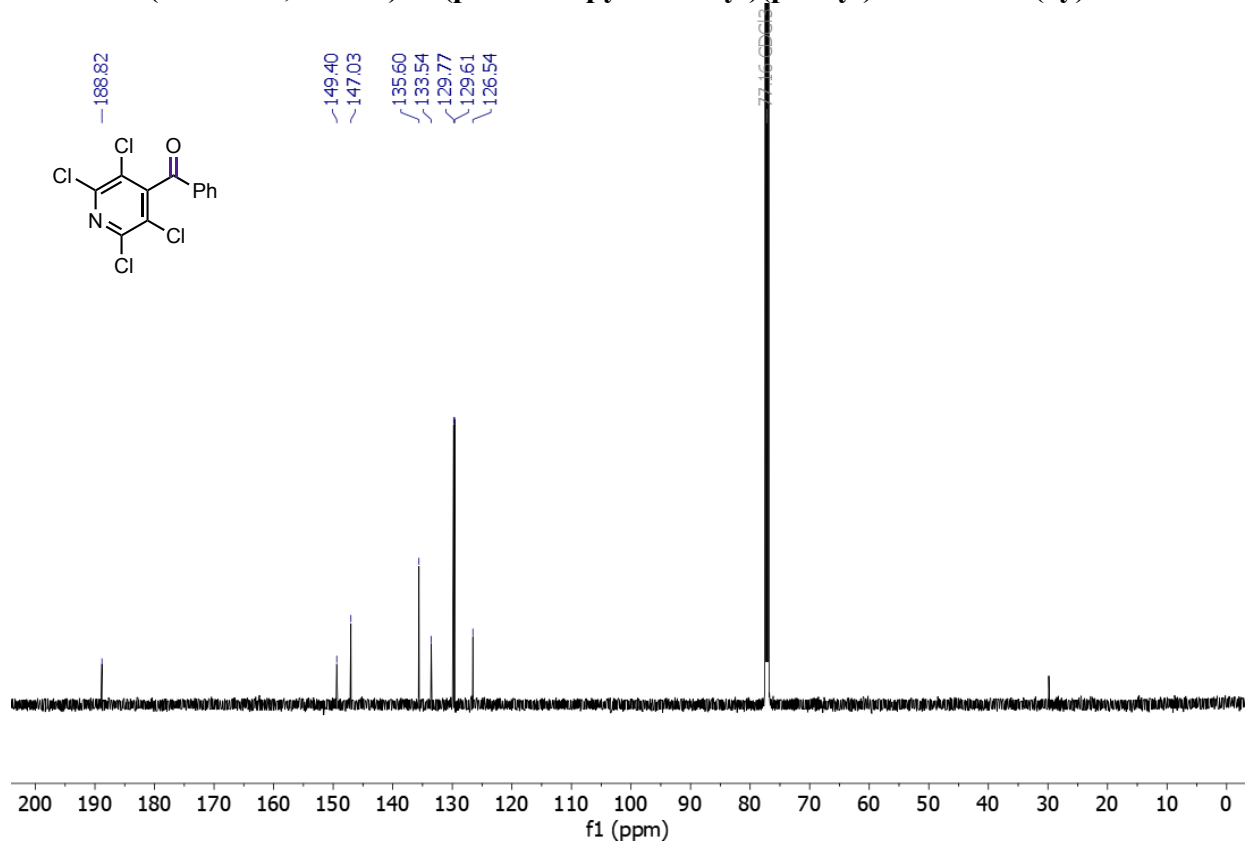
¹³C NMR (126 MHz, CDCl₃) of (2,3-diphenylquinoxalin-6-yl)(phenyl)methanone (3x)



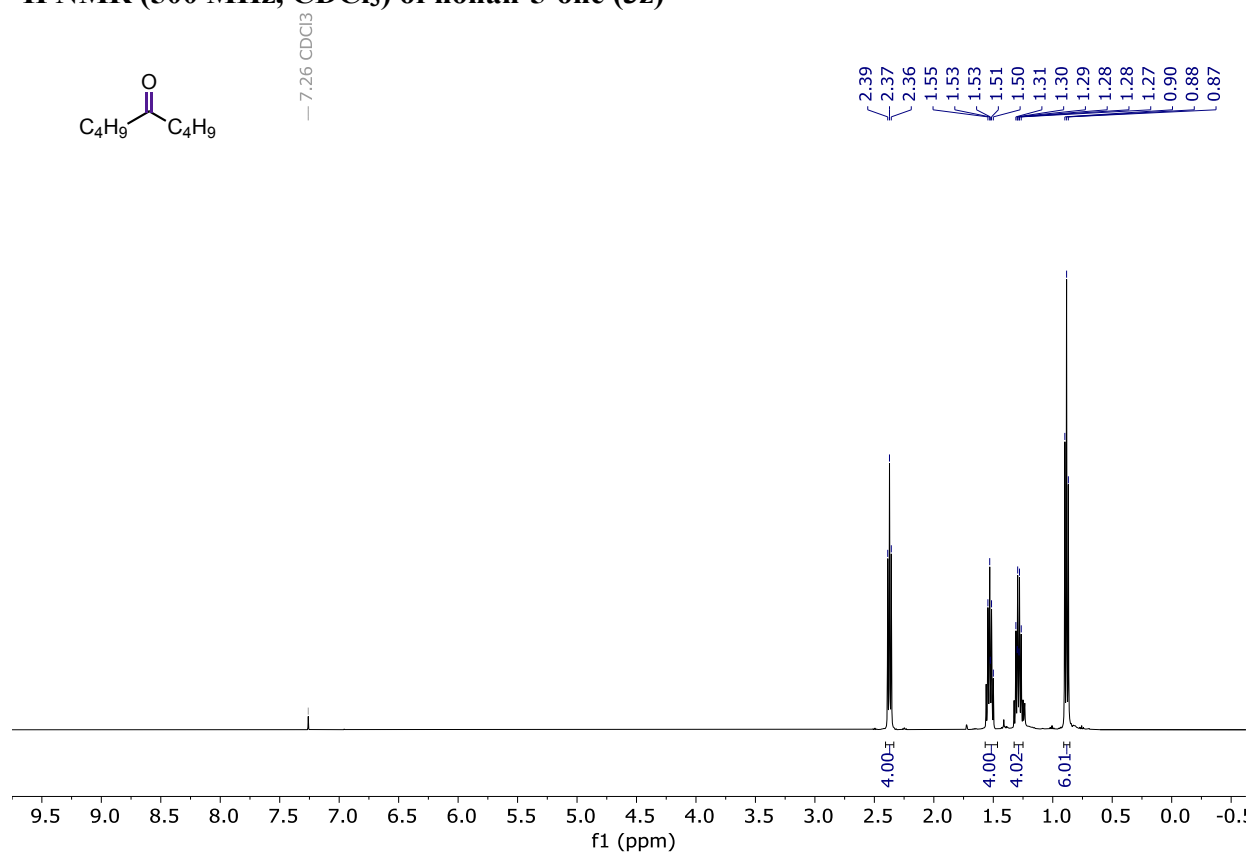
¹H NMR (600 MHz, CDCl₃) of (perchloropyridin-4-yl)(phenyl)methanone (3y)



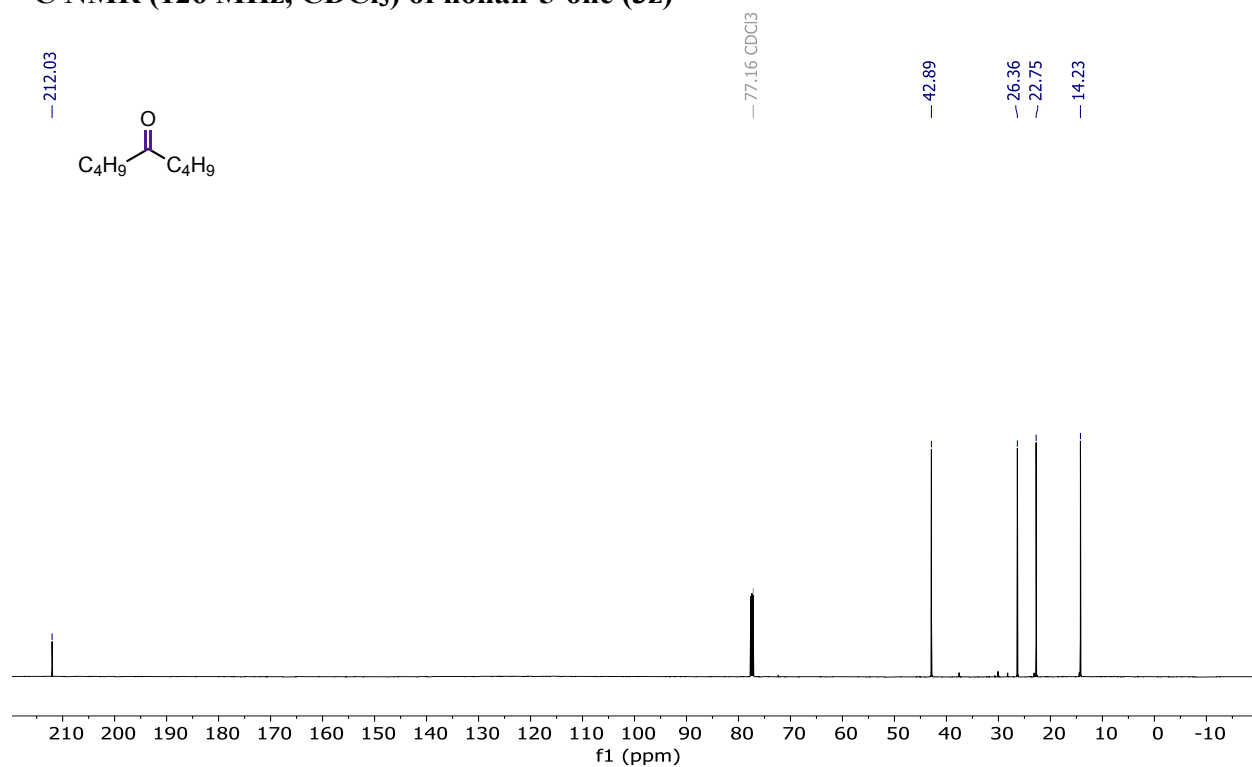
¹³C NMR (126 MHz, CDCl₃) of (perchloropyridin-4-yl)(phenyl)methanone (3y)



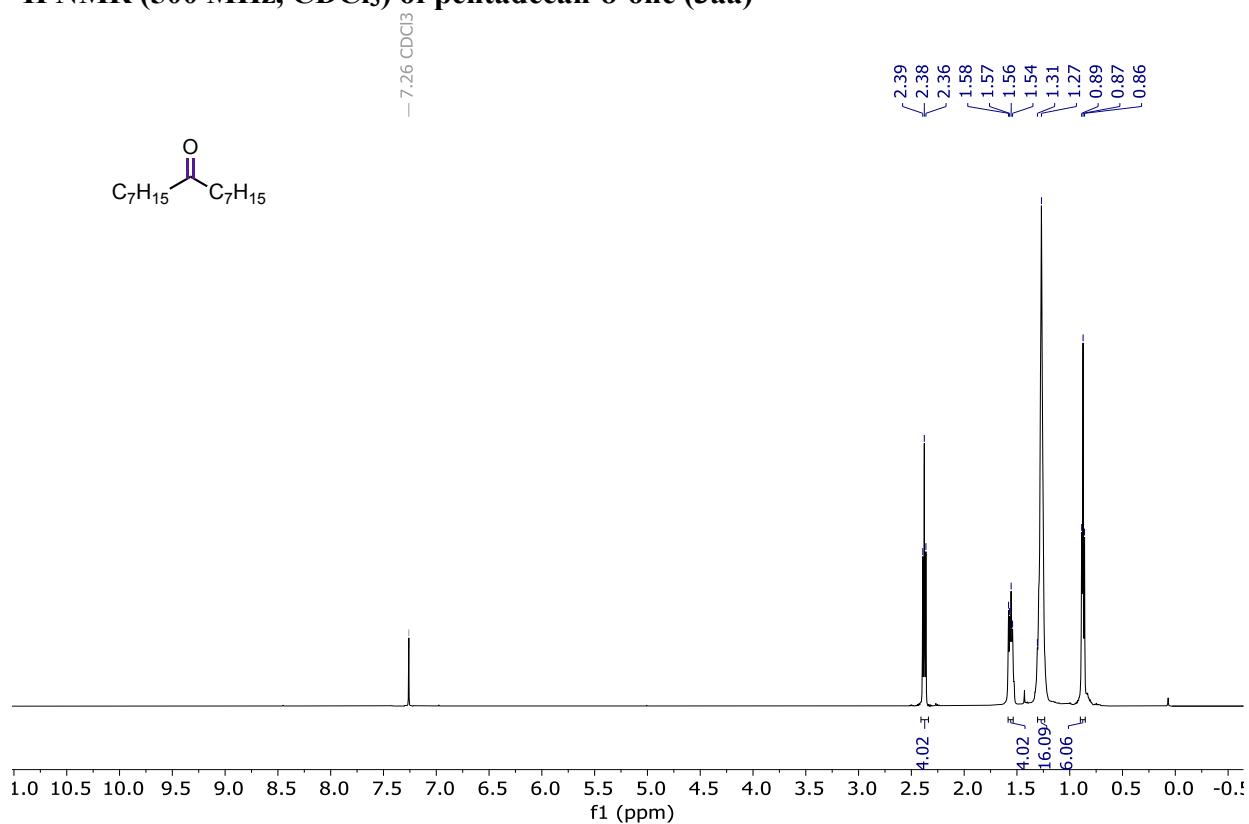
^1H NMR (500 MHz, CDCl_3) of nonan-5-one (3z)



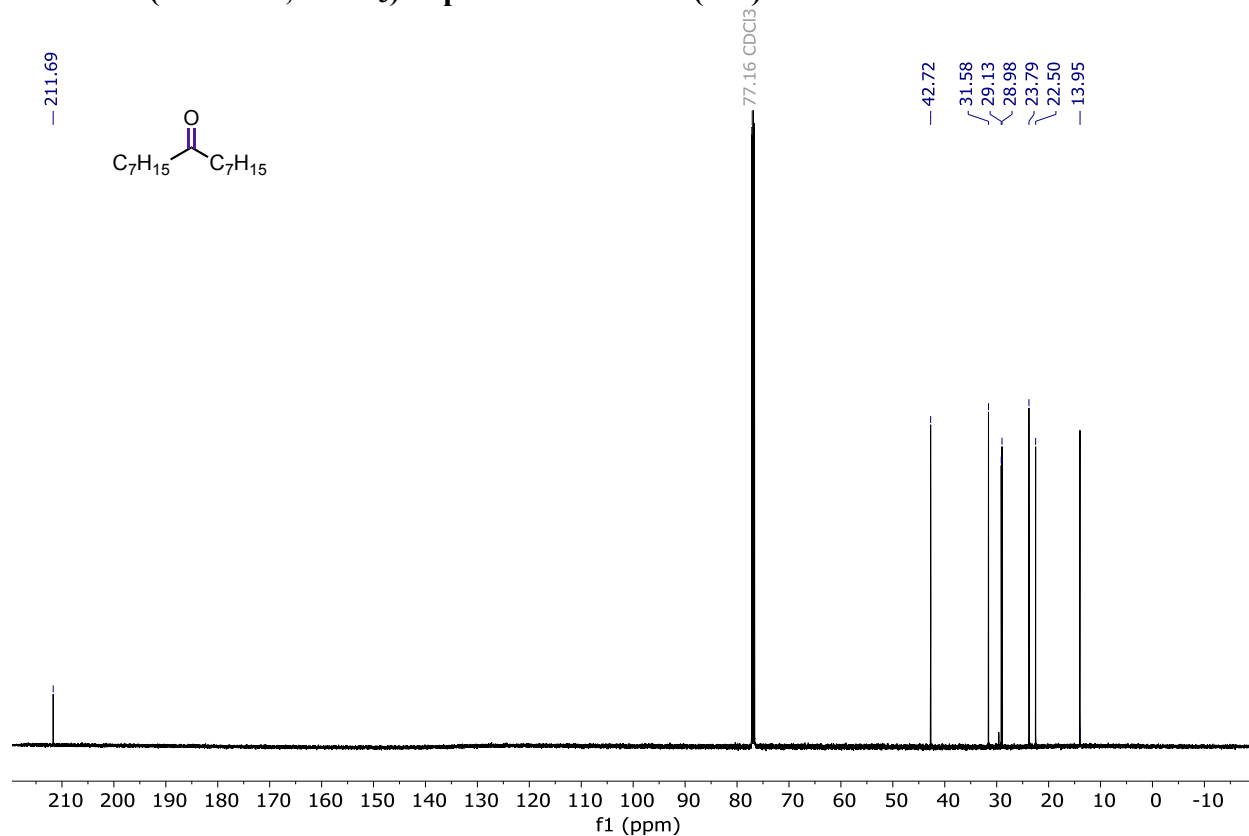
^{13}C NMR (126 MHz, CDCl_3) of nonan-5-one (3z)



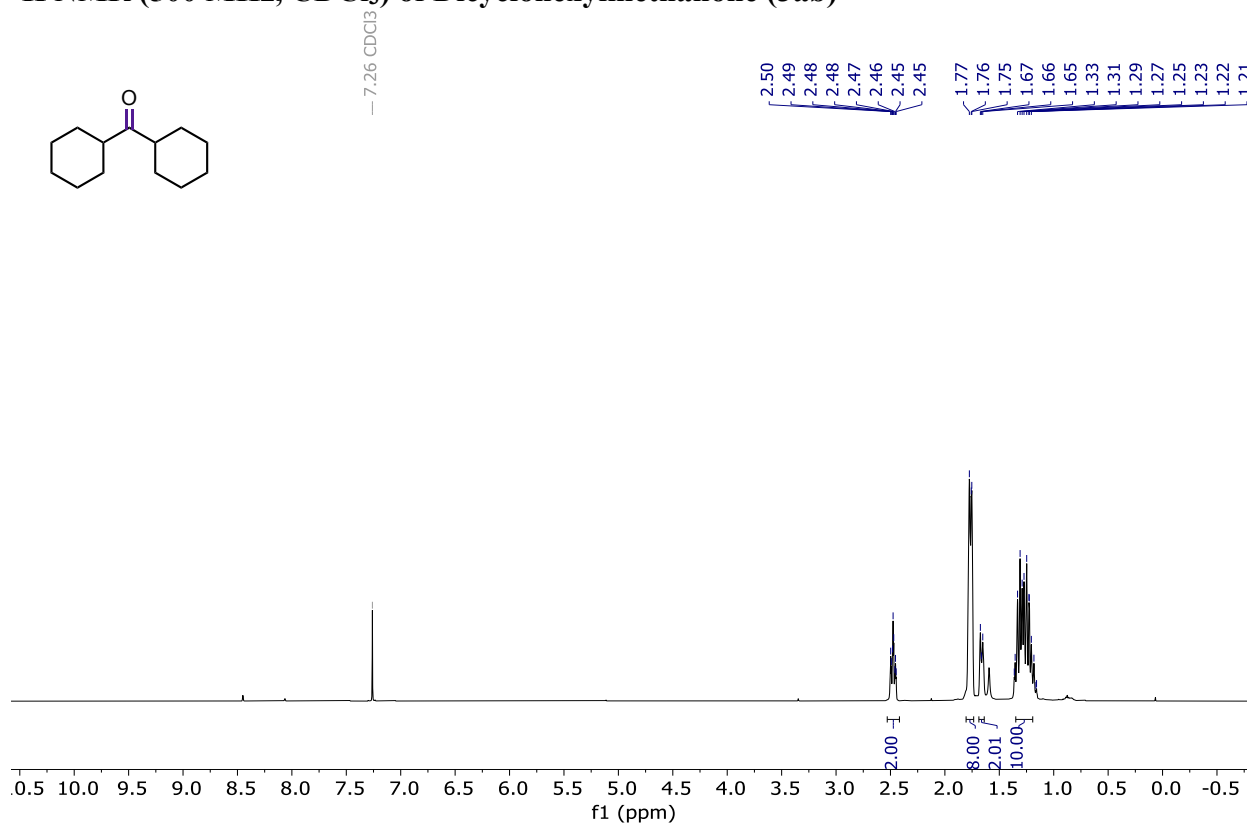
^1H NMR (500 MHz, CDCl_3) of pentadecan-8-one (3aa)



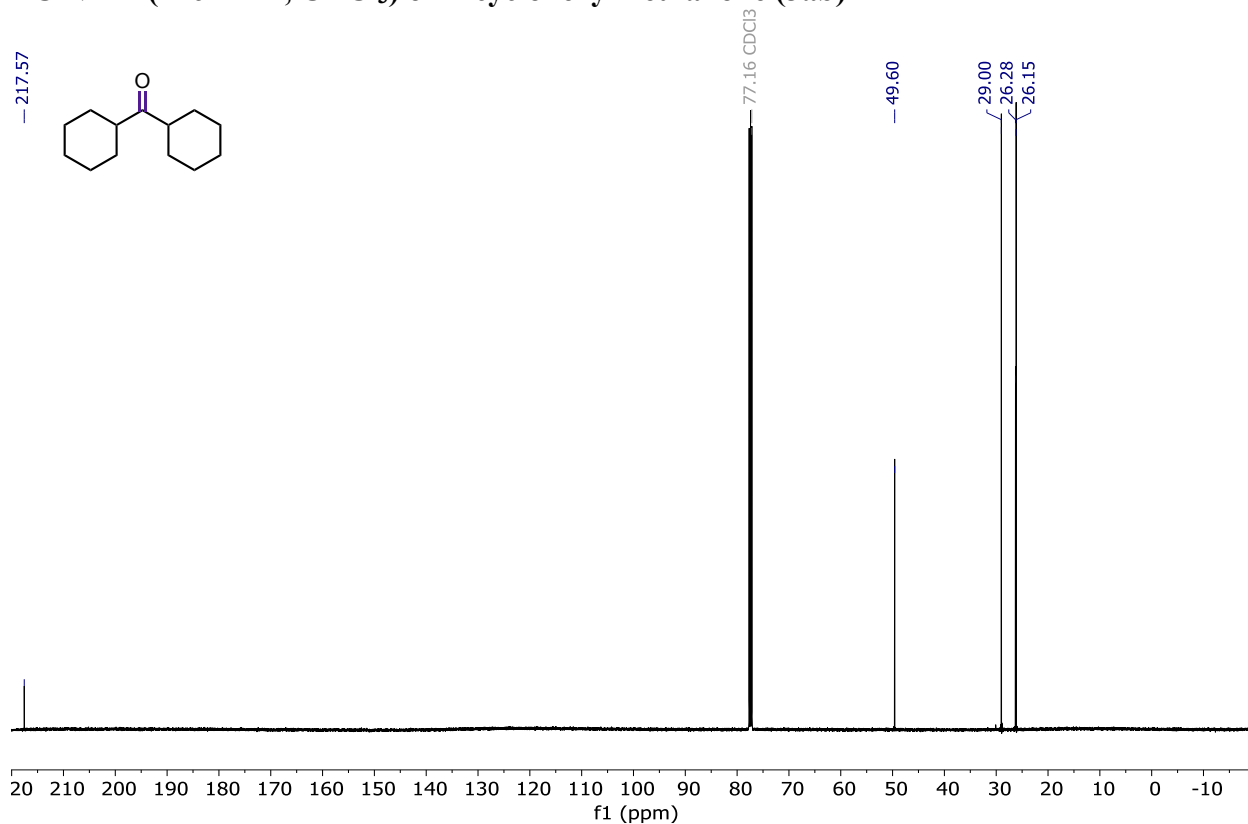
^{13}C NMR (126 MHz, CDCl_3) of pentadecan-8-one (3aa)



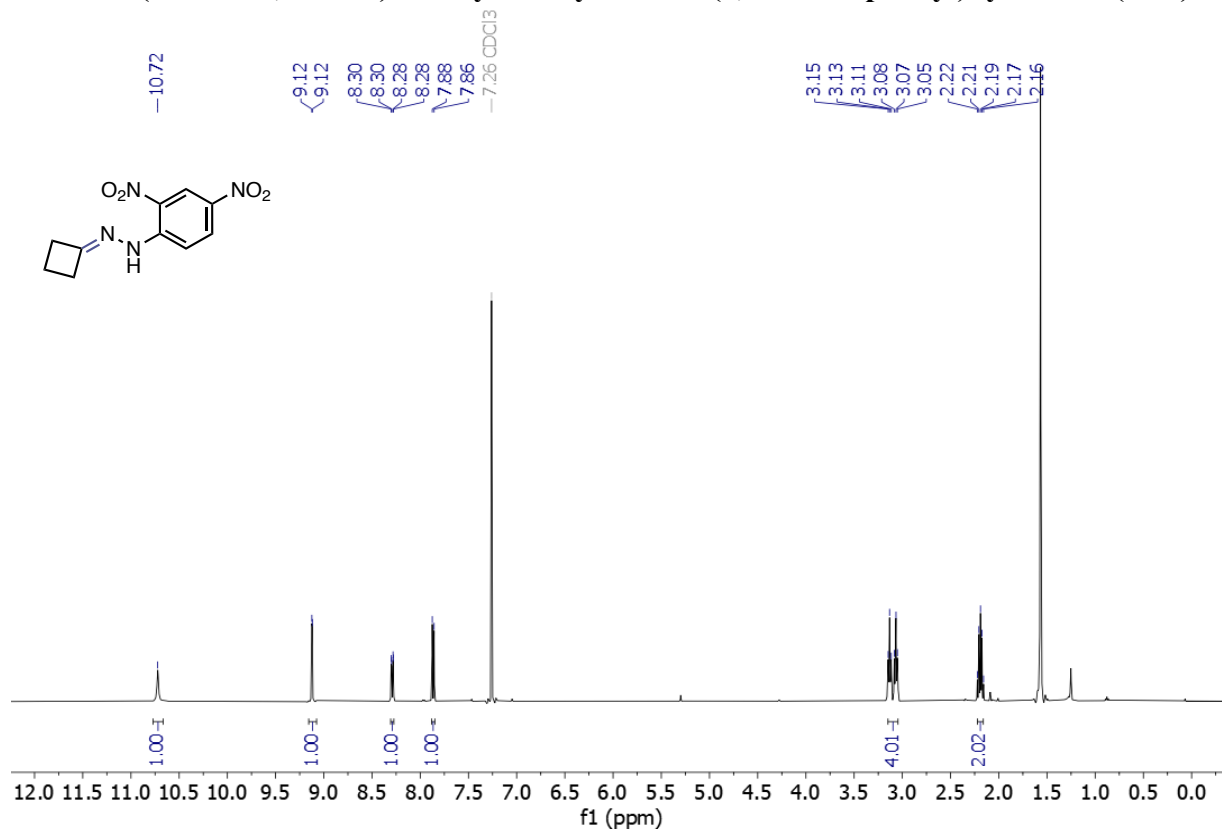
¹H NMR (500 MHz, CDCl₃) of Dicyclohexylmethanone (3ab)



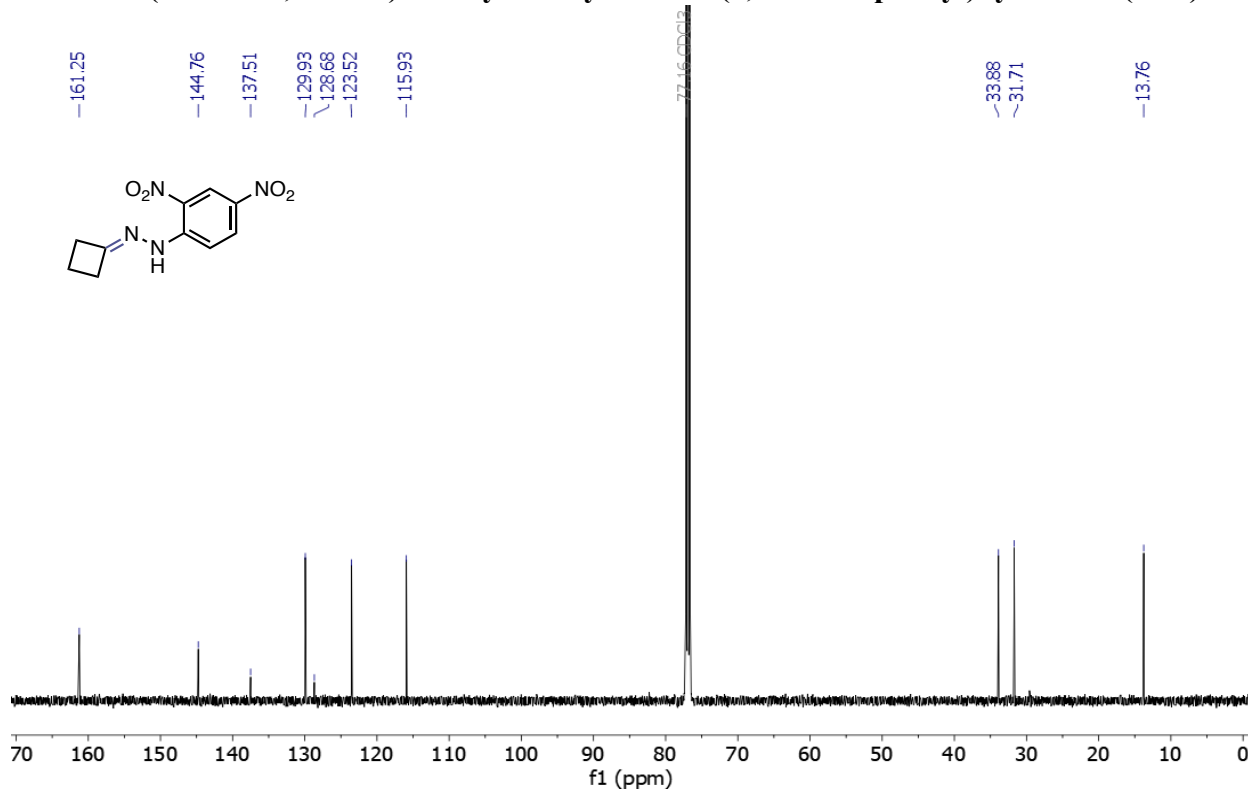
¹³C NMR (126 MHz, CDCl₃) of Dicyclohexylmethanone (3ab)



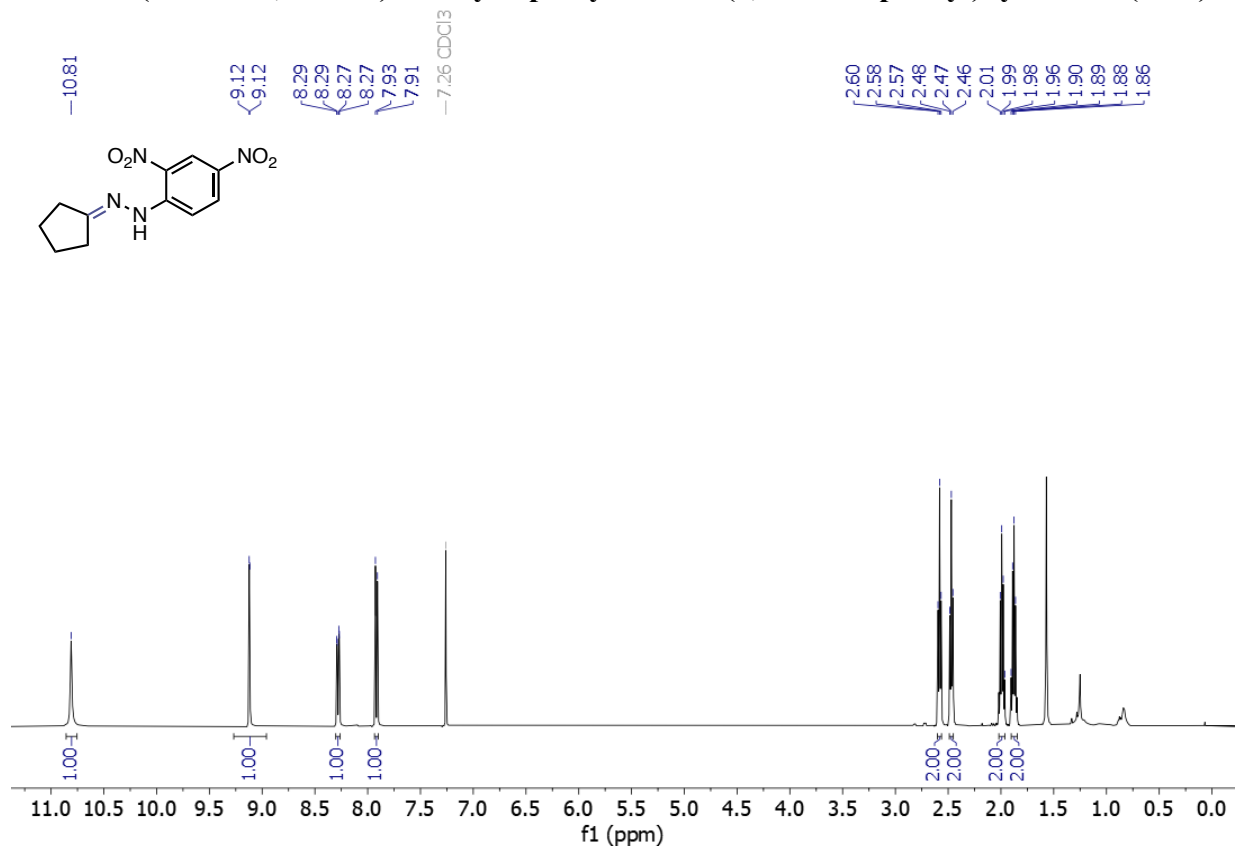
¹H NMR (400 MHz, CDCl₃) of 1-cyclobutylidene-2-(2,4-dinitrophenyl)hydrazine (3ac')



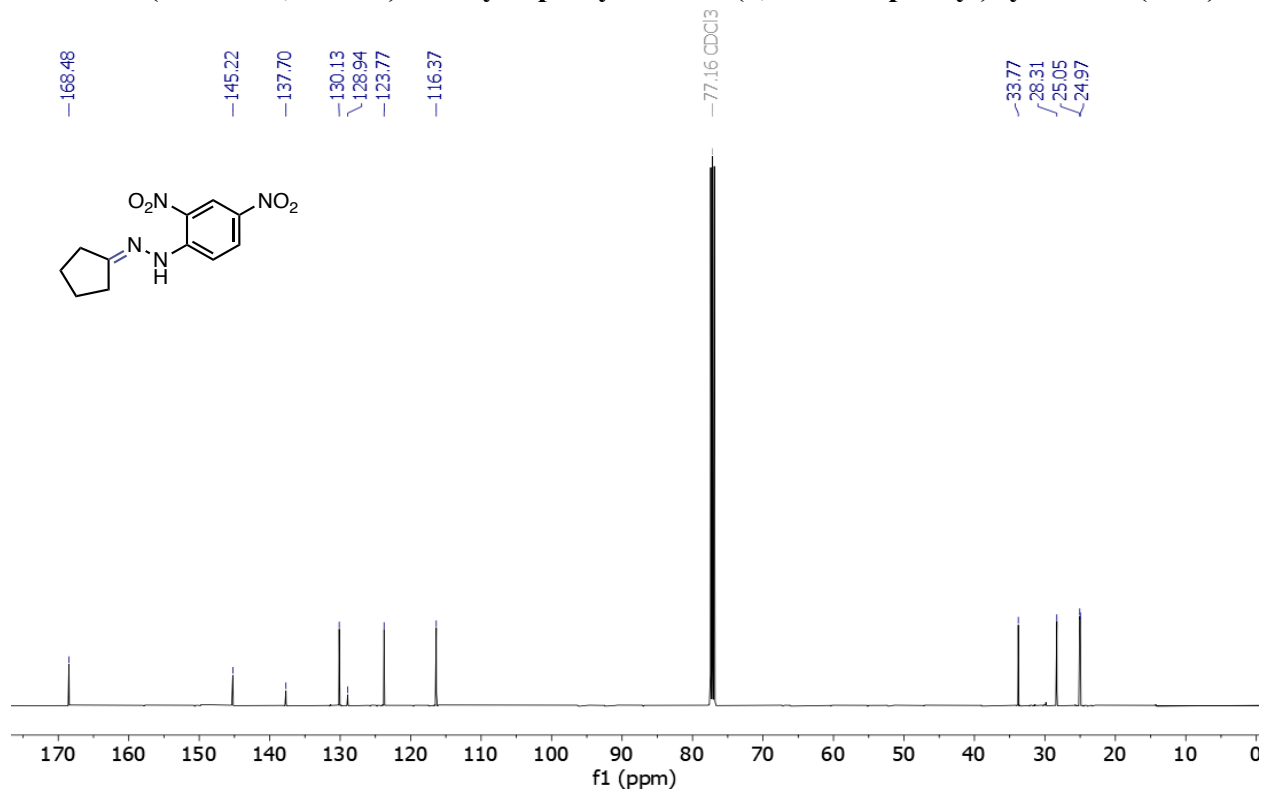
¹³C NMR (101 MHz, CDCl₃) of 1-cyclobutylidene-2-(2,4-dinitrophenyl)hydrazine (3ac')



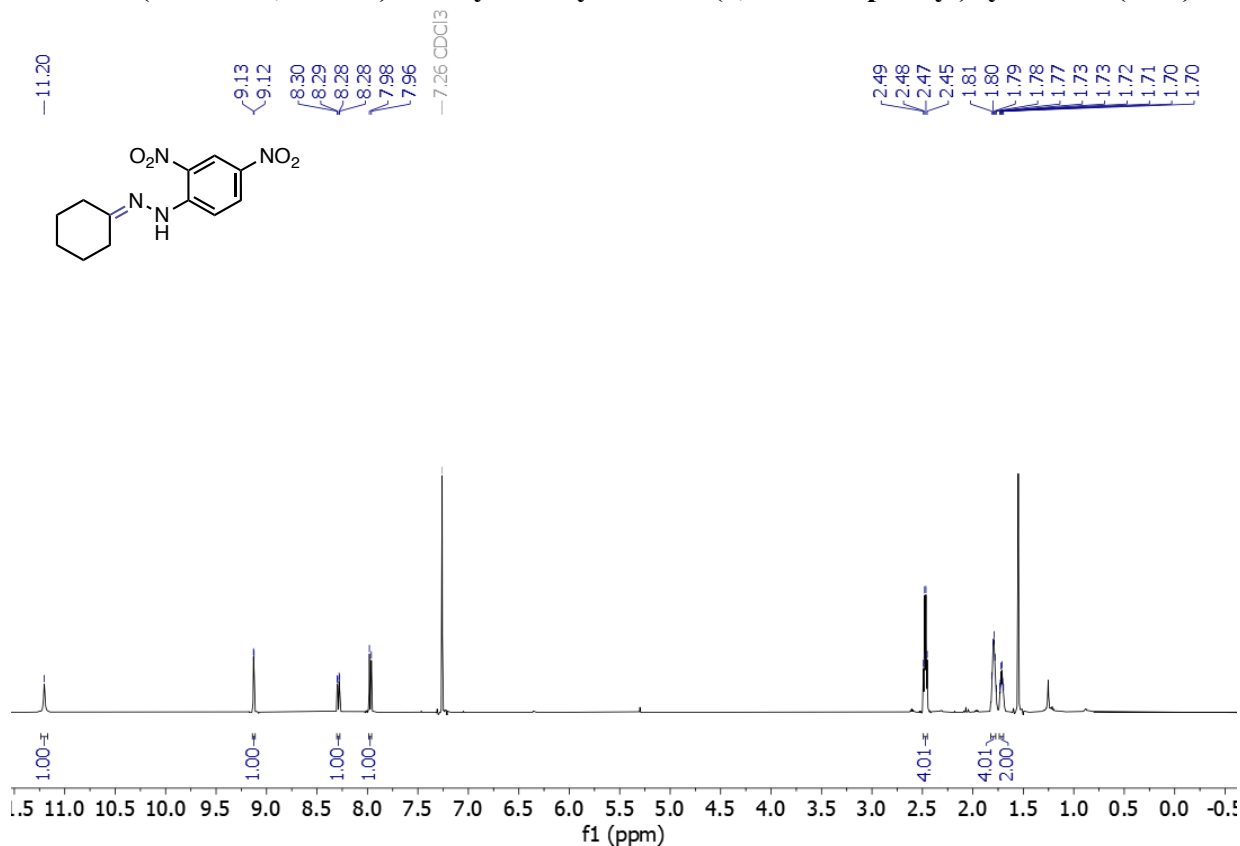
¹H NMR (400 MHz, CDCl₃) of 1-cyclopentylidene-2-(2,4-dinitrophenyl)hydrazine (3ad')



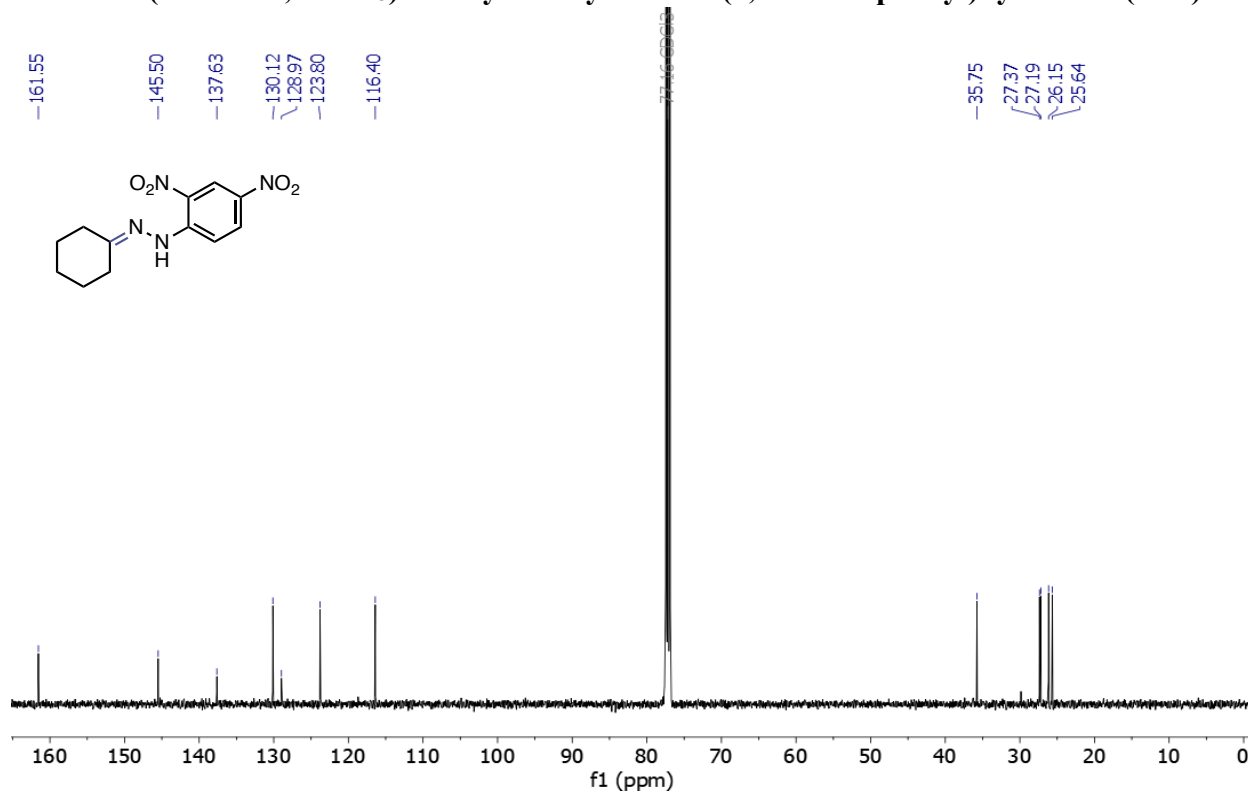
¹³C NMR (101 MHz, CDCl₃) of 1-cyclopentylidene-2-(2,4-dinitrophenyl)hydrazine (3ad')



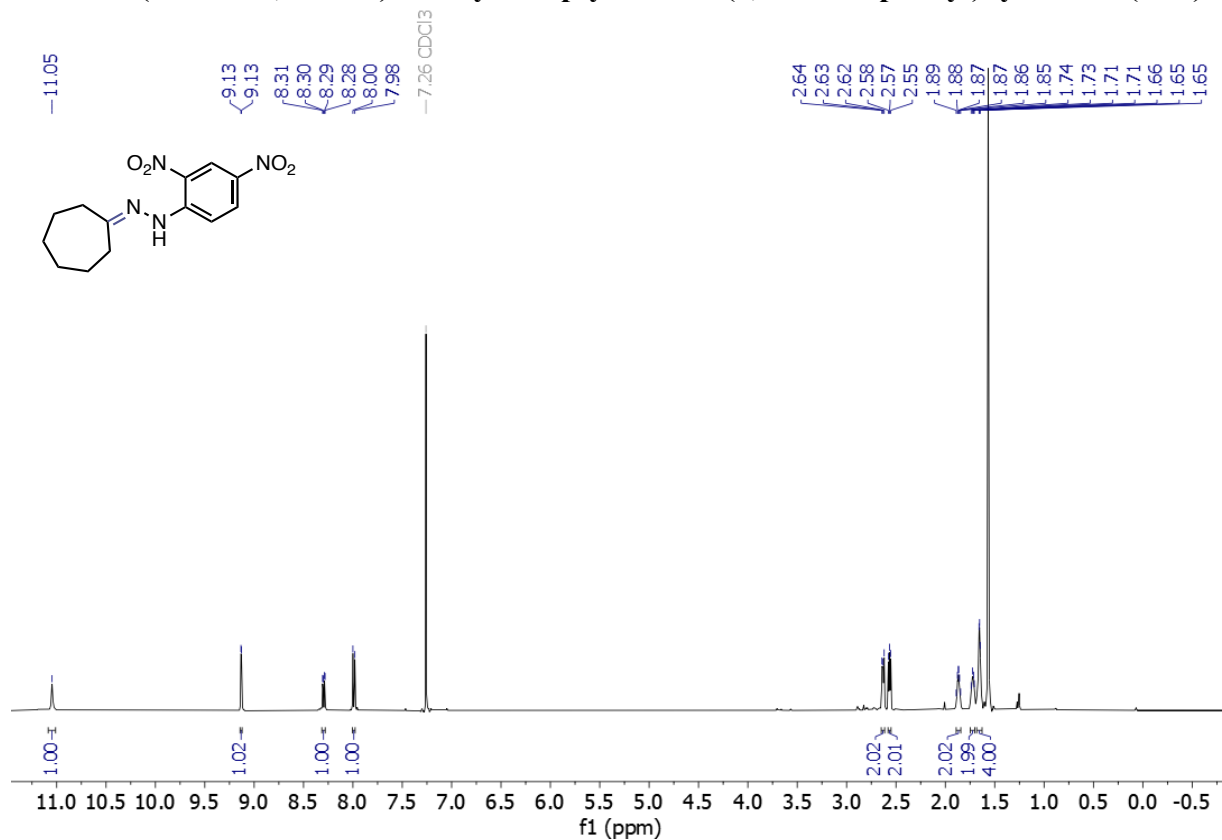
¹H NMR (400 MHz, CDCl₃) of 1-cyclohexylidene-2-(2,4-dinitrophenyl)hydrazine (3ae')



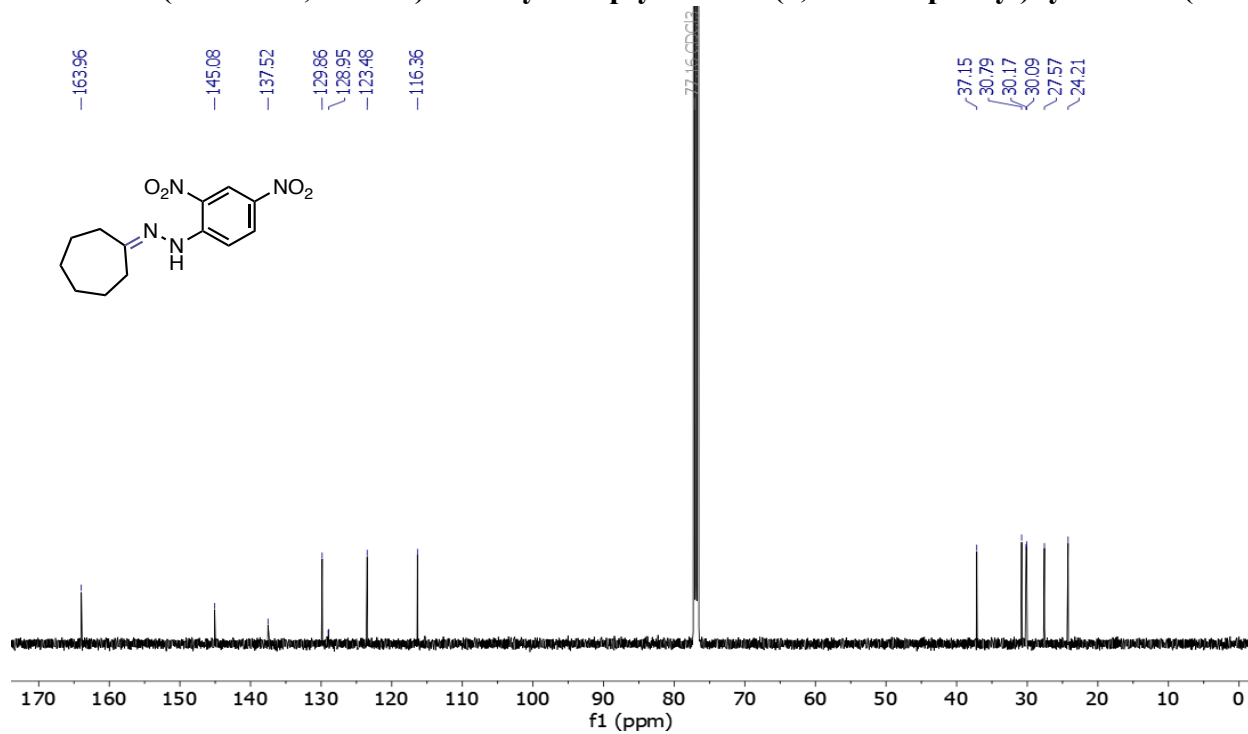
¹³C NMR (101 MHz, CDCl₃) of 1-cyclohexylidene-2-(2,4-dinitrophenyl)hydrazine (3ae')



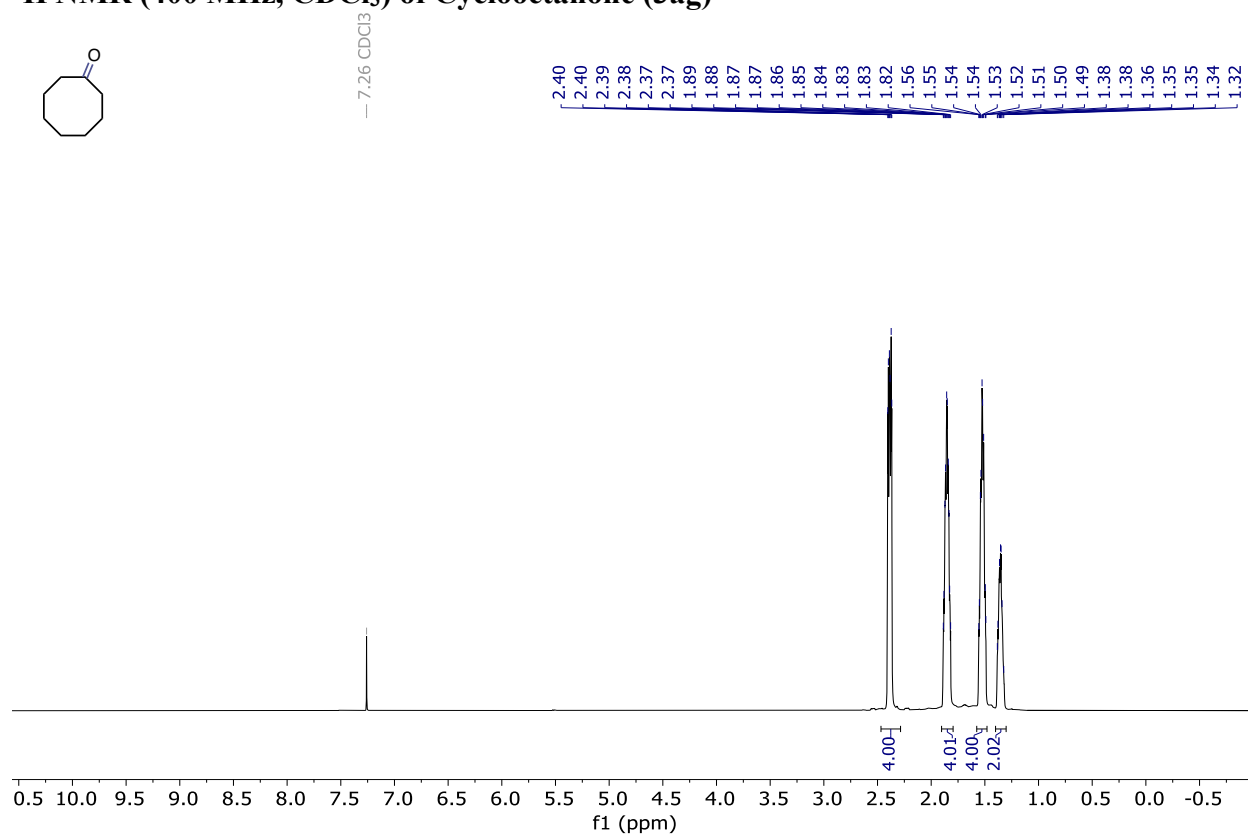
¹H NMR (400 MHz, CDCl₃) of 1-cycloheptylidene-2-(2,4-dinitrophenyl)hydrazine (3af')



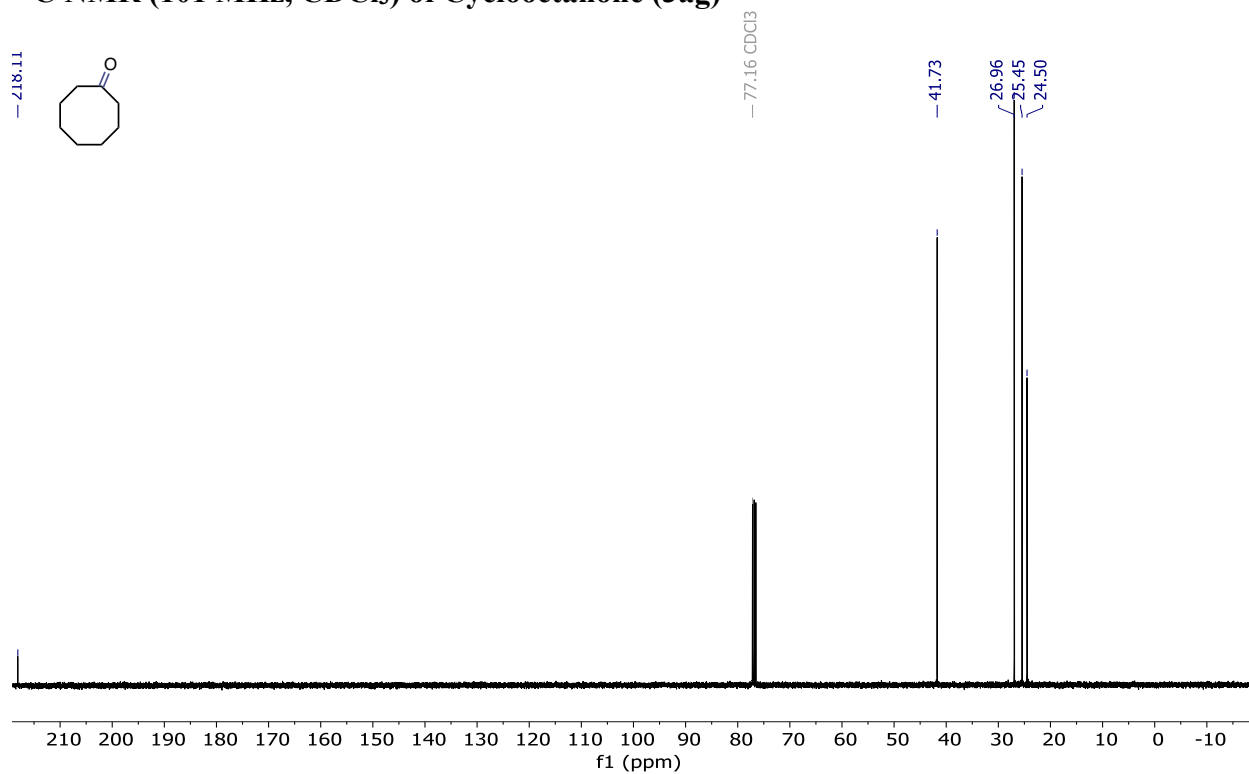
¹³C NMR (101 MHz, CDCl₃) of 1-cycloheptylidene-2-(2,4-dinitrophenyl)hydrazine (3af')



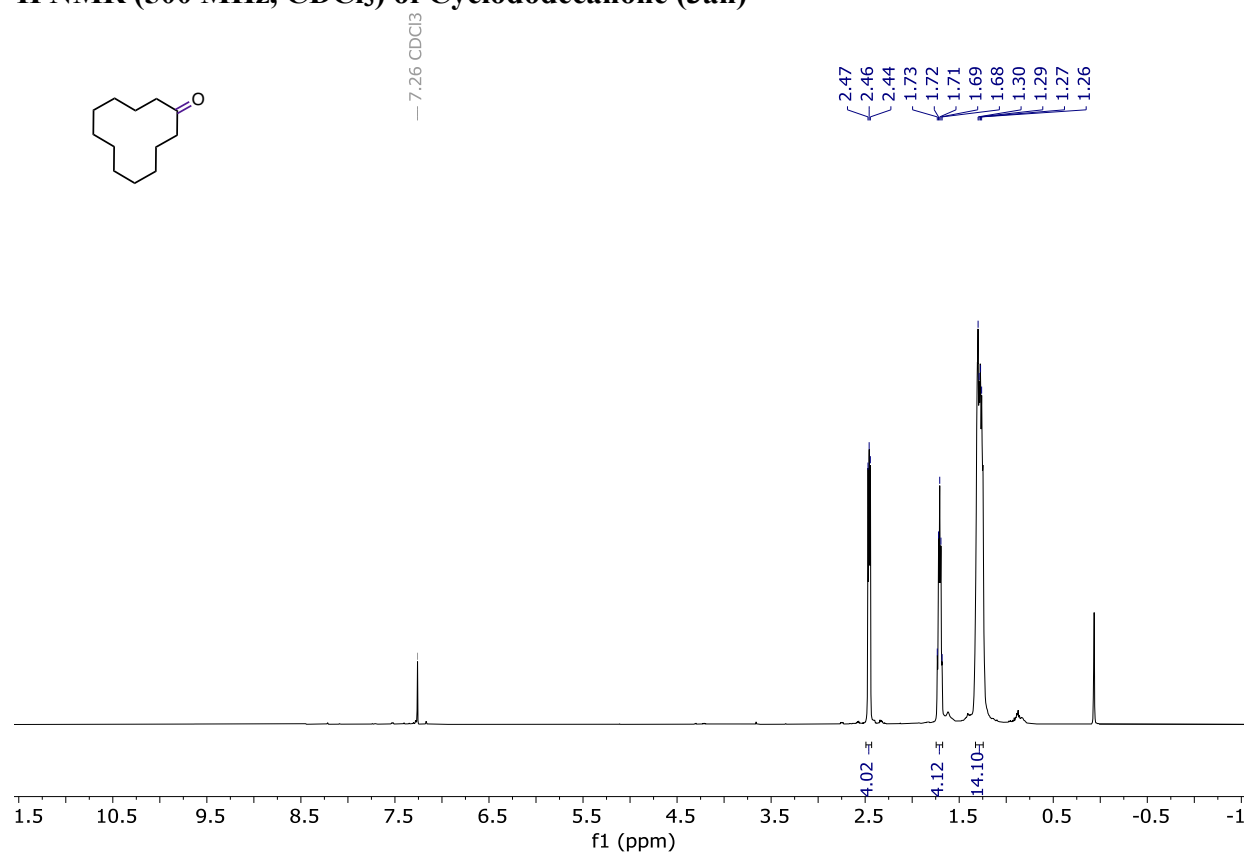
¹H NMR (400 MHz, CDCl₃) of Cyclooctanone (3ag)



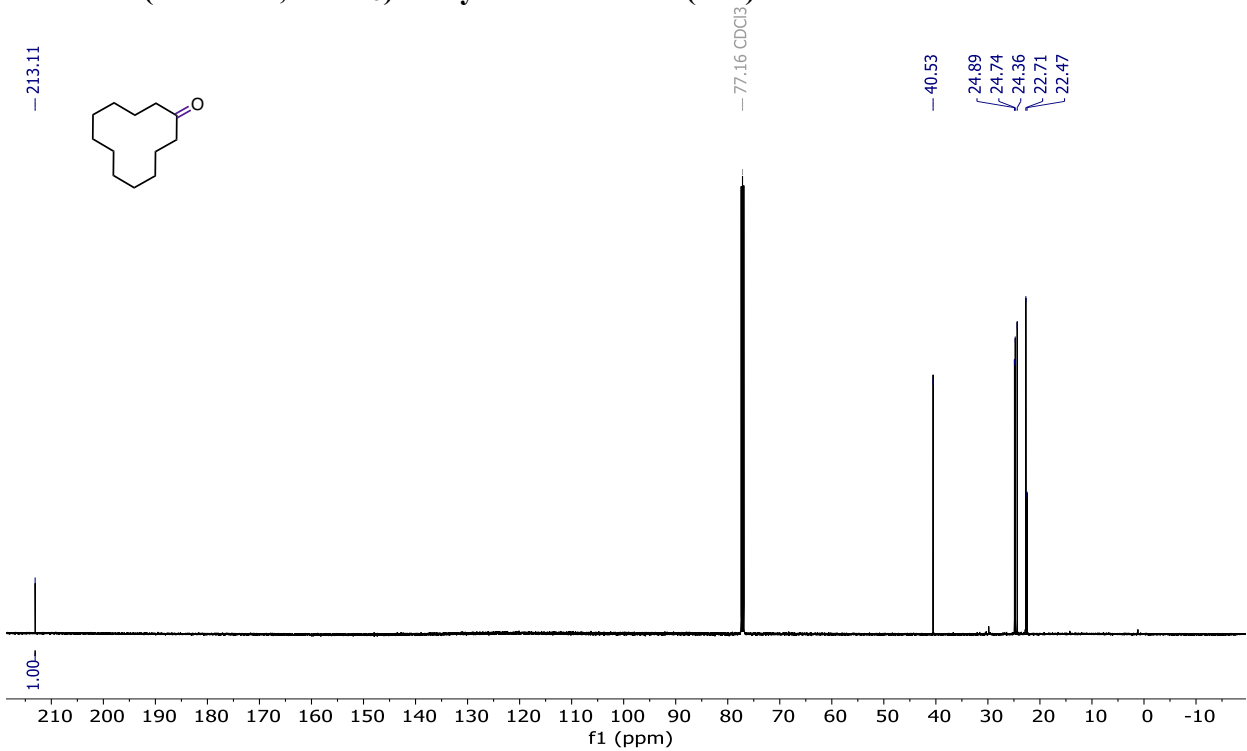
¹³C NMR (101 MHz, CDCl₃) of Cyclooctanone (3ag)



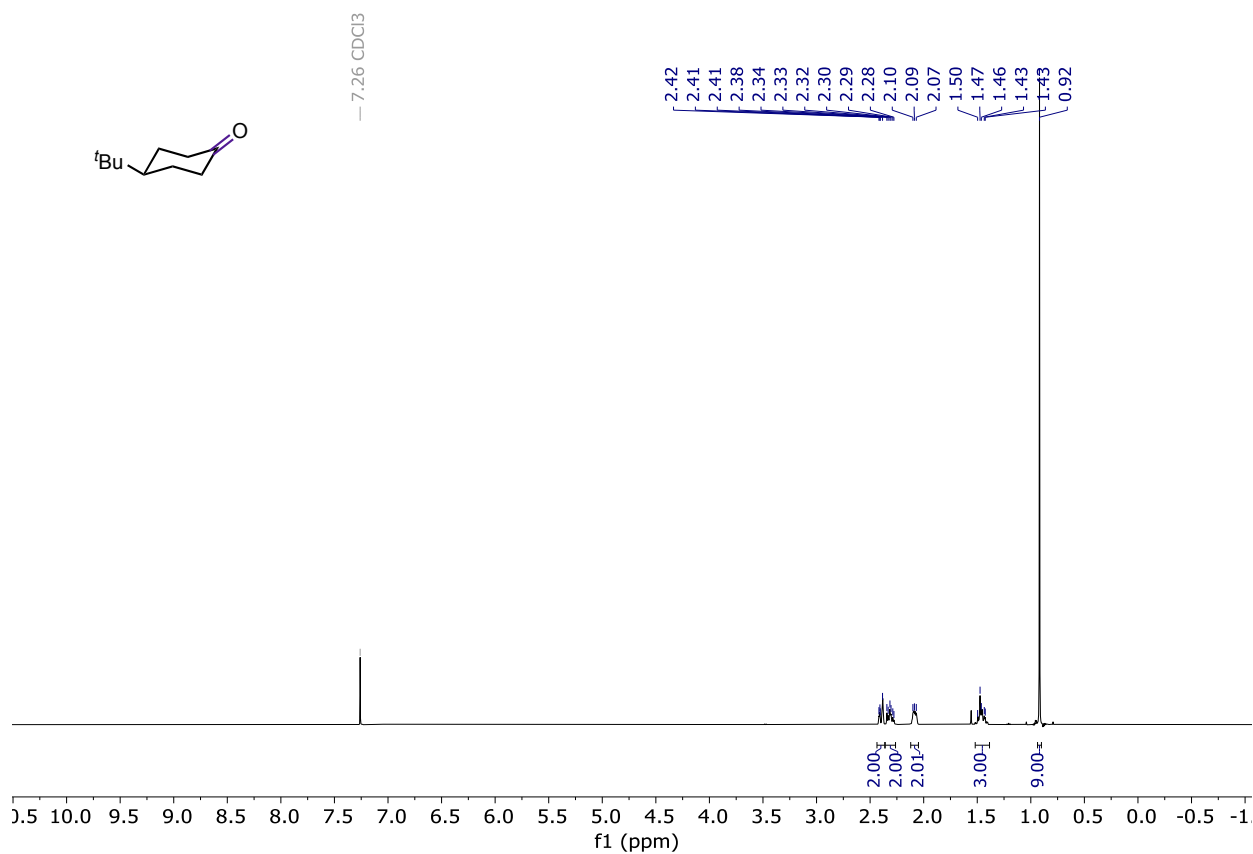
¹H NMR (500 MHz, CDCl₃) of Cyclododecanone (3ah)



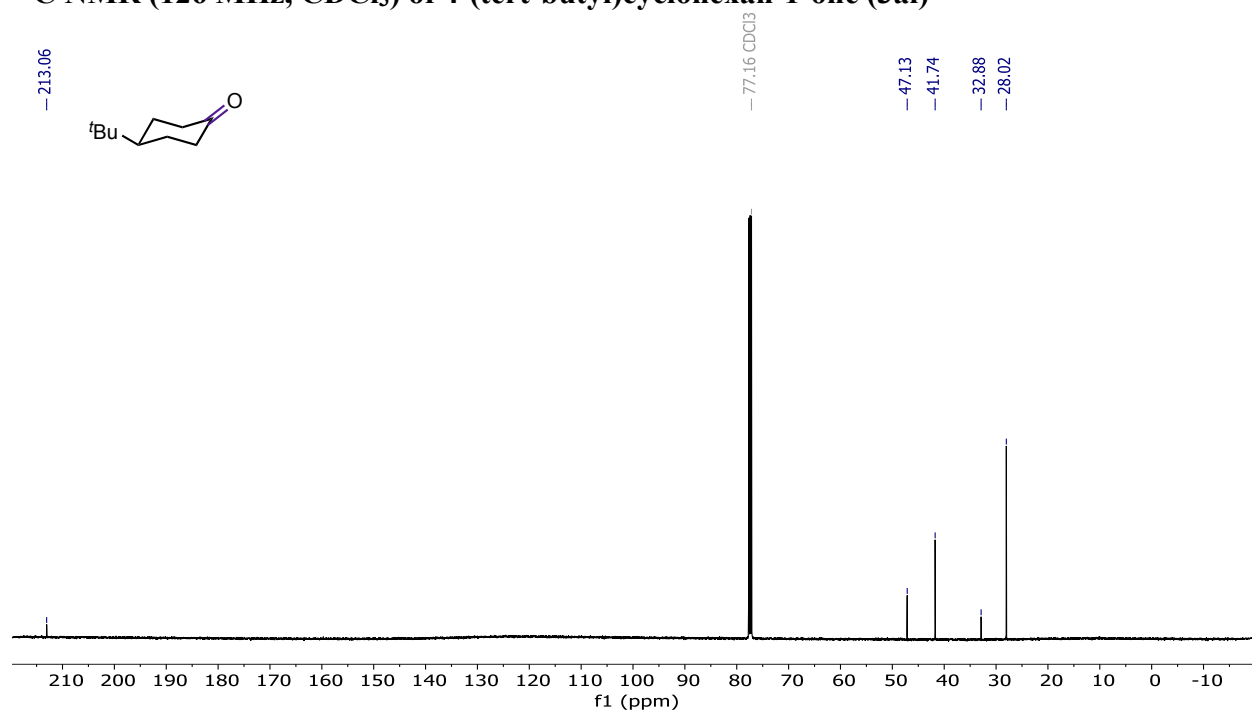
¹³C NMR (101 MHz, CDCl₃) of Cyclododecanone (3ah)



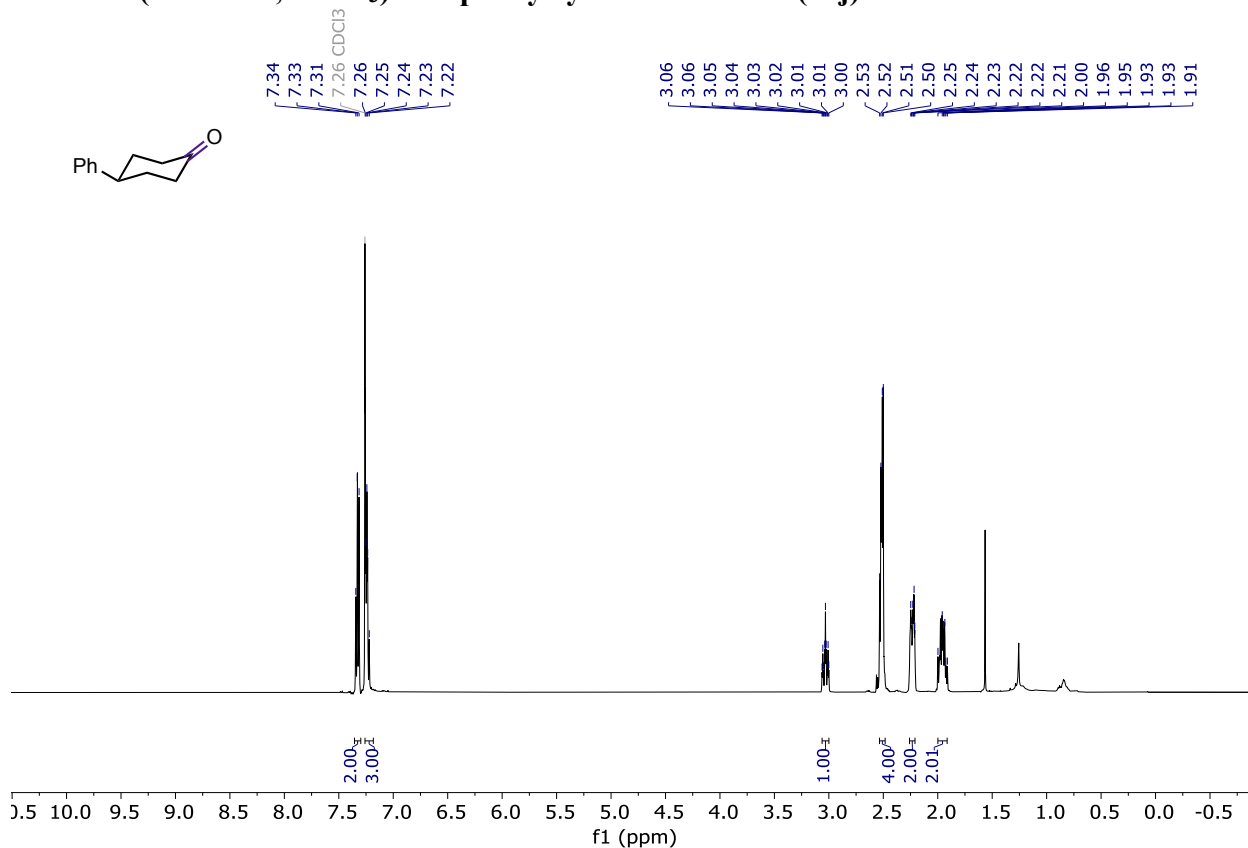
¹H NMR (500 MHz, CDCl₃) of 4-(tert-butyl)cyclohexan-1-one (3ai)



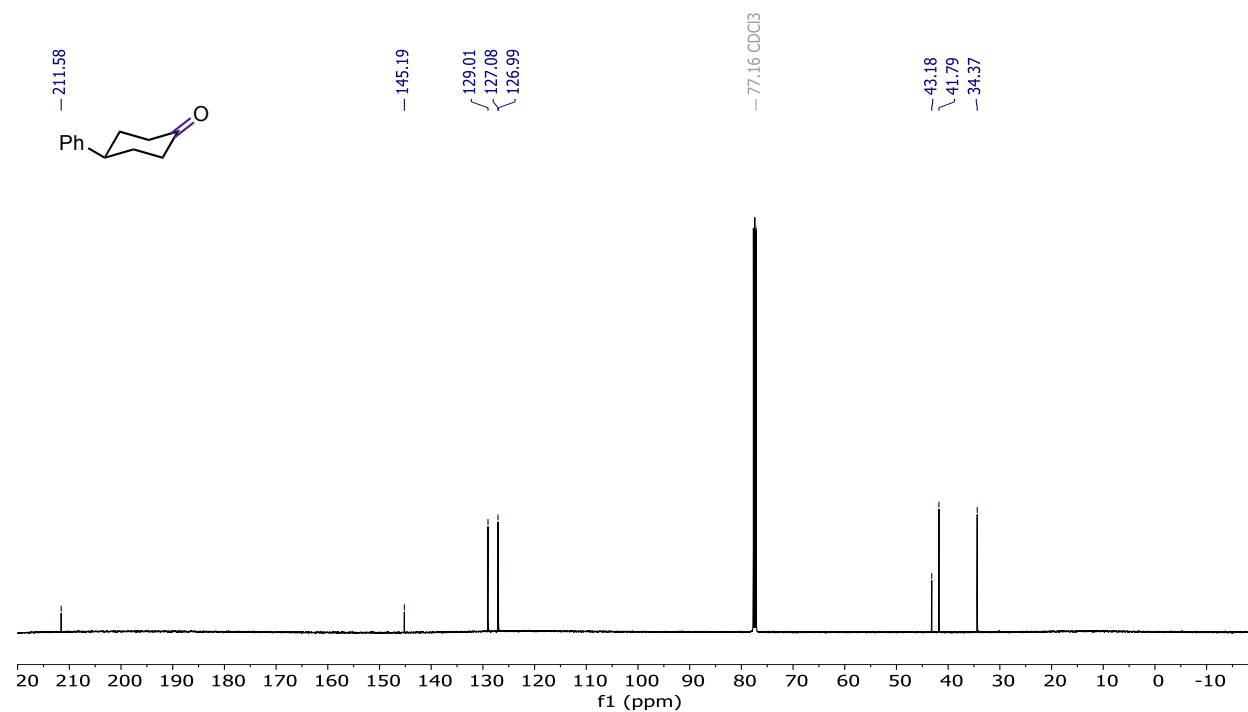
¹³C NMR (126 MHz, CDCl₃) of 4-(tert-butyl)cyclohexan-1-one (3ai)



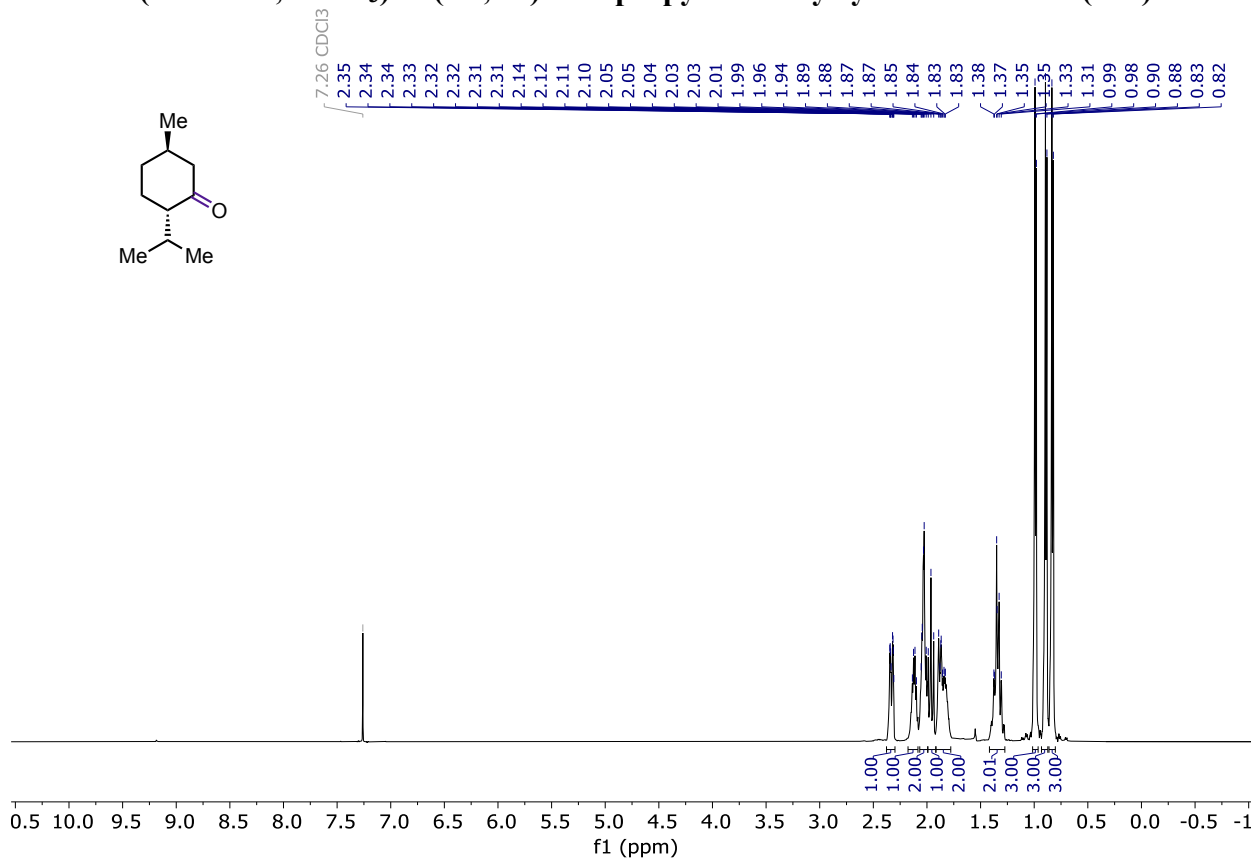
¹H NMR (500 MHz, CDCl₃) of 4-phenylcyclohexan-1-one (3aj)



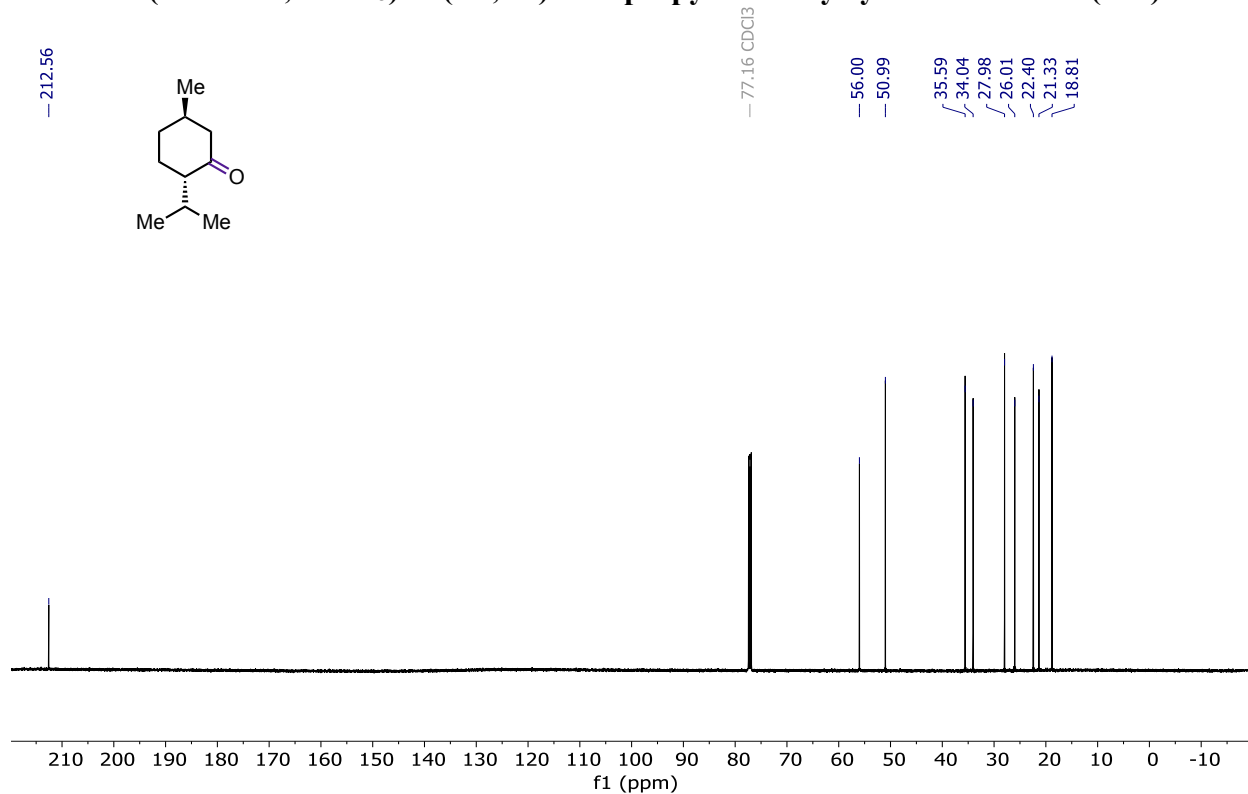
¹³C NMR (126 MHz, CDCl₃) of 4-phenylcyclohexan-1-one (3aj)



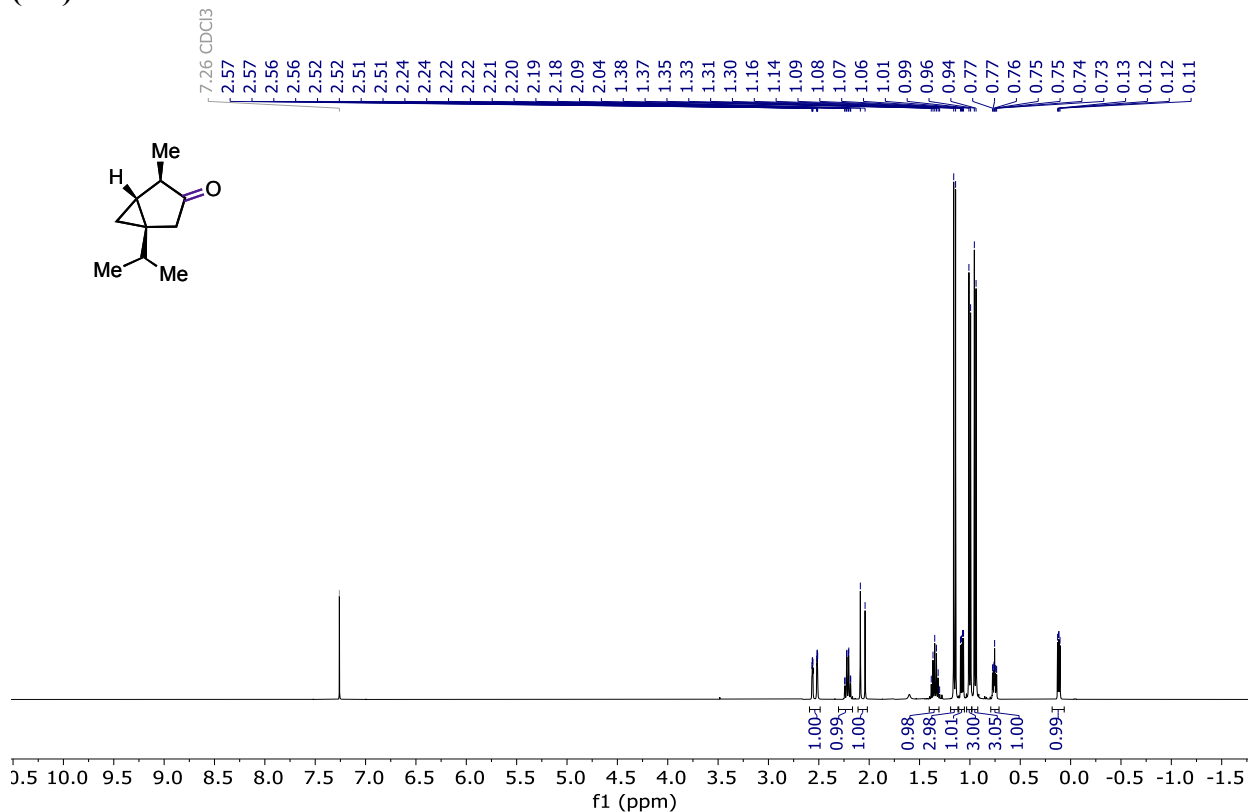
¹H NMR (500 MHz, CDCl₃) of (2R,5R)-2-isopropyl-5-methylcyclohexan-1-one (3ak)



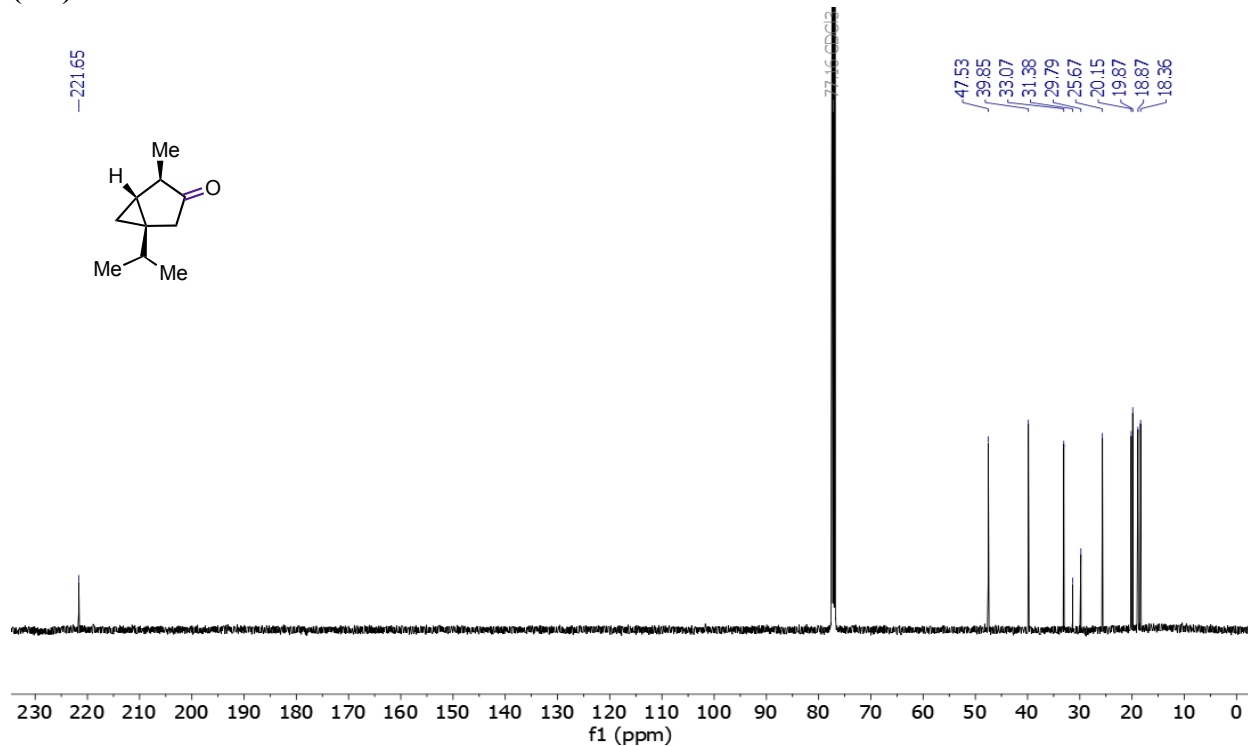
¹³C NMR (126 MHz, CDCl₃) of (2R,5R)-2-isopropyl-5-methylcyclohexan-1-one (3ak)



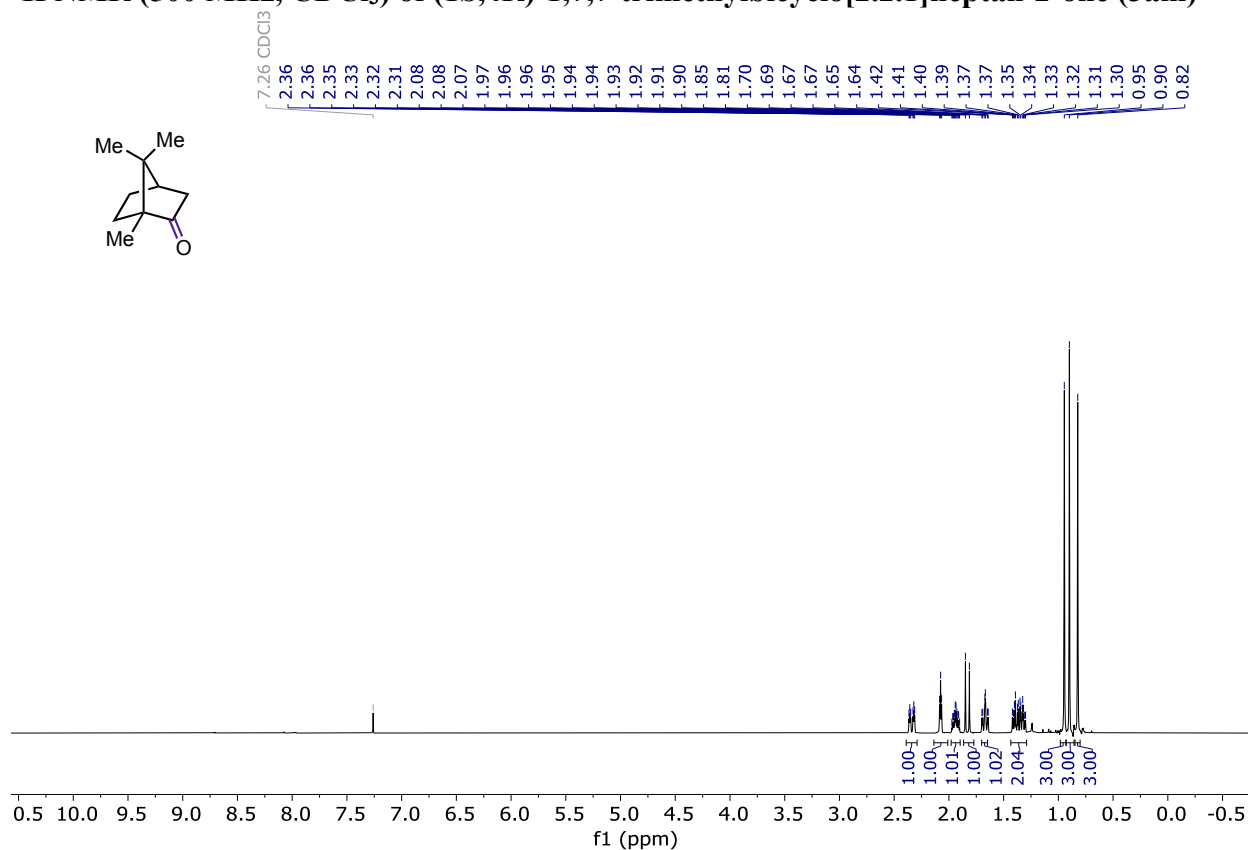
¹H NMR (400 MHz, CDCl₃) of (1S,4R,5R)-1-isopropyl-4-methylbicyclo[3.1.0]hexan-3-one (3a)



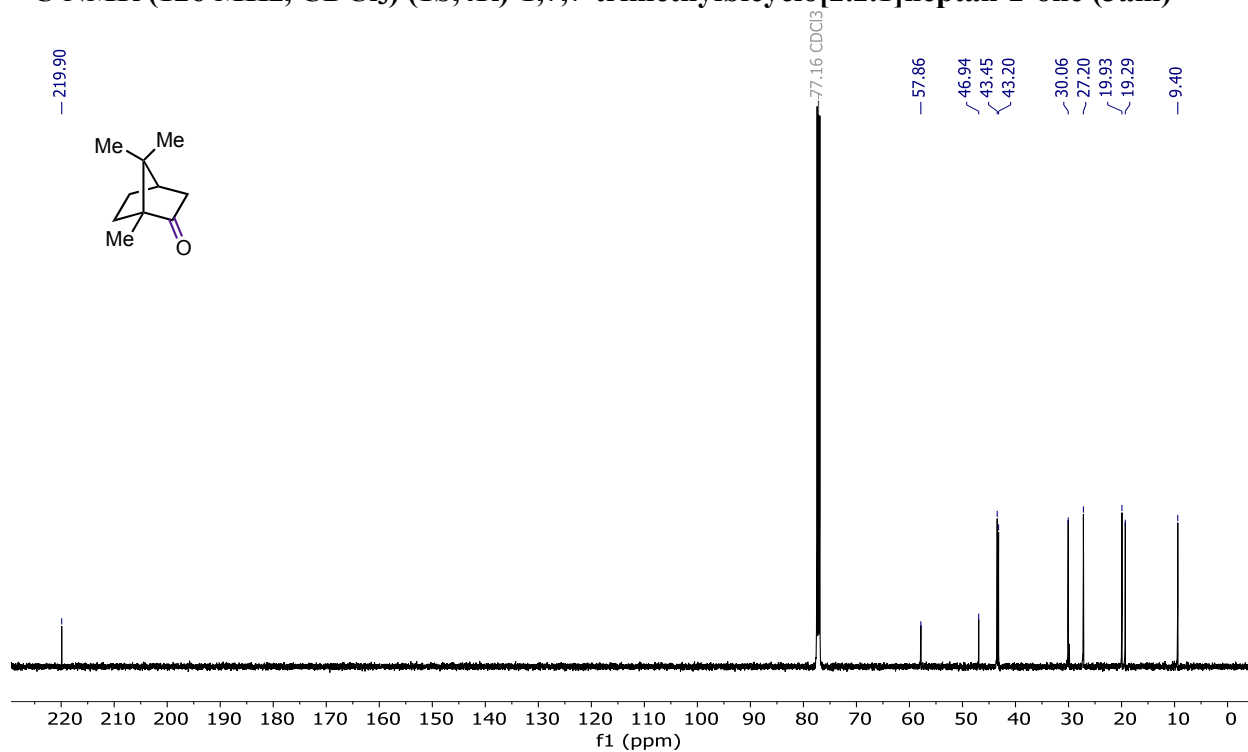
¹³C NMR (126 MHz, CDCl₃) of (1S,4R,5R)-1-isopropyl-4-methylbicyclo[3.1.0]hexan-3-one (3a)



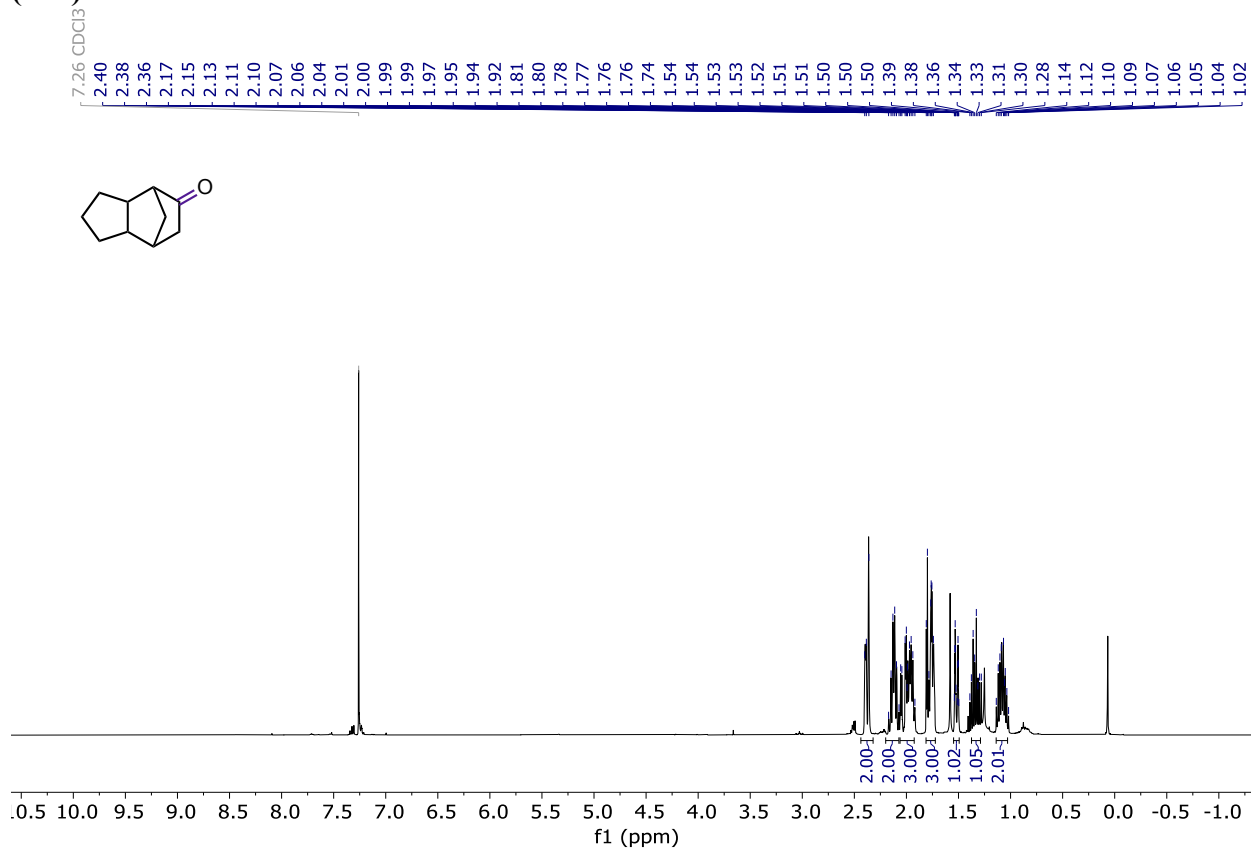
¹H NMR (500 MHz, CDCl₃) of (1S,4R)-1,7,7-trimethylbicyclo[2.2.1]heptan-2-one (3am)



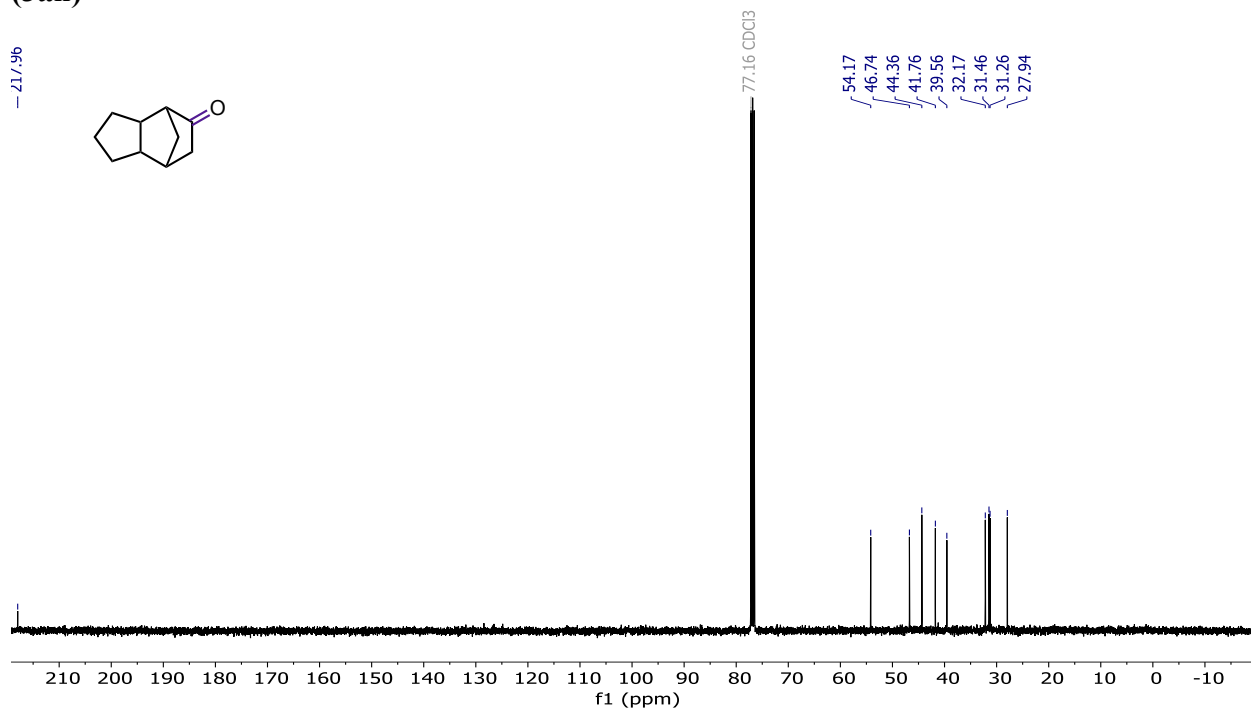
¹³C NMR (126 MHz, CDCl₃) (1S,4R)-1,7,7-trimethylbicyclo[2.2.1]heptan-2-one (3am)



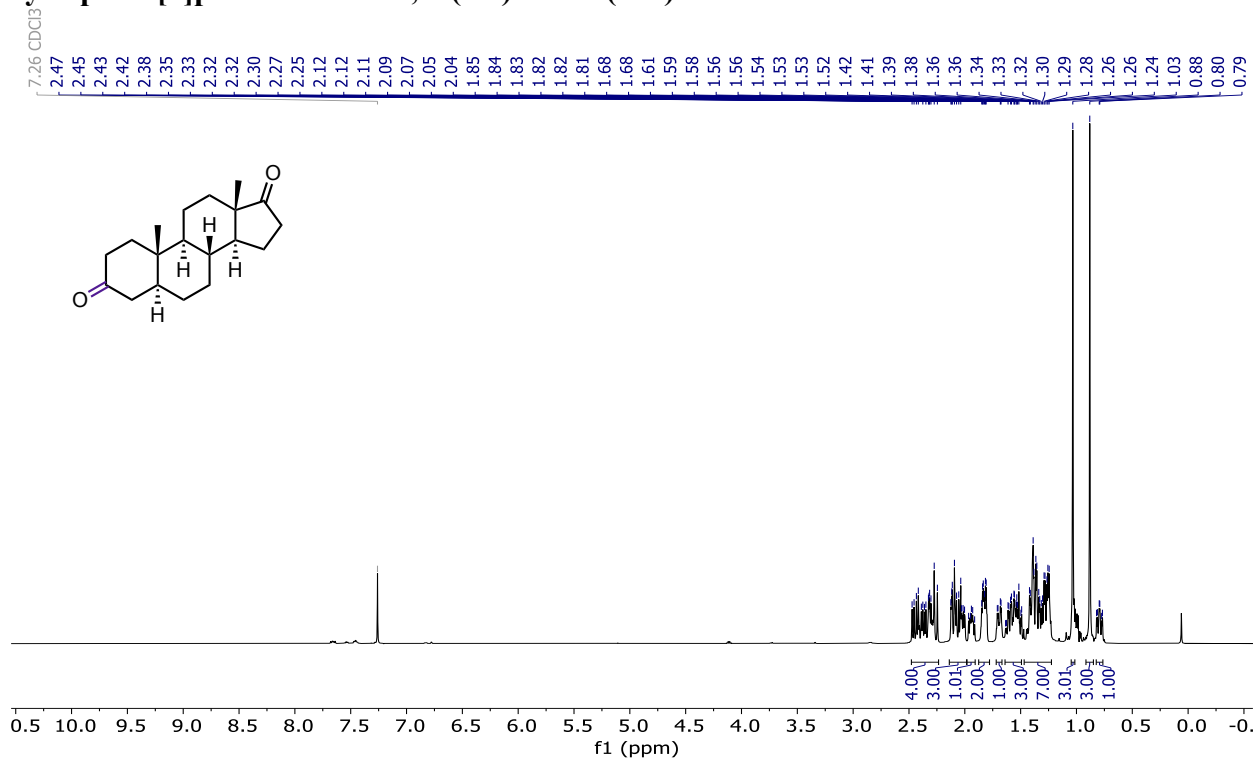
¹H NMR (400 MHz, CDCl₃) of (3aR,4S,7S,7aR)-octahydro-5H-4,7-methanoinden-5-one (3an)



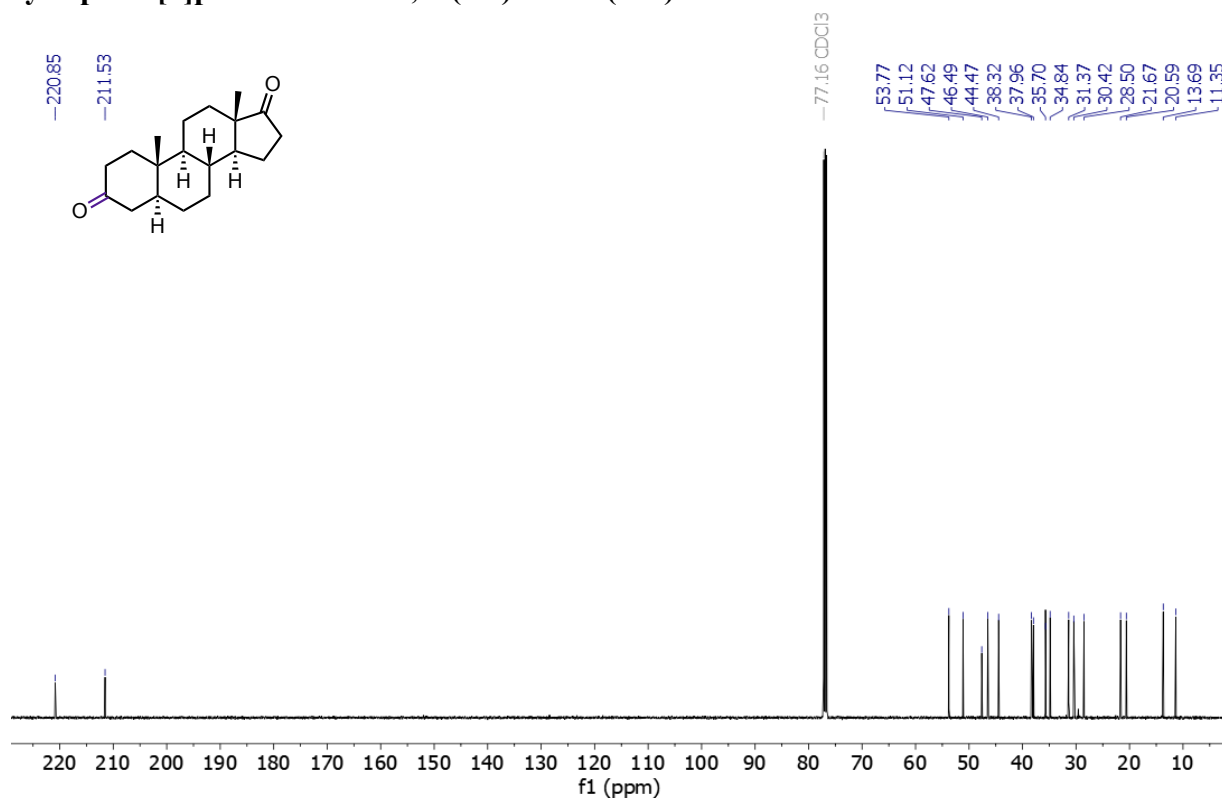
¹³C NMR (101 MHz, CDCl₃) of (3aR,4S,7S,7aR)-octahydro-5H-4,7-methanoinden-5-one (3an)



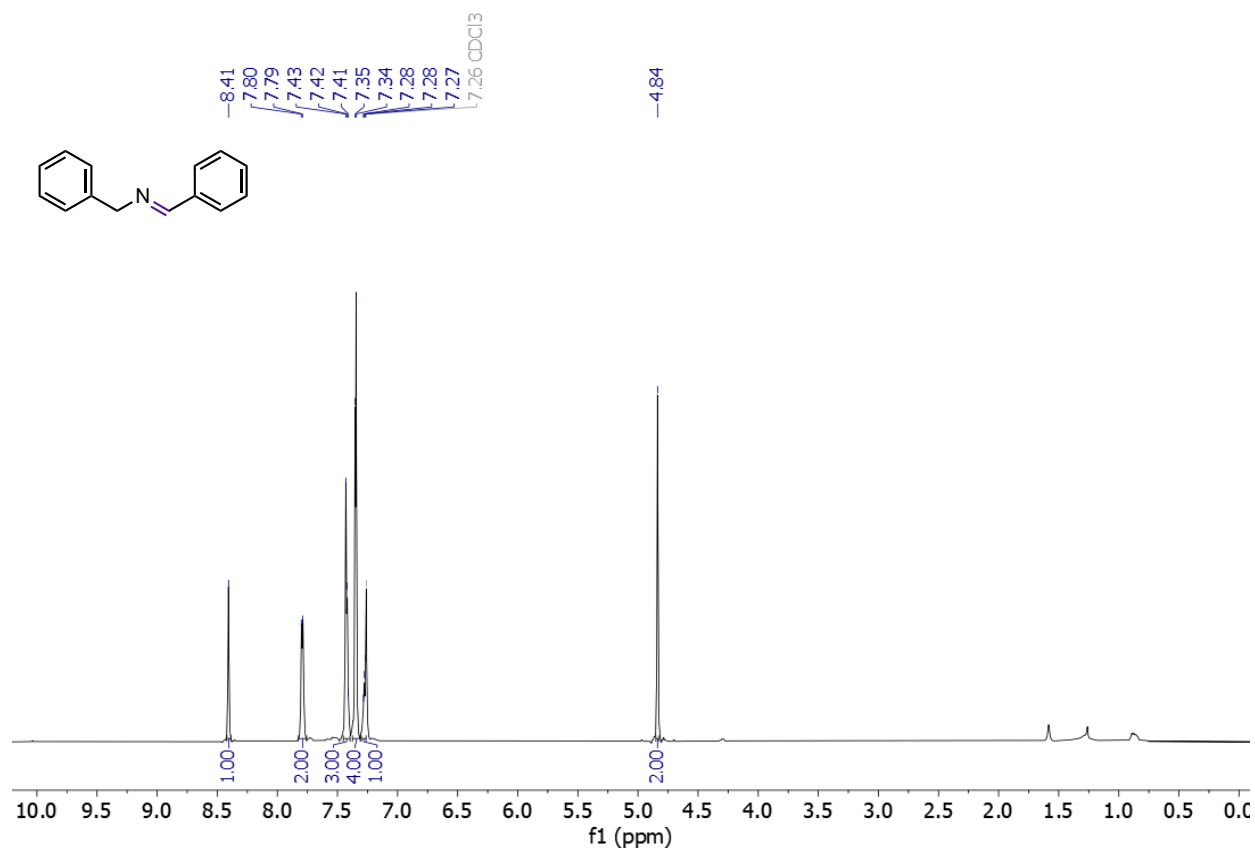
¹H NMR (500 MHz, CDCl₃) of (8*R*,9*S*,10*S*,13*S*,14*S*)-10,13-dimethyltetradecahydro-3*H*-cyclopenta[*a*]phenanthrene-3,17(2*H*)-dione (3a)



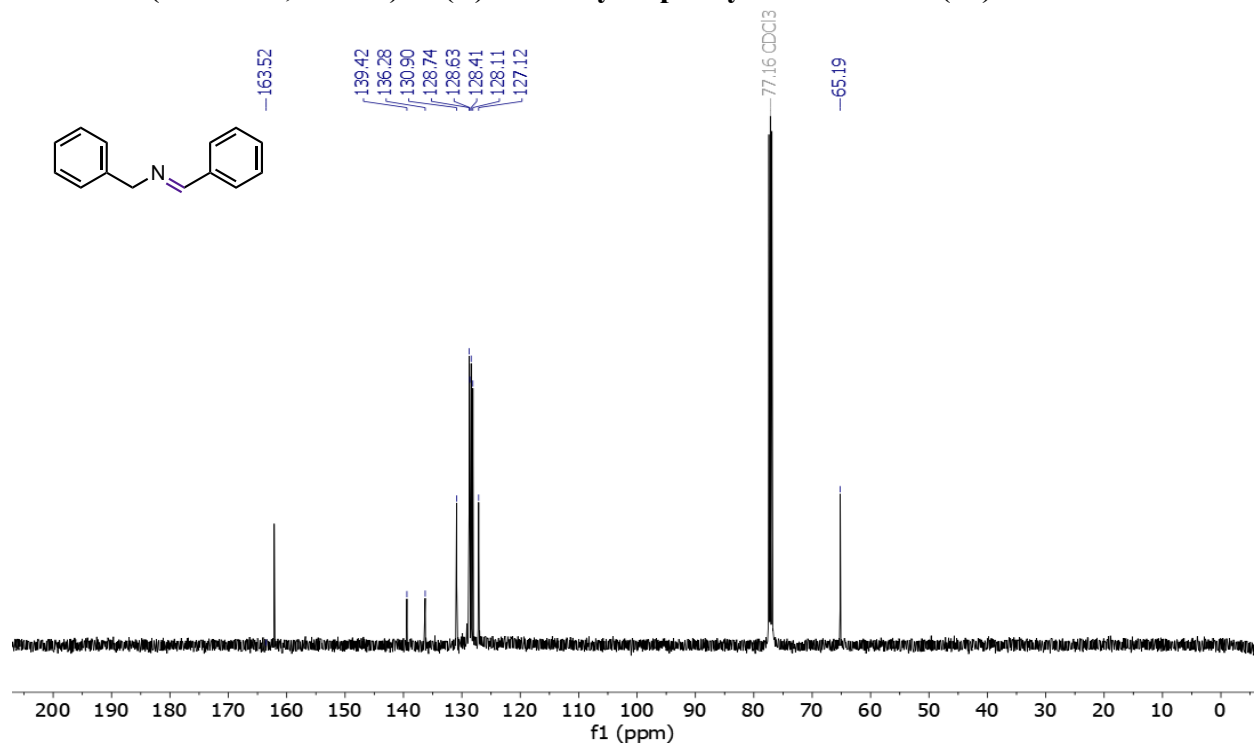
¹³C NMR (126 MHz, CDCl₃) of (8*R*,9*S*,10*S*,13*S*,14*S*)-10,13-dimethyltetradecahydro-3*H*-cyclopenta[*a*]phenanthrene-3,17(2*H*)-dione (3a)



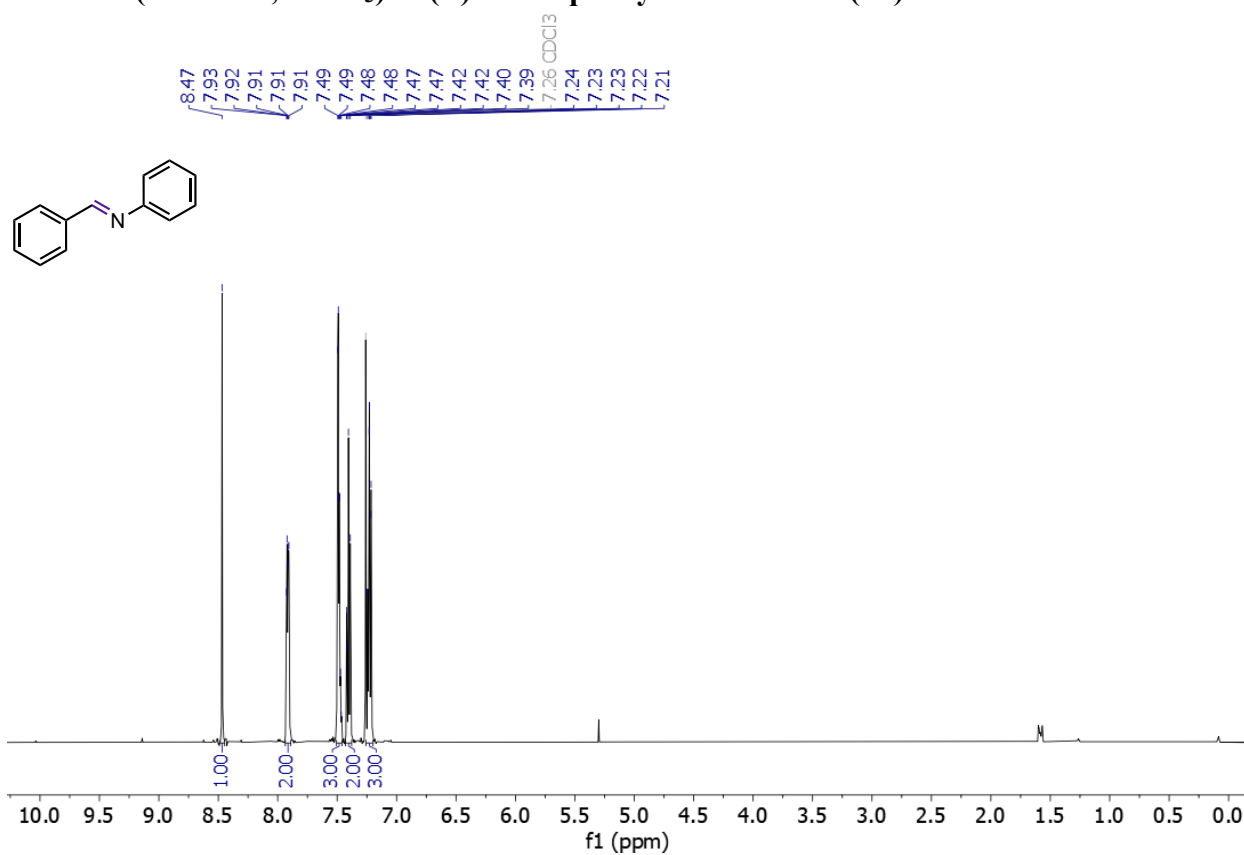
¹H NMR (500 MHz, CDCl₃) of (*E*)-*N*-Benzyl-1-phenylmethanimine (4a)



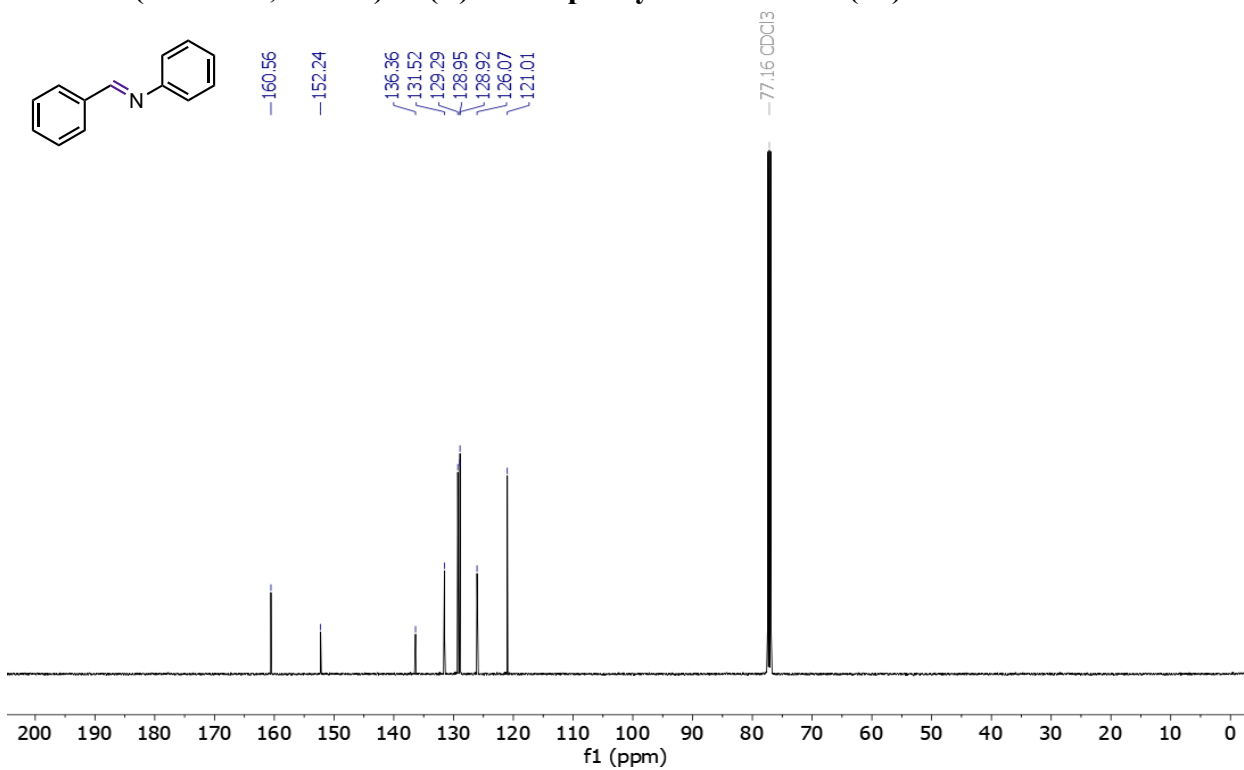
¹³C NMR (126 MHz, CDCl₃) of (*E*)-*N*-Benzyl-1-phenylmethanimine (4a)



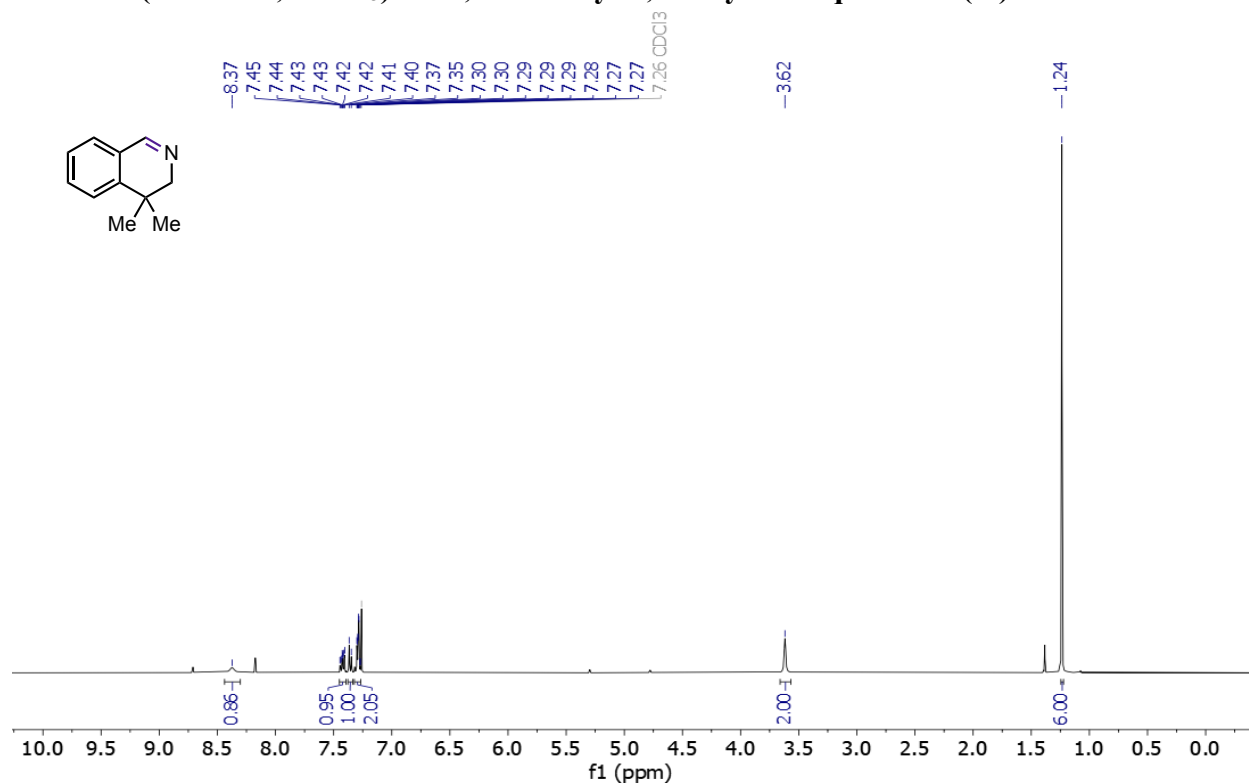
¹H NMR (500 MHz, CDCl₃) of (*E*)-*N*-1-diphenylmethanimine (4b)



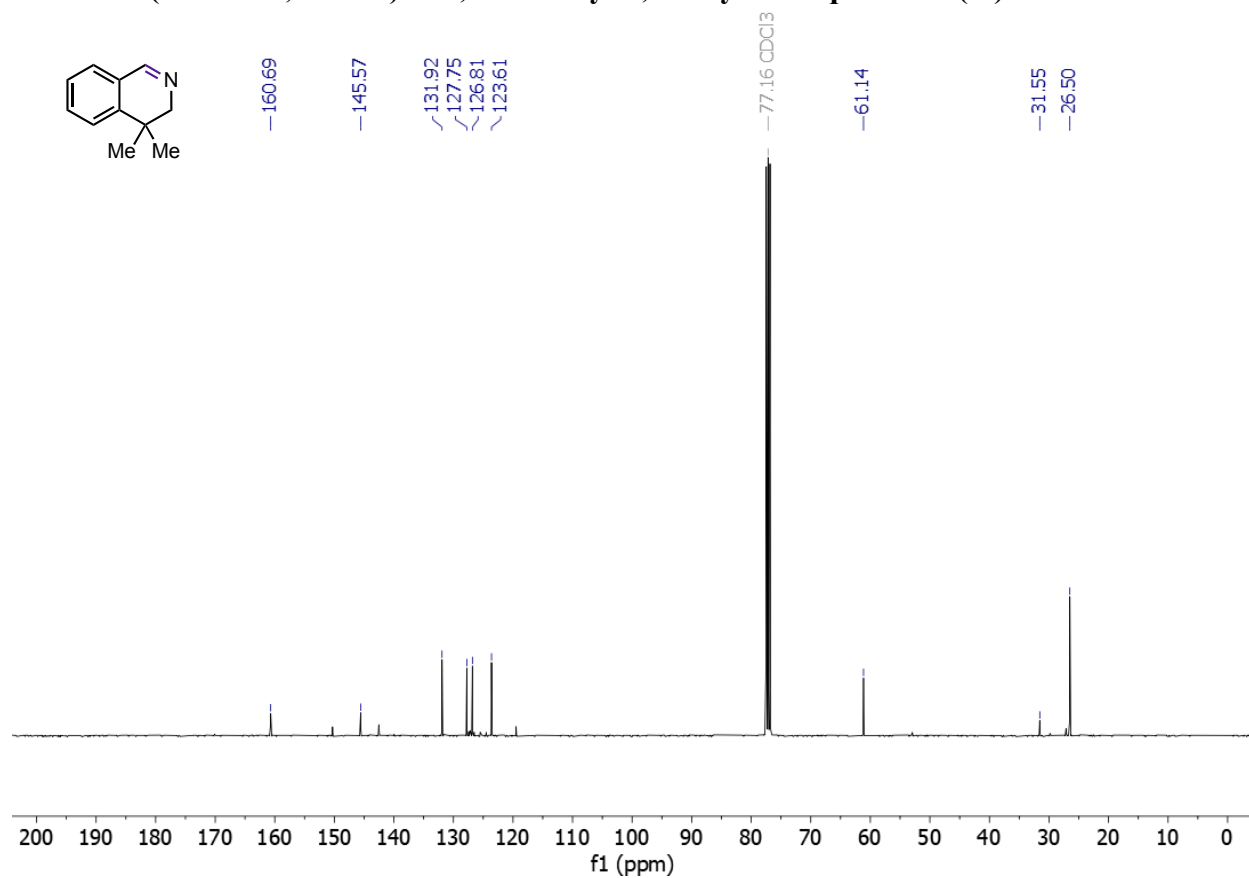
¹³C NMR (126 MHz, CDCl₃) of (*E*)-*N*-1-diphenylmethanimine (4b)



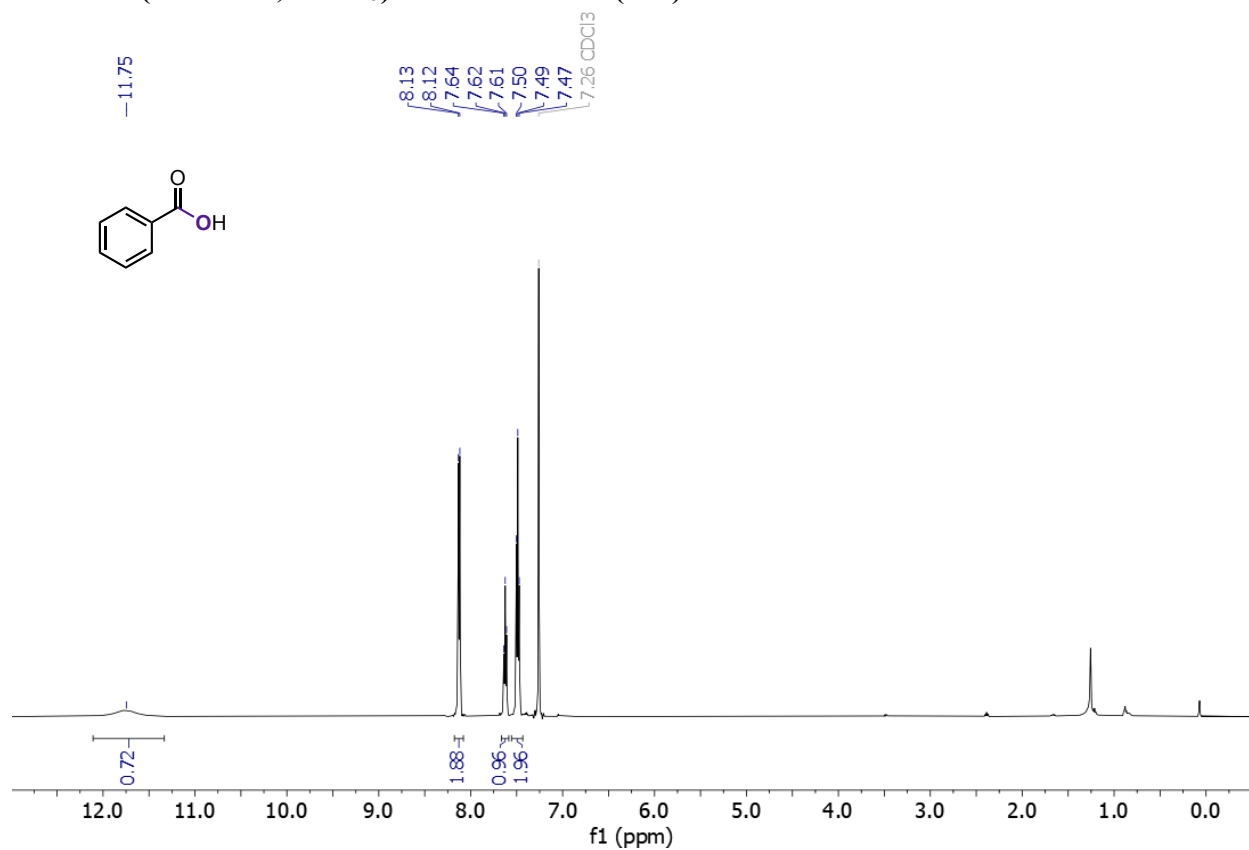
¹H NMR (500 MHz, CDCl₃) for 4,4-dimethyl-3,4-dihydroisoquinoline (4f)



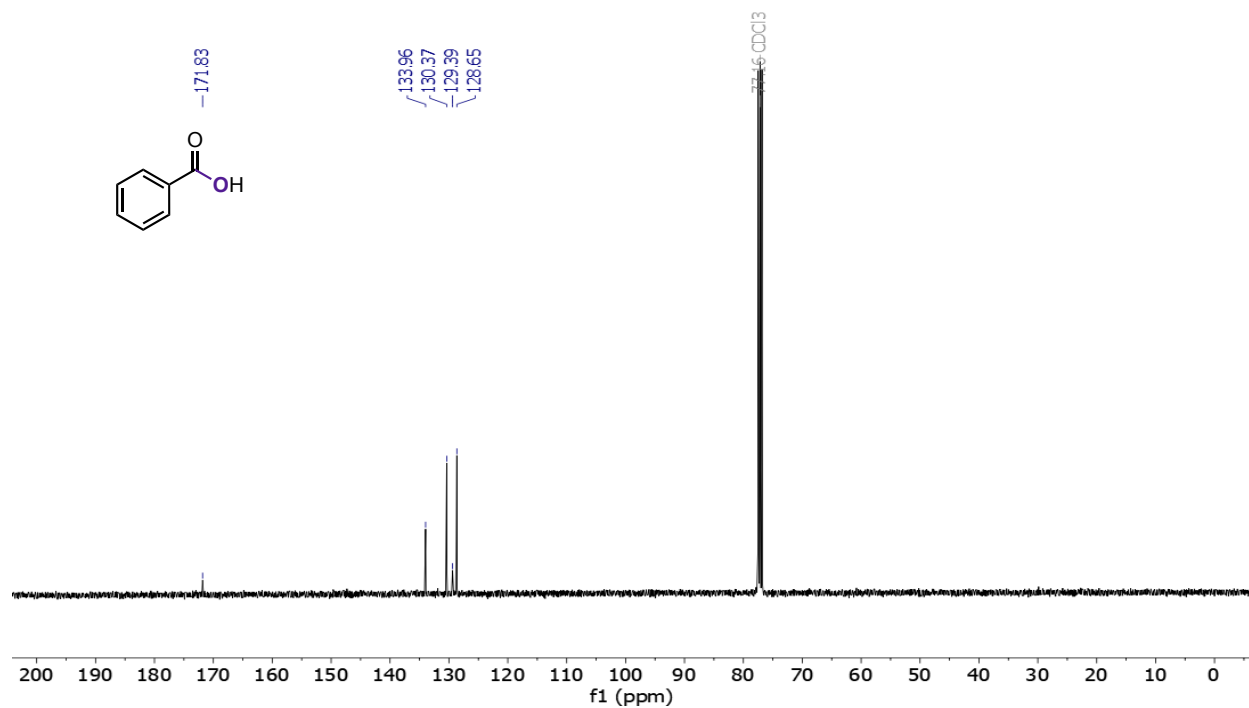
¹³C NMR (101 MHz, CDCl₃) of 4,4-dimethyl-3,4-dihydroisoquinoline (4f)



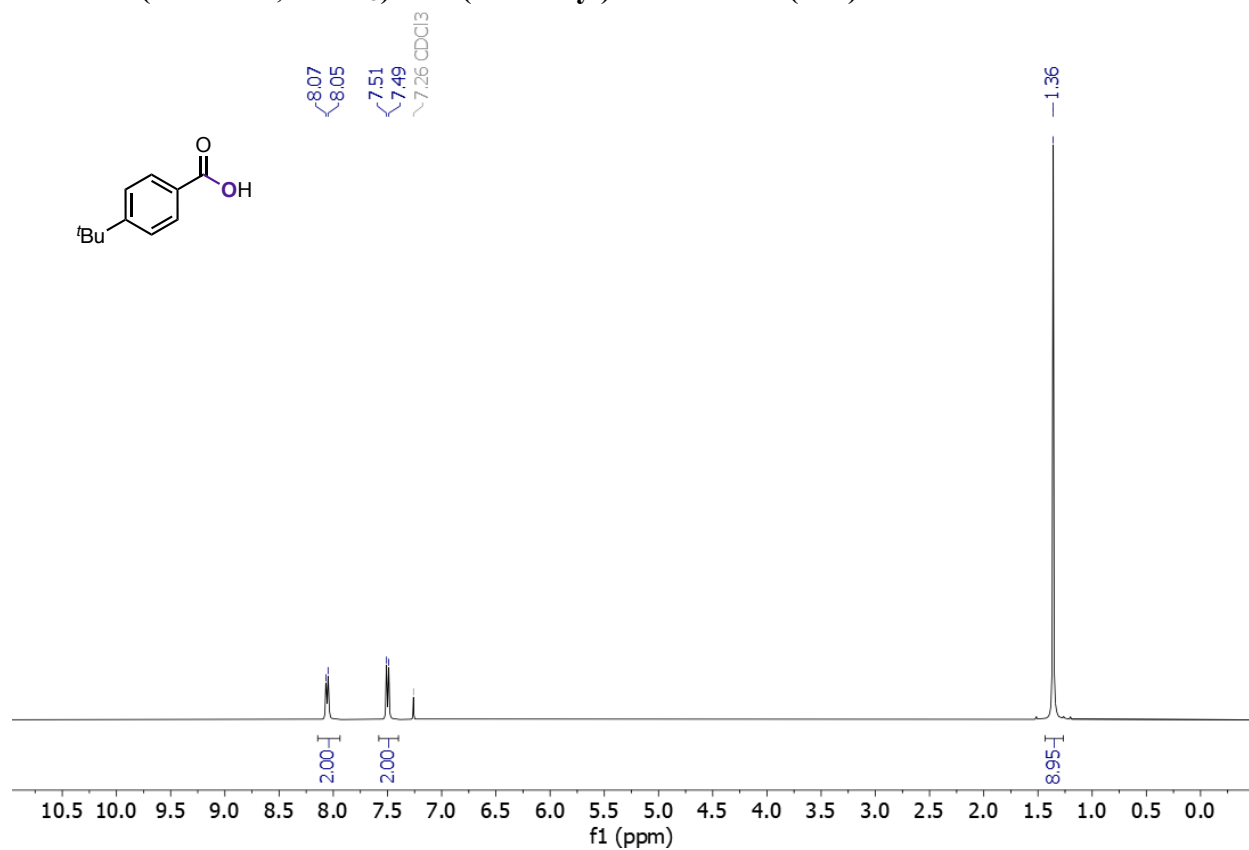
¹H NMR (500 MHz, CDCl₃) of Benzoic acid (10a)



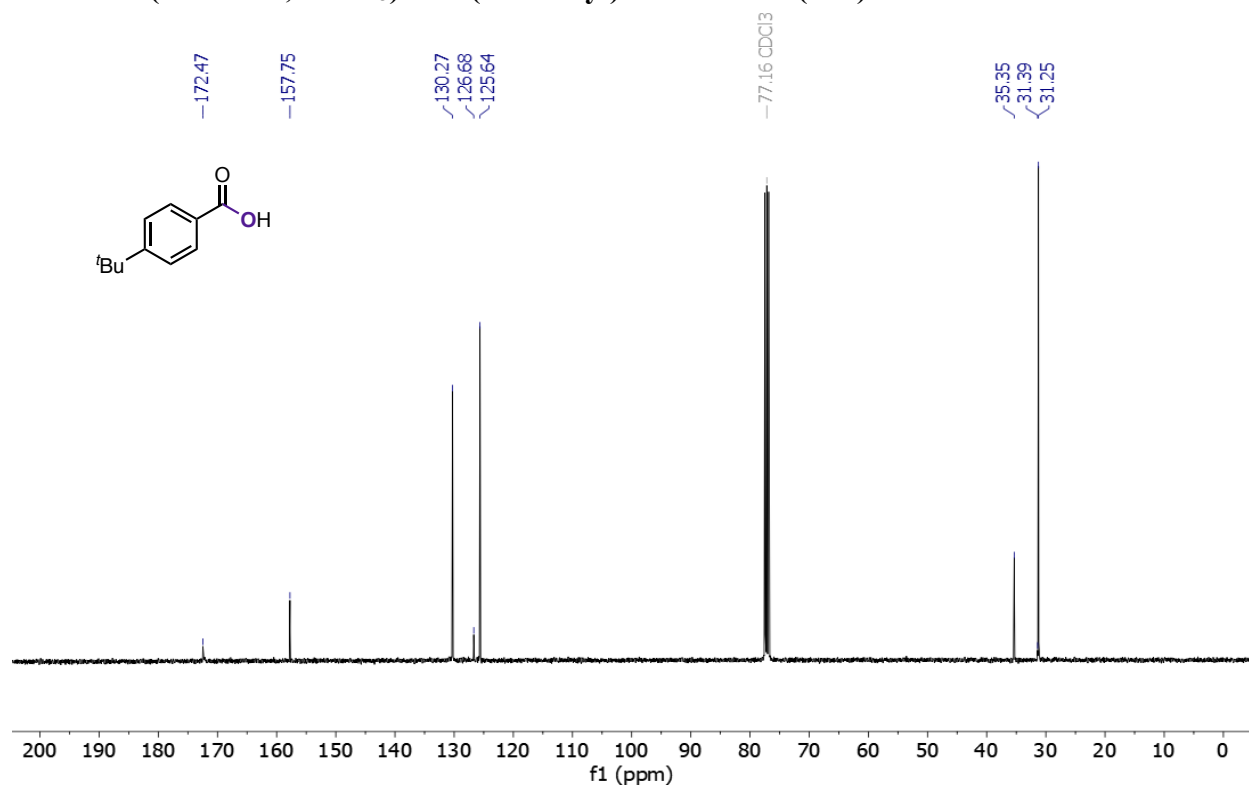
¹³C NMR (101 MHz, CDCl₃) of Benzoic acid (10a)



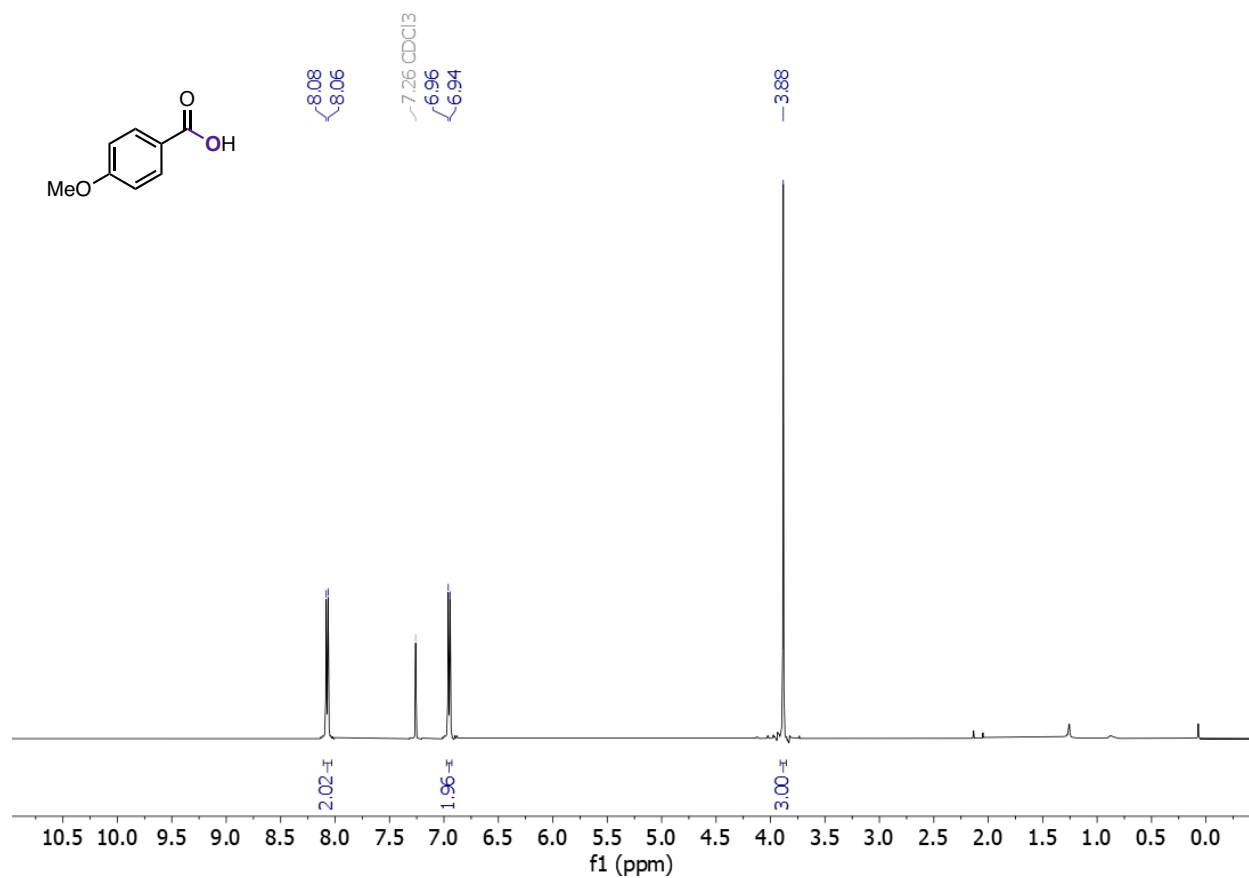
¹H NMR (400 MHz, CDCl₃) of 4-(*tert*-Butyl)benzoic acid (10b)



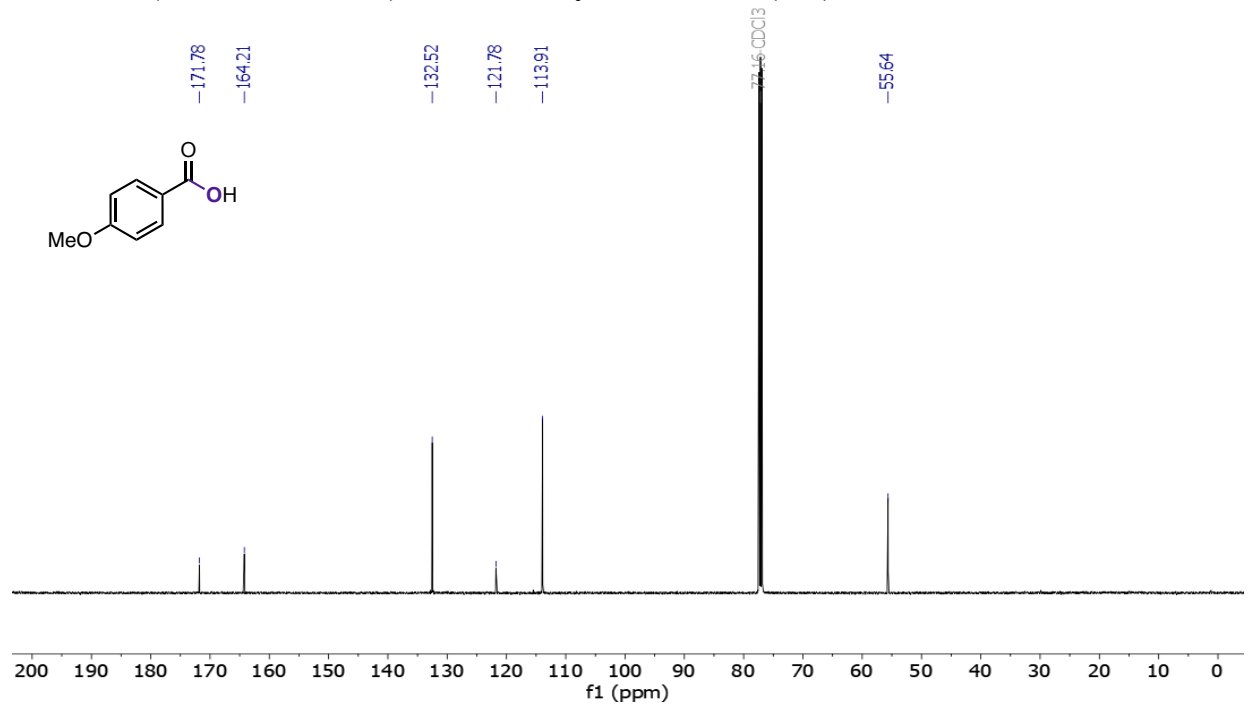
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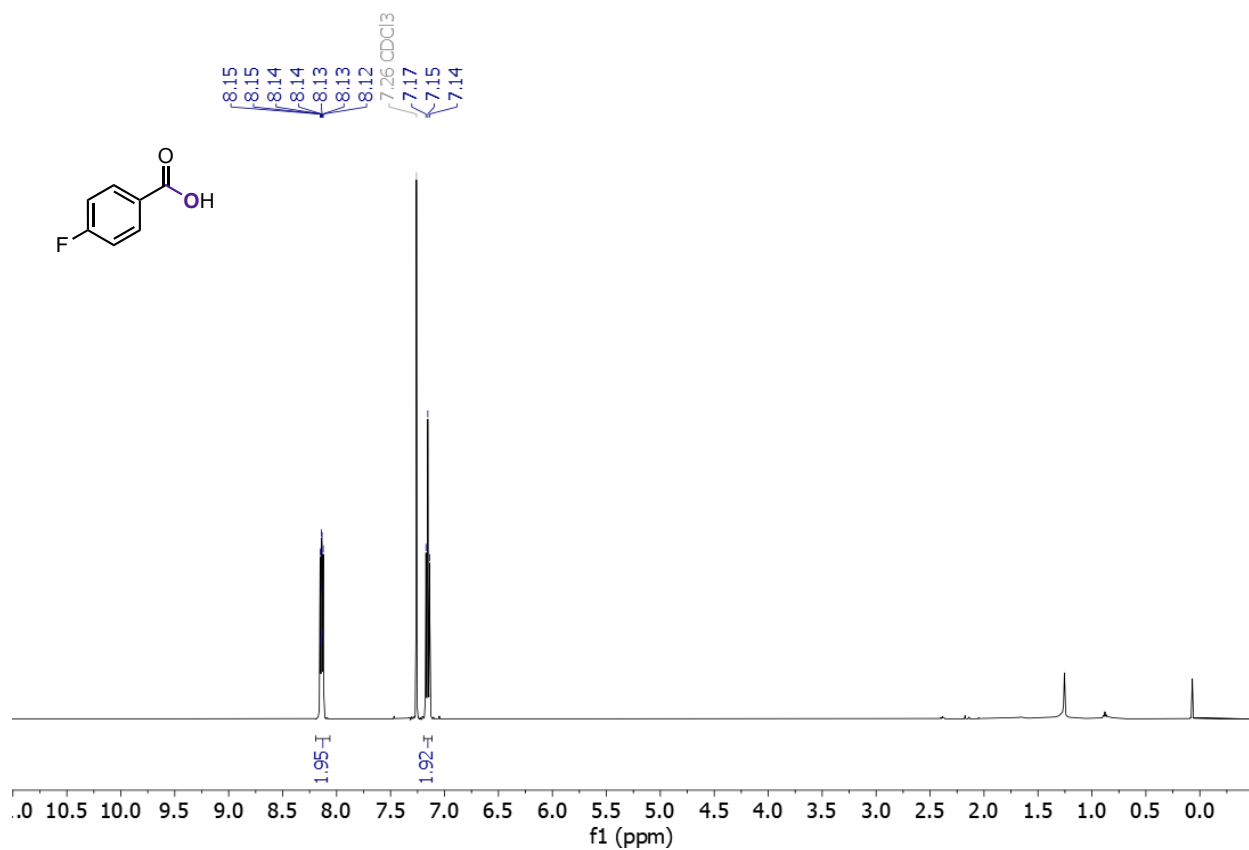
¹H NMR (500 MHz, CDCl₃) of 4-Methoxybenzoic acid (10c)



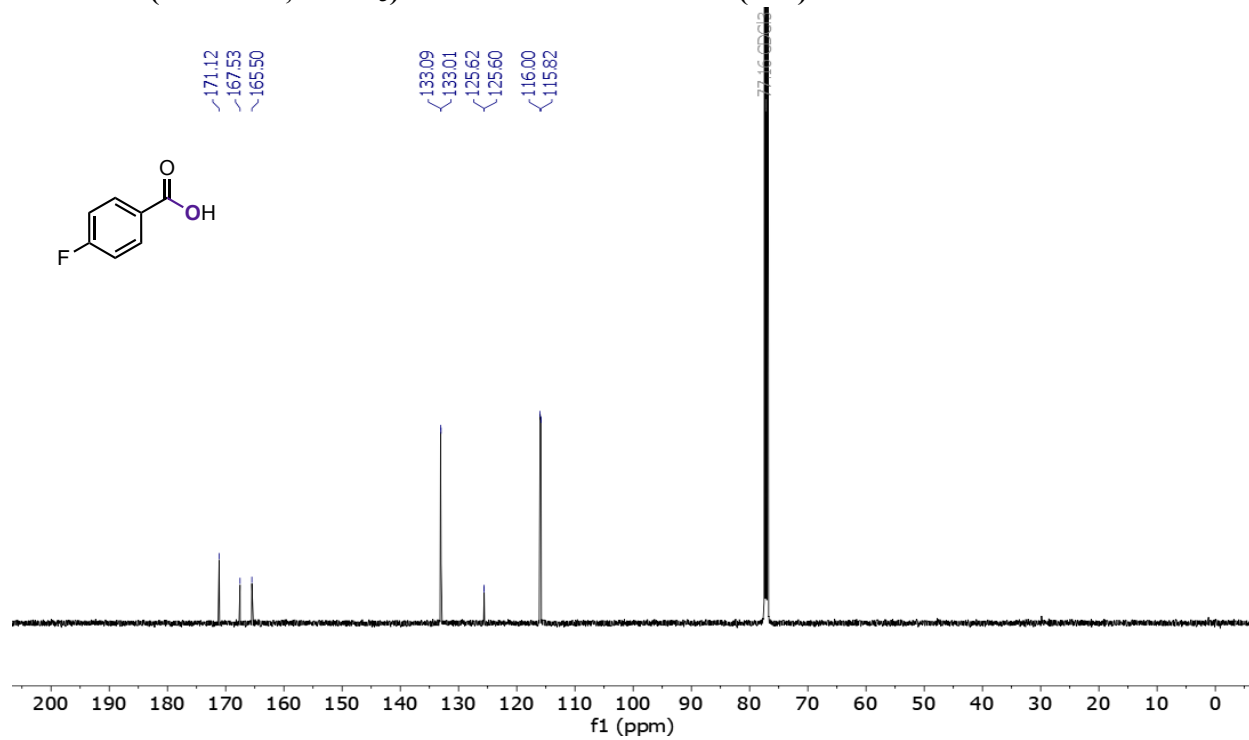
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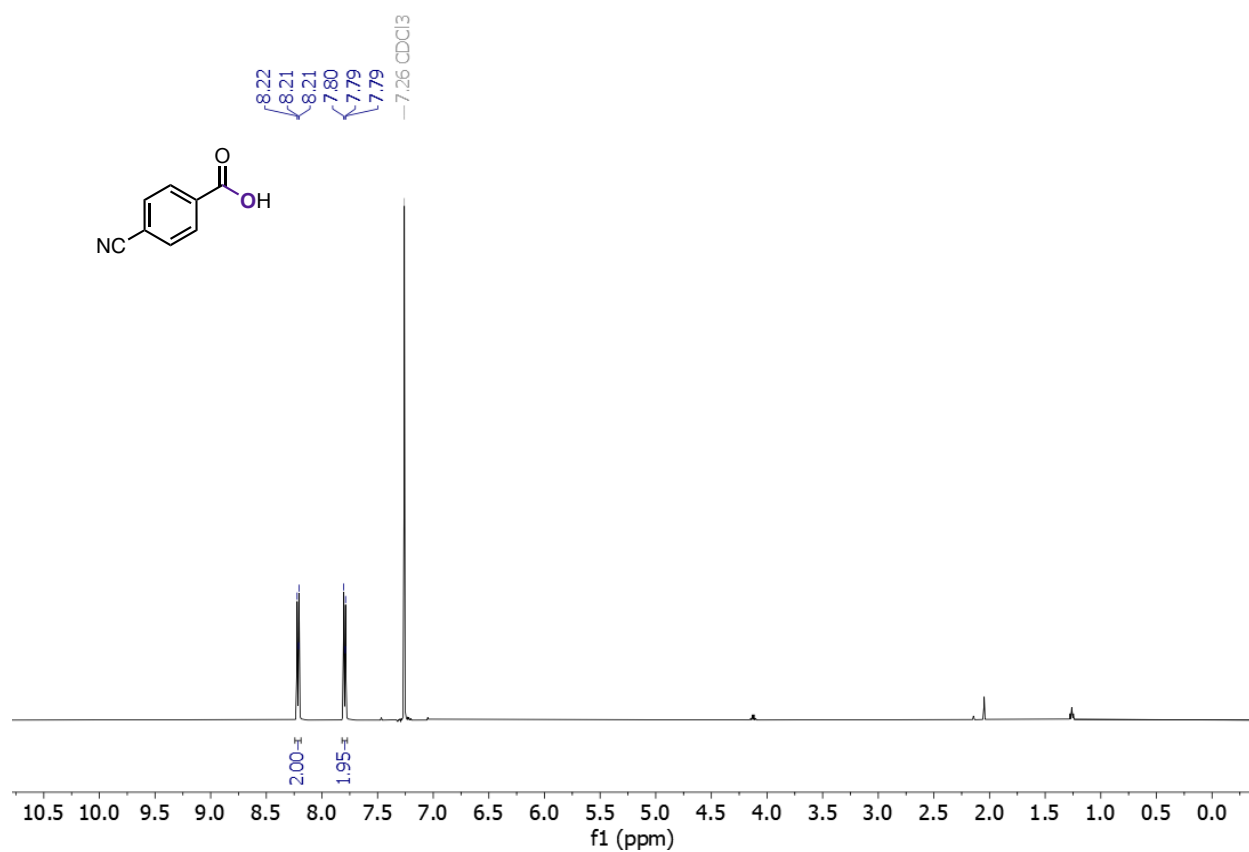
¹H NMR (500 MHz, CDCl₃) of 4-Fluorobenzoic acid (10d)



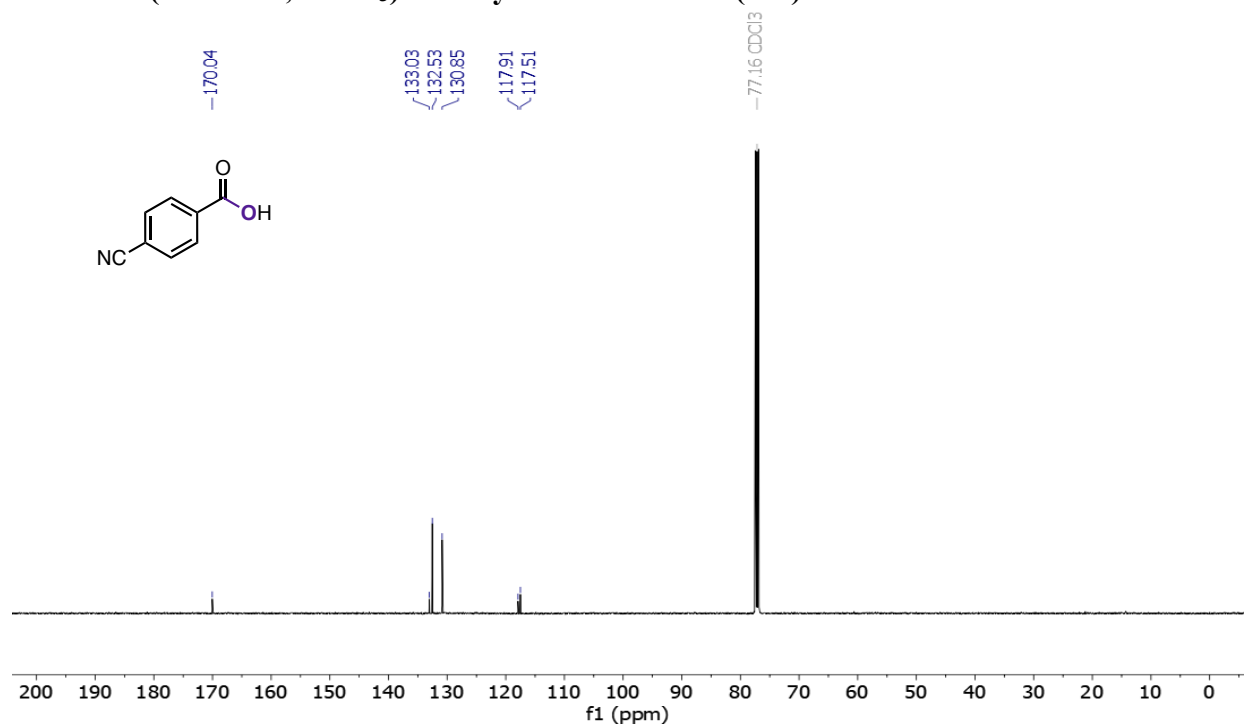
¹³C NMR (126 MHz, CDCl₃) of 4-Fluorobenzoic acid (10d)



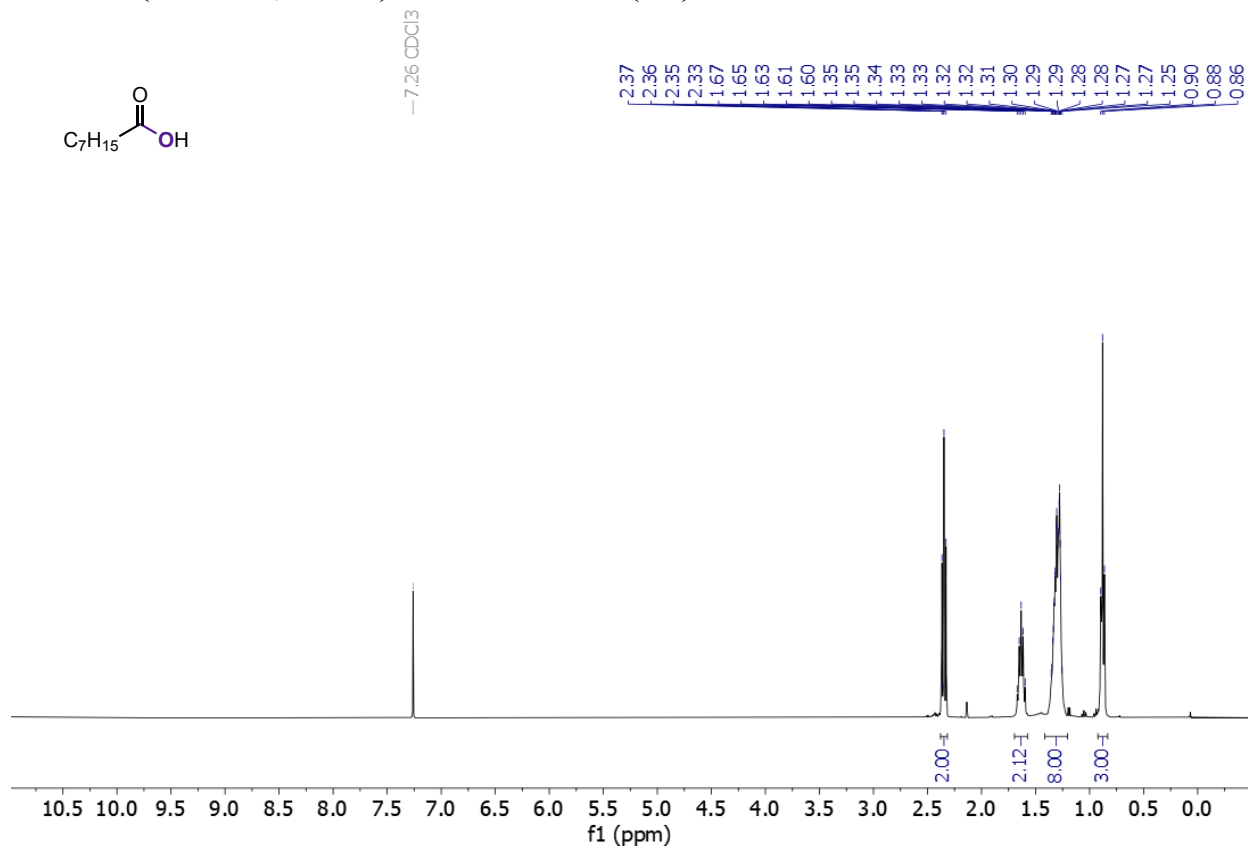
¹H NMR (500 MHz, CDCl₃) of 4-Cyanobenzoic acid (10e)



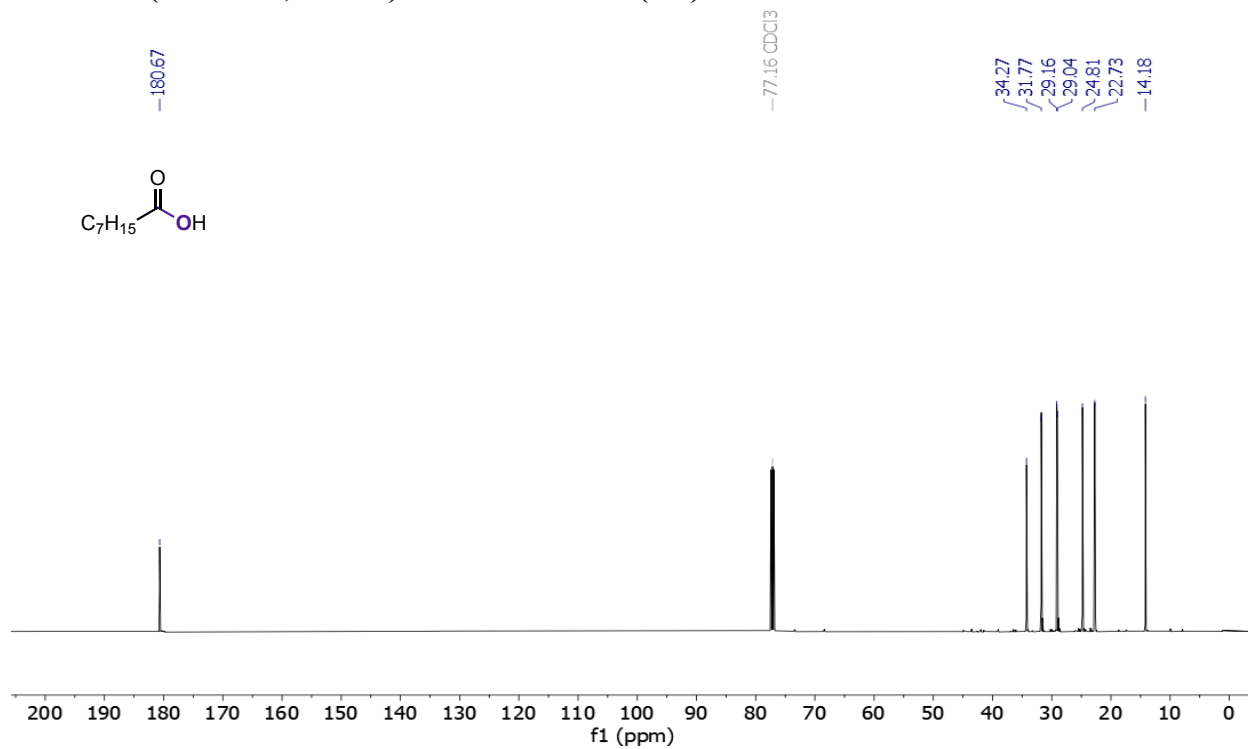
¹³C NMR (126 MHz, CDCl₃) of 4-Cyanobenzoic acid (10e)



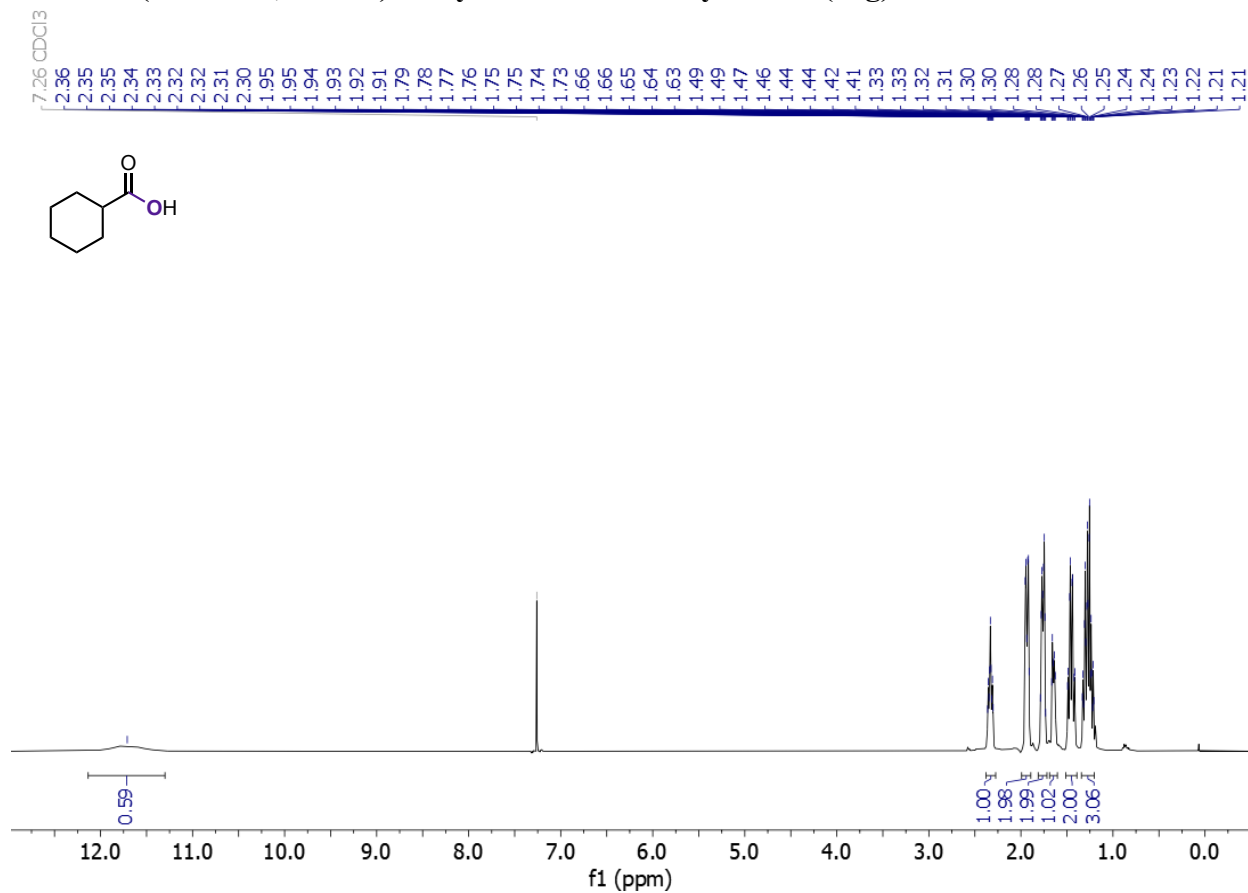
^1H NMR (400 MHz, CDCl_3) of Octanoic acid (10f)



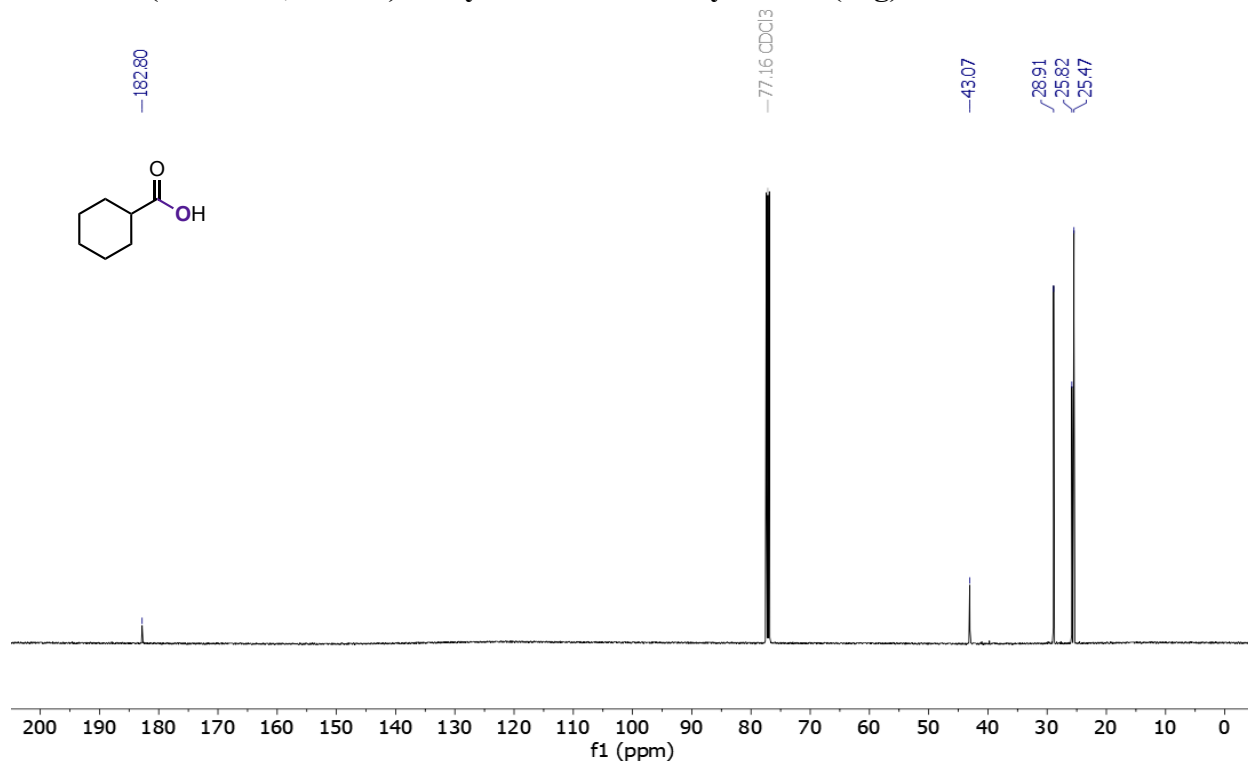
^{13}C NMR (126 MHz, CDCl_3) of Octanoic acid (10f)



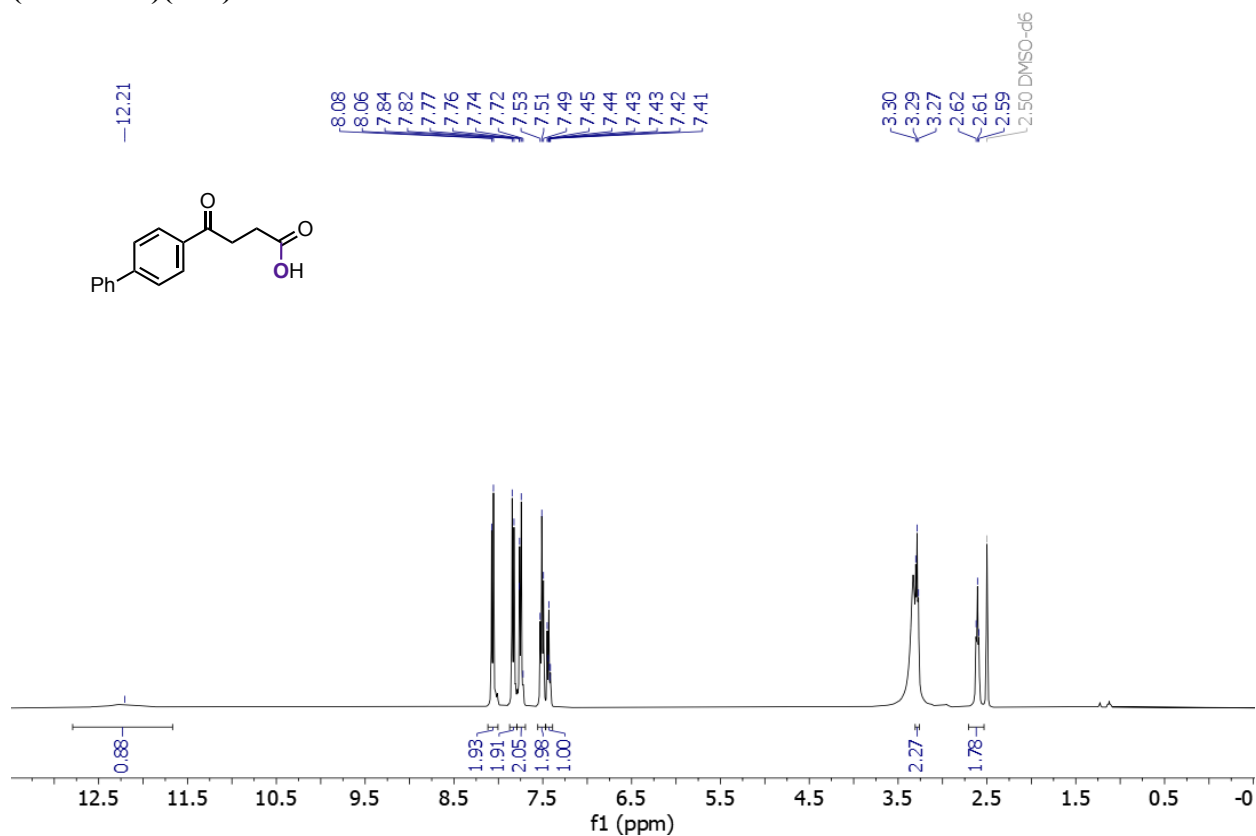
¹H NMR (500 MHz, CDCl₃) of Cyclohexanecarboxylic acid (10g)



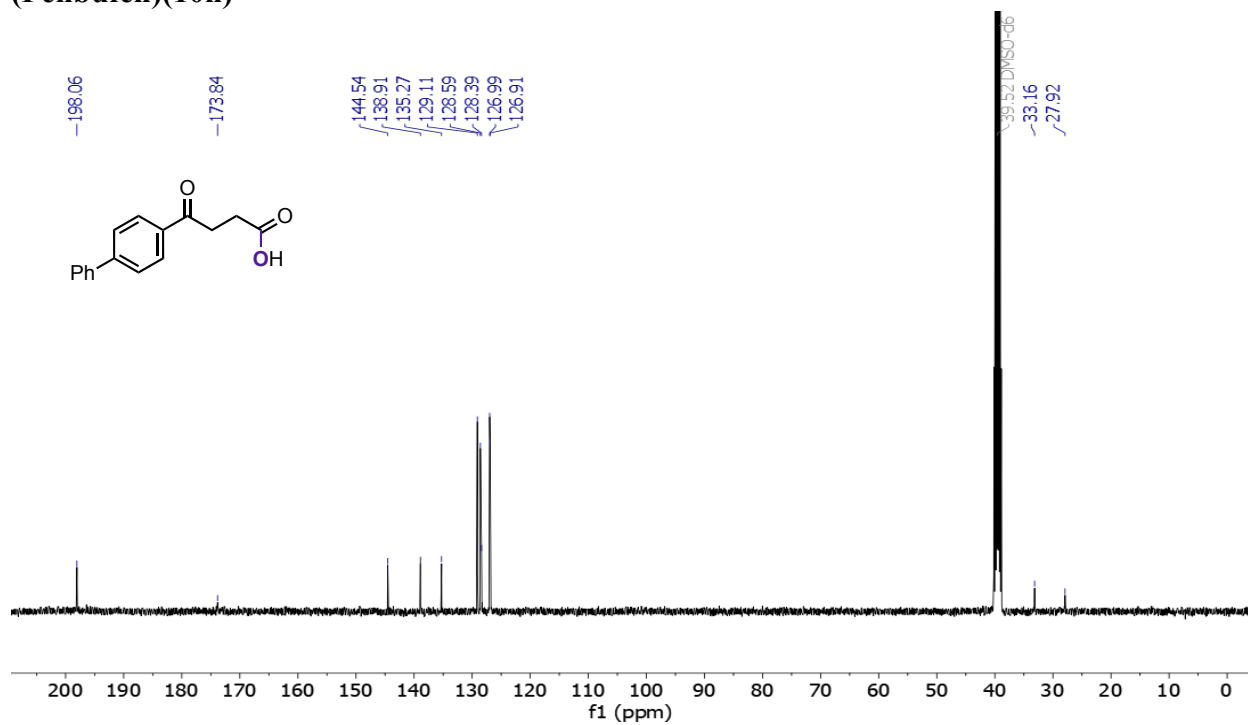
¹³C NMR (126 MHz, CDCl₃) of Cyclohexanecarboxylic acid (10g)



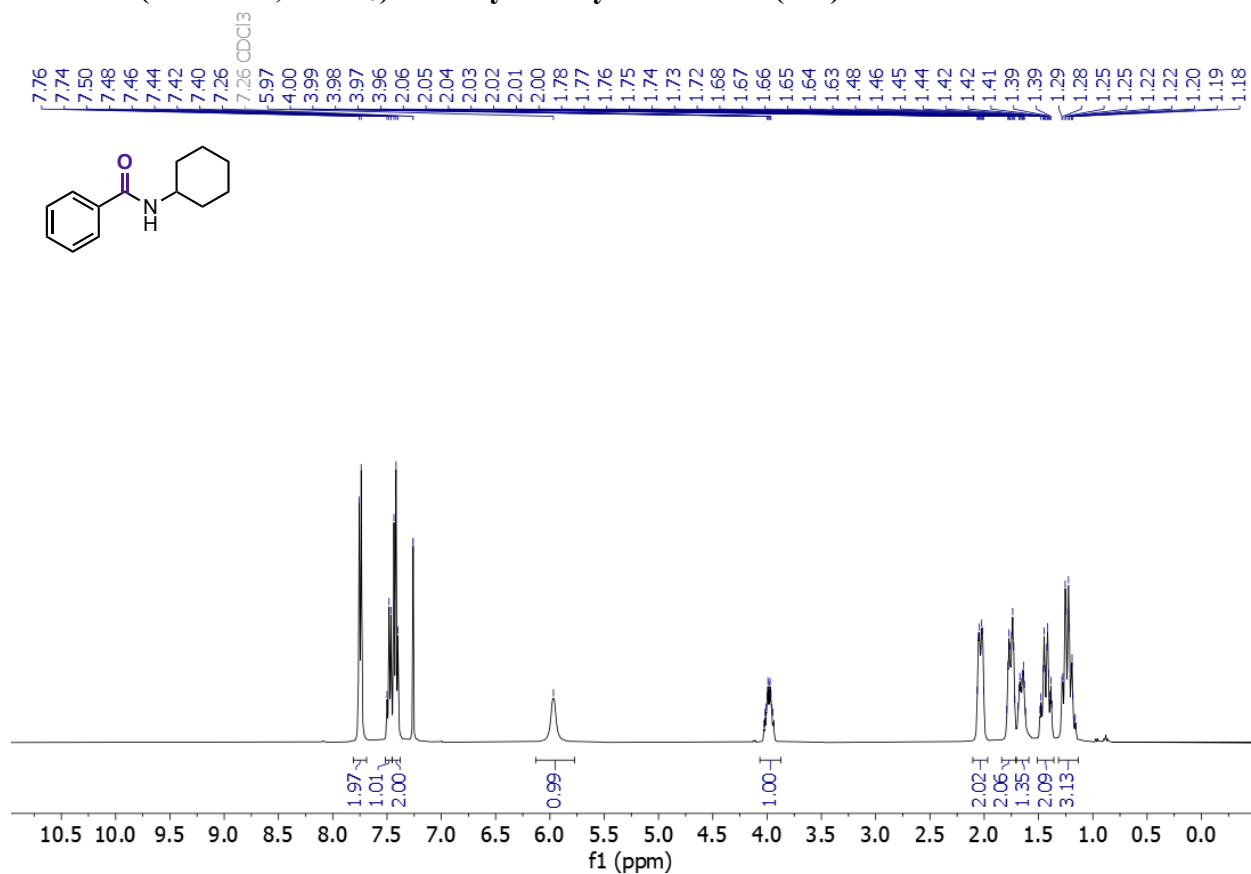
¹H NMR (400 MHz, DMSO-*d*₆) of 4-([1,1'-Biphenyl]-4-yl)-4-oxobutanoic acid (Fenbufen)(10h)



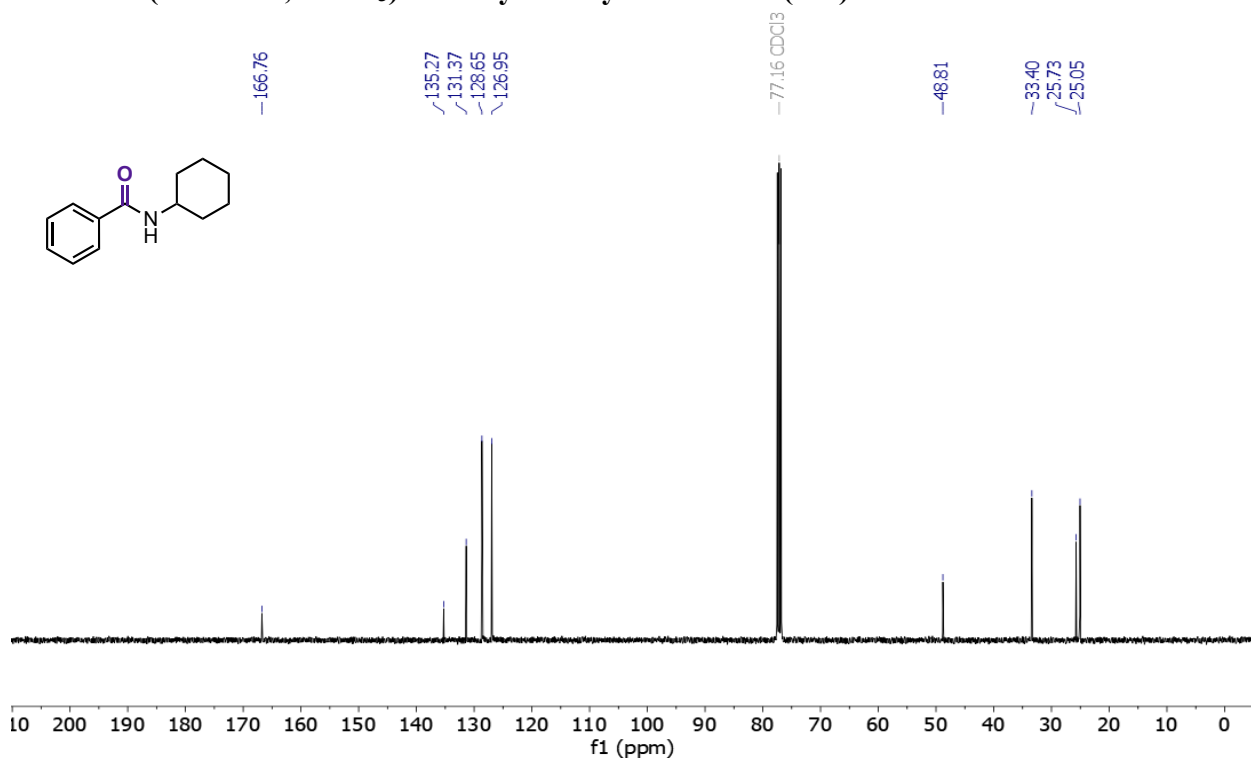
¹³C NMR (101 MHz, DMSO-*d*₆) of 4-([1,1'-Biphenyl]-4-yl)-4-oxobutanoic acid (Fenbufen)(10h)



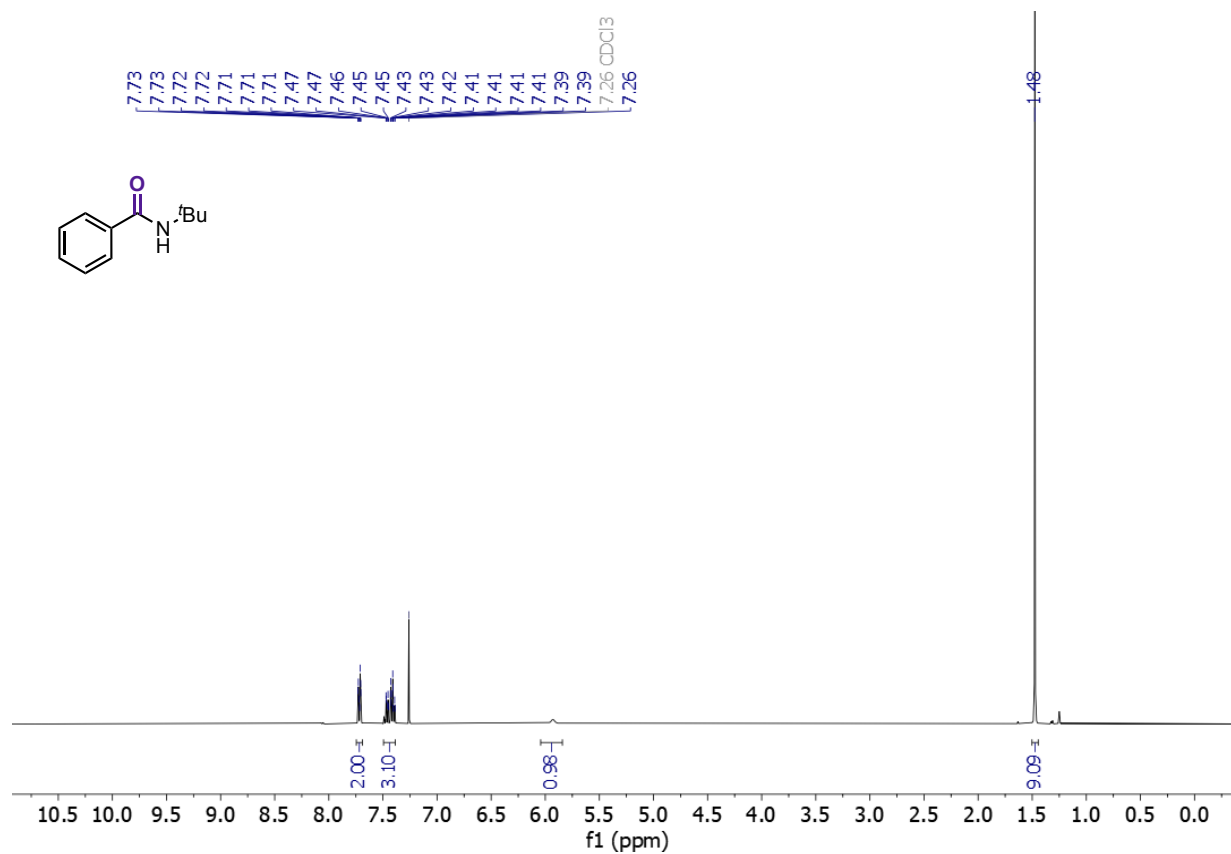
¹H NMR (400 MHz, CDCl₃) of *N*-Cyclohexylbenzamide (11a)



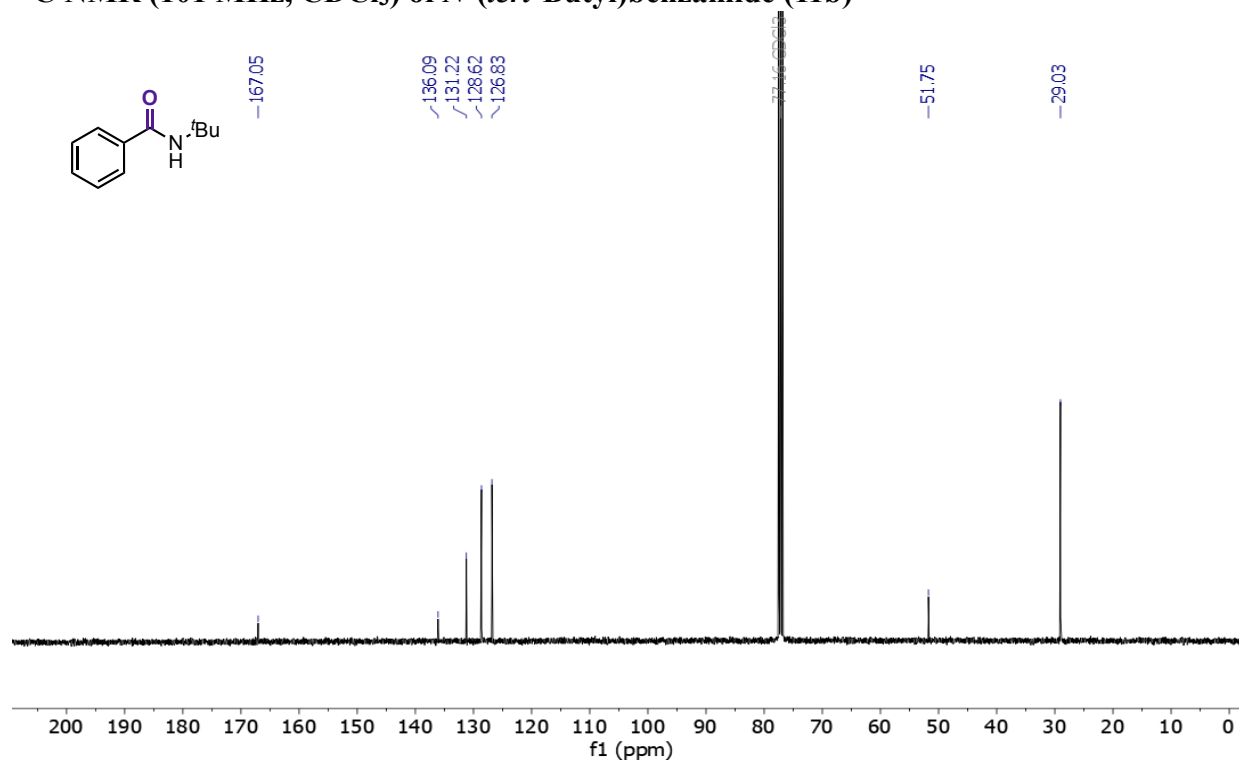
¹³C NMR (101 MHz, CDCl₃) of *N*-Cyclohexylbenzamide (11a)



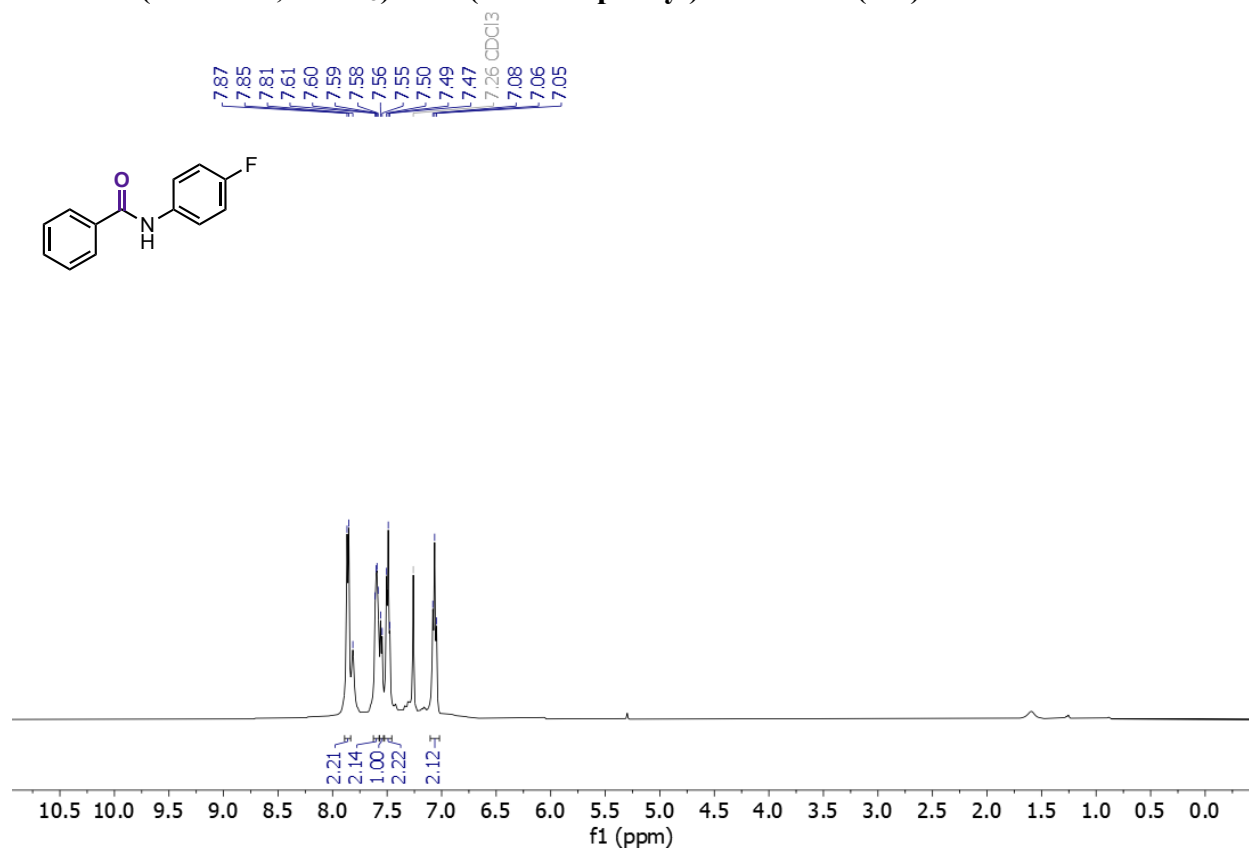
¹H NMR (400 MHz, CDCl₃) of *N*-(*tert*-Butyl)benzamide (11b)



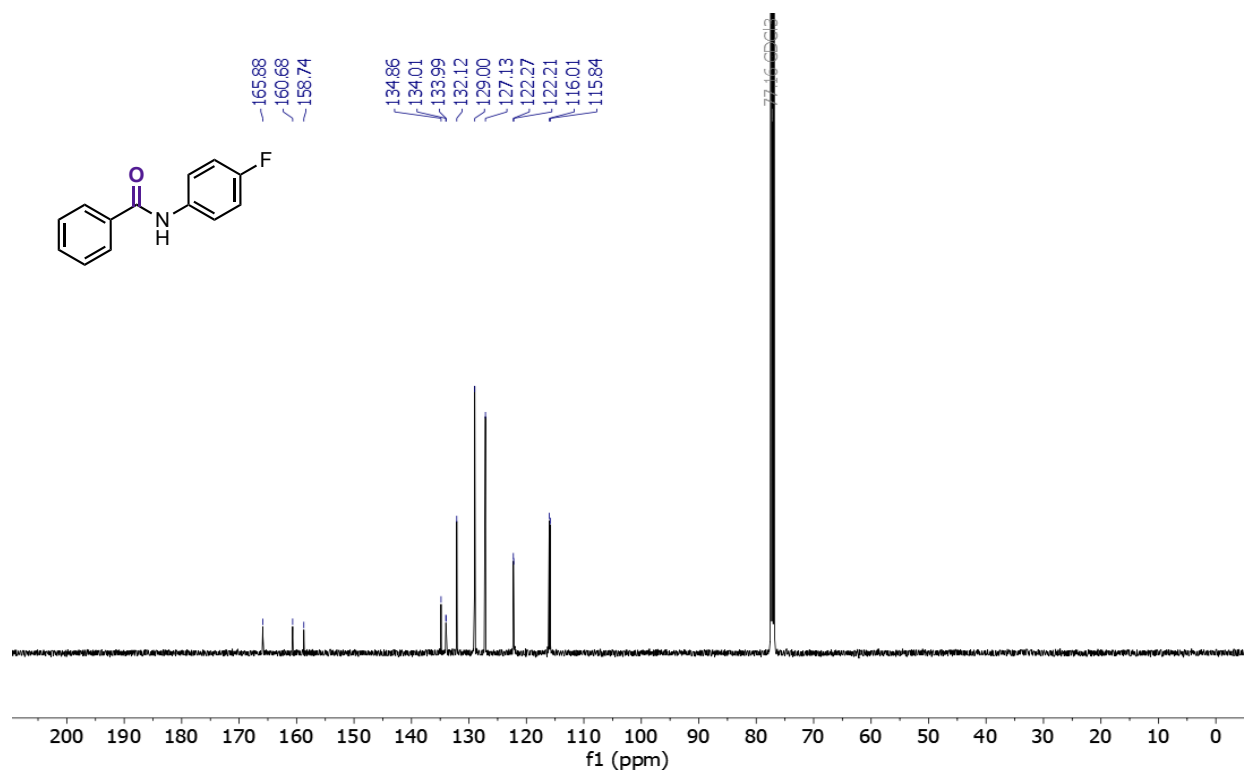
¹³C NMR (101 MHz, CDCl₃) of *N*-(*tert*-Butyl)benzamide (11b)



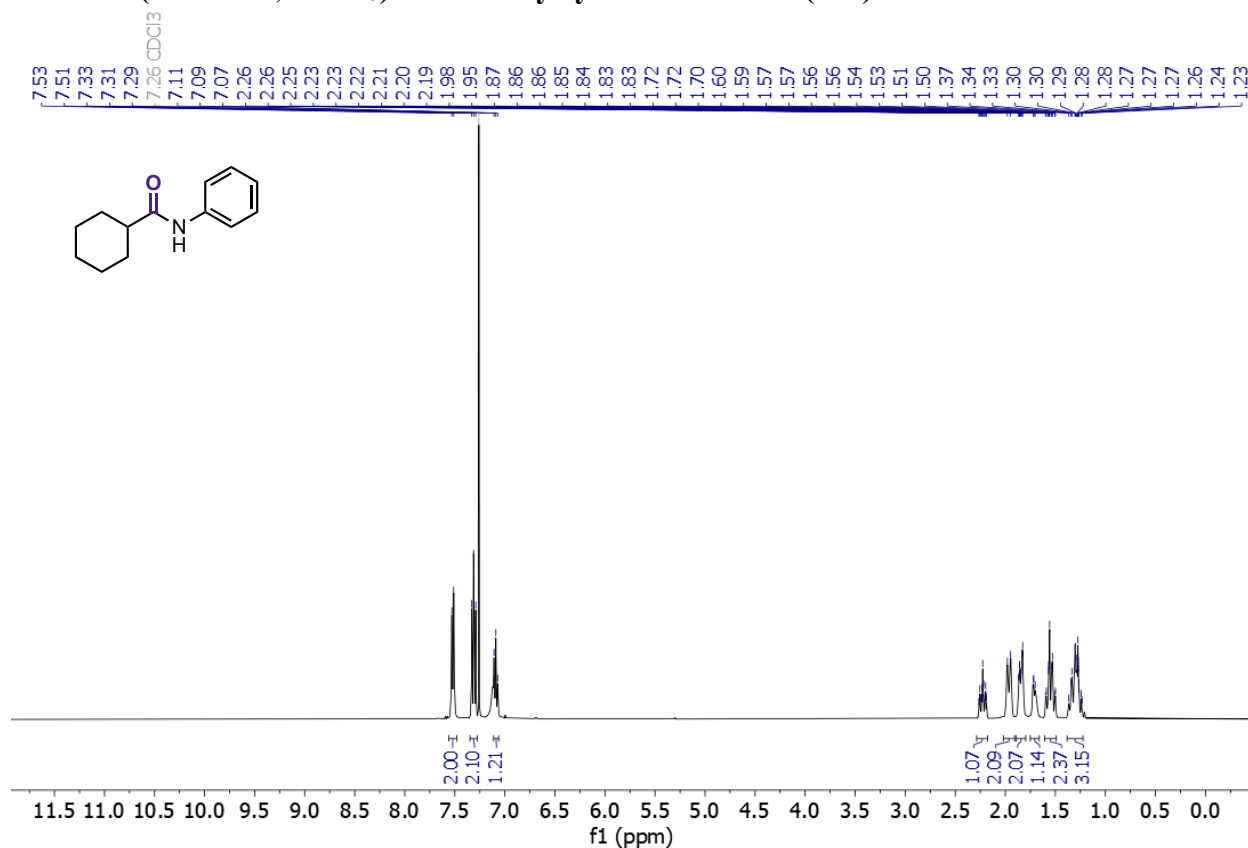
¹H NMR (500 MHz, CDCl₃) of *N*-(4-Fluorophenyl)benzamide (11c)



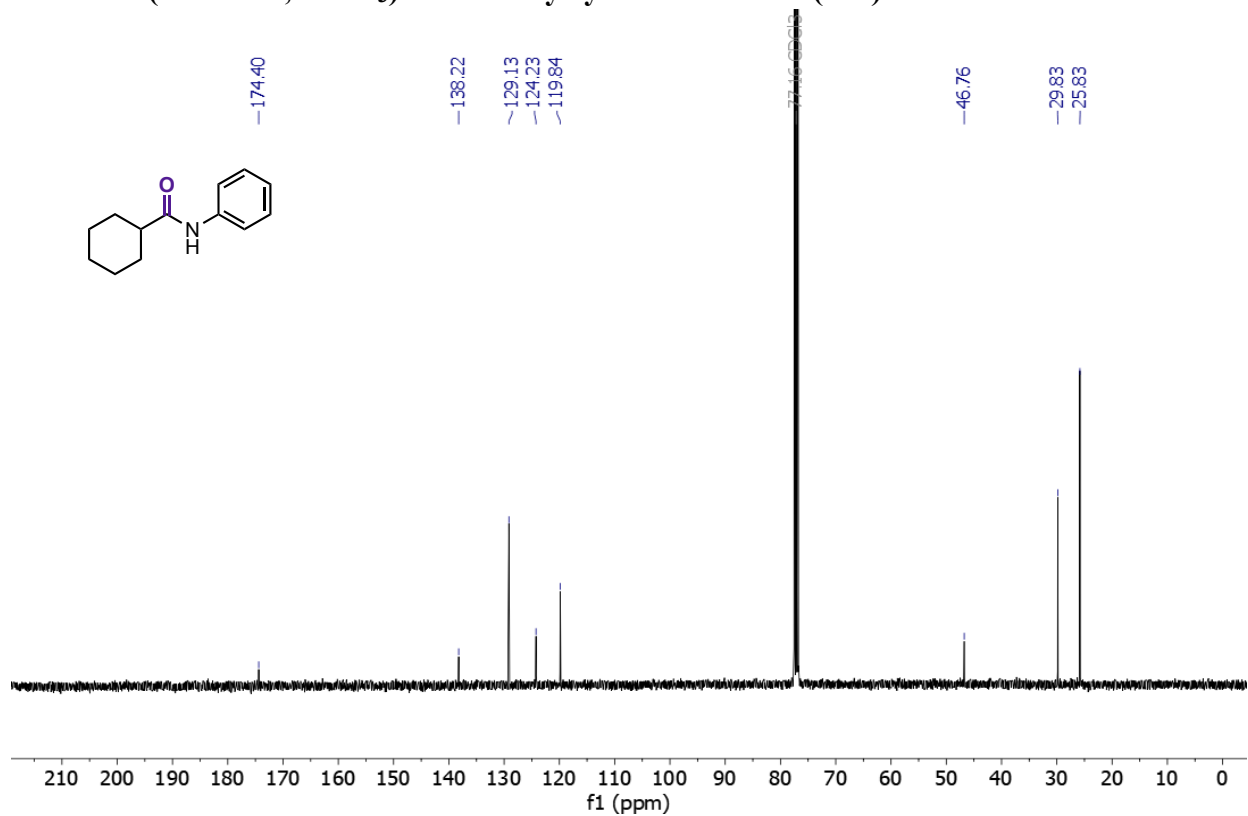
¹³C NMR (126 MHz, CDCl₃) of *N*-(4-Fluorophenyl)benzamide (11c)



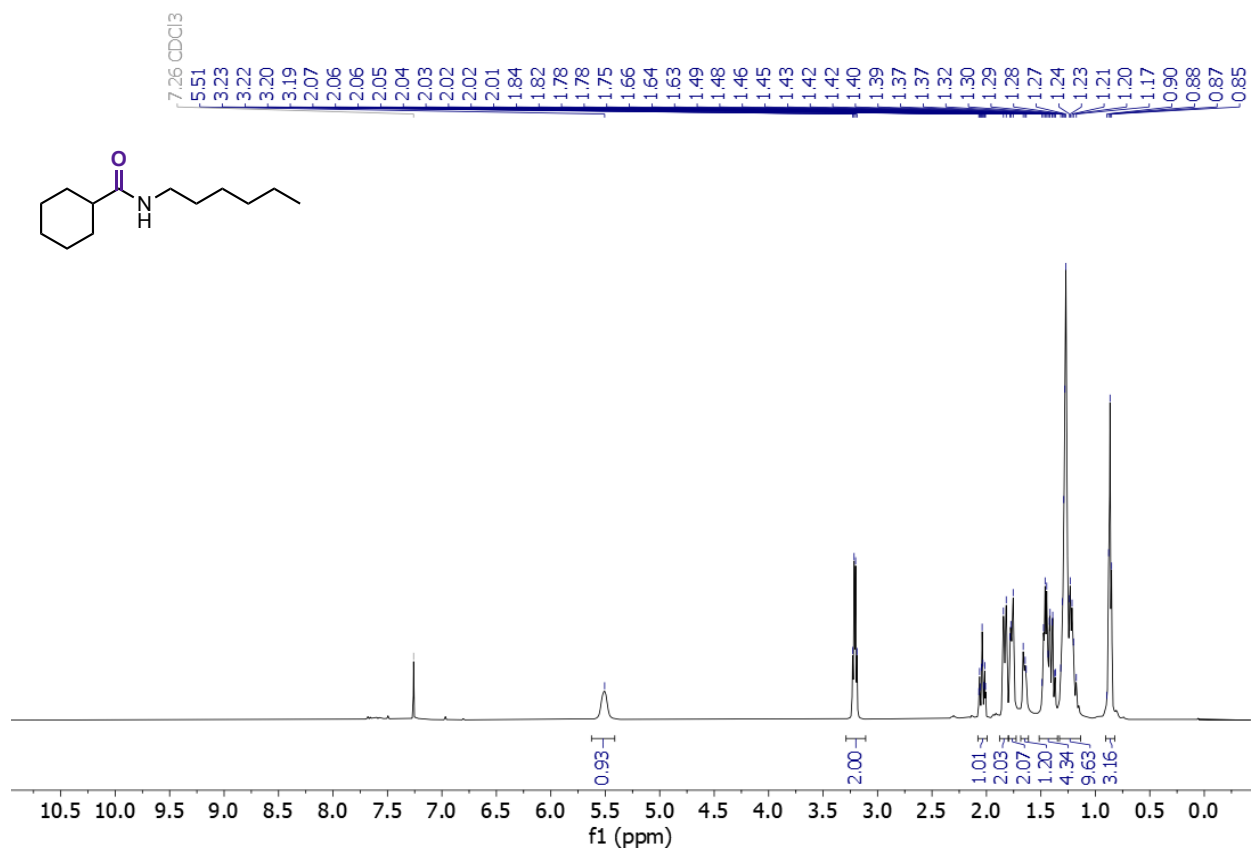
¹H NMR (400 MHz, CDCl₃) of *N*-Phenylcyclohexanamide (11d)



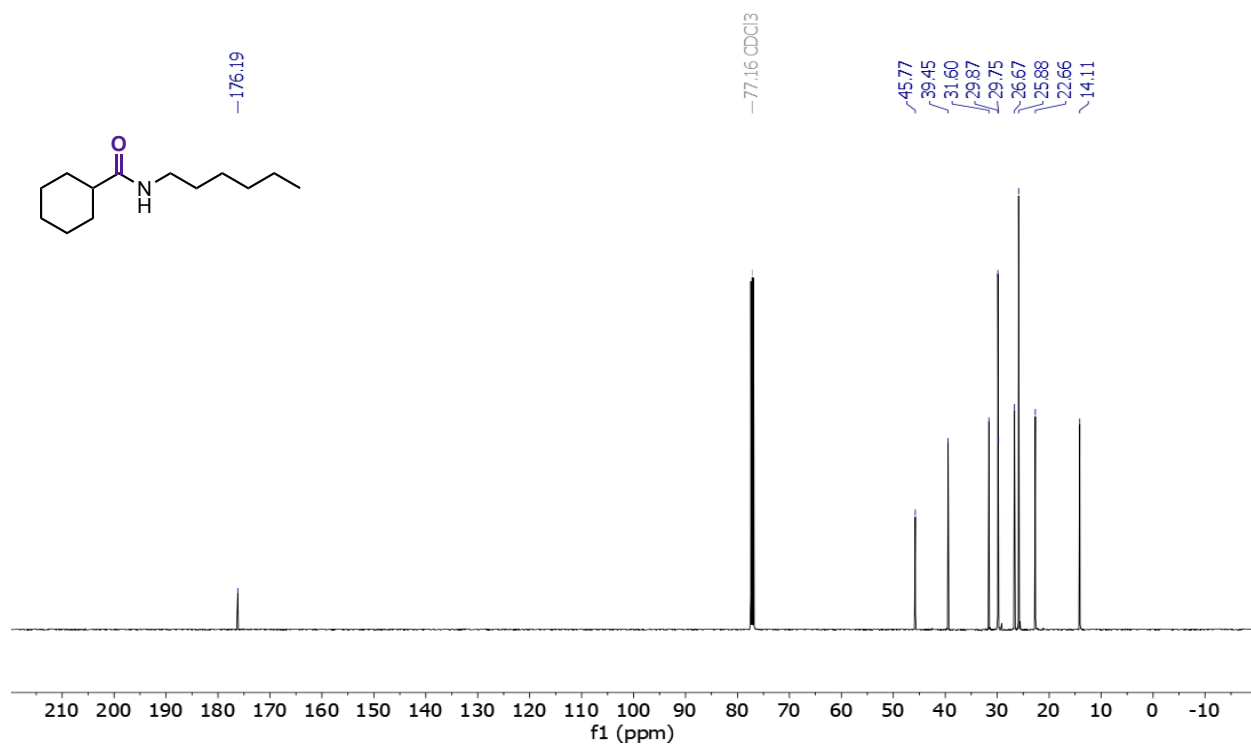
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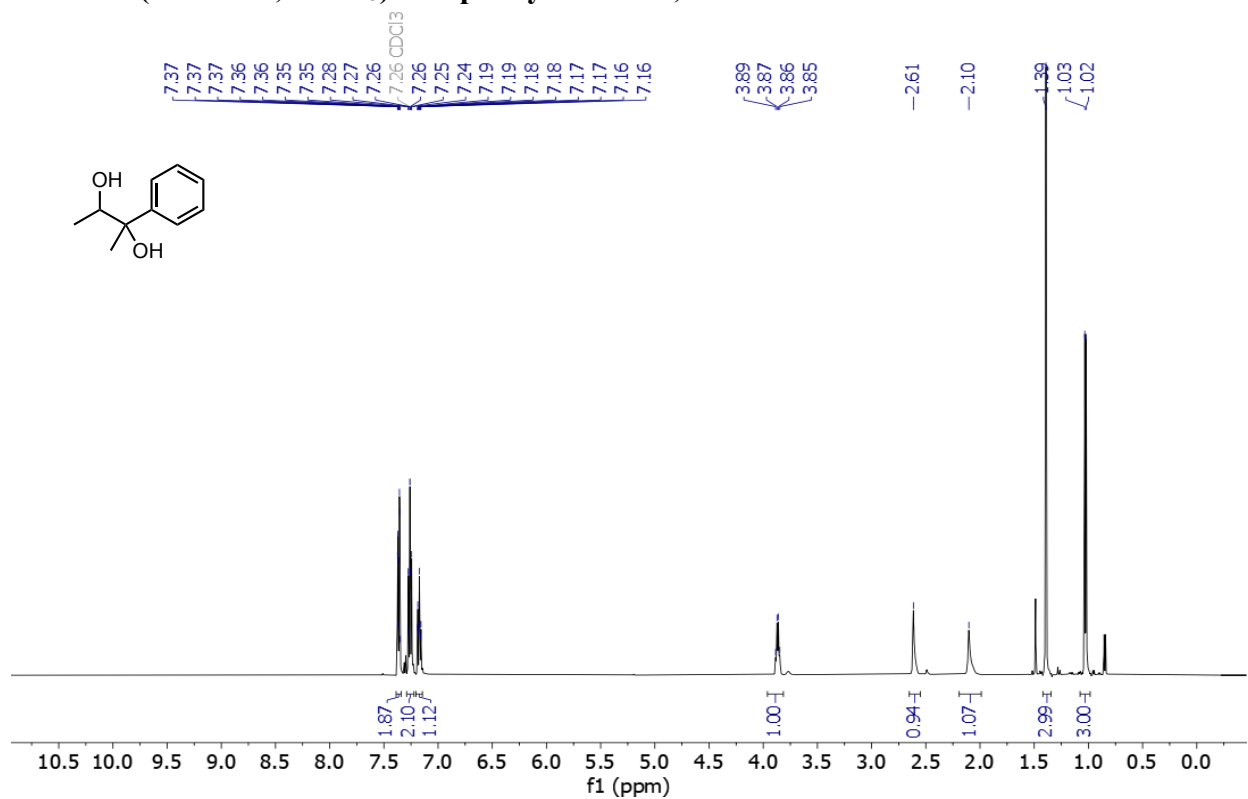
¹H NMR (500 MHz, CDCl₃) of N-Hexylcyclohexanecarboxamide (11e)



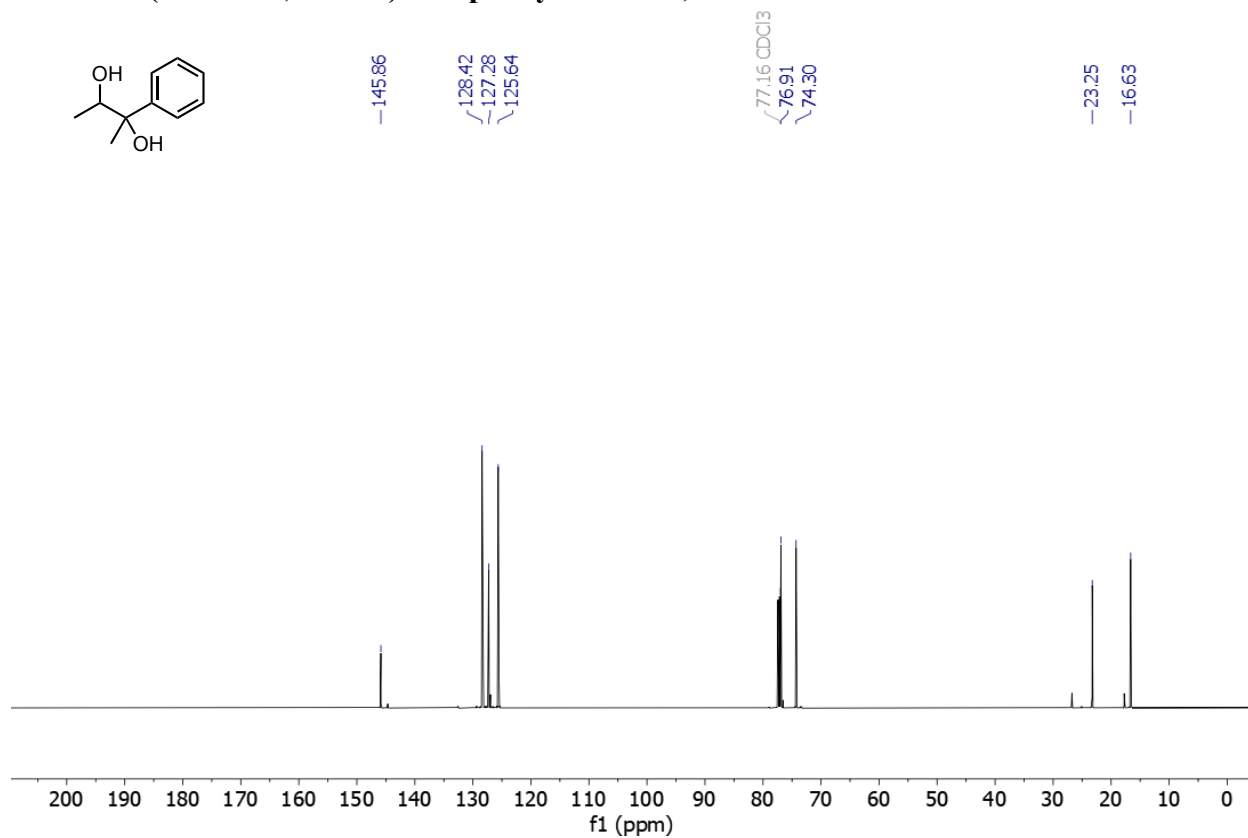
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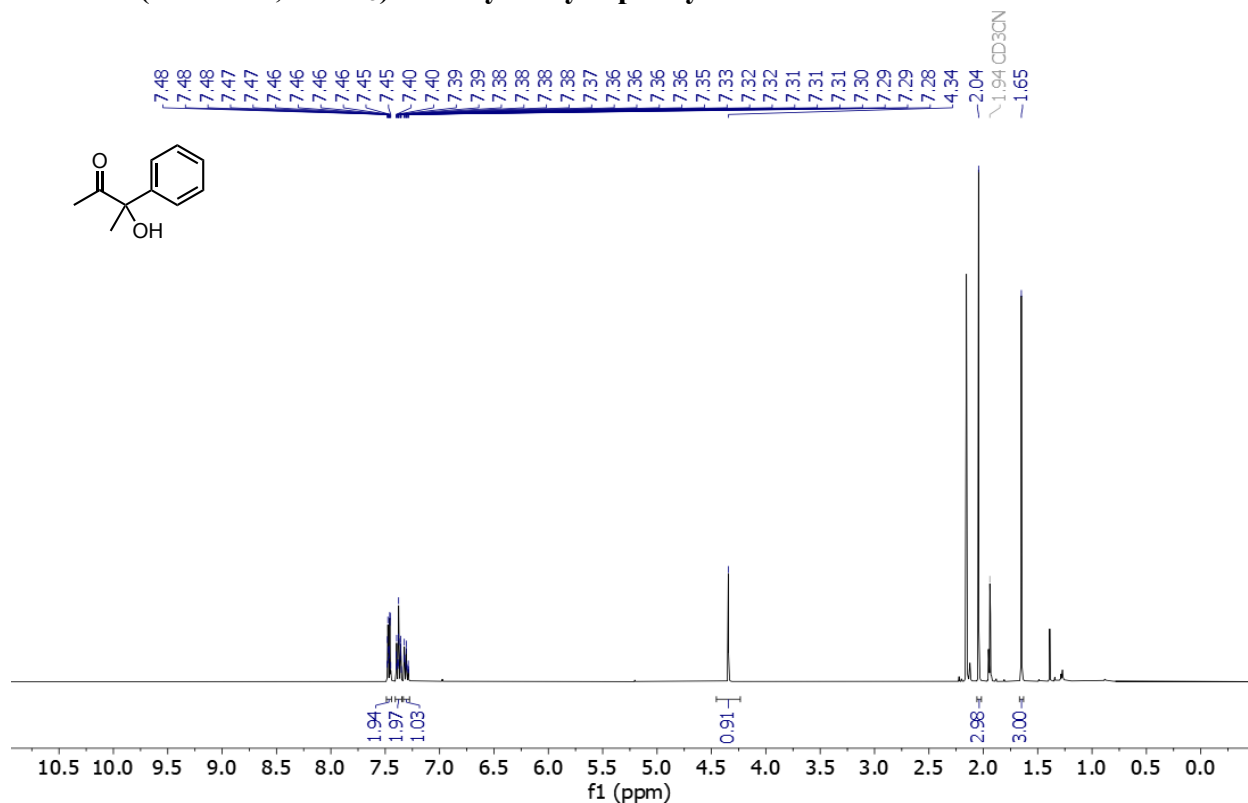
^1H NMR (500 MHz, CDCl_3) of 2-phenylbutane-2,3-diol



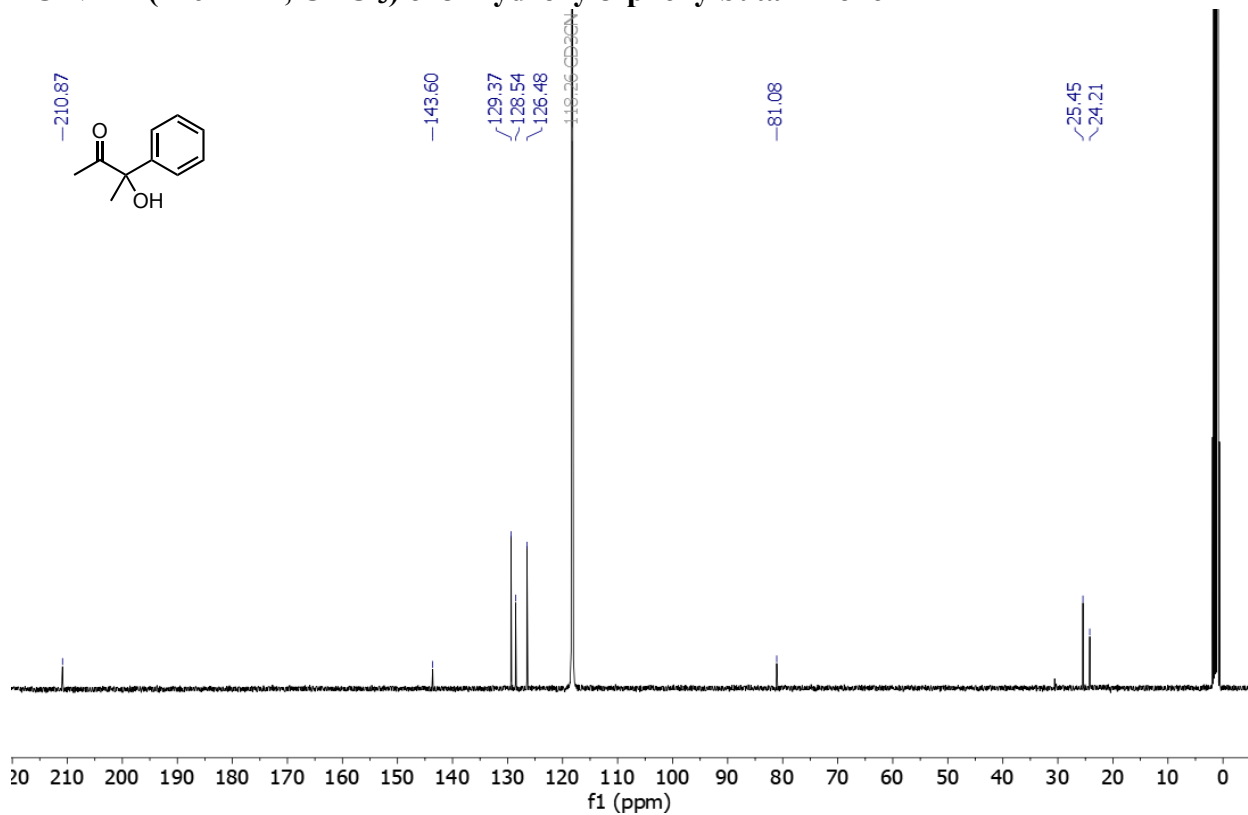
^{13}C NMR (126 MHz, CDCl_3) of 2-phenylbutane-2,3-diol



¹H NMR (500 MHz, CDCl₃) of 3-Hydroxy-3-phenylbutan-2-one

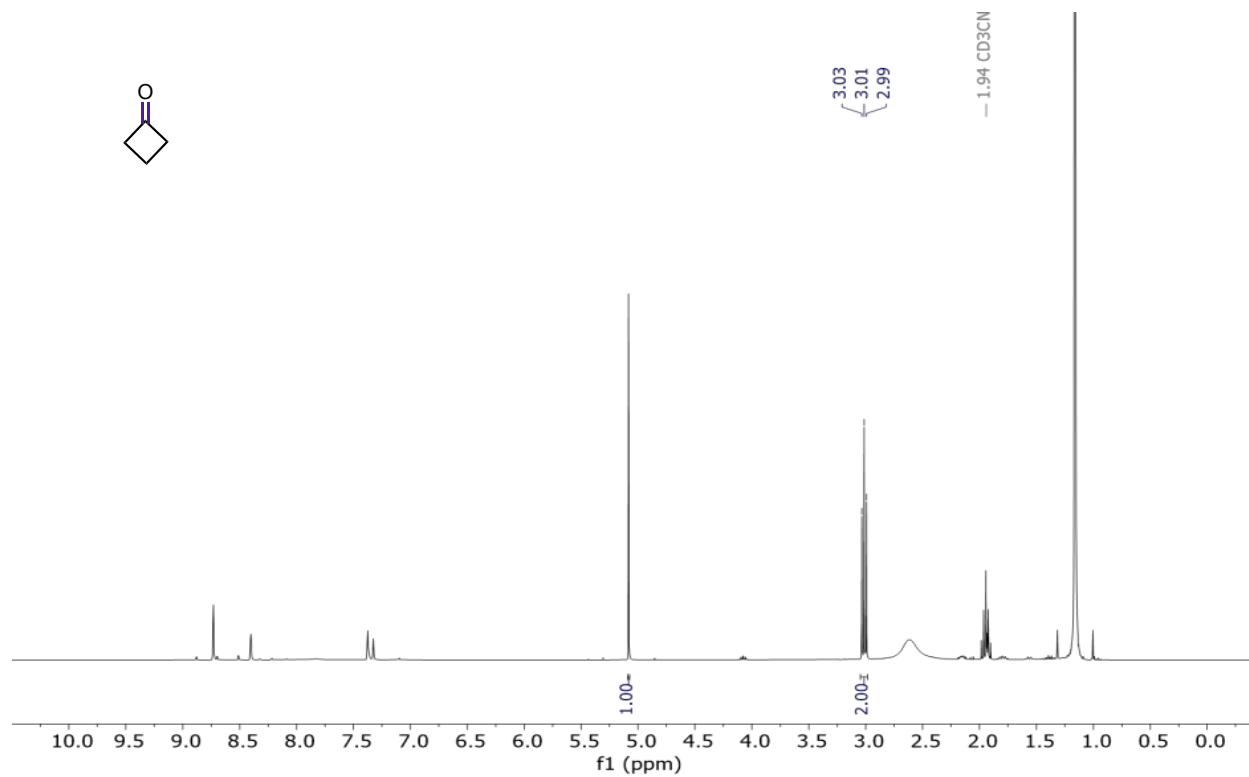


¹³C NMR (126 MHz, CDCl₃) of 3-Hydroxy-3-phenylbutan-2-one

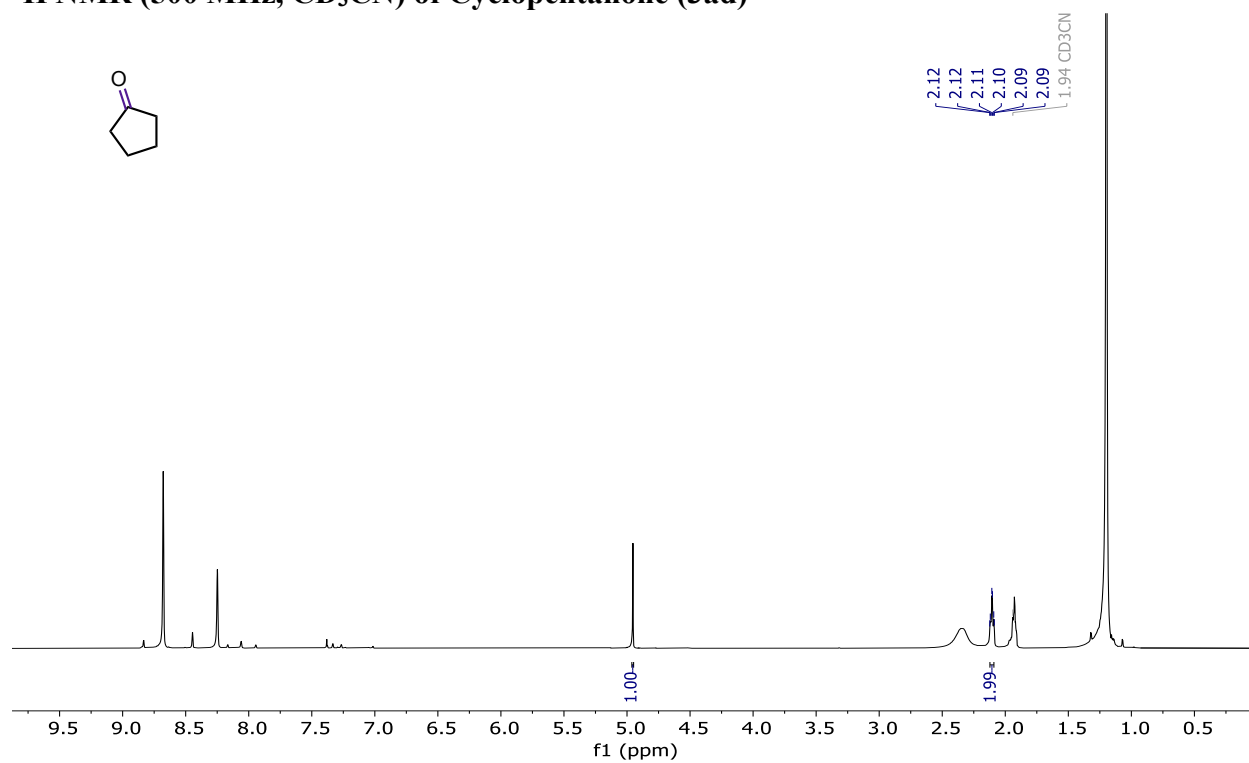


NMR yield spectra

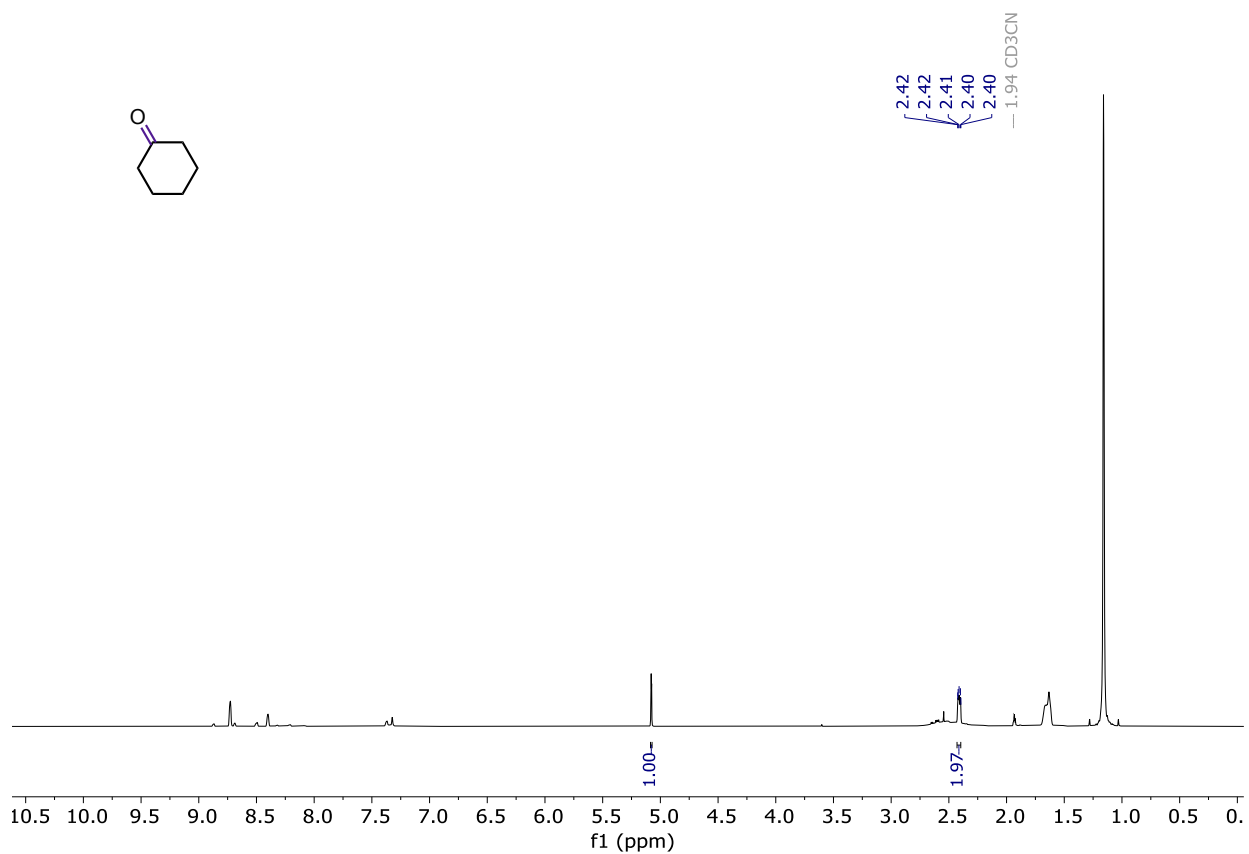
^1H NMR (500 MHz, CD_3CN) of Cyclobutanone (3ac)



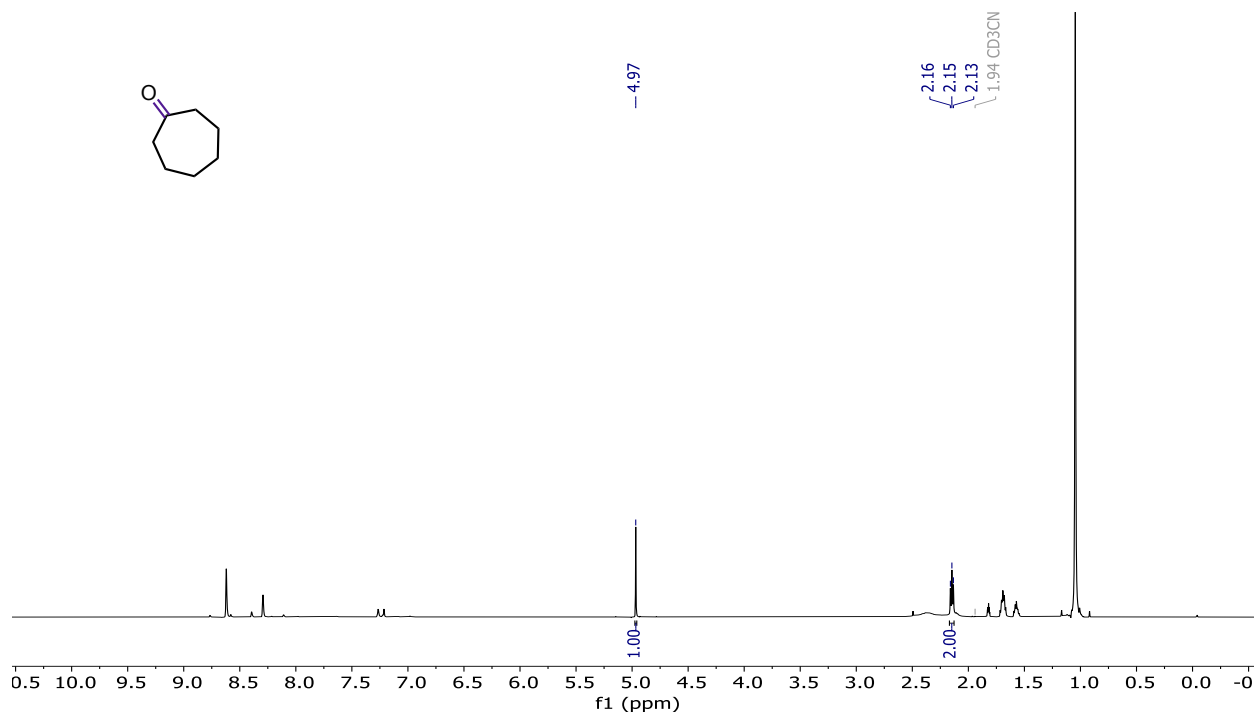
^1H NMR (500 MHz, CD_3CN) of Cyclopentanone (3ad)



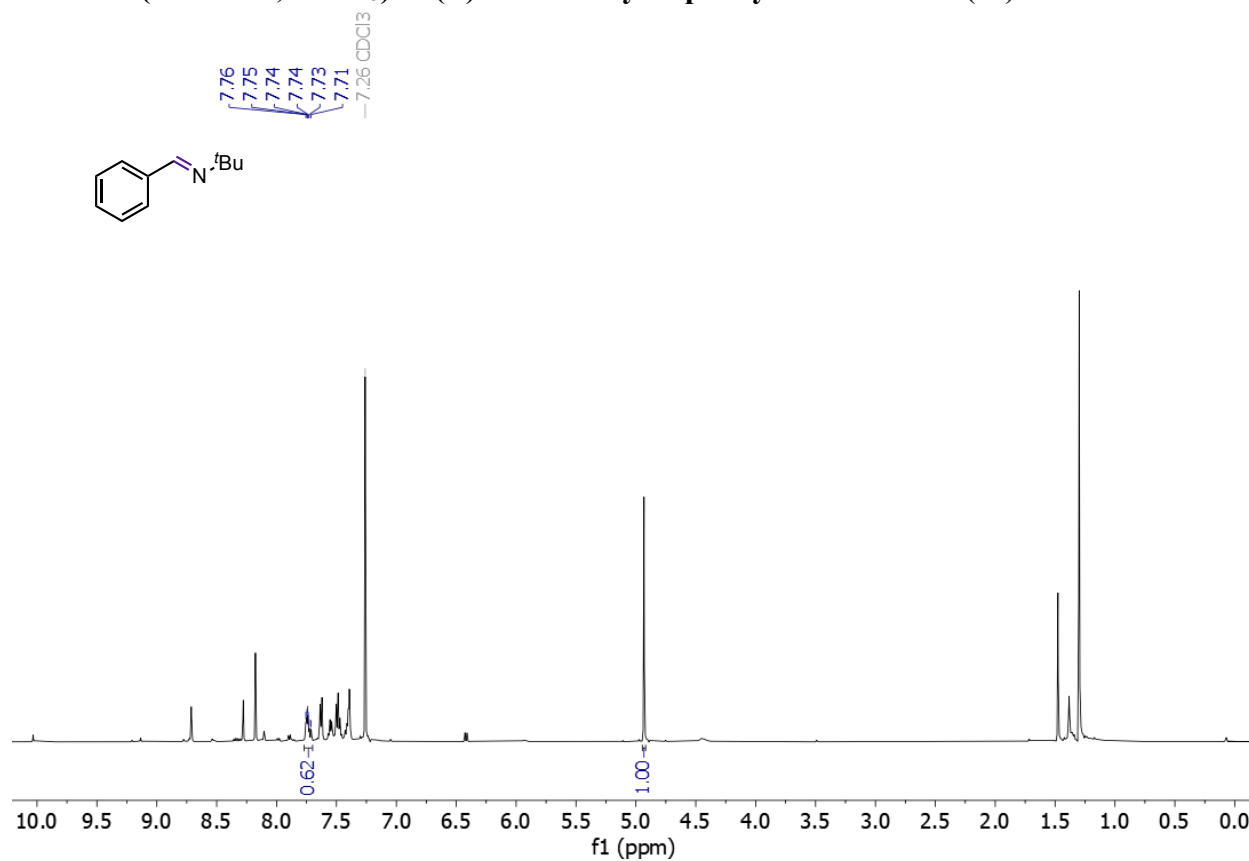
¹H NMR (500 MHz, CD₃CN) of Cyclohexanone (3ae)



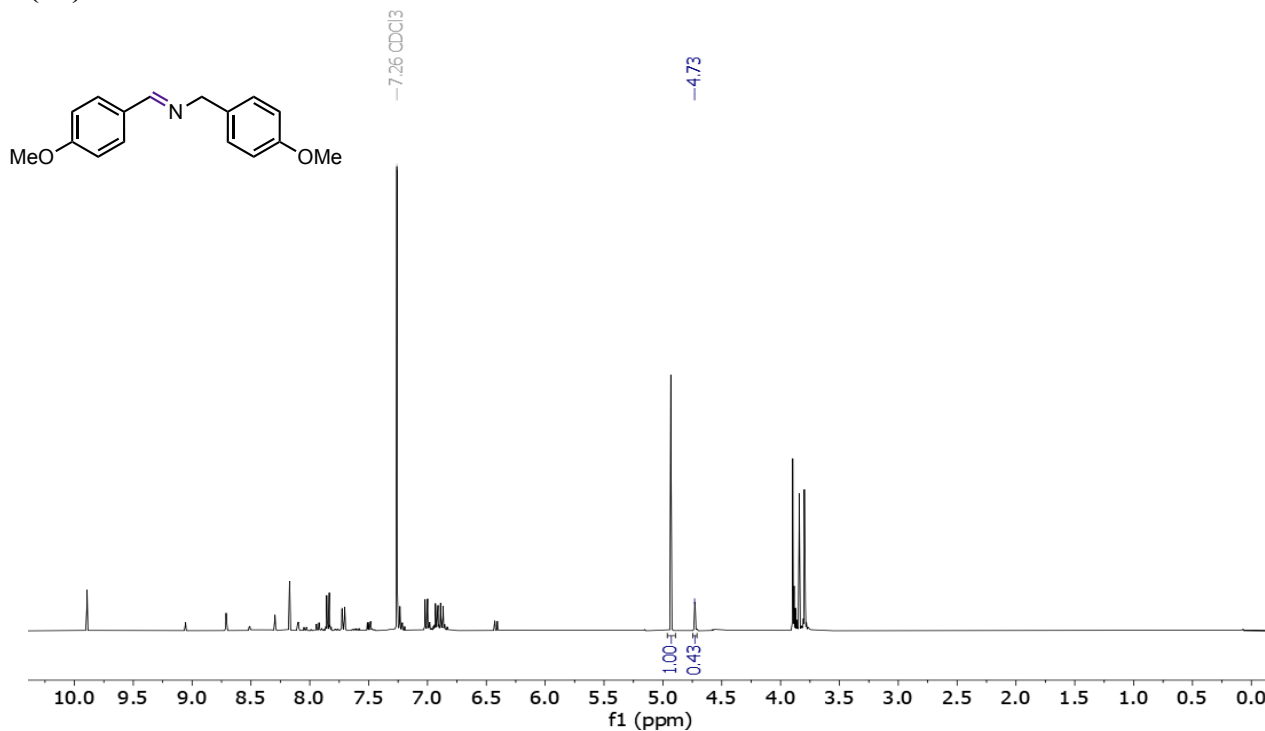
¹H NMR (500 MHz, CD₃CN) of Cycloheptanone (3af)



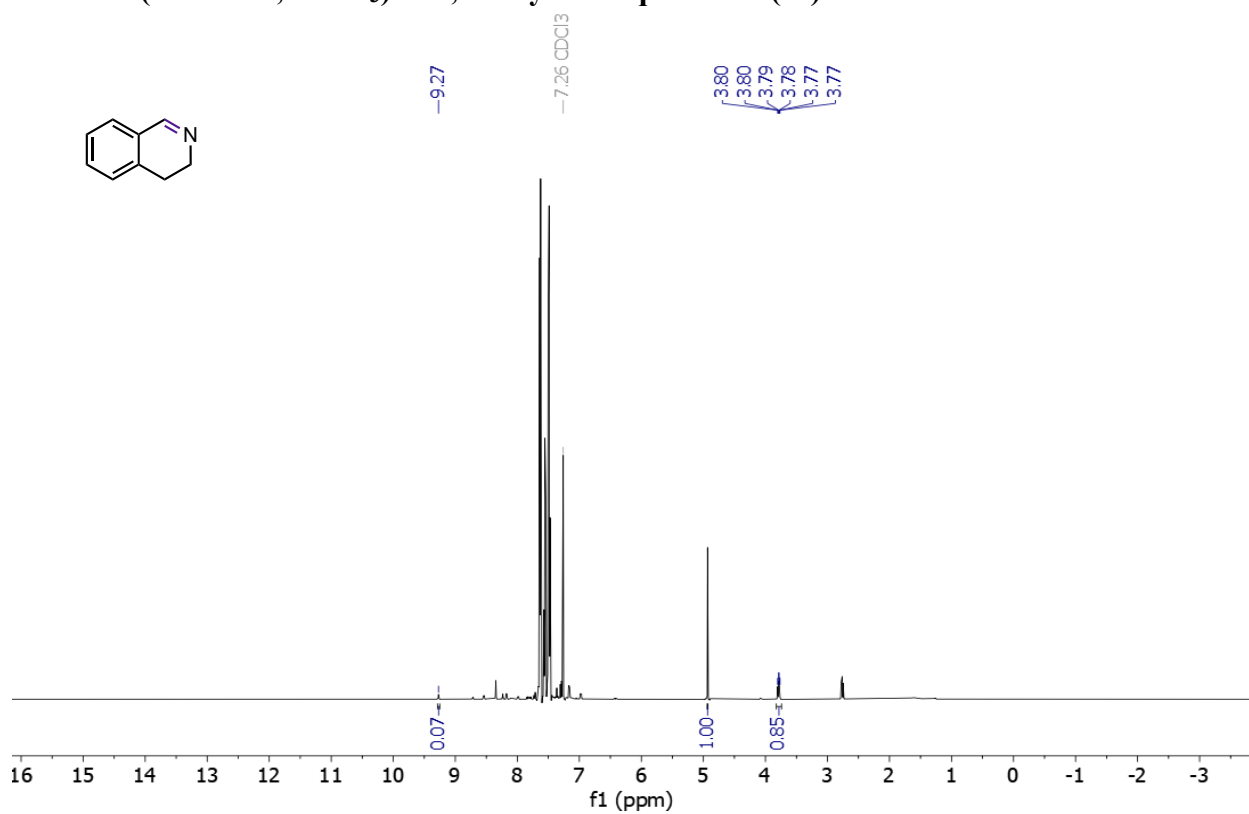
¹H NMR (500 MHz, CDCl₃) of (*E*)-*N*-*tert*-butyl-1-phenylmethanimine (4c)



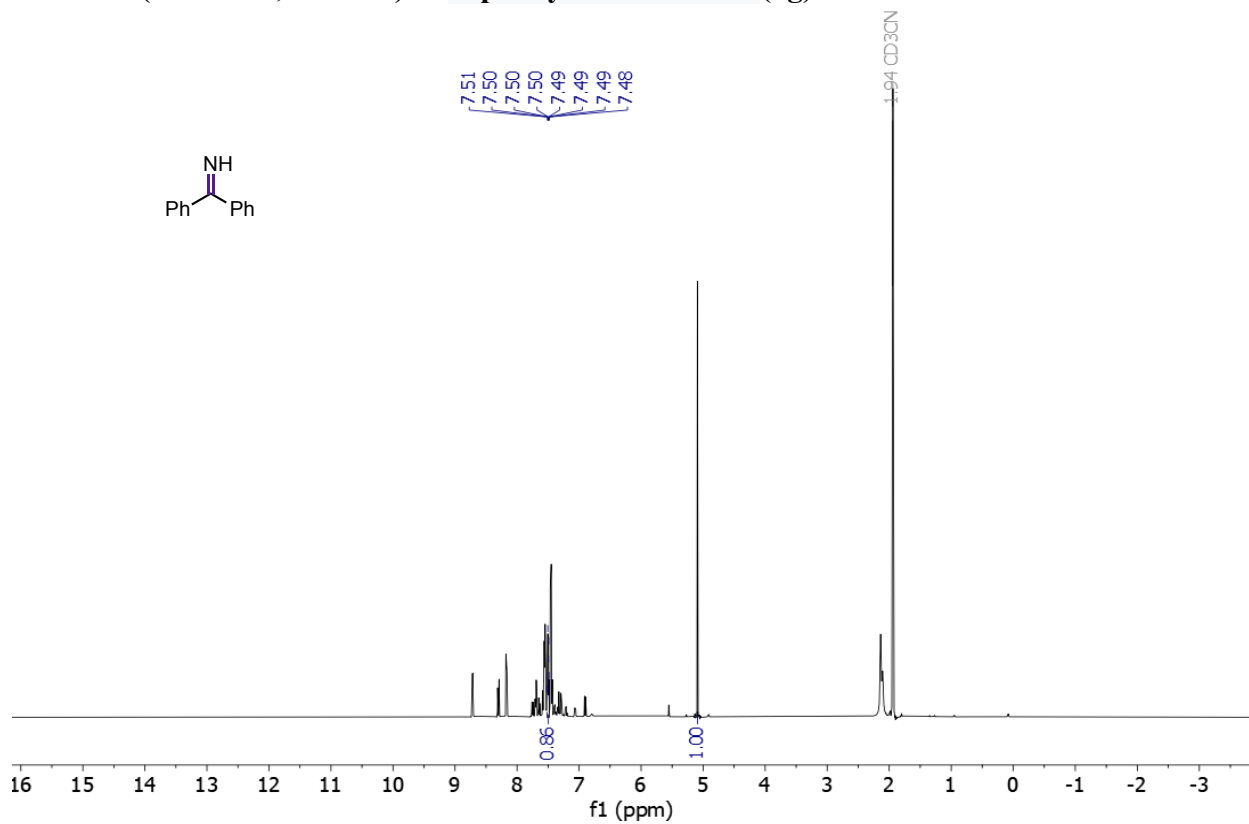
¹H NMR (500 MHz, CDCl₃) of (*E*)-*N*-(4-methoxybenzyl)-1-(4-methoxyphenyl)methanimine (4d)



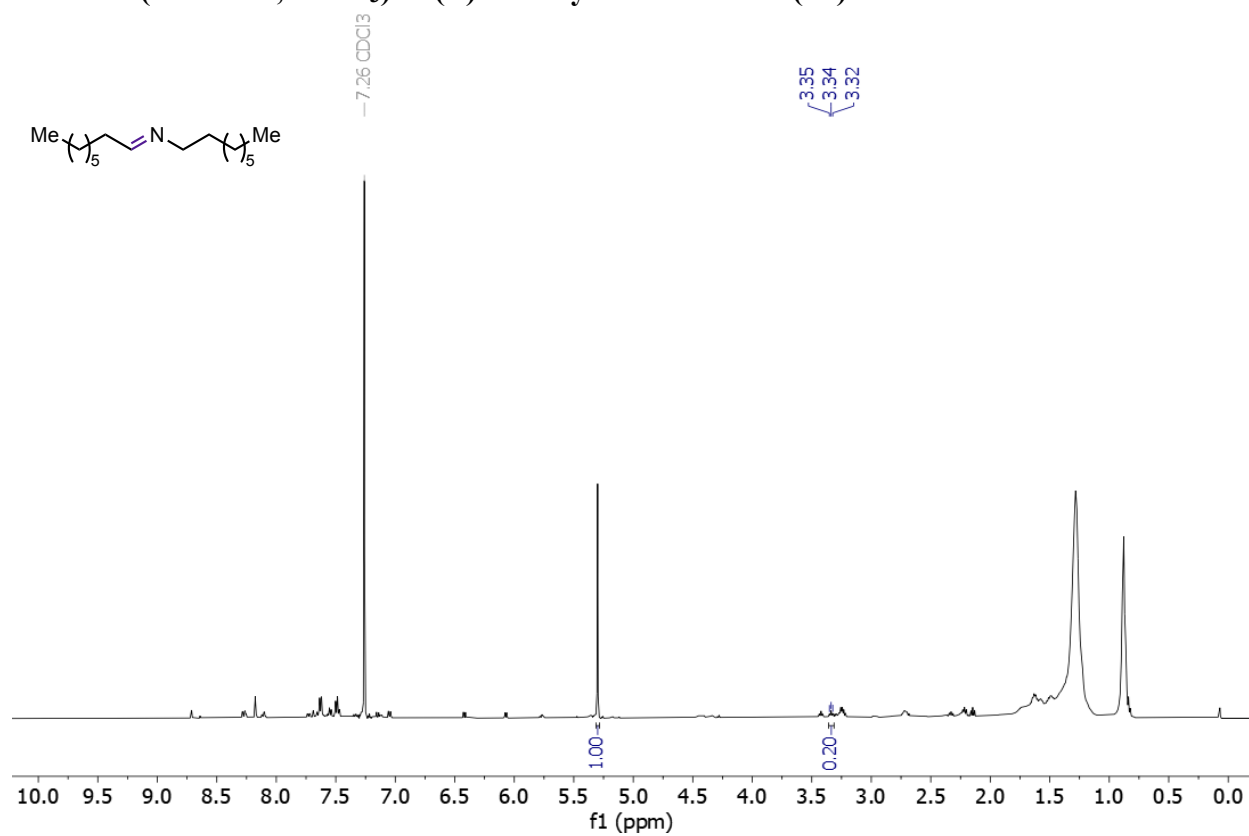
^1H NMR (500 MHz, CDCl_3) of 3,4-dihydroisoquinoline (4e)



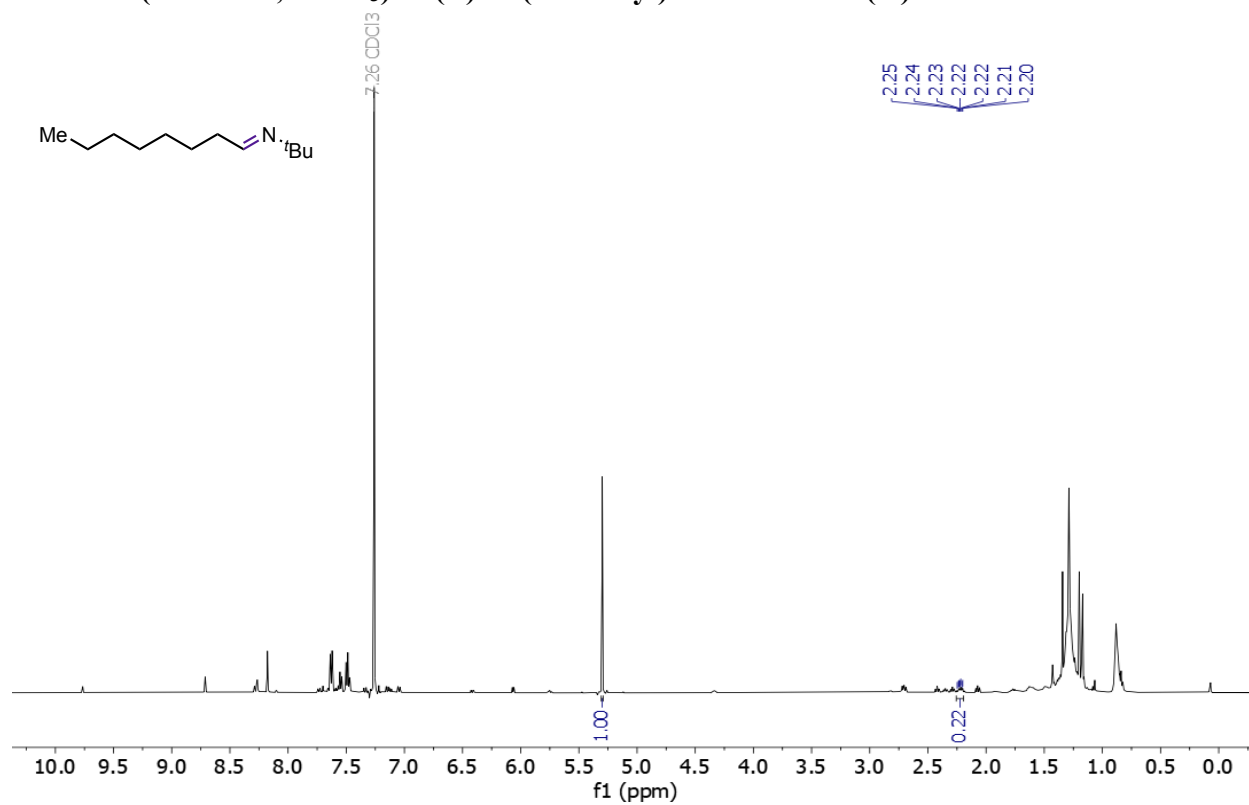
^1H NMR (500 MHz, CD_3CN) of Diphenylmethanimine (4g)



¹H NMR (500 MHz, CDCl₃) of (*E*)-*N*-Octyloctan-1-imine (4h)

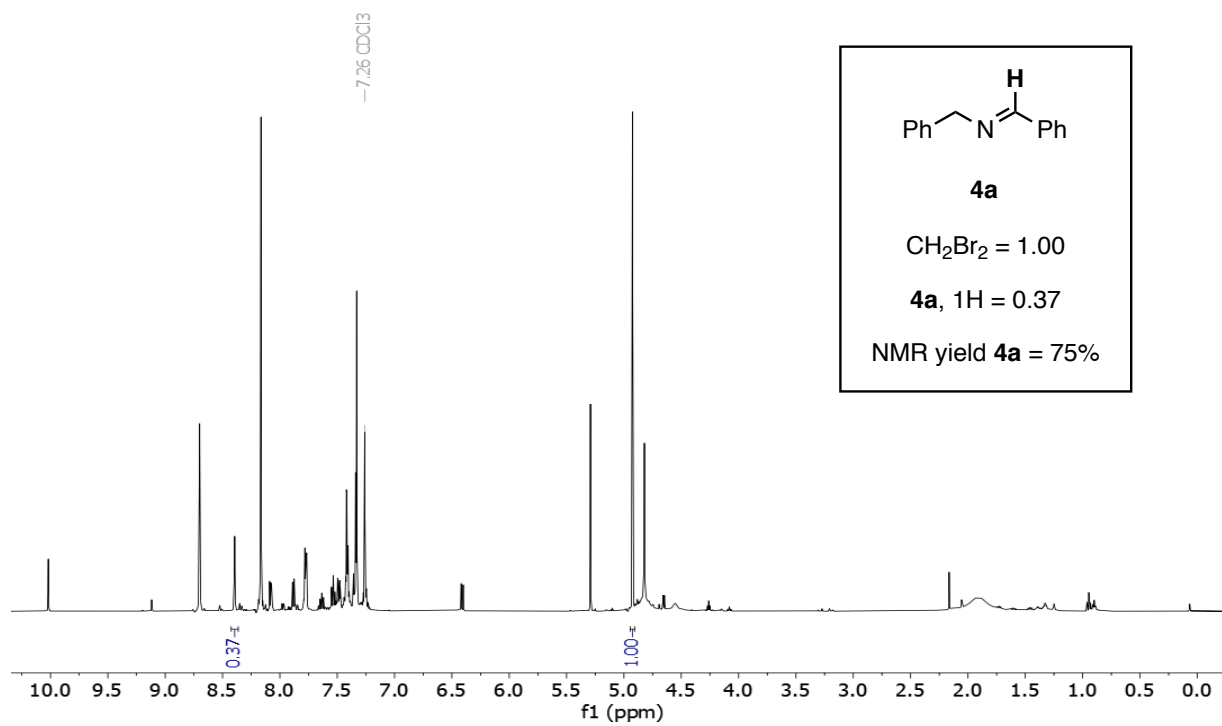


¹H NMR (500 MHz, CDCl₃) of (*E*)-*N*-(*tert*-butyl)octan-1-imine (4i)

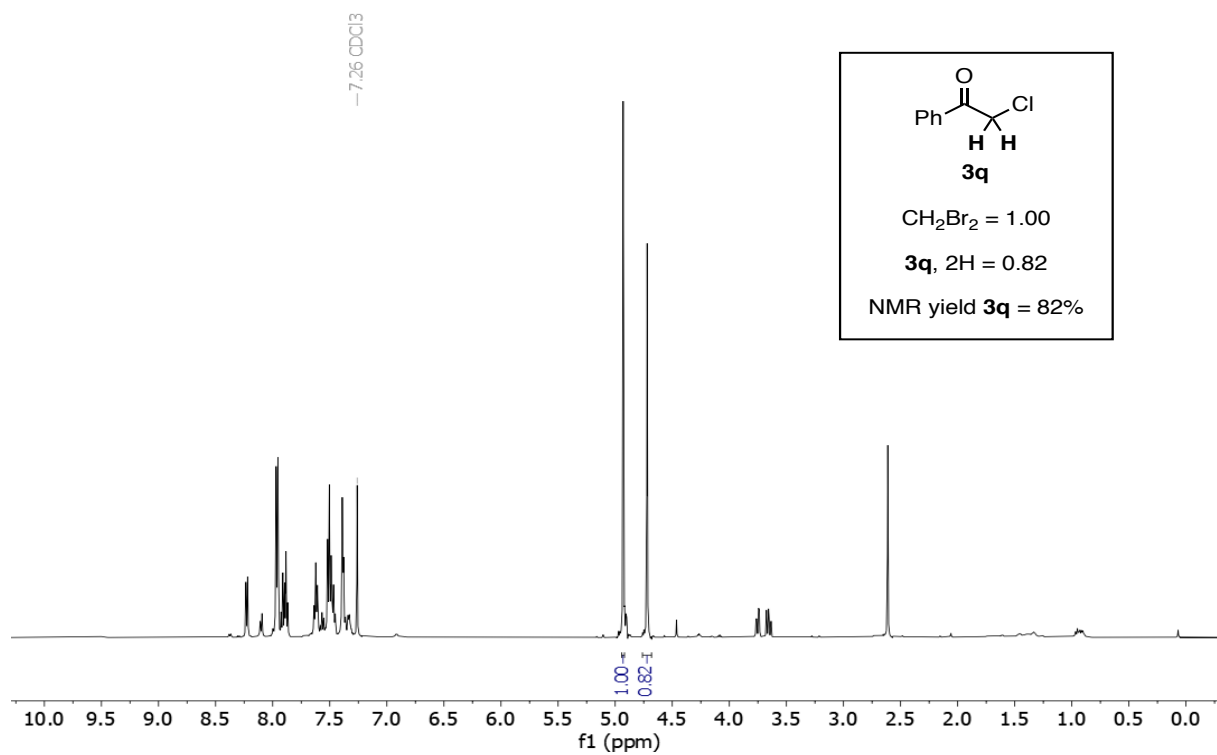


Continuous-Flow NMR yield spectra

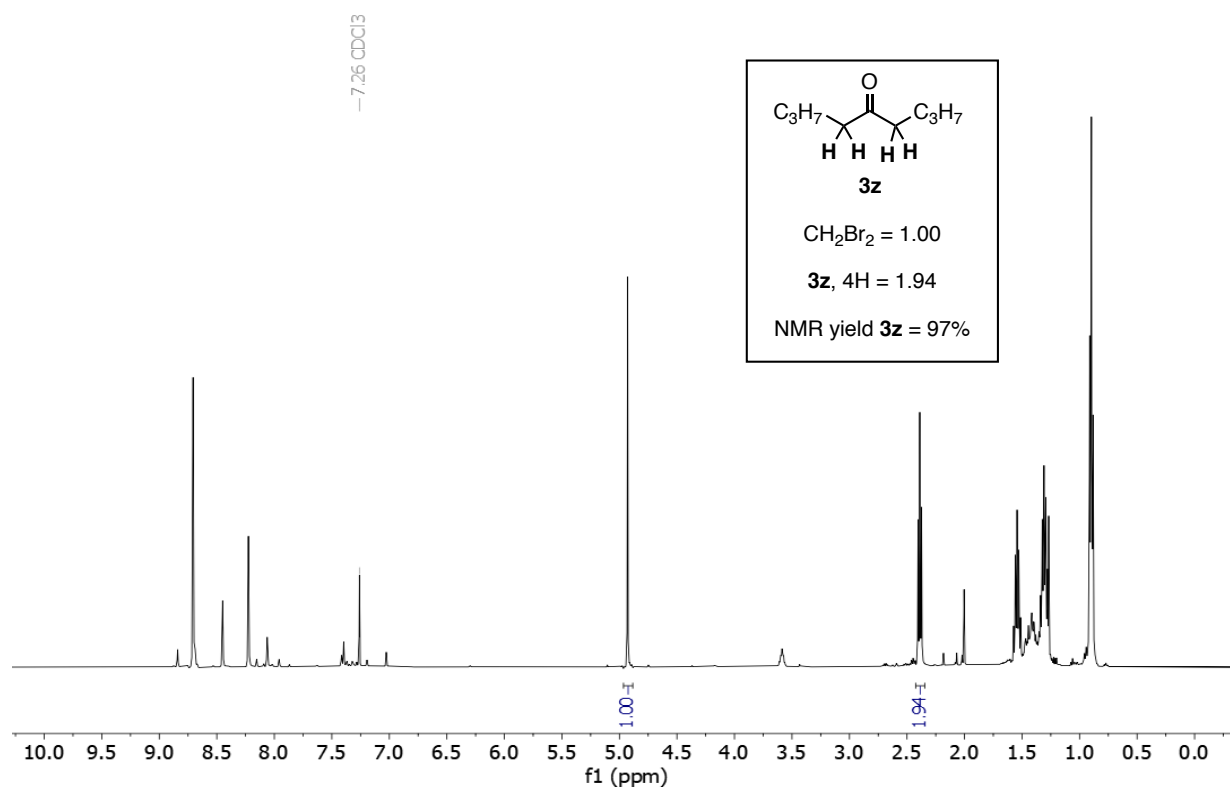
^1H NMR (400 MHz, CDCl_3) of (*E*)-*N*-benzyl-1-phenylmethanimine (**4a**)



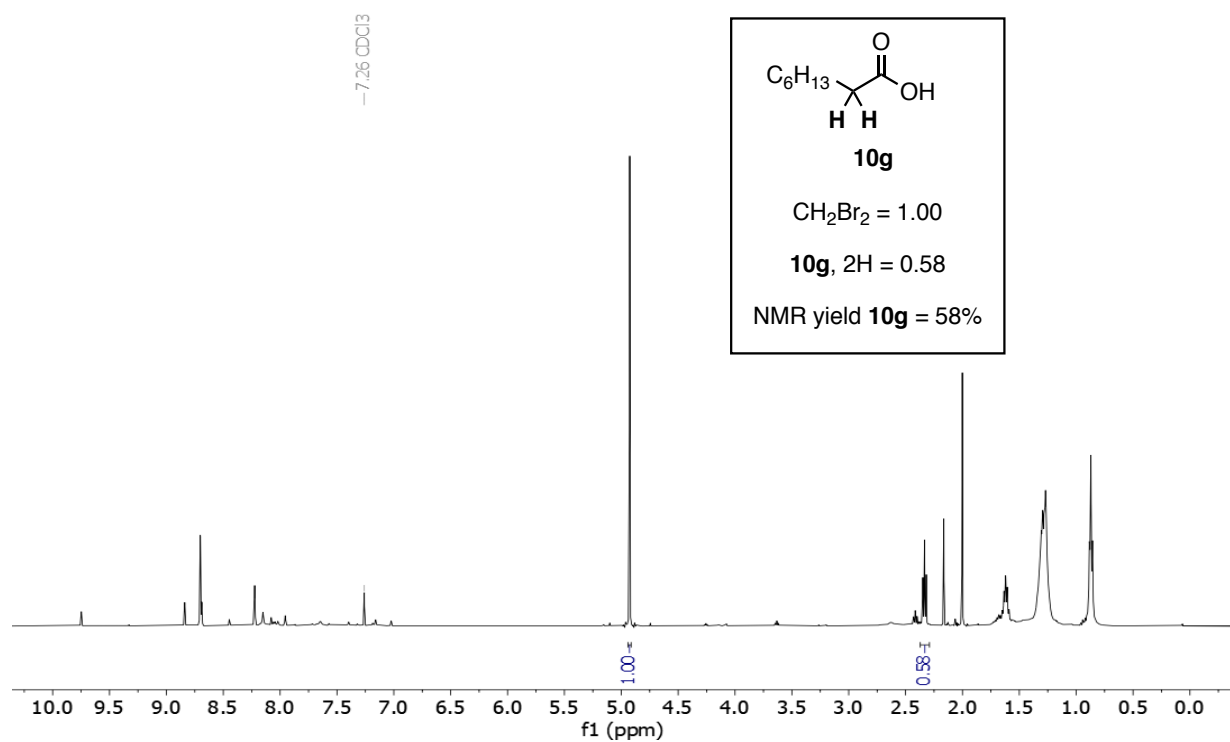
^1H NMR (400 MHz, CDCl_3) of 2-chloro-1-phenylethan-1-one (**3q**)



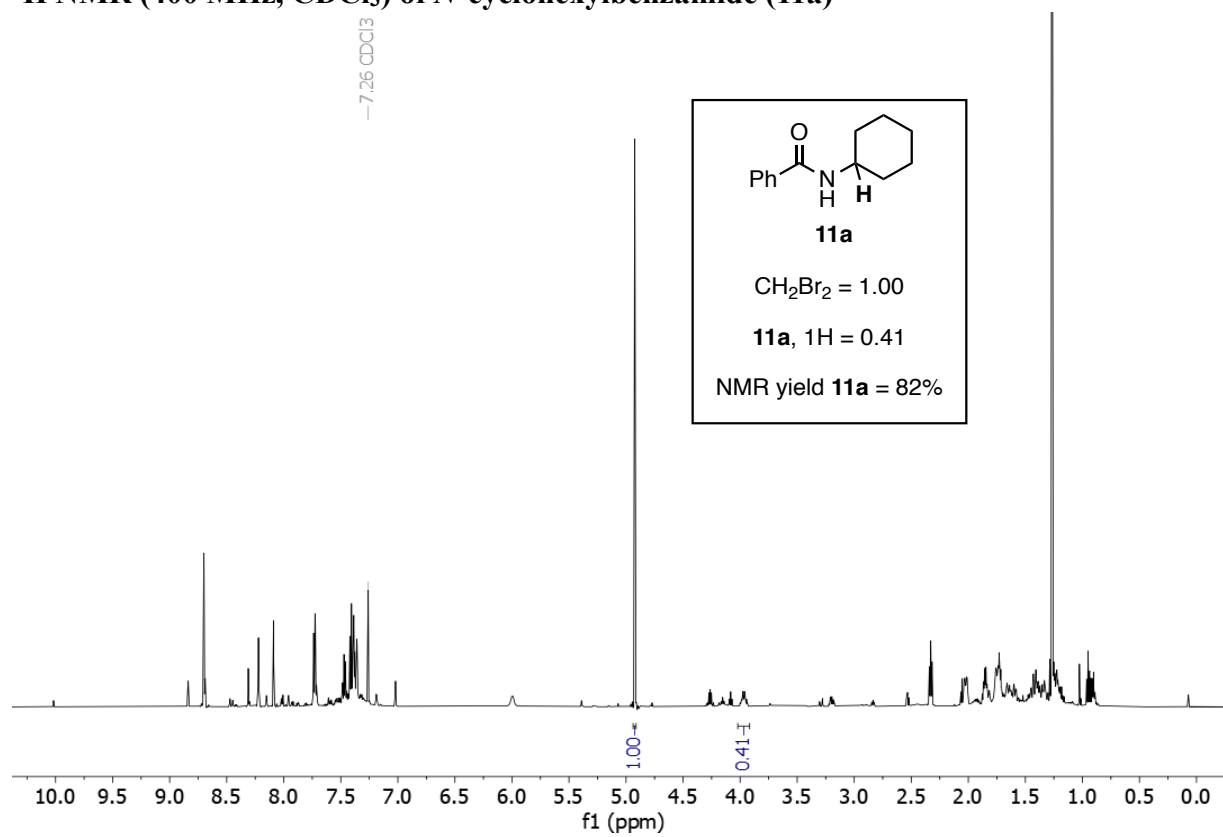
^1H NMR (400 MHz, CDCl_3) of 5-nonanone (3z**)**



^1H NMR (400 MHz, CDCl_3) of Octanoic acid (10g**)**



¹H NMR (400 MHz, CDCl₃) of *N*-cyclohexylbenzamide (11a)



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