SUPPLEMENTARY MATERIALS FOR

Single nucleus transcriptomics of ventral midbrain identifies glial activation associated with chronic opioid use disorder

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Table S1: Summary of Case and Control Cohort (Demographics and postmortem confounds)

Supplementary References

 0.00

 0.25

 0.50

 0.75

 1.00

Figure S1: Quality controls for VM single nuclei RNA-seq profiling. (A) Dexmulet genotyping confirms 100% match by donor by pool against the background of all 95 donors. Y-axis represents the library pools for 95 individuals. X axis represents the individual, prefix of which denoting the expected library for each subject. Heat bar, percentage of nuclei for specific individual in the library, as indicated. (B-D) Violin plots, showing separately for 45 individuals who died by opioid overdose (blue) and 50 controls (red), (B) the distribution of numbers of nuclei per subject (1 sample = 1 subject), (C) the average number of genes per nucleus for each subject, and (D) the average numbers of reads per sample (1 sample = 1 subject). (E) UMAP plot for VM from 95 samples (present) study, (F, G) automatic annotation of major VM cell types from previous study in a smaller reference sample¹.

Figure S2: Distribution of single nuclei with astrocyte, oligodendrocyte (ODC) and neuronal gene

expression. UMAP plots showing (A) separation of astrocyte and ODC nuclei, (B,C) expression of key (B) ODC marker genes, essentially limited to nuclei in ODC cluster, (C) astrocyte marker genes essentially limited to nuclei in the astrocyte cluster, (D,E) separation of neuronal nuclei into dopaminergic and non-dopaminergic subtypes which can be further split into subpopulations of nuclei expressing gabaergic (*GAD1, SLC32A1*) and glutamatergic (*SLC17A6*) neuron marker genes (see text).

Figure S3

Figure S3: Cell type-specific expression of opioid receptor genes in the VM. Violin plots depicting nuclear RNA levels for each of the 4 opioid receptor genes, *OPRM1, OPRK1, OPRD1 and OPRL*, for each of the 10 major VM cell types (x-axes) as indicated (left) at the single nucleus level (Seurat, default parameters) and (right) as cell type by subject, data shown as at normalized CPM, log2 transformed $(+1).$

Figure S4: **Differential gene expression across cohorts and cell types**. (A) Z-score (disease control differential) correlations comparing (y-axis, Miami cohort; x-axis Detroit cohort) cohorts for 4 major glial populations, as indicated, and non-dopaminergic neurons in the VM. (B) Proportional representation of DEGs shared between specific pairs of glial subtypes, as indicated on x-axis, based on shared vs. opposite directionality.

PCC celltype Astrocyte
DA
Microglia Microglia

Microglia

Microglia

Non DA

ODC

ODC

ODC

OPC

Pericyte

Pericyte 0.5 Microglia_2 $\overline{0}$ Microglia 0.5 Microglia_0 Pericyte

DA

Non_DA

Microglia_0 Pericyte Z

Microglia

Non_DA

 ODC_0

OPC Astrocyte Microglia_2

ODC ODC_1

#Nuclei of Opioid and Control for each subcluster

Figure S5: **Opioid exposure affects the transcriptome of multiple subtypes of oligodendrocytes**

and microglia. (A, B) UMAP subclusterings for (A) oligodendrocyte and microglial populations. (B) (left) Differential genes for ODC subclusters '0'and '1, as indicated, (right) Violin plots summarizing *OPALIN, S100B* and *RBFOX1* marker gene expression levels by subject (midbrain sample) and oligodendrocyte subtype, as indicated (C) Correlational matrix for differential gene expression (diseased versus control individuals) across all glial and neuronal populations, including subtypes (D) population structure of ODC single nuclei, diseased individuals vs. controls, as indicated.

Astrocyte 1

nucleobase-containing compound catabolic.

Astrocyte \downarrow

● vascular associated smooth muscle cell.

negative regulation of oligodendrocyte. positive regulation of astrocyte differentiation neuronal stem cell population maintenance **Outflow tract morphogenesis** chordate embryonic development neural tube development negative regulation of nervous system. negative regulation of neurogenesis regulation of nervous system development hindlimb morphogenesis **O** embryonic hindlimb morphogenesis **O** extracellular matrix constituent secretion regulation of extracellular matrix constituent. regulation of glial cell differentiation Cregulation of oligodendrocyte differentiation oligodendrocyte differentiation gliogenesis glial cell differentiation **O** response to amphetamine nucleotide-sugar metabolic process **O** apical protein localization bssification regulation of cell shape **O** exit from mitosis **Onegative regulation of transcription from RNA.** regulation of erythrocyte differentiation cellular response to oxidative stress cellular response to chemical stress ♦ regulation of microvillus organization **Internal metabolism O** regulation of cell projection size **Onegative regulation of stress-activated protein.** negative regulation of stress-activated MAPK. placenta blood vessel development Clabyrinthine layer development **Operipheral nervous system axon ensheathment O** myelination in peripheral nervous system Schwann cell development peripheral nervous system development Schwann cell differentiation ensheathment of neurons myelination axon ensheathment protein localization to peroxisome protein targeting to peroxisome establishment of protein localization to. peroxisomal transport peroxisome organization Microglia **↓ ● putrescine biosynthetic process ODNA** methylation involved in gamete generation O positive regulation of NMDA glutamate receptor.

- **O**inositol trisphosphate metabolic process **O**regulation of bone development
- \bigcirc negative regulation of inclusion body assembly
- negative regulation of transcription from RNA poly.
- \bigcirc N-acetylneuraminate catabolic process
- N-acetylneuraminate metabolic process
- response to dietary excess
- diet induced thermogenesis
- \bullet requlation of adipose tissue development
- **•** positive regulation of adipose tissue development
- **Positive regulation of glial cell migration**
- **•** regulation of glial cell migration
- negative regulation of interleukin-1 production
- negative regulation of interleukin-1 beta production
- **Omyeloid leukocyte differentiation** eukocyte differentiation
- regulation of cell activation
- positive regulation of myeloid cell differentiation
- **O** positive regulation of monocyte differentiation
- positive regulation of hemopoiesis
- positive regulation of leukocyte differentiation
- **O**positive regulation of nucleotide-binding oligo.
- **O** positive regulation of nucleotide-binding oligo.
- regulation of nucleotide-binding oligo.
- **D** regulation of nucleotide-binding olig.

embryonic oligodendrocyte morphogenesis constituent

number of genes

p.adjust

0.025

0.050

0.075

cellular response cascade stress

labyrinthine layer

blood vessel

myelination peripheral axon ensheathment

establishment peroxisomal targeting peroxisome

N-acetylneuraminate metabolic process activity

diet adipose dietary tissue

glial interleukin-1 migration production

leukocyte myeloid

activation differentiation

number of genes

p.adjust

nucleotide-binding oligomerization domain containing

10 15

Figure S6: Cell-type specific pathway enrichments of differentially expressed genes in astrocytes and microglia. GO pathway enrichments for Biological Process, shown separately for upand down-regulated DEGs, as indicated. See also Figure 3 and Data S5, including gene ratios (Data S5, column E).

ODC 1

Non DA neuron ↑

histone H3-K9 modification

ODC **L**

O leukemia inhibitory factor signaling pathway **D** pentose-phosphate shunt, non-oxidative branch \bullet · glomerulus morphogenesis ● · · regulation of chloride transport **O** negative regulation of T cell cytokine production ← positive regulation of peptidyl-serine phosphorylation. keratan sulfate metabolic process **C** keratan sulfate biosynthetic process **D**organelle fission nuclear division mitotic nuclear division mitotic spindle organization regulation of mitotic nuclear division

regulation of centrosome cycle

Non DA neuron J

glomerulus inhibitory factor branch

regulation chloride transport

negative cell peptidylserine cytokine

keratan sulfate biosynthetic process

mitotic nuclear centrosome division

 \bigcirc 10

number of genes

 10.0

 \bullet 5.0

p.adjust

0.005 0.010 0.015 0.020

rane

sis ht

ntal lved **Figure S7: Cell-type specific pathway enrichments of differentially expressed genes in oligodendrocytes** (ODC) **and non-dopaminergic neurons** (Non-DA)**.** GO pathway enrichments for Biological Process, shown separately for up- and down-regulated, as indicated. See also Figure 3 and Data S5, including gene ratios (Data S5, column E).

Figure S8

z-score cell type specific nuclei

Figure S8: **Correlations between prior bulk RNA-seq study and the current RNA-seq analysis with**

single cell resolution. Z-score (disease vs. control differential) correlations comparing (y-axis, bulk tissue RNA-seq study; x-axis, cell type specific RNA-seq at single nuclei resolution, current study) for 7 cell types, as indicated.

Figure S9

Figure S9: **DEGs linked to TWAS**. (A) Percentages representing, for each cell type the normalized proportion of DEGs overlapping with PhenomeXcan TWAS for substance use and medical or neurological traits, as indicated. We focused on Substantia Nigra (SN) gene expression and populationscale SUD phenotypes in PhenomeXcan. We included a total of 1,260 PhenomeXcan genes for a total of 40 SUD-related traits from the following categories of traits or diseases: alcohol, caffeine, marijuana, and smoking (Data S6). We also analyzed a recent study² that had called addiction risk genes via TWAS-guided integration of GTEX and PsychENCODE expression quantitative trait loci (see Data S7,S8). For GTEx, the study conducted TWAS analyses using MetaXcan via integration of eQTL from 13 brain regions and identified a total of 351 addiction risk factor genes (FDR<10%). For PsychENCODE, using the frontal and temporal cortex, TWAS analysis using S-PrediXcan identified a total of 410 addiction risk genes with FDR<10%². (B) Cell-type specific counts of genes called as DEG in current study *and* linked to substance use trait(s) in PhenomeXcan (see Data S7).

Figure S10: Tissue and demographic variables impacting single nuclei transcriptomes (A) Ventral midbrain region-of-interest. Drawing from representative coronally cut VM tissue block, showing the substantia nigra and adjacent portion of ventral tegmental area, the region-of-interest prepared by dissection and then further processed with the nuclei extraction protocol (see Methods). (B, C) Effects of tissue quality and demographic variables on single nuclei transcriptomes. (B) Frequency histogram, showing the fraction of mitochondrial genes/nucleus. Note fraction of <1% for the overwhelming majority, or 93.7% of nuclei. Note also that there are 1.8% of nuclei with >2% mitochondrial, 0.1% of nuclei with >10% mitochondrial and 0.025% of nuclei had >20% mitochondrial reads. (C) Linear regression testing for association of (left to right) nuclei number/donor, number of reads/nuclei, number of genes/nucleus and fraction of mitochondrial reads/nucleus with (top to bottom) donor age, brain tissue pH, postmortem interval (PMI), genotype principal component PC1 and PC2, and sex, *(**) P< 0.05, 0.01.

Supplemental References

- 1. Kamath T*, et al.* Single-cell genomic profiling of human dopamine neurons identifies a population that selectively degenerates in Parkinson's disease. *Nat Neurosci* **25**, 588- 595 (2022).
- 2. Hatoum AS*, et al.* Multivariate genome-wide association meta-analysis of over 1 million subjects identifies loci underlying multiple substance use disorders. *Nat Ment Health* **1**, 210-223 (2023).