

Supplementary Materials for
PARP14 is a PARP with both ADP-ribosyl transferase and hydrolase activities

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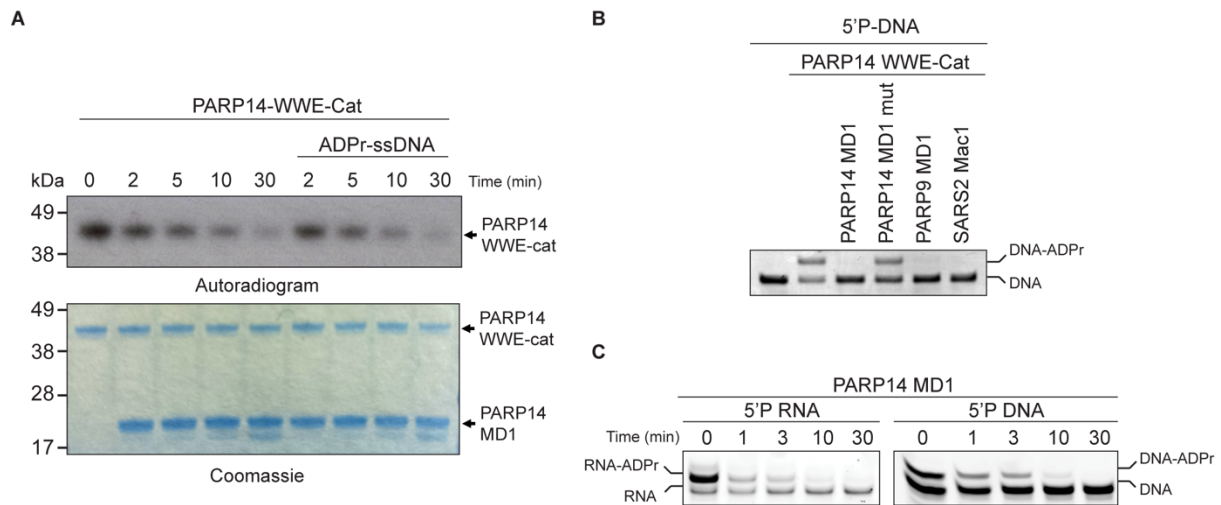
The PDF file includes:

Figs. S1 to S5
Tables S1 to S5
Legends for data S1 to S3

Other Supplementary Material for this manuscript includes the following:

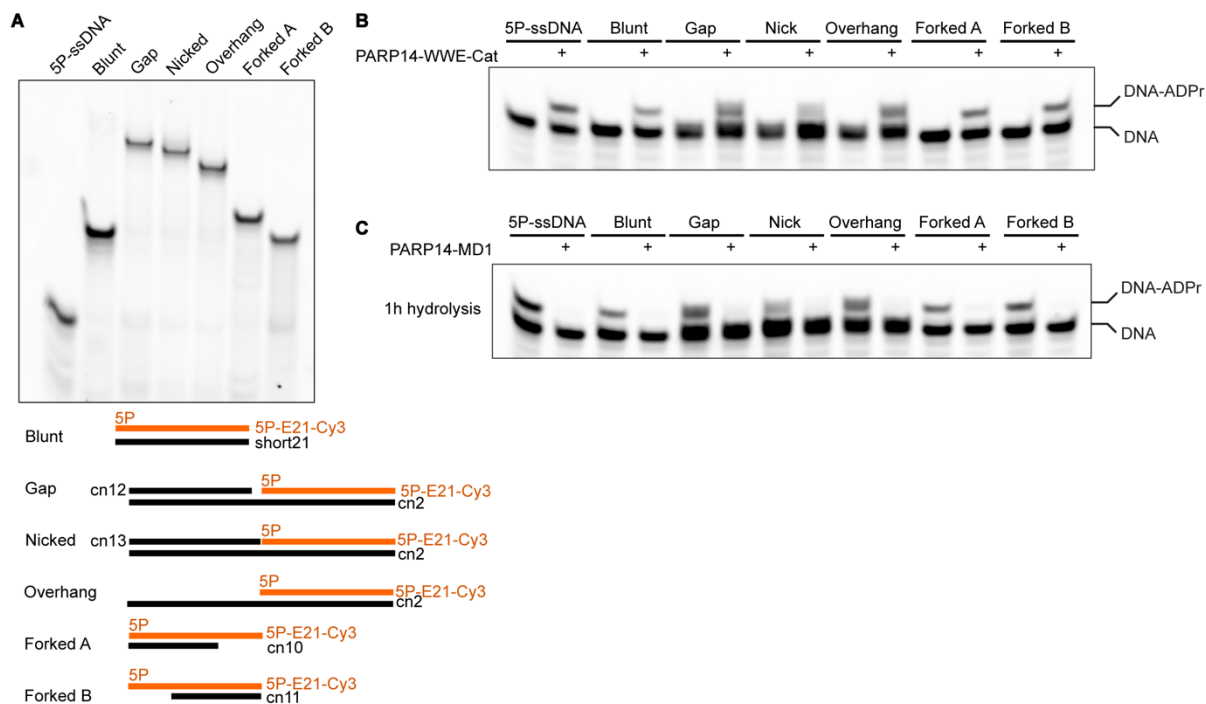
Data S1 to S3

Supplementary Figures:

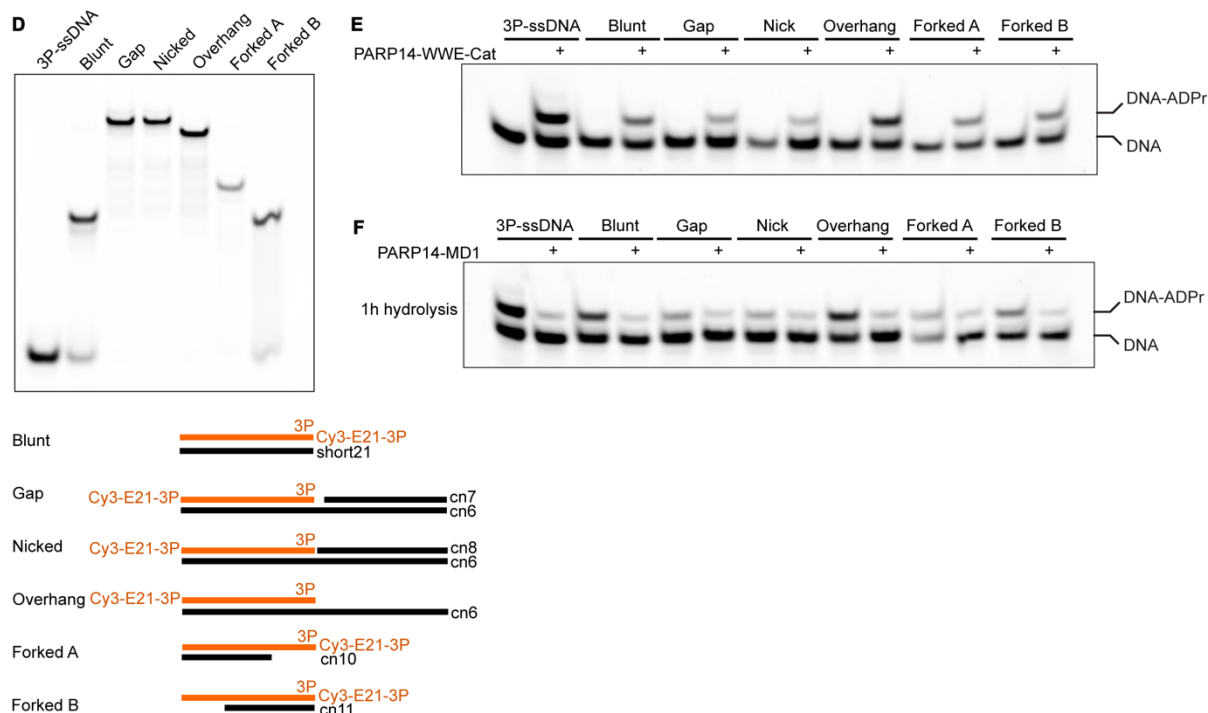


Supplementary Figure 1: PARP14 macrodomain 1 reverses protein, DNA and RNA ADP-ribosylation. (A) Time-course reversal of ADP-ribosylated PARP14 WWE-cat by PARP14 MD1 in the presence or absence of excess ADPr-ssDNA. ADP-ribosylated PARP14 WWE-cat was incubated with or without PARP14 MD1 in presence or absence of eight fold molar excess of ADP-ribosylated ssDNA for the indicated times. (B) PARP14 MD1 reverses ADP-ribosylation of 5'P-ssDNA. ssDNA with 5' phosphate and 3' Cy3 were ADP-ribosylated using PARP14 WWE-cat. Subsequently, the ADP-ribosylated DNA was purified and used as a substrate for de-ADP-ribosylation reactions with PARP14 MD1, MD1 mut, MD2, MD3 and SARS2 Mac1 (C) Time-course reversal of ADP-ribosylated 5' phosphate ssRNA or 5' phosphate ssDNA using PARP14 MD1. ADP-ribosylated 5' and 3' phosphorylated ssRNA and ssDNA were treated with PARP14 MD1 and samples were taken at the indicated timepoints.

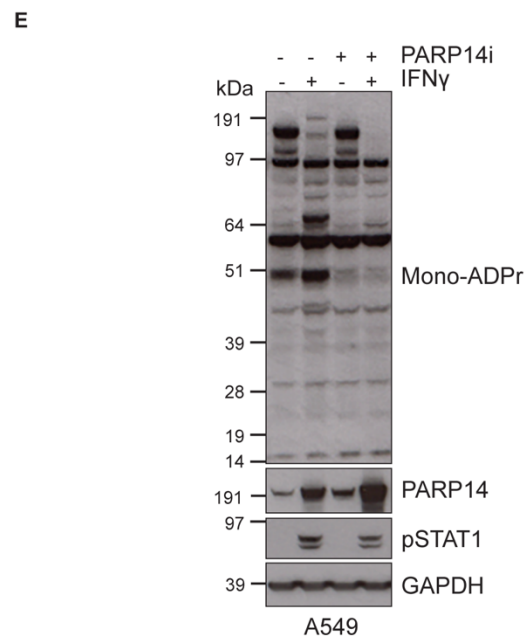
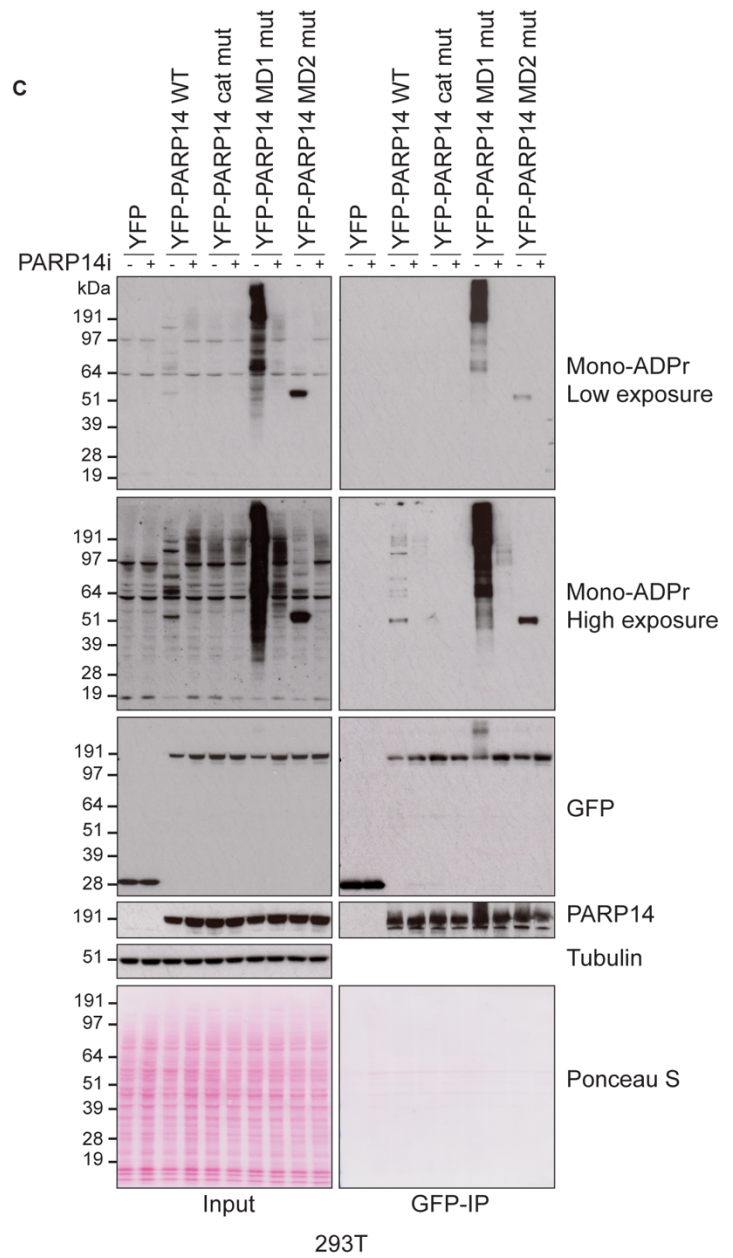
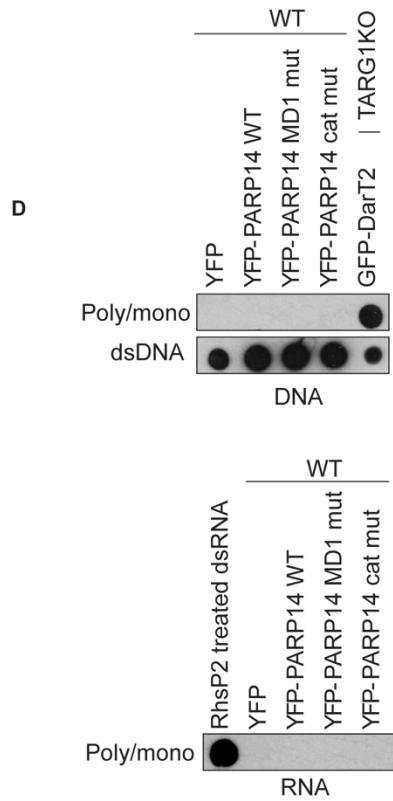
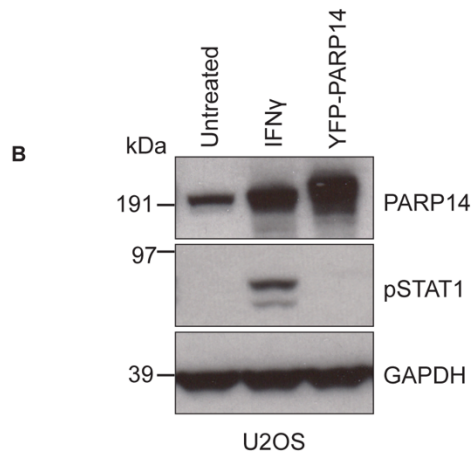
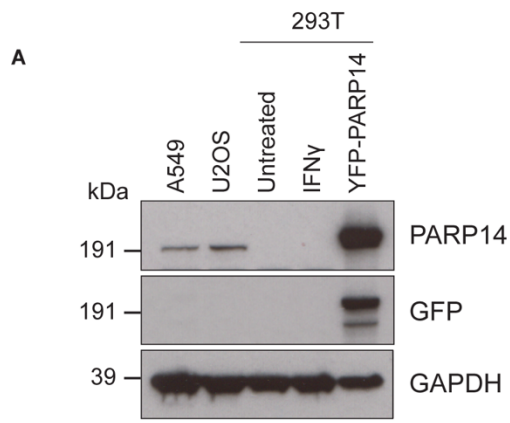
5'P-dsDNA set



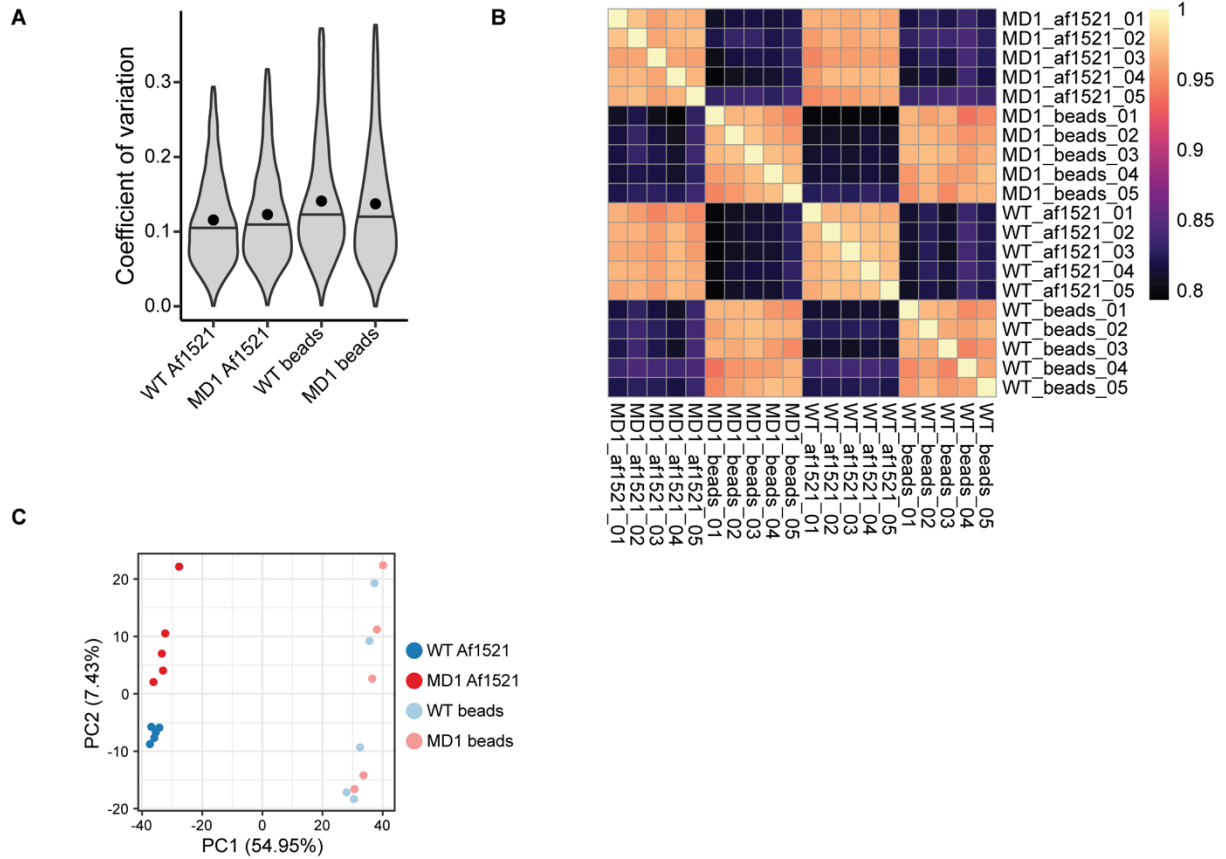
3'P-dsDNA set



Supplementary Figure 2: PARP14 macrodomain 1 can remove ADPr from various dsDNA substrates. (A and D) Quality control of annealed 5'P-dsDNA and 3'P dsDNA analysed by native polyacrylamide gel electrophoresis. (B and E) Annealed 5'P-dsDNA and 3'P dsDNA were ADP-ribosylated with 5 μ M PARP14 WWE-cat and analysed by urea polyacrylamide gel electrophoresis (C and F) ADP-ribosylated 5'P-dsDNA from B and ADP-ribosylated 3'P dsDNA from E were treated with 4 μ M PARP14 MD1 and analysed by urea polyacrylamide gel electrophoresis.

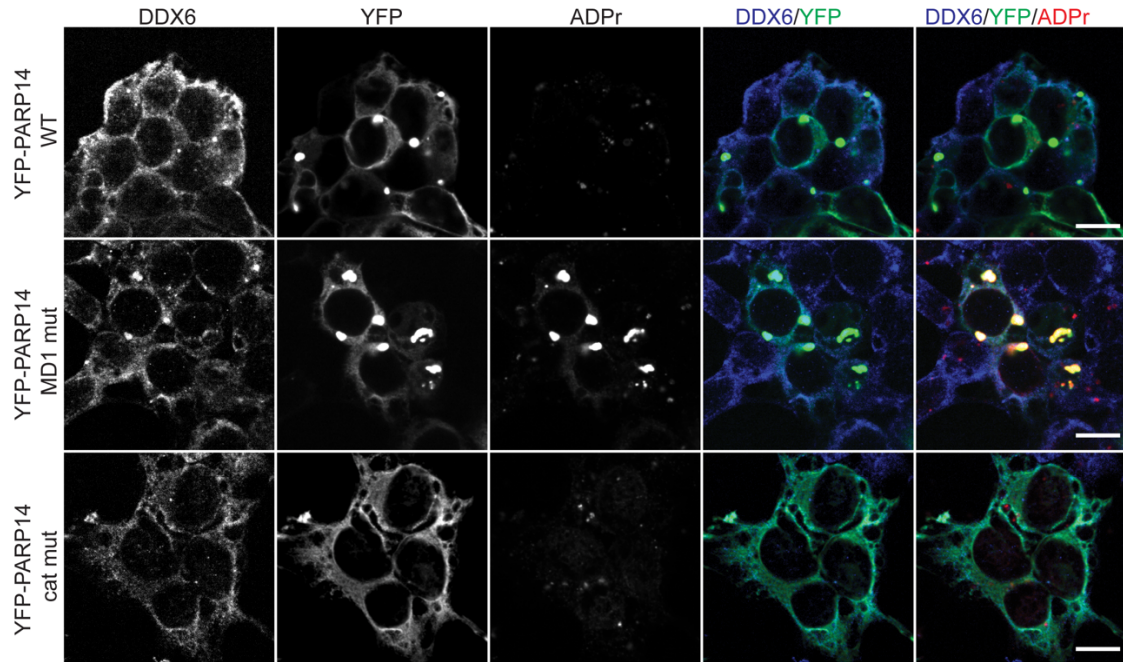


Supplementary Figure 3: PARP14 protein levels in cells and its activities on protein and nucleic acid substrates in cells. (A) A549, U2OS and 293T cells were examined for their PARP14 levels. 293T cells were stimulated with IFN γ (100 ng/mL) for 24 h or transfected with YFP-PARP14 WT. GFP was used to show expression of the YFP-tagged PARP14. GAPDH was used as a loading control. (B) Levels of overexpressed PARP14 are similar to levels of endogenous PARP14 after induction with IFN γ in U2OS cells. U2OS cells were stimulated with IFN γ (100 ng/mL) for 24 h or transfected with YFP-PARP14 WT. Phospho-STAT1 was used to demonstrate IFN γ stimulation. GAPDH was used as a loading control. (C) PARP14 is highly active and ADPr is reversed by its macrodomain 1. 293T cells were transfected with the indicated plasmids in the presence or absence of PARP14 inhibitor (PARP14i). Cell lysates and GFP-immunoprecipitations (GFP-IP) were examined by western blotting using the indicated antibodies. Tubulin and Ponceau S were used as a loading controls. (D) PARP14 activity on nucleic acid substrates is not detectable in cells. U2OS cells were transfected with the indicated plasmids. gDNA or total RNA was spotted on membranes to examine the levels of ADPr. (E) Endogenous PARP14 ADP-ribosylates protein substrates in both unstimulated and IFN γ stimulated cells. A549 cells, in the presence or absence of PARP14i, were stimulated or not with IFN γ (100 ng/mL). Cell lysates were probed with the indicated antibodies. Phospho-STAT1 was used to demonstrate IFN γ stimulation. GAPDH was used as a loading control.

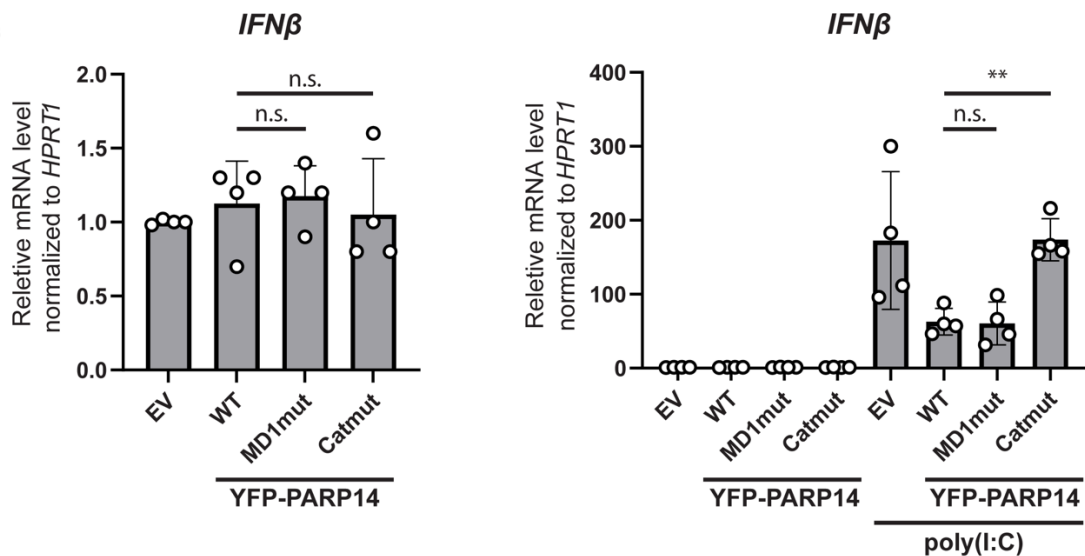


Supplementary Figure 4: Mass-Spec identification of ADP-ribosylated proteins regulated by PARP14 macrodomain 1. (A) Violin plots depicting the distribution of the coefficient of variation (CV) within the different experimental conditions. The flat line corresponds to the median CV, while the dots marks the mean CV of the distributions. Outliers were visually removed from the plot. Rows were considered to be outliers if their CV's exceed 1.5 times the inter-quantile-range (IQR) of the distribution. (B) Heatmap showing the inter-experiment Pearson correlation coefficients of the samples (C). Linear dimensionality reduction of the dataset by principal component analysis (PCA), displaying the clustering of each experiment. The x- and y-axis display the first two principal components, which capture 54.95% and 7.43% of the total variance of the dataset, respectively.

A



B



Supplementary Figure 5: PARP14 regulates various cellular functions. (A) PARP14 MD1 mutant colocalise with DDX6 and ADP-ribosylation. 293T cells were transfected with the indicated plasmids and examined by immunofluorescence with DDX6 (Blue), or ADPr (poly/mono) (red) antibodies. YFP expression is shown in Green. Scale bar: 10 μm. (B) The relative gene expression analysis of *IFNβ* in unstimulated and Poly(I:C) stimulated 293T cells transfected with PARP14 WT, PARP14 MD1 mutant or PARP14 catalytic mutant as determined by RT-qPCR normalized to the expression of *HPRT1*. Left panel: Unstimulated 293T cells. Right: 293T cells stimulated with Poly(I:C) for 24 hours. Data collected from cells without Poly(I:C) treatment is recapitulated to show the increase in expression. EV: empty vector (YFP), WT: YFP-PARP14 WT, MD1 mut: YFP- PARP14 MD1 mutant, Catmut: YFP-PARP14 catalytic domain mutant. Error bars indicate average S.D. from four independent replicates. Asterisks indicate statistical significance compared with the control, as determined by Welch's t-test (ns: not significant, ** : $p < 0.01$, Two-tailed P value, WT vs Catmut, $p = 0.0012$).

Supplementary Tables:

Supplementary Table 1. Mono-ADP-ribosylated peptides used in this study.

| Peptide | Sequence |
|----------------|--------------------------------------|
| Arginine-ADPr | Ac-GR(ADPr)LIFAG-OH |
| Serine-ADPr | Ac-PAKS(ADPr)APAPKKG-NH ₂ |
| Glutamate-ADPr | H-AAPVE(ADPr)VVAPR-NH ₂ |

Supplementary Table 2. Oligos used in this study.

| Name | Sequence (5'-3') |
|-----------------------------------|--|
| 5P RNA 3Cy3 | [Phos] GUGGCGCGGAGACUUAGAGAA [Cy3] |
| 5Cy3 RNA 3P | [Cy3] GUGGCGCGGAGACUUAGAGAA [Phos] |
| 5P DNA3 Cy3 | [Phos] GTGGCGCGGAGACTTAGAGAA [Cy3] |
| Short21 | TTCTCTAAGTCTCCGCGCCAC |
| cn2 | TTCTCTAAGTCTCCGCGCCACTAAACCGCGCCCCTTAAGG |
| cn6 | GGAATTCCCCGCGCCAAATTTCTCTAAGTCTCCGCGCCAC |
| cn7 | TTTGGCGCGGGGAATTCC |
| cn8 | ATTTGGCGCGGGGAATTCC |
| cn10 | AAGAACAAGTCTCCGCGCCAC |
| cn11 | TTCTCTAAGTCTCCGAGACGA |
| cn12 | CCTTAAGGGGCGCGGTTT |
| cn13 | CCTTAAGGGGCGCGGTTTA |
| ssDNA for competition assay | [Phos]GTGGCGCGGAGACTT |
| 5P E21 ssDNA | [Phos] GTGGCGCGGAGACTTAGAGAA [Cy3] |
| 3P E21 ssDNA | [Cy3] GTGGCGCGGAGACTTAGAGAA [Phos] |
| RNA oligo for RhsP2 | AUCUACGGUACCUCUGGCUACGACGACAGGCGCUAAUCAGACUCCGACUG |

Supplementary Table 3: Primers used in this study.

| Name | Sequence (5'-3') |
|-----------------------|-------------------------------------|
| PARP14 R1699A Forward | TTTAACGCCAGCTATGCCGGAAGAATGCTGTG |
| PARP14 R1699A Reverse | GTCAATCGAAATGGCTTTAACGCCAGCTAT |
| PARP14 G832E Forward | CTTAAGCATTATGGTGTAGCTGGCCGCTGCGCTCT |
| PARP14 G832E Reverse | AGAGCGCAGCGGCCAGCTCACCATAATGCTTAAG |
| PARP14 G1044E Forward | CTCGTGCTTAGTAGAGAGCCTCTTTCTAAGTCCC |
| PARP14 G1044E Reverse | GGGACTTAGAAAGAGGCTCTCTACTAAGCACGAGA |
| PARP9 G113E Forward | GCATGGGGGAGAAGTGGCCCTGG |
| PARP9 G113E Reverse | AGAAGATCTTCATTGGCTGCATTC |

Supplementary Table 4: Antibodies used in this study.

| Target | Host | Company | Reference | Dilution in WB | Dilution in IF |
|-------------------------------|--------|--------------------------|---------------------------|----------------|----------------|
| GFP | Rabbit | Abcam | Ab290 | 1:3000 | - |
| PARP14 | Rabbit | Abcam | Ab229756 | 1:1000 | - |
| β -tubulin | Rabbit | Abcam | Ab6046 | 1:2000 | - |
| DDX3 | Mouse | Proteintech | 67915-1-Ig | 1:1000 | - |
| HDAC2 | Mouse | Santa Cruz Biotechnology | sc-9959 | 1:1000 | - |
| DDX6 | Rabbit | Proteintech | 14632-1-ap | 1:1000 | - |
| GAPDH | Mouse | Merck | MAB374 | 1:3000 | - |
| RPA2 | Rabbit | Cambridge Bioscience | A300-244A | 1:2000 | - |
| Histone H3 | Rabbit | Sigma-Aldrich | 06-755 | 1:10000 | - |
| FLAG | Rabbit | Sigma-Aldrich | F7425 | 1:2000 | - |
| autoanti-dsDNA | Mouse | DSHB | AB_10805293 | 1:200 | - |
| Phosphor-STAT1 (Tyr701) | Rabbit | Cell Signaling | 7649 | 1:1000 | |
| DDX6 | Mouse | Abnova | h00001656-m01 | - | 1:500 |
| GFP | Goat | Abcam | Ab5450 | - | 1:500 |
| Poly/mono | Rabbit | Cell Signaling | 83732 | 1:1000 | 1:500 |
| Anti-mouse Alexa Fluor 405 | Donkey | Thermo Fisher Scientific | A48257 | - | 1:500 |
| Anti-goat Alexa Fluor 488 | Donkey | Thermo Fisher Scientific | A11055 | - | 1:500 |
| Anti-rabbit Alexa Fluor 647 | Donkey | Thermo Fisher Scientific | A32795 | - | 1:500 |
| HRP conjugated anti-Mono-ADPr | - | - | Gift from Ivan Matic (53) | 1:1000 | - |
| HRP-conjugated anti-mouse | Goat | Agilent | P0447 | 1:3000 | - |
| HRP-conjugated anti-rabbit | Swine | Agilent | P0399 | 1:3000 | - |

Supplementary Table 5: RT-qPCR primers used in this study.

| Gene | Forward (5'-3') | Reverse (5'-3') |
|-------------|-----------------------|-----------------------|
| IFN β | TAGCACTGGCTGGAATGAG | GTTTCGGAGGTAACCTGTAAG |
| HPRT1 | GCGTCGTGATTAGCGATGATG | CTCGAGCAAGTCTTTCAGTCC |

Other Supplementary Material for this manuscript includes:

Supplementary Data 1: Table with Af1521-enriched proteins

Supplementary Data 2: Table with MD1-specific ADPr-targets

Supplementary Data 3: Table with gene set enrichment analysis statistics