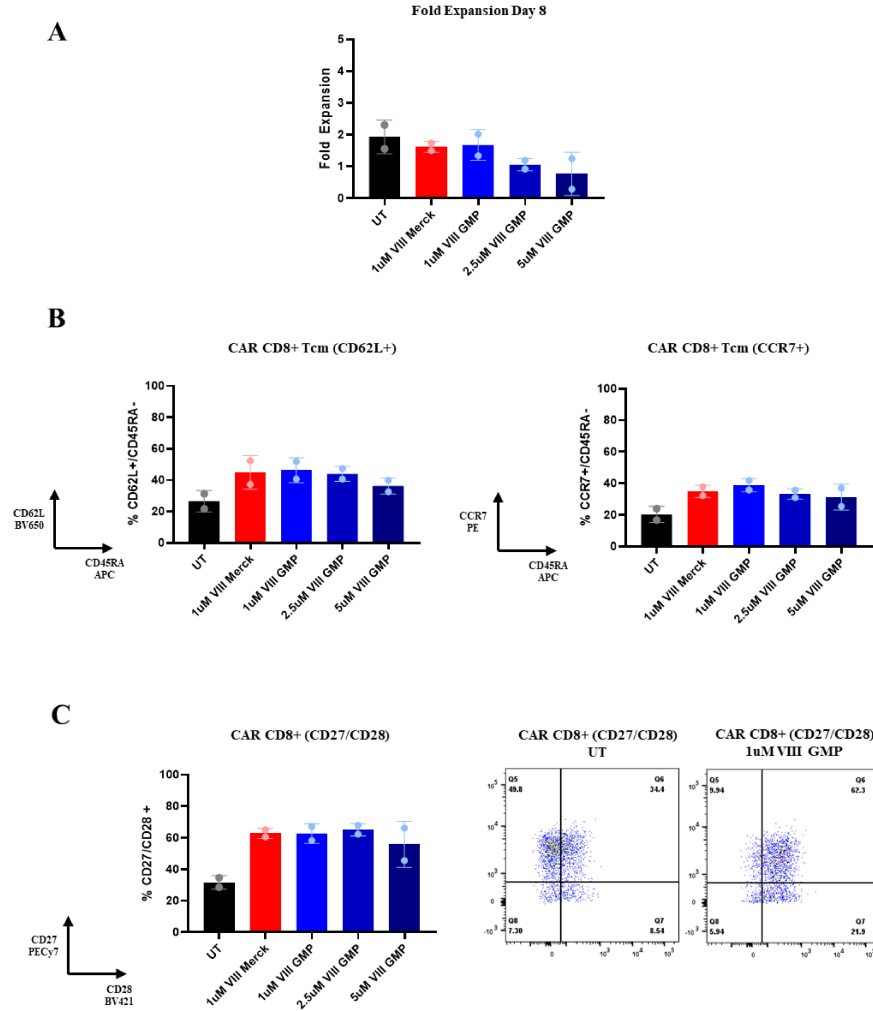
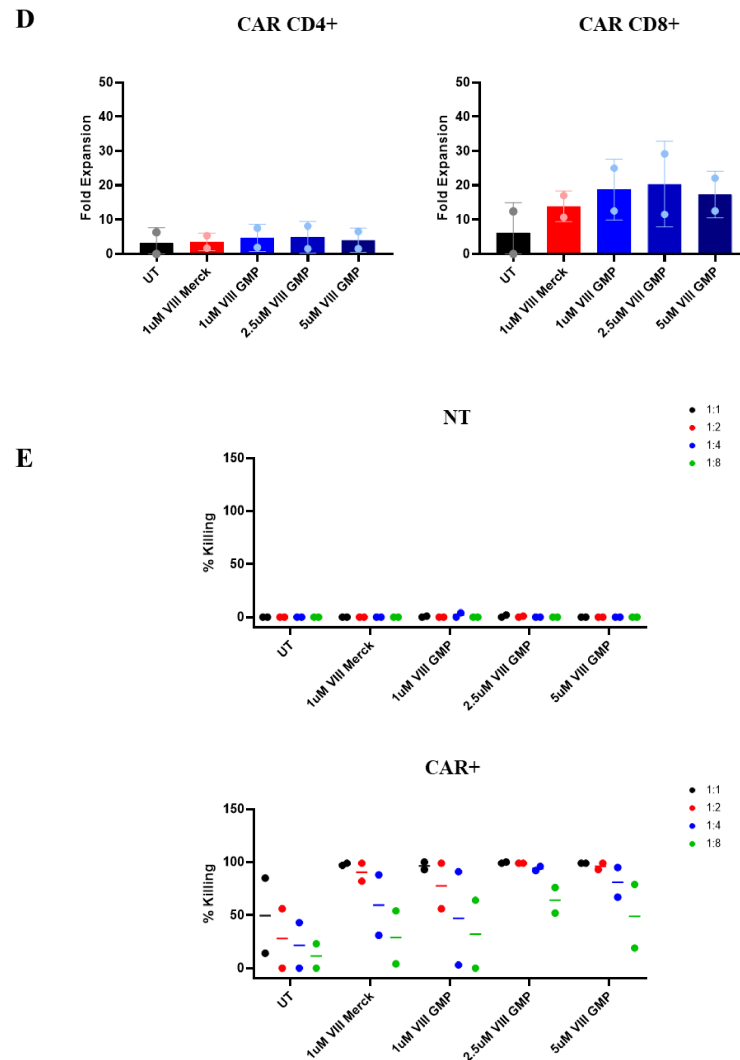


**Supplementary Figure 1**



## Supplementary Figure 1

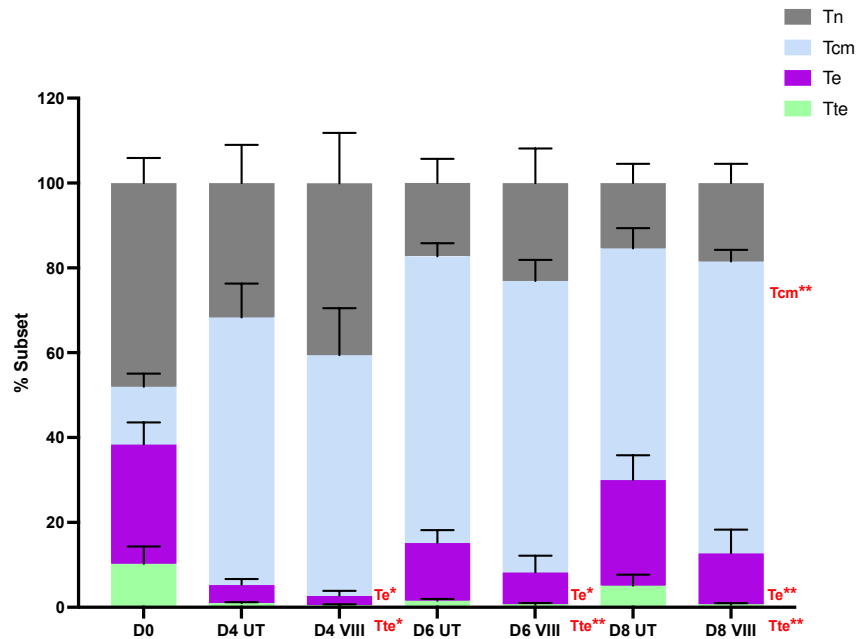


**Supplementary Figure 1. Comparison of VIII (Merck) vs VIII-GMP (Ardena) and titration of VIII-GMP.**

(A) Total T-cell fold expansion by the end of manufacture at Day 8 (B) % Tcm subset in CAR CD8s at Day 8, determine by flowcytometry as (CD62L+/CD45RA-) and (CCR7+/CD46RA-). (C) Extended phenotyping depicting % CD27+/CD28+ cells characterised by flowcytometry. Representation flowcytometry plots from one donor in the UT and 1µM VIII-GMP are depicted to the right. (D) CD4/CD8 CAR fold expansion following a 7-day co-culture with irradiated RAJI-19WT

target cell lines. (E) Graphs depicting % killing of RAJI-19GFP target cells post rechallenge of NT or CAR T-cells in a 72-hour killing assay. Results from all conditions were normalised to the untreated non-transduced (NT) condition at each E:T ratio. All data sets compare 1 $\mu$ M VIII-Merck and 1 $\mu$ M VIII-GMP. VIII GMP is further titrated to 2.5 $\mu$ M and 5 $\mu$ M at n=2 per condition  $\pm$  SD and/or individual data points.

## Supplementary Figure 2

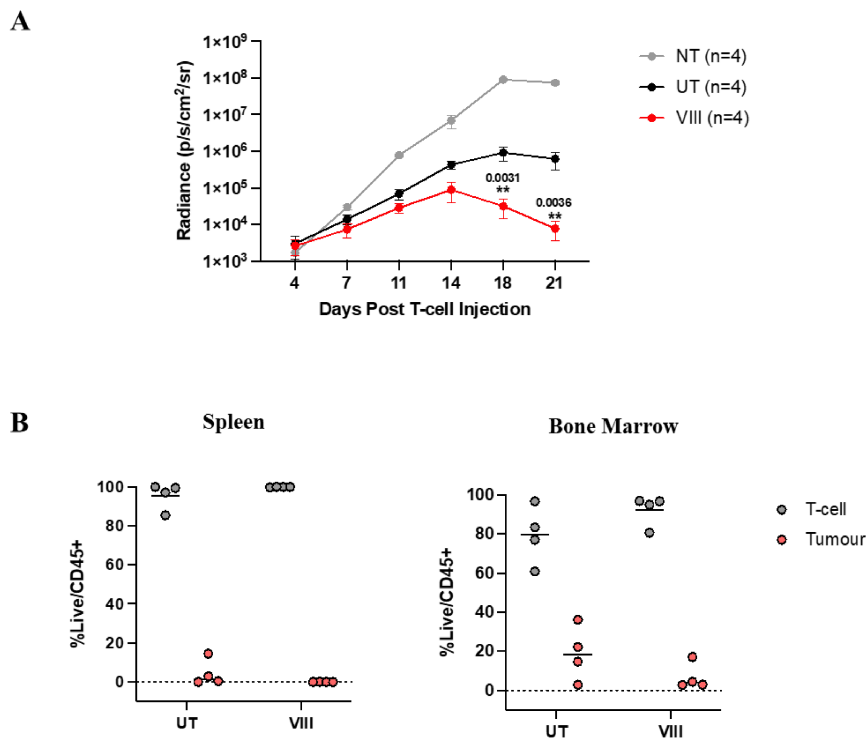


**Supplementary Figure 2. Summary of CAR-T phenotype throughout manufacture.**

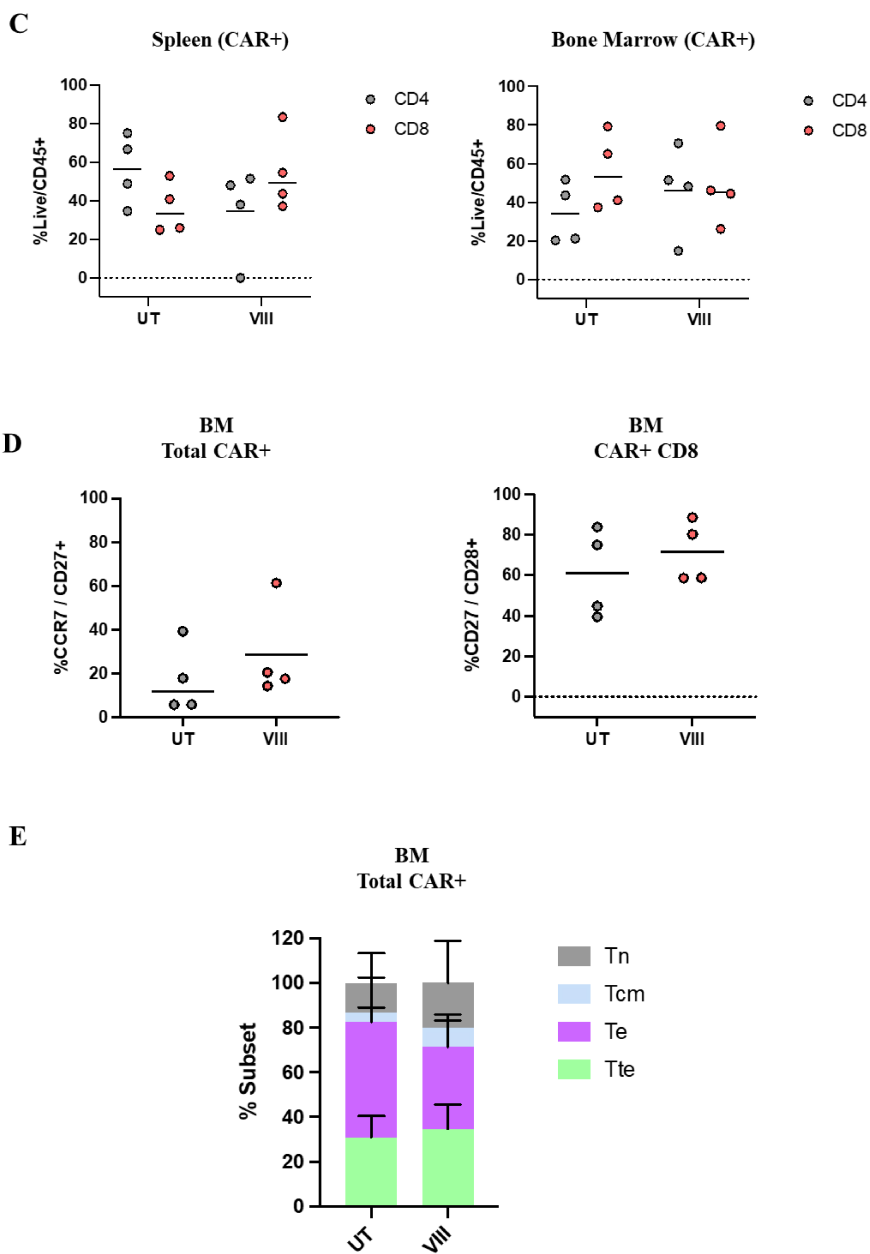
Graphical representation of Tn (CCR7+/CD45RA+), Tcm (CCR7+/CD45RA-), Te (CCR7-/CD45RA-) and Tte (CCR7-/CD45RA+) subsets in total CAR T-cells determined by flowcytometry,  $\pm$  SD at baseline and throughout manufacture. Statistical comparisons were made against paired UT sample in each subset at matched timepoints, n=6. Two tailed Mann-Whitney U test, ns  $P > 0.05$ , \* $P < 0.05$  and \*\* $P < 0.01$ .

**Method:** Systemic leukaemia was established via intravenous injection of  $5 \times 10^5$  NALM6-FLUC followed by  $5 \times 10^5$  non-transduced (NT) or CAR T-cells 4 days later. Tumour burden was measured bi-weekly via bioluminescent imaging (BLI) using the IVIS spectrum in-vivo imaging system (Perkin Elmer) following intraperitoneal (IP) injection of 2mg D-luciferin in 200 $\mu$ l PBS. Photon emission from NALM6 cells was measured as photons/sec/cm<sup>2</sup>/steradian. Mice were humanely euthanised Day 21 and spleen/bone marrow samples were harvested in cold Hanks Balanced Salt Solution (HBSS) (Sigma Aldrich) for downstream analysis. Spleens were gently pressed over a 70 $\mu$ m and subsequently a 30 $\mu$ m cell strainer to dissociate. Bone marrow was flushed with PBS from femoral shafts over a 70 $\mu$ m and subsequently a 30 $\mu$ m cell strainer. CountBright™ beads were added to samples to determine absolute cell numbers, and flowcytometry was performed on the BD LSRFortessa™ and analysed using FlowJo v10 software.

Supplementary Figure 3



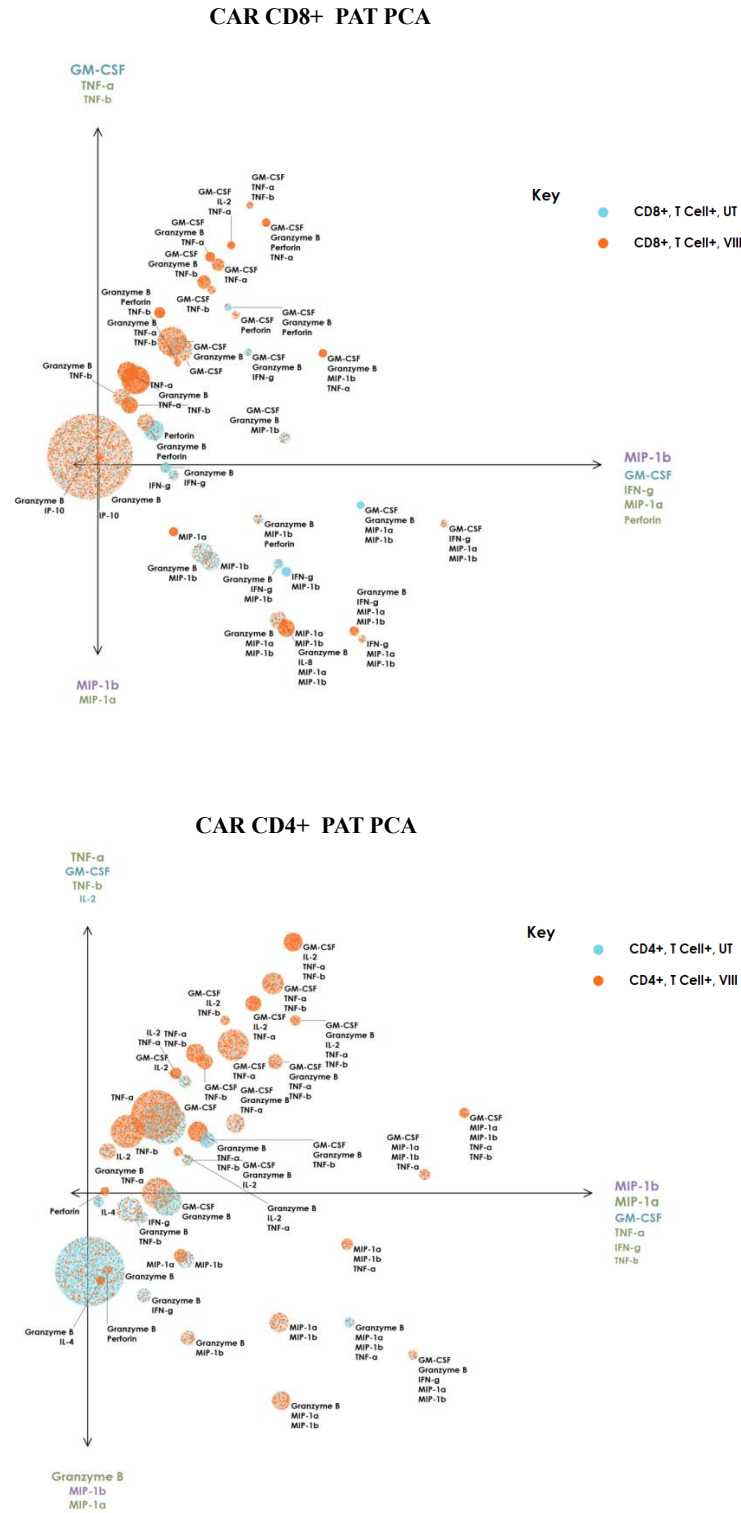
**Supplementary Figure 3**



***Supplementary Figure 3. In-vivo model and CAR T-cell analysis from mouse spleen and bone marrow at Day 21.***

(A) Tumour burden measured by bioluminescent imaging (BLI) in NALM6 tumour established mice treated with non-transduced (NT) T-cells or CAR T-cells manufactured with (VIII) or without (UT)  $\pm$  SEM (B) Residual percentages of total T-cell and tumour cells in mouse spleen and bone marrow at Day 21, depicting individual data points (C) Percentage of CAR+ CD4/8 in spleen and bone marrow at Day 21 in NALM6 tumour established mice treated with UT/VIII CAR T-cells, depicting individual data points. (D) Frequencies of CCR7+/CD27+ in total CAR T-cells and CD27+/CD28+ in CD8 CAR T-cells in bone marrow of mice treated with UT/VIII CAR T-cells, depicting individual data points. (E) Graphical representation of Tn (CCR7+/CD45RA+), Tcm (CCR7+/CD45RA-), Te (CCR7-/CD45RA-) and Tte (CCR7-/CD45RA+) subsets in total CAR T-cells from bone marrow determined by flowcytometry,  $\pm$  SD. (A-E) Cells derived from one healthy donor, n=4 mice per group. (A) Two-way ANNOVA corrected for multiple comparisons by Tukey's test on log transformed data, ns P>0.05 and \*\* P<0.01. (B-C) Two-way ANNOVA corrected for multiple comparisons by Bonferroni's test, ns P>0.05. (D-E) Two tailed Mann-Whitney U test, ns P>0.05.

Supplementary Figure 4

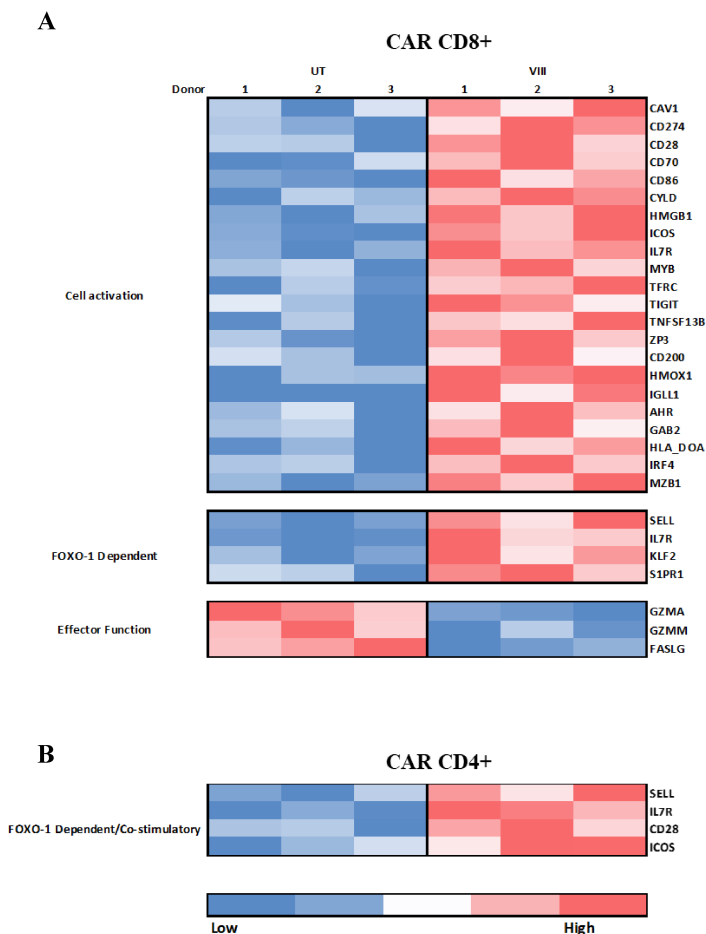




***Supplementary Figure 4. PAT PCA of polyfunctional profiles.***

Polyfunctionality Activity Topography (PAT), Principal Component Analysis (PCA) of CD4 and CD8 T-cell subsets demonstrating primary polyfunctional profiles where radius is proportional to secretion frequency. Determined using Isoplexis™ platform, following 20-hour 2:1 RAJI-19WT:CAR stimulation, n=4.

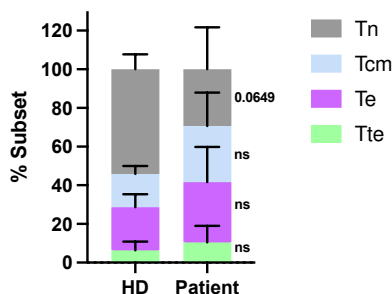
## Supplementary Figure 5

*Supplementary Figure 5. CAR T-cell transcriptome signature*

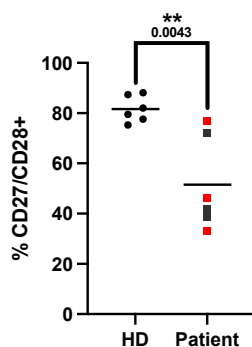
(A-B) Heat map representation of transcripts specific to cell activation, FOXO1 and effector function in CD8 CAR T-cells and FOXO1/co-stimulatory transcripts in CD4 CAR T-cells. Colour represents low to high transcript expression. Rows are specific to each gene and columns highlights expression in each donor following UT/VIII manufacture. Only significantly differentially expressed pathways and genes are included in figures at a  $p < 0.05$  cut off.

## Supplementary Figure 6

A



B

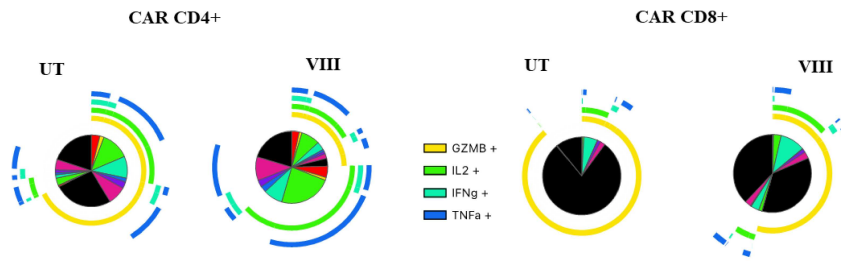


**Supplementary Figure 6. Phenotype analysis of baseline T-cells derived from healthy donor and B-ALL patients.**

(A) Graphical representation of Tn (CCR7+/CD45RA+), Tcm (CCR7+/CD45RA-), Te (CCR7-/CD45RA-) and Tte (CCR7-/CD45RA+) subsets in total T-cells from healthy donor (HD) and B-ALL patients at baseline determined by flowcytometry,  $\pm$  SD. (B) Percentage CD27+/CD28+ in total T-cells from healthy donor (HD) and B-ALL patients at baseline, depicting individual data points. Black squares in patient group represent patients in remission and red squares represent patients with CD19+ relapse. (A-B)  $n=6$ , Two tailed Mann-Whitney U test, ns  $P>0.05$  and \*\*  $P<0.01$ .

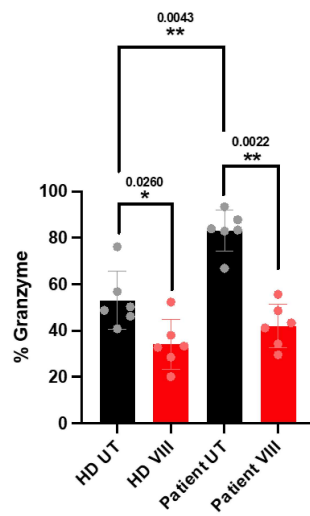
**Supplementary Figure 7**

**A**

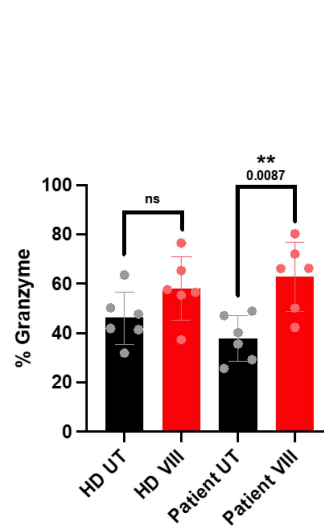


**B**

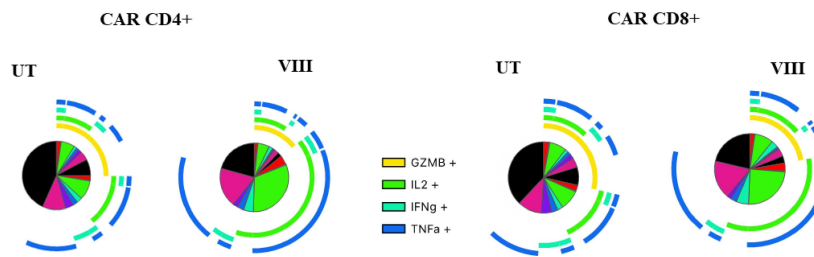
**End of Manufacture  
B-ALL Patients Small-scale  
GZMB**



**Rechallenge  
B-ALL Patients Small-scale  
GZMB**

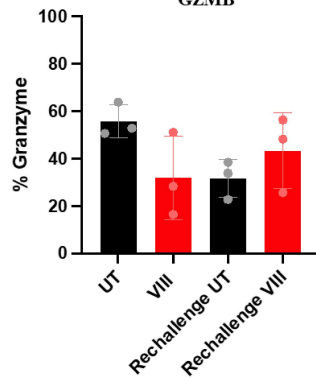


**C**



**D**

**End of Manufacture and Rechallenge  
B-ALL Patients Prodigy-scaled  
GZMB**



**Supplementary Figure 7. Intracellular cytokine production**

(A) Pie chart of proportion of intracellular cytokines measured by flowcytometry following RAJI-19WT stimulation in B-ALL patient CAR-T manufactured at small-scale. Arcs represent proportion of single or polyfunctional secretion. (B) Percentage of total CAR-T producing GZMB in healthy donor (HD) or B-ALL patient in small-scale assessments at the end of manufacture or at rechallenge using CAR-T co-cultured with RAJI-19WT targets 7-days prior,  $\pm$  SD. (C) Pie chart of proportion of intracellular cytokines measured by flowcytometry following RAJI-19WT stimulation in B-ALL patient CAR-T manufactured at Prodigy scale. Arcs represent proportion of single or polyfunctional secretion. (D) Percentage of total CAR-T producing GZMB in B-ALL patient Prodigy-scale assessments at the end of manufacture or at rechallenge using CAR-T co-cultured with RAJI-19WT targets 7-days prior,  $\pm$  SD. (A-B) n=6. (C) n=3 (B) Two tailed Mann-Whitney U test, ns  $P>0.05$ . \* $P<0.05$  and \*\*  $P<0.01$ . (D) n=3, Two tailed Mann-Whitney U test, ns  $P>0.05$ .