

## Supplementary Information

# Structural insights into functional properties of the oxidized form of cytochrome *c* oxidase

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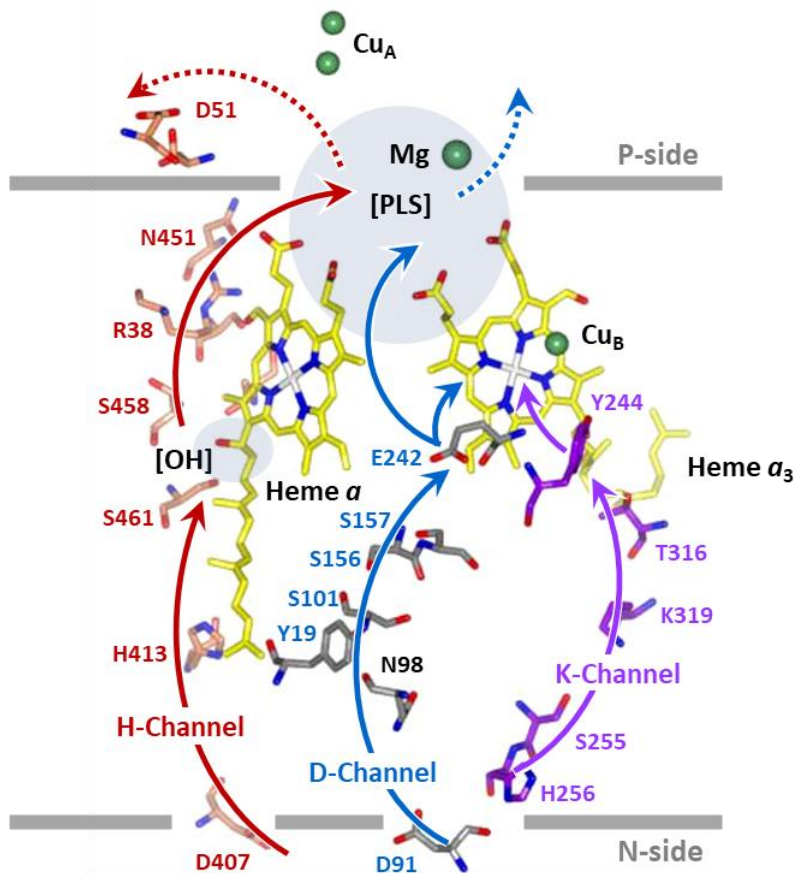
The PDF file includes:

Supplementary Figures 1 to 3  
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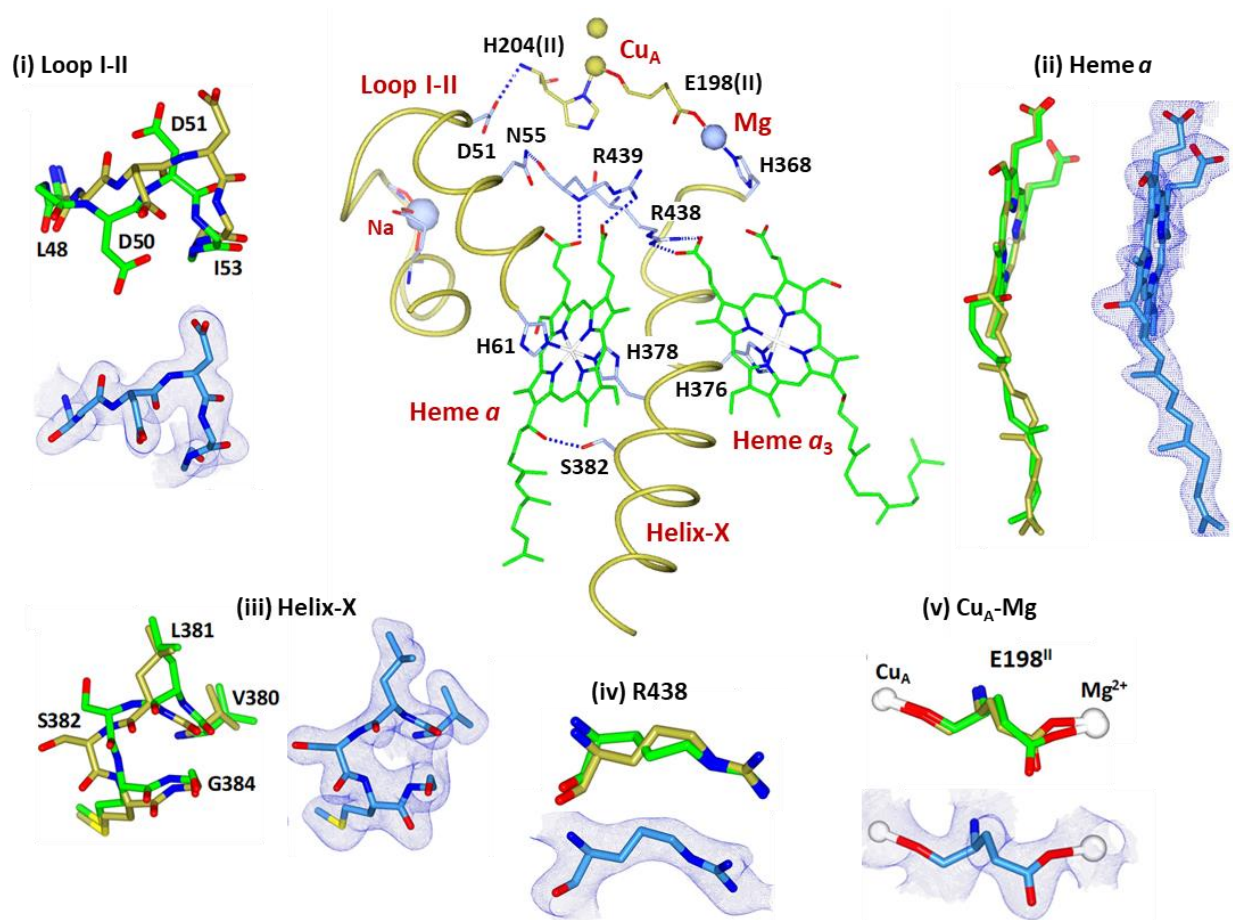
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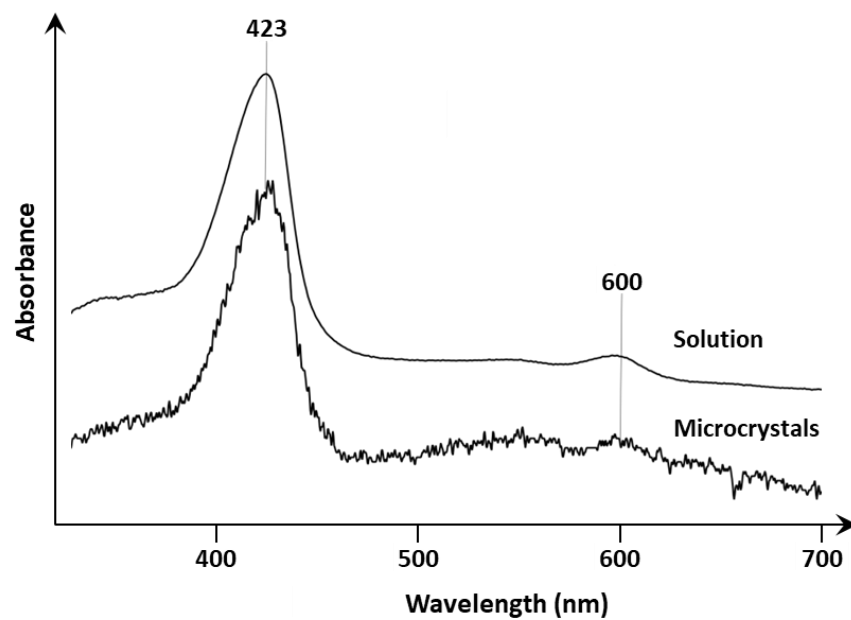
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**Supplementary Fig. 1. Proton transfer channels in bCcO.** Based on structural and mutagenesis studies of bacterial enzymes, two proton transfer channels, the D and K-channels, have been proposed in CcO<sup>1</sup>. The D-channel, starting at D91 (bCcO numbering) and ending at E242 (which serves as a diverter valve), delivers either substrate protons to the BNC during the oxidative phase of the catalytic cycle, or pumped protons to the PLS for subsequent translocation to the P-side of the membrane. The K-channel, starting at H256 and ending at Y244, delivers substrate protons to the BNC during the reductive phase of the catalytic cycle. An additional channel, the H-channel, starting at D407 and ending at D51, was proposed in bCcO, which translocates pumped protons to the P-side of the membrane *via* the PLS, as gated by either a water pool<sup>2</sup> or the hydroxide group of the farnesyl sidechain (indicated as [OH])<sup>3</sup> of heme *a*. The role of the H-channel, however, remains controversial as it is not conserved in bacterial CcOs<sup>4,5</sup>.



**Supplementary Fig. 2. SFX structure of the O derivative of bCcO in the redox sensitive regions.** Inset (i)-(v) show the structural comparison of the oxidized enzyme determined with synchrotron radiation (**O\***) (in yellow) and reduced enzyme (**R**) (in green) in the redox sensitive regions. The  $2F_o - F_c$  electron density map of **O**, contoured at  $1.5 \sigma$ , is shown in each inset to demonstrate that the structure of **O**, consistent with that of **O\***, is in the oxidized state. The PDB IDs of the **O\*** and **R** are 7TIE and 7THU, respectively.



**Supplementary Fig. 3. UV-Vis absorption spectra of oxidized bCcO in the free solution phase and in the microcrystalline state.** The Soret maximum at 423 nm indicates that the bCcO samples are in the same fast (active) form.<sup>6</sup>

**Supplementary Table 1.** Crystallographic data collection and refinement statistics (molecular replacement).

	8GCQ
<b>Data collection</b>	
Wavelength(Å)	1.24
Temperature (K)	293
beamline	SLAC LCLS MFX
Space group	P 2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>
Cell dimensions	
<i>a</i> , <i>b</i> , <i>c</i> (Å)	178.60, 189.50, 211.10
α, β, γ (°)	90.00, 90.00, 90.00
Resolution (Å)	33.00-2.38(32.80-2.38) *
<i>R</i> <sub>sprit</sub>	0.2296(0.6388)
<i>I</i> / σ <i>I</i>	4.59(1.78)
Completeness (%)	100(100)
Redundancy	2742(2286)
<b>Refinement</b>	
Resolution (Å)	35.00-2.38
No. reflections	270,854
<i>R</i> <sub>work</sub> / <i>R</i> <sub>free</sub>	0.232 / 0.266
No. atoms	31,465
Protein	28,514
Ligand/ion	2,350
Water	601
<i>B</i> -factors	34.79
Protein	32.42
Ligand/ion	64.39
Water	31.74
R.m.s. deviations	
Bond lengths (Å)	0.0114
Bond angles (°)	1.742
Ramachandran statistics (%)	
Favored	89.24
Allowed	9.73
Outliers	1.03

\*Values in parentheses are for highest-resolution shell.

## Supplementary References

- 1 Wikstrom, M., Krab, K. & Sharma, V. Oxygen Activation and Energy Conservation by Cytochrome c Oxidase. *Chemical reviews* **118**, 2469-2490, doi:10.1021/acs.chemrev.7b00664 (2018).
- 2 Shimokata, K. *et al.* The proton pumping pathway of bovine heart cytochrome c oxidase. *Proceedings of the National Academy of Sciences of the United States of America* **104**, 4200-4205, doi:10.1073/pnas.0611627104 (2007).
- 3 Ishigami, I. *et al.* Snapshot of an Oxygen Intermediate in the Catalytic Reaction of Cytochrome c Oxidase. *Proc Nat Acad Sci (USA)* **116**, 3572-3577 (2019).
- 4 Qin, L. *et al.* Redox-dependent conformational changes in cytochrome C oxidase suggest a gating mechanism for proton uptake. *Biochemistry* **48**, 5121-5130, doi:10.1021/bi9001387 (2009).
- 5 Yoshikawa, S. & Shimada, A. Reaction mechanism of cytochrome C oxidase. *Chemical reviews* **115**, 1936-1989, doi:10.1021/cr500266a (2015).
- 6 Baker, G. M., Noguchi, M. & Palmer, G. The reaction of cytochrome oxidase with cyanide. Preparation of the rapidly reacting form and its conversion to the slowly reacting form. *The Journal of biological chemistry* **262**, 595-604 (1987).