SUPPLEMENTAL INFORMATION

SINGLE-CELL MULTIOMICS OF HUMAN FETAL HEMATOPOIESIS DEFINES A DEVELOPMENTAL SPECIFIC POPULATION AND A FETAL SIGNATURE

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- Supplemental Figures 1–6
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SUPPLEMENTAL FIGURES



Supplemental Figure 1

Individual donor contribution and verification of cluster identities

(A) Sorting strategy of CS22 LIN⁻CD45⁺CD34⁺ FL cells. Viable cells (7-AAD⁻) were selected based on size (scatter) and gated negative for mature lineage markers (CD3, CD235a). Further gating is indicated in the figure. (B) UMAP of LIN⁻CD45⁺CD34⁺ FL cells at CS22, as per donor (*left*) and bar plot of donor contribution per cluster (percent; *right*). (C) Immunophenotypic defined HSPCs from *Ranzoni et al* [12] were projected on the CS22 UMAP in *Figure 1B*. Distribution per cluster of each population is shown. In total 87-97% of the cells from the different populations mapped.



Integration of ADTs and transcriptome data and immunophenotypic gating strategy (A) The ADTs and the transcriptome data were integrated using WNN. UMAPs showing WNN clusters (*left*) and the FL clusters defined in *Figure 1B* (*right*). (B) Bead plot of z-scored ADT expression values per cluster, where size of the circle represents fraction of cells with expression and the color correlates with expression level. (C) Gating of conventional immunophenotypic populations using ADT signals (cells were sorted LIN⁻CD45⁺CD34⁺ as shown in *Supplemental Figure 1A*). Top row CLP gating, middle row myeloid progenitors (CMP, GMP, MEP) and bottom row primitive populations (HSC, MPP, LMPP). (D) Transcriptional cell states captured for each immunophenotypic gated population are shown within the contours on the FL CS22 UMAP. Pie charts show percentage of the different molecular clusters (as per *Figure 1B*) within each immunophenotypic gated progenitor. (E) PC2 vs PC3 of pseudo-bulked ADT gated cell populations using top 500 variably expressed genes (2 replicates/donor). (F) Combined PCA of pseudo-bulked ADT gated cell populations and clusters (defined in *Figure 1B*), top 500 variably expressed genes. PC2 vs PC3 is shown. (For immunophenotypic definition of populations *see Materials and Methods*.)



Projection of primitive FL and CB cells on adult BM reference

(A) UMAP of LIN⁻CD34⁺ adult BM cells with cluster annotations from [19]. (B) PCA of pseudo-bulked immunophenotypic populations from FL CS22 and adult BM using 500 top variably expressed genes. PC1 vs PC2 (*left*) and PC2 vs PC3 (*right*). (C) Projection of FL CS16, FL CS22, FL 9 pcw and CB on the adult BM UMAP. Size of dots represents mapping-score

and color of dots represents BM clusters. (D) Quantification of projected cells (*Supplemental Figure 3C*) in all mapped developmental stages. (E) Contribution (percent) of individual donors to the mapping in *Supplemental Figure 3C-D*. (F) Classification of FL CS22 derived clusters (x-axis) on adult BM.



Integration of FL CS22 and adult BM transcriptome data

(A) UMAP of cellular states after integration of FL CS22 and adult BM using Harmony. (B) Harmony UMAP of individual donors (*left*) and contribution of individual donors per cluster (percent, *right*). (C) Harmony UMAP displaying only the CS22 FL cells with Harmony clusters

(*left*) and FL clusters as per *Figure 1B* (*mid*). Bar plot (*right*) shows distribution of FL clusters per Harmony cluster. (D) Harmony UMAP displaying only the adult BM cells with Harmony clusters (*left*) and BM clusters as per *Supplemental Figure 3A* (*mid*). Bar plot (*right*) shows distribution of BM clusters per Harmony cluster. (E) Predicted cell cycle phase for each FL CS22 cluster. (F) GSEA of pseudo-bulked clusters comparing MPP-I to HSC and MPP-II respectively. Color code according to NES (Normalized Enrichment Score) value; red; upregulated and blue; downregulated in MPP-I. Selected gene sets with FDR q-value of <0.05 are shown.



Differential gene expression analysis comparing CS22 FL and adult BM counterparts

(A-B) PCA of pseudo-bulked clusters from FL CS22 and adult BM counterparts using 500 top variably expressed genes. Colors represent clusters and shapes represents sample type (*circle*: FL CS22 and *triangle*: adult BM). (A) PC1 vs PC2; (B) PC2 vs PC3. (C) Venn diagrams of downregulated genes in FL CS22 compared to adult BM counterparts defined by log2 fold change < -1 and adj. P value <0.05. (D) Validation of the *fetal core signature* using data set [15]. *Left*: PCA of 500 top variably expressed genes; *mid*: as left but eliminating *fetal core signature* genes; *right*: PCA of *fetal core signature* genes. (E) Venn diagrams showing overlap between *fetal core signature* and DEGs detected without pseudo-replicates. (F) PCA of gene set generated without pseudo-replicates using data set [15]. (G) Venn diagrams showing overlap between *fetal core signature* and DEGs detected using a single cell method (log2 fold change <[0.5]). (H) PCA of gene set generated with single cell method using data set [15].



Fetal core signature in B-ALL patient samples

(A) PCA of B-ALL samples [18] before batch correction (*left*) and after batch correction (*right*), of top 500 variably expressed genes. Color coded according to sequencing type. (B) PC1 vs PC2 of *KMT2A::AFF1* (*MLL::AF4*) B-ALL using the *fetal core signature*. Age group color

code is shown to the right. (C) PCA of individual leukemia sub-types using the *fetal core signature* (*top*) and representative PCA plot using random genes (*bottom*). PC1 vs age. (D) Box plots showing z-scored mean expression of the *fetal core signature* per age group. Up and down regulated gene sets are shown separately. High hyperdiploid (*top*), *BCR::ABL1* (*bottom*), adj. P values <0.05 indicated in the figure. The boxes include first to third quartile and the line indicates the median. (E) Heatmap of the genes marking the top 25 highest and 25 lowest values of PC1 in *KMT2A::AFF1* B-ALL. Samples were hierarchically clustered based on their gene expression. Top rows describe sex and age, and gene names are shown to the right (*blue label: fetal signature; red label: adult signature*).

Sample	Sex	# Sorted cells	# Cells pre- filtering	# Cells used in analysis
FL CS16 Donor1	Female	906	210	109
FL CS16 Donor2	Male	244	63	34
FL CS22 Donor1	Female	10 000	4536	3692
FL CS22 Donor2	Male	10 000	3797	3040
FL 9pcw Donor1	Male	2 213	458	297
FL 9pcw Donor2	Male	4 946	1057	842

Supplemental Table 1: Samples included in the study.

Reactivity	Surface marker	Product	Clone	Isotype	Cat number	Lot	Company
Anti human	CD38	Totalseq- A0059	HIT2	mIgG1 k	94672	B254149	Biolegend
Anti human	CD90	Totalseq- A0060	5E10	mIgG1 k	94673	B254151	Biolegend
Anti human	CD117 (cKIT)	Totalseq- A0061	104D2	mIgG1 k	94674	B254153	Biolegend
Anti human	CD10	Totalseq- A0062	HI10a	mIgG1 k	94675	B254154	Biolegend
Anti human	CD45RA	Totalseq- A0063	HI100	mIgG2b k	94676	B254155	Biolegend
Anti human	CD123	Totalseq- A0064	6H6	mIgG1 k	94677	B254157	Biolegend
Anti human	CD127 (IL7RA)	Totalseq- A0065	A019D5	mIgG1 k	94678	B254159	Biolegend
Anti human	CD7	Totalseq- A0066	CD7-6B7	mIgG2a k	94679	B254161	Biolegend
Anti human	CD71	Totalseq- A0067	CY1G4	mIgG2a k	94680	B254162	Biolegend
Anti human	CD105	Totalseq- A0068	43A3	mIgG1 k	94681	B254163	Biolegend
Anti human	CD201 (EPCR)	Totalseq- A0069	RCR-401	mIgG1 k	94682	B254164	Biolegend
Anti human	CD49f	Totalseq- A0070	GoH3	rIgG2a k	94683	B254165	Biolegend
Anti human	BAH1 (cMLP)	Totalseq- A0071	BAH1	mIgG1 k	94684	B254166	Biolegend
Anti human	CD18	Totalseq- A0385	TSI/18	mIgG1 k	302121	B270525	Biolegend
Anti human	CD25	Totalseq- A0085	BC96	mIgG1 k	302643	B271394	Biolegend
Anti human	CD196	Totalseq- A0143	Go34E3	mIgG2b k	353437	B270546	Biolegend
Anti human	CD62L	Totalseq- A0147	DREG-56	mIgG1 k	304847	B272608	Biolegend
Anti human	CD4	Totalseq- A0072	RPA-T4	mIgG1 k	300563	B268766	Biolegend
Anti human	CD26	Totalseq- A0396	BA5b	mIgG2a k	302720	B261042	Biolegend
Anti human	CD32	Totalseq- A0142	FUN-2	mIgG2b k	303223	B263613	Biolegend
Anti human	CD42b	Totalseq- A0216	HIP1	mIgG1 k	303937	B271616	Biolegend
Anti human	CD45RO	Totalseq- A0087	UCHL1	mIgG1 k	304255	B269954	Biolegend
Anti human	CD9	Totalseq- A0579	H19a	mIgG1 k	312119	B271599	Biolegend
Anti human	Integrin B7	A0214	FIB504	rIgG2a k	321227	B269330	Biolegend
Anti human	CD93	Totalseq- A0446	VIMD2	mIgG1 k	336121	B270926	Biolegend
Anti human	CD61	Totalseq- A0372	VI-PL2	mIgG1 k	336423	B271605	Biolegend
Anti human	CD95	Totalseq- A0156	DX2	mIgG1 k	305649	B269386	Biolegend
Anti human	CD135	Totalseq- A0351	BV10A4H2	mIgG1 k	313317	B272437	Biolegend
Anti human	CD11c	Totalseq- A0053	S-HCL-3	mIgG2b k	371519	B270802	Biolegend
Anti human	CD52	Totalseq- A0033	HI186	mIgG2b k	316017	B260031	Biolegend

Supplemental Table 2: CITE-seq antibodies used.

		Totalseq-					
Anti human	CD36	A0407	5-271	mIgG2a k	96419	B267789	Biolegend
		Totalseq-					
Anti human	CD41	A0353	HIP8	mIgG1 k	303737	B271597	Biolegend
		Totalseq-					
Anti human	CD48	A0029	BJ40	mlgG1 k	336709	B260028	Biolegend
	GD1074	Totalseq-	11440	LOLI	220647	D070007	D. 1 1
Anti human	CD10/A	A0155	H4A3	mlgG1 k	328647	B273007	Biolegend
Anti humon	CD25	l otalseq-	E11	mIaC11	222407	D260227	Dialagand
Anti numan	CD35	A010/	EII	niigot k	555407	B209337	Biolegena
Anti human	CD155	A 0023	SKII A	mIaG1 k	337673	B260344	Biolegand
Anti numan	CD155	Totalseq_	SKIL4	Inigot k	337023	D207344	Diolegenu
Anti human	CD79b	A0187	CB3-1	mIøG1 k	341415	B270535	Biolegend
	CDTYO	Totalseq-	000 1	inigorik	511115	B2/0000	Diologena
Anti human	CD33	A0052	P97.6	mIgG1 k	366629	B259970	Biolegend
-		Totalseq-					6
Anti human	CD56	A0084	QA17A16	mIgG1 k	392421	B273249	Biolegend
		Totalseq-					
Anti human	CD44	A0125	BJ18	mIgG1 k	338825	B270931	Biolegend
		Totalseq-					
Anti human	CD11a	A0185	TS2/4	mIgG1 k	350615	B260299	Biolegend
		Totalseq-					
Anti human	CD54	A0217	HA58	mIgG1 k	353123	B269343	Biolegend
		Totalseq-					
Anti human	CD70	A0027	113-16	mlgG1 k	355117	B263558	Biolegend
	XX 1. 4	Totalseq-	LNH-	LOLI	204607	D0(170)	D. 1 1
Anti human	Hashtag 4	A0254	94/2M2	mlgG1 k	394607	B264726	Biolegend
A (1)	II 14 5	l otalseq-	LNH-	LC11	204600	D2(4725	D' 1 1
Anti human	Hashtag 5	A0255	94/2M2	migGTK	394609	B264/25	Biolegend
Anti human	Hashtag 6	1 otaiseq-	LINH- 04/2M2	mIaG1 k	304611	B264724	Biolegand
Anu numan	riasinag o	A0230	J NU	niigot k	394011	D204/24	Biolegenu
Anti human	Hashtag 7	A0257	04/2M2	mIaG1 k	304613	B264723	Biolegend
	Tasinag /	Totalseq.	I NH-	migurk	574015	D204723	Diolegenu
Anti human	Hashtag 8	A0258	94/2M2	mIgG1 k	394615	B264722	Biolegend

	CS22				
Gated populations	FL1	FL2			
HSC ^{CD49F}	17	83			
HSC ^{CD90}	44	56			
MPP	40	60			
LMPP	45	55			
CMP ^{CD123}	60	40			
CMP ^{CD135}	60	40			
GMP ^{CD123}	54	46			
GMP ^{CD135}	53	47			
MEP ^{CD123}	60	40			
MEP ^{CD135}	61	39			
CLP ^{CD10}	80	20			
CLP ^{CD7}	65	35			
CLP ^{IL7R}	55	45			

Supplemental Table 3: Contribution of individual donors (percent) to the gated populations (related to Figure 2C and Supplemental Figure 2D).

Supplemental Table 4: Fetal core signature upregulated genes (Fetal signature) (related to Figure 4D).

Fetal signature									
Primitive clusters	Ly cl	usters	My cl	My clusteres					
ANGPT1	AASDHPPT	KLHL9	ALKBH1	IGF2BP3	ANGPT1				
ARID3A	ACY3	KNOP1	ANGPT1	IGFBP2	CD7				
BAMBI	ADGRG5	KNSTRN	ANKRD46	IL6ST	CHD7				
CALCRL	AL590226.1	KPNA2	ARID3B	IPCEF1	CRNDE				
CCT8	ANGPT1	LCK	ASPM	ITGA4	CTHRC1				
CD200	ANKRD46	LIN28B	AURKA	KIF15	DLK1				
CD7	ARID3A	LTB	BACH1	KIF2C	FBLN1				
CHD7	ASPM	MAD2L1BP	BAMBI	KNL1	HSPA1A				
CLCN3	BACH1	MARCKS	BLM	KPNA2	HSPA1B				
CRNDE	BET1	MARCKSL1	BUB1	LAMC1	HSPA6				
CSF1R	BRCA2	MED21	BUB1B	LDAH	IGF2BP1				
CTHRC1	CD164	MOB4	CA8	LEF1	IGF2BP3				
DLK1	CD244	MS4A1	CACNA2D3	LIN28B	ITGA4				
FAM43A	CD3D	MT2A	CD200	LTB	LIN28B				
FBLN1	CD7	MTERF3	CD300C	MAPK6	TPM2				
GNG11	CDK1	МҮС	CD7	MARCKS					
GOLM1	CHD7	N4BP2	CDCA2	MCM10					
GSTM3	COMMD10	NOC3L	CDCA8	MDK					
GSTM5	CRNDE	NUP37	CDK1	MEG3					
HMGA2	CTHRC1	PDE3B	CHD7	MIR99AHG					
HSPA1A	CXORF21	PPP1R14A	CHORDC1	NCAPG					
HSPA1B	CYBB	PRKDC	CIP2A	PPAT					
HSPA6	DICER1	PSMB5	CRNDE	PPIL4					
IGF2BP1	DLGAP5	RBM7	CTHRC1	PRKCA					
IGF2BP3	DLK1	RIF1	DHRS9	PRSS21					
IGFBP2	DNMT3B	RIOK2	DIAPH3	PSAT1					
ITGA4	EIF5A	RMI1	DICER1	PUDP					
KDM5B	FAM110A	RNF146	DLAT	PWWP2A					
LIN28B	FBLN1	RNF219	DLK1	QPRT					
MARCKSL1	FBXO5	RRAS2	DNAJB1	RAB7B					
MYCN	FKBP4	RSRC2	DNMT3B	RDX					
NEK6	FSCN1	SAMSN1	DSCC1	RNF146					
NRIP1	GMNN	SCN3A	DTL	RNF219					
RGS10	GPR174	SLC39A10	FBLN1	RRM2					
SUPT16H	HACD3	SMARCAD1	FBLN2	SMARCA1					
SUSD3	HEMGN	SS18	FBXO17	SMARCAD1					

TPM2	HIST1H1D	ST18	FBXO5	STIP1	
TRH	HMGA1	SUPT16H	GNG11	SUSD3	
	HMMR	TAF2	GNPDA1	TPM2	
	HSPA1A	TESC	GOLM1	TRH	
	HSPA1B	TPM2	GSTM3	TRIP13	
	HSPA6	TPM4	HACD3	ZBTB16	
	HSPB1	TRMT11	HEATR1	ZGRF1	
	HSPD1	WDHD1	HIST1H1A		
	HSPE1	WDR43	HIST1H1B		
	HSPH1	ZC3H8	HIST1H1D		
	IFI16	ZNF217	HIST1H2BH		
	IGF2BP1	ZNF644	HMGA1		
	IGF2BP2		HMGA2		
	IGF2BP3		HPGDS		
	IPO7		HSPA1A		
	ITGA4		HSPA1B		
	ITM2A		HSPA6		
	KCNQ10T1		HSPH1		
	KDM5B		IGF2BP1		

Supplemental Table 5: Fetal core signature downregulated genes (Adult signature)(related to Supplemental Figure 5C).

Adult signature									
Primitive	e clusters	Ly clusters My clusters			1	All clusters			
ABCA2	PCDH9	ABHD15	KLF10	A1BG	GBP1	RUNX2	AC006480.2		
AC002454.1	PKN1	ABTB1	KLF13	A1BG-AS1	GBP2	S100A10	AC007952.4		
AC004130.1	PPP1R15A	AC004687.1	KLF4	AC004069.1	GLI4	S100A6	AC103591.3		
AC006480.2	PRKG2	AC006480.2	KLF6	AC004540.2	GOLGA8B	SARM1	AC114760.2		
AC007952.4	PRNP	AC007952.4	KLF7	AC004687.1	GSAP	SCAND1	AC245014.3		
AC020916.1	PROM1	AC008429.1	L3MBTL4	AC004865.2	HAGHL	SCD5	AKR1C3		
AC026369.3	PRR34-AS1	AC016866.1	LINC00226	AC006480.2	HDAC5	SEPT1	AL731577.1		
AC103591.3	PSMB8-AS1	AC087239.1	LINC00623	AC007325.4	HLA-A	SERPINB9	ANAPC2		
AC114760.2	PSMB9	AC103591.3	LINC01089	AC007952.4	HLA-B	SERPINE2	ANXA1		
AC245014.3	PTGER2	AC114760.2	LITAF	AC020916.1	HLA-C	SH3D21	AP002387.2		
AKR1C3	PTGER4	AC245014.3	LPXN	AC044849.1	HLA-DMA	SH3PXD2A	AREG		
AL136454.1	PTMS	ACSL1	LRRC26	AC092069.1	HLA-DMB	SLFN11	ARMH1		
AL731577.1	RARRES3	ACSM3	LRRK1	AC103591.3	HLA-DPA1	SMIM3	AVP		
ANAPC2	RENBP	ACTN1	MANIAI	AC114760.2	HLA-DPB1	SMIM6	B2M		
ANKRD28	RIPOR2	AHNAK	MAP2K2	AC133919.1	HLA-DQB1	SMKR1	CD74		
ANXA1	RNF24	AJ009632.2	MAP7	AC245014.3	HLA-DRA	SOCS3	CLEC2B		
AP002387.2	S100A11	AKR1C3	MCUB	AC245297.3	HLA-DRB1	SPATC1L	CRHBP		
AREG	S100A6	AL355075.4	MFSD6	ACTL10	HLA-F	SPRY1	CXCR4		
ARMH1	SCAMP5	AL592183.1	MIB2	ACTN1	HOTAIRM1	SQOR	DNTT		
ATF3	SCAND1	AL731577.1	MILR1	ADARB1	HOXA10	SRGN	DUSP1		
ATP1B1	SCD5	ALDH1A1	MME	ADGRG6	HOXA3	SRSF5	EMP1		
ATP2B1-AS1	SCN9A	ANAPC2	MRPL23	AHNAK	HOXA7	STAT4	EPDR1		
AVP	SDK2	ANXA1	MSRB3	AIG1	HSBP1L1	STK17B	FAAP20		
B2M	SEPT1	ANXA2	MT-ND3	AKR1C3	IDS	SUN2	FAM30A		
BTG2	SERPINE2	AP001453.2	MXD4	AL121944.1	IF127L2	SYNE3	FHL1		
C160RF54	SLC39A3	AP002387.2	MYLK	AL136454.1	IFI44	TAP1	FLT3		
C1ORF21	SLC40A1	AREG	MZB1	AL355075.4	IGHM	TCF7	FOS		
C22ORF34	SMIM6	ARHGAP45	N4BP2L1	AL731577.1	IGSF10	TGFB1	FOSB		
C3ORF80	SOCS2	ARL4C	NBEAL1	ALDH1A1	IL17RC	TMEM102	HLA-A		
C90RF139	SOCS3	ARMH1	NEAT1	ALDH2	IL18	TMEM107	HLA-B		
CARD16	SOD2 1	ASB13	NEGR1	ANAPC2	ISG20	TMEM14A	HLA-C		
CAT	SPARC	ATP10D	NFIC	ANKH	JAKMIP2	TMEM160	HLA-DMA		
CAVINI	SPNS3	ATP6V0E2	NFKBID	ANKRD28	JAM2	TNFRSF14	HLA-DMB		
CBX6	SPPL2B	AVP	NPC2	ANXA1	JUND	TNFSF13B	HLA-DPA1		
CD69	SQOR	B2M	NPDC1	ANXA2	KLF10	TNNT3	HLA-DPB1		
CD74	STK17R	R4D	ND1112	ANVAD	VIE12	TOPI	HIA DORI		

Adult signature

CD9	SUN2	BLNK	NR4A2	AP002387.2	KLF2	TPD52	HLA-DRA
CDCP1	TGFB1	BLVRA	OPTN	AP002748.3	KLF4	TPGS1	HLA-DRB1
CEBPA	TMEM160	BSPRY	PARP10	AREG	KLF6	TRADD	HLA-F
CLEC2B	TMEM246	BST1	PBXIP1	ARL4A	KLF9	TRBC2	IDS
CLEC9A	TNFAIP3	BTG2	PDE4B	ARMH1	KRT18	TRIM47	IFI44
CLIC2	TNFRSF14	C160RF86	PHACTR1	ASB13	L3MBTL4	TSC22D3	IGHM
CRHBP	TNFSF13B	CIORF162	PKN1	ATL1	LGALS3BP	TSPAN13	ISG20
CTSW	TOB1	C1QTNF4	PLK2	ATP10D	LHFPL6	TSTD1	JAM2
CXCR4	TPD52	CAPN2	PNRC1	ATP2B1-AS1	LINC00623	TTC32	KLF10
DNTT	TPGS1	CAST	PPP1R14B	AVP	LINC01637	VAMP2	KLF6
DUSP1	TRBC2	CAVIN2	PPP1R15A	AZUI	LINC02256	VIM	LITAF
EMP1	TSC22D3	CBR3	PRKCH	B2M	LINC02573	WAC-ASI	MRPL23
EMP3	TSPAN13	CCND3	PROM1	BCAS4	LITAF	WDR49	MSRB3
EPDR1	TSPO	CCZ1B	PRR34-AS1	BEX5	LRCH4	Z93241.1	MZB1
EREG	TSTD1	CD69	PSMB8-AS1	BHLHE40	LRRC26	ZBTB20	N4BP2L1
FAAP20	TUBA1A	CD74	PSMB9	BLVRA	LRRK1	ZC3HAV1	NEAT1
FAM30A	VIM	CD9	PTGER2	BSPRY	LTBP3	ZDHHC2	NFIC
FCER1A	YBX3	CDKN2D	PTGER4	BST2	LTC4S	ZFP36	NPC2
FHL1	ZBTB20	CIITA	PTMS	BTN3A3	MAFF	ZNF385C	NPDC1
FLT3	ZBTB7A	CLEC2B	RABL6	C160RF45	MAP3K8	ZNF395	NR4A2
FOS	ZDHHC2	CLIP4	RARRES3	C160RF54	MAP7	ZNF90	PKN1
FOSB	ZFP36	CLN8	RBM38	C1ORF21	MARCH9		PPP1R15A
FUT7	ZNF90	COBL	REL	C1QTNF4	MCL1		PROM1
GUCY1B1		CPT1A	RENBP	C21ORF2	МСИВ		PRR34-AS1
HLA-A		CRHBP	REPIN1	C22ORF34	MED17		PSMB8-AS1
HLA-B		CRTAP	RETREG1	C3ORF80	MESP1		PSMB9
HLA-C		CSRNP1	RETREG2	C90RF139	METTL7A		PTGER2
HLA-DMA		CXCR4	RGS1	C9ORF43	MFSD6		PTGER4
HLA-DMB		DACH1	RGS2	CARMIL2	MGLL		PTMS
HLA-DPA1		DDIT4	RHPN1	CAST	MIF		RARRES3
HLA-DPB1		DNTT	RIPOR1	CAT	MLXIP		RENBP
HLA-DQA1		DUOX1	RNF43	CAVINI	MRPL23		S100A6
HLA-DQB1		DUSP1	RUBCN	CBR1	MSRB3		SCAND1
HLA-DRA		EAF2	RUBCNL	CBR3	MT-ND1		SCD5
HLA-DRB1		EIF4E3	RUNX2	CCDC57	MT-ND3		SEPT1
HLA-F		EMP1	S100A10	CCL28	MT1F		SERPINE2
ID2		ENTPD6	S100A11	CD151	MTURN		SQOR
IDS		EPDR1	S100A6	CD300LF	MVP		STK17B
IER2		FAAP20	SCAND1	CD38	MYCT1		SUN2
IFI27L2		FAM215B	SCD5	CD59	MYO1G		TGFB1
IFI44		FAM30A	SCHIP1	CD74	MZB1		TMEM160
IFI6		FBXW5	SDK2	CDCP1	N4BP2L1		TNFSF13B

IGFBP7	FCMR	SEPT1	CDK14	NAALADL1	TOB1
IGHM	FHL1	SERPINB9	CDKN2C	NBEAL1	TPD52
IL18	FKBP5	SERPINE2	CHST2	NDRG1	TPGS1
IMPA2	FLOT1	SMIM3	CIITA	NEAT1	TSC22D3
IRF2BP2	FLT3	SOD2 1	CITED4	NECTIN1	TSPAN13
ISG20	FOS	SPARC	CLEC12A	NFIA	VIM
JAM2	FOSB	SPATC1L	CLEC2B	NFIC	ZBTB20
JUN	GBP1	SPINK2	CLGN	NFKBIZ	ZFP36
JUNB	GBP2	SPRY1	CLIC2	NPC2	ZNF90
KCNAB2	GSAP	SQOR	CLIP4	NPDC1	
KLF10	H1FX	SRSF5	COG3	NPW	
KLF6	HAGHL	SRSF6	CPT1A	NR4A1	
LAT2	HHIP-AS1	STK17B	CRHBP	NR4A2	
LINC01122	HLA-A	SUN2	CRYL1	NUCB2	
LINC01637	HLA-B	TAP1	CST7	OSBPL10	
LITAF	HLA-C	TESPA1	CTSF	OSM	
LKAAEAR1	HLA-DMA	TGFB1	CTSW	P4HTM	
LRCH4	HLA-DMB	THEMIS2	CXCR4	PARD6A	
MAP3K8	HLA-DOA	TIGD3	CYTH1	PARP10	
MAP4K1	HLA-DPA1	TMEM107	DACH1	PER1	
MCL1	HLA-DPB1	TMEM160	DDO	PHKG1	
MESP1	HLA-DQA1	TMEM220	DGCR6	PIEZO1	
MGLL	HLA-DQB1	TNFRSF21	DNAJC4	PKN1	
MIF	HLA-DRA	TNFSF13B	DNTT	PLXND1	
MRPL23	HLA-DRB1	TNK2	DUSP1	PODXL2	
MSRB3	HLA-E	TOB1	EDARADD	PPP1R15A	
MT-ND2	HLA-F	TP53I13	EIF4E3	PROM1	
MX1	HOXA10	TPD52	ELANE	PRR34-AS1	
MYO5C	HPCAL1	TPGS1	EMP1	PRR7	
MZB1	ID2	TRIM8	EPDR1	PRRG4	
N4BP2L1	IDS	TSC22D3	EREG	PSMB8	
NAALADL1	IER2	TSPAN13	FAAH	PSMB8-AS1	
NEAT1	IFI44	TTC32	FAAP20	PSMB9	
NEGR1	IFI44L	TTPAL	FAM117A	PTGER2	
NFIA	IGHD	UBE2J1	FAM30A	PTGER4	
NFIC	IGHM	UBR5-AS1	FBXW5	PTMS	
NFKBIZ	IL17RC	UCP2	FCER1A	PYROXD2	
NPC2	IMPA2	VAMP2	FHL1	RAB40B	
NPDC1	ISG20	VIM	FKBP5	RARRES3	
NPW	ITM2C	WAC-AS1	FLNA	RENBP	
NR1H2	JAKMIP2	YPEL3	FLT3	RGCC	
NR4A2	JAM2	ZBTB20	FOS	RGS1	

NT5M	JUN	ZFP36	FOSB	RHPN1	
OSM	JUNB	ZNF90	FRY	RNASE2	
PARD6A	JUND		GAS6-AS1	RNF130	