nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

Please do not complete any field with "not applicable" or n/a. Refer to the help text for what text to use if an item is not relevant to your study. For final submission: please carefully check your responses for accuracy; you will not be able to make changes later.

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For	all statistical an	alyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.	
n/a	a Confirmed		
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement		
	🔽 A stateme	ent on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly	
\checkmark		tical test(s) used AND whether they are one- or two-sided non tests should be described solely by name; describe more complex techniques in the Methods section.	
\checkmark	A descript	cion of all covariates tested	
	🗸 A descript	cion of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons	
\checkmark	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)		
\checkmark	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.		
\checkmark	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings		
\checkmark	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes		
\square Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated			
Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.			
Software and code			
Poli	cy information a	about <u>availability of computer code</u>	
Da	ata collection	ChIP-seq libraries were sequenced on Illumina HiSeq2500 or Illumina NovaSeq 6000 platforms.	
Da	ata analysis	Bowtie2 2.4.2, deepTools 3.5.0, pyGenomeTracks 3.6, SAMtools 1.15.1, Quast Genome assembly Quality (Galaxy Version 5.0.2+galaxy4), SICER2, Trim Galore! (Galaxy Version 0.6.7+galaxy0), FASTQ joiner (Galaxy Version 2.0.1.1 + galaxy0), minimap2 (Galaxy Version 2.26+galaxy0), IGV 2.9.2, BLAT (v. 36), MultAlin	
		g custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and	

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Original data produced in this work are available at NCBI Bioproject database (PRJNA945609 and PRJNA949688) and NCBI Nucleotide database (OQ679756).

In this work we also used publicly available WGS (SRR6374293), ChIP-seq (SRX2789367, SRX2789358, SRX2789324, SRX2789325, SRX2789347, SRX2789336, SRX2789370, SRX2789369, SRX6609390, SRX6609391, SRX6609392 and SRX6609393) and PacBio reads (SRR6374292).

Research inv	olving human participants, th	neir data, or biological material
	pout studies with <u>human participants or hur</u> on and <u>race, ethnicity and racism</u> .	man data. See also policy information about sex, gender (identity/presentation),
Reporting on sex	nd gender	
Reporting on race other socially rele groupings		
Population charac	teristics	
Recruitment		
Ethics oversight		
Note that full information	ion on the approval of the study protocol must als	so be provided in the manuscript.
Field-spe	cific reporting	
Please select the on	e below that is the best fit for your research	. If you are not sure, read the appropriate sections before making your selection.
🖊 Life sciences	Behavioural & social sciences	Ecological, evolutionary & environmental sciences
For a reference copy of th	e document with all sections, see <u>nature.com/document</u>	ts/nr-reporting-summary-flat.pdf
Life scien	ces study design	
All studies must disc	lose on these points even when the disclosu	ire is negative.
Sample size	Details are reported in Methods section referring to experir	mental design
Data exclusions	No data was excluded from this study	
Replication	All attempts to replicate this study were successful. A large body of previous data from our lab support pipelines and methodologies	
Randomization	Randomization was not used	
Blinding	Blinding was not used	
Behaviou	ral & social sciences	s study design
All studies must disclose on these points even when the disclosure is negative.		
Study description		
Research sample		
Sampling strategy		
Data collection		

Timing

Data exclusions

Non-participation

Randomization

	these points even when the disclosure is negative.
Study description	
Research sample	
Sampling strategy	
Data collection	
Timing and spatial scale	
Data exclusions	
Reproducibility	
Randomization	
Blinding	
Field conditions	
Access & import/export	
Disturbance	
We require information from a	r specific materials, systems and methods uthors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each materia rant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response. Intal systems Methods
n/a Involved in the study	n/a Involved in the study
	I IIM I CHESCU
Antibodies Like Li	Flow cytometry
n/a Involved in the study	☐ ChIP-seq

Antibody validation was described in our previous publication: Cappelletti et al. Scientific Reports 2019, PMID: 31676881

Antibodies

Antibodies used

Validation

Sheep serum agains horse CENP-A protein

Eukaryotic cell line	S	
Policy information about <u>cell</u>	lines and Sex and Gender in Research	
Cell line source(s)	Fibroblast cell lines were established in our lab or in other labs of the FAANG consortium	
Authentication	Morphological, cytogenetic and genome sequence analysis	
Mycoplasma contamination	All the cell lines tested negative for mycoplasma	
Commonly misidentified lin (See ICLAC register)	None None	
Palaeontology and	Archaeology	
Specimen provenance		
Specimen deposition		
Dating methods		
Tick this box to confirm	that the raw and calibrated dates are available in the paper or in Supplementary Information.	
Ethics oversight		
Note that full information on the	approval of the study protocol must also be provided in the manuscript.	
Animals and other	research organisms	
Policy information about <u>studies involving animals</u> ; <u>ARRIVE guidelines</u> recommended for reporting animal research, and <u>Sex and Gender in Research</u>		
Laboratory animals		
Wild animals		
Reporting on sex		
Field-collected samples		
Ethics oversight		
Note that full information on the	approval of the study protocol must also be provided in the manuscript.	
Clinical data		
Policy information about clini	ical studies	
	ith the ICMJE guidelines for publication of clinical research and a completed CONSORT checklist must be included with all submissions.	
Clinical trial registration		
Study protocol		
Data collection		
Outcomes		

Dual use research of concern

Policy information about <u>dual use research of concern</u>

Hazards

Could the accidental, deliberate or reckless misuse of agents or technologies generated in the work, or the application of information presented in the manuscript, pose a threat to:

I		
No Yes		
X Public health		
X National security		
X Crops and/or livest	ock	
X Ecosystems		
X Any other significa	nt area	
Experiments of concer	n	
Does the work involve an	y of these experiments of concern:	
No Yes		
X Demonstrate how	to render a vaccine ineffective	
X Confer resistance t	o therapeutically useful antibiotics or antiviral agents	
X Enhance the virule	nce of a pathogen or render a nonpathogen virulent	
X Increase transmiss	bility of a pathogen	
X Alter the host rang	e of a pathogen	
X Enable evasion of a	liagnostic/detection modalities	
X Enable the weapor	ization of a biological agent or toxin	
X Any other potentia	lly harmful combination of experiments and agents	
Plants		
Seed stocks		
Novel plant ganetypes		
Novel plant genotypes		
Authentication		
ChIP-seq		
Data deposition		
•	and final processed data have been deposited in a public database such as GEO.	
	e deposited or provided access to graph files (e.g. BED files) for the called peaks.	
	deposited of provided access to graph files (e.g. beb files) for the called peaks.	
Data access links May remain private before public	NCBI Bioproject database (https://www.ncbi.nlm.nih.gov/bioproject/): PRJNA945609 and PRJNA949688	
Files in database submiss	On Accession numbers are reported in Methods section	
Genome browser session (e.g. <u>UCSC</u>)	The relevant horse genome regions were visualized by IGV 2.9.2	
Methodology		
Replicates	eplicates Fibroblast cell lines from 11 individuals and 4 tissues from 4 individuals were analyzed	
Sequencing depth	About 40 million 150 bp paired-end reads for ChIP samples and about 20 million 150 bp paired-end reads for Input samples. Detailed information in Methods.	
Antibodies	Sheep anti-horse-CENP-A serum	
Peak calling parameters	Normalization of read coverage of the ChIP datasets against the input datasets was performed using bamCompare available in the deepTools 3.5.0 using RPKM normalization in subtractive mode. Peaks were obtained with pyGenomeTracks 3.6. Peak calling was performed with SICER2 using -w 200 and -g 1000 paramete	
Data quality	The quality of raw reads was assessed by FastQC. Per base mean sequence quality was always higher than 30 (base accuracy > 99.9%.	

Bowtie2 2.4.2, deepTools 3.5.0, pyGenomeTracks 3.6, SAMtools 1.15.1, Quast Genome assembly Quality (Galaxy Version 5.0.2+galaxy4), SICER2, Trim Galore! (Galaxy Version 0.6.7+galaxy0), FASTQ joiner (Galaxy Version 2.0.1.1 + galaxy0), minimap2 (Galaxy Version 2.26+galaxy0), IGV 2.9.2, BLAT (v. 36), MultAlin

Software

Flow Cytometry	
The axis scales are clearly visib	er and fluorochrome used (e.g. CD4-FITC). ole. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers). h outliers or pseudocolor plots. of cells or percentage (with statistics) is provided.
Methodology	
Sample preparation	
Instrument	
Software	
Cell population abundance	
Gating strategy	
Tick this box to confirm that a	figure exemplifying the gating strategy is provided in the Supplementary Information.
Magnetic resonance in	naging
	<u>laging</u>
Experimental design Design type	
Design type Design specifications	
Behavioral performance measure	
bellavioral performance measure	
Imaging type(s)	
Field strength	
Sequence & imaging parameters	
Area of acquisition	
Diffusion MRI Used	☐ Not used
Preprocessing	
Preprocessing software	
Normalization	
Normalization template	
Noise and artifact removal	
Volume censoring	
Statistical modeling & inferer	nce
Model type and settings	
Effect(s) tested	
Specify type of analysis: Wh	oole brain ROI-based Both

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Statistic type for inference		
(See Eklund et al. 2016)		
Correction		
Models & analysis		
n/a Involved in the study		
Functional and/or effective co	onnectivity	
Graph analysis		
Multivariate modeling or pred	lictive analysis	
Functional and/or effective connect	tivity	
Graph analysis		

Multivariate modeling and predictive analysis