

**Supplementary Figure 1.** Results of sensitivity analysis for model training. PRCC values for influential initial concentrations (A) and parameters (B). Y axes indicate PRCC values.



**Supplementary Figure 2.** Results of identifiability analysis. identifiability analysis identifies the parameters that have one unique model output for each parameter value. In this method, pair-wise correlation coefficients between parameters were calculated. The identifiable parameters having correlations with all other parameters between -0.9 and 0.9 are labeled blue. The unidentifiable parameters having correlations of > 0.9 or < -0.9 with at least one other parameter are labeled yellow.



**Supplementary Figure 3.** Monte Carlo simulations for training and validation data for IL-6 alone and in combination with sIL-6R stimulation. 50 ng/ml IL-6 with or without additional 100 ng/ml sIL-6R induced relative pSTAT3 (A), ppAkt (B), and pERK (C). Varying concentrations of IL-6 alone induced relative pSTAT3 (D) and with additional 100 ng/ml sIL-6R induced relative pSTAT3 (E). The circles are experimental data. Bars are mean ± SEM. Curves are the mean values of the 1,000 Monte Carlo simulations. Shaded regions show 95% confidence intervals. Light gray: 50 ng/ml IL-6 (A-C), 0 – 50 ng/ml IL-6 (D), and 10 ng/ml IL-6 (F-H) stimulation; Dark gray: 50 ng/ml IL-6 + 100 ng/ml sIL-6R (A-C), 0 – 50 ng/ml IL-6R (F-H) stimulation.



**Supplementary Figure 4.** Predicted AUC of pSTAT3, pAkt, and pERK responses. AUC of pSTAT3 (A), pAkt (B), and pERK (C) in response to IL-6 concentrations varying from 0 to 5 nM without sIL-6R. In the absence of IL-6R, 0.2 nM IL-6 in combination with sIL-6R concentrations varying from 0 to 100 nM induced AUC of pSTAT3 (D), pAkt (E), and pERK (F). Curves are the mean values of the 12 best fits. Shaded regions show 95% confidence intervals of the fits. Orange: classic signaling responses; Yellow: trans-signaling responses.



Supplementary Figure 5. Predicted time courses of pSTAT3, pAkt, and pERK following stimulation by 2 nM IL-6 alone with a mean value of 6.4 nM IL-6R (orange), 2 nM IL-6 in combination with a mean value of 6.4 nM sIL-6R in the absence of IL-6R (yellow), and 2 nM IL-6 with a mean value of 6.4 nM of both IL-6R and sIL-6R (gray) when IL-6R and sIL-6R are set at the same level and remain constant within four-hour simulation time (A), when kinetic rates governing R1 and R2 to be the same as the corresponding kinetic rates for R3 and R4 (B), and when both IL-6R was set as a constant input and kinetic rates governing R1 and R2 to be the same as the corresponding kinetic rates for R3 and R4 (C). Curves are the mean values of the 12 best fits. Shaded regions show 95% confidence intervals of the fits. Orange: classic signaling responses; Gray: overall responses. R1: IL-6 + IL-6R  $\leftrightarrow$  IL-6:IL-6R; R2: 2 IL-6:IL-6R + 2 gp130  $\leftrightarrow$  Rcomplex; R3: IL-6 + sIL-6R  $\leftrightarrow$  IL-6:SIL-6R; R4: 2 IL-6:SIL-6R + 2 gp130  $\leftrightarrow$ 



Supplementary Figure 6. Dissociation constants (Kd) of ligand-receptor binding. R1: IL-6 + IL-6R  $\leftrightarrow$  IL-6:IL-6R; R2: 2 IL-6:IL-6R + 2 gp130  $\leftrightarrow$  Rcomplex; R3: IL-6 + sIL-6R  $\leftrightarrow$  IL-6:sIL-6R; R4: 2 IL-6:sIL-6R + 2 gp130  $\leftrightarrow$  Rcomplex. Each circle represents one fit. Orange: classic signaling responses; Yellow: trans-signaling responses.



Supplementary Figure 7. Dynamics of relevant species involved in ligand-receptor binding following stimulation by 2 nM IL-6 alone with a mean value of 6.4 nM IL-6R (orange) (A-B), 2 nM IL-6 in combination with a mean value of 6.4 nM sIL-6R in the absence of IL-6R (yellow) (C-D), and 2 nM IL-6 with a mean value of 6.4 nM of both IL-6R and sIL-6R (gray) (E-G). Curves are the mean values of the 12 best fits. Shaded regions show 95% confidence intervals of the fits.



**Supplementary Figure 8.** Results of sensitivity analysis for the trained and validated model. PRCC values that are greater than 0.5 or less than -0.5 for influential initial concentrations pSTAT3 (A), pAkt (B), and pERK, and influential parameters to pSTAT3 (D) and pAkt (E). Y axes indicate PRCC values. Note, no parameters were identified as influential for pERK.

## MATLAB.m file containing model code

function dsdt = coreFile\_IL6(s,y,params)

IL6 =	у(	1	);		
IL6R =	у(	2	);		
IL6_IL6R	=	у(	3	);	
gp130 =	y(	4	);		
Rcomplex =	y(	5	);		
sIL6R =	y(	6	);		
IL6_sIL6R	=	y(	7	);	
Rcomplex_a :	=y(	8	);		
SOCS3	=	y(	9	);	
Rcomplex_a_	Ras_G	DP	=	y(	10);
STAT3 =	y(	11	);		-
pSTAT3	=	y(	12	);	
SOCS3mRN/	A 1	=	y(	13	);
SOCS3mRN/	A_2	=	ý(	14	);
SOCS3mRN/	<u>م</u> _	y(	15	);	,.
SOCS3_1	=	ý(	16	);	
SOCS3_2	=	ý(	17	);	
Rcomplex_a_	pPI3K =	=ý(	18	);	
Ras_GDP		ý(	19	);	
Rcomplex a	pPI3K	PIP2 =	y(	20	);
Ras GTP		v(	21	);	
PI3K = y(	22	);		,.	
MEK =	y(	23	);		
ERK =	ý(	24	);		
MEKpp	=	y(	25	);	
ERKpp=	y(	26	);		
MEKpp_ERK	=	y(	27	);	
pERK =	y(	28	);		
pERK_ppMEl	K=	y(	29	);	
pMEK =	y(	30	);	,.	
Ptase2 =	y(	31	);		
Ptase3=	y(	32	);		
	- •				

ppMEK_Ptase	2	=	у(	33)			
pMEK_Ptase2	2 =	у(	34	);			
ppERK_Ptase	3	=	у(	35	);		
pERK_Ptase3	5 =	y(	36	);			
aRas_GTP	=	y(	37	);			
Raf =	y(	38	);				
Raf_Ras_GTF	<b>D</b> =	у(	39	);			
aRaf =	у(	40	);				
Ptase1 =	y(	41	);				
aRaf_Ptase1	=	у(	42	);			
MEK_aRaf	=	y(	43	);			
pMEK_aRaf	=	y(	44	);			
PIP3 =	у(	45	);				
PTEN =	у(	46	);				
PIP3_PTEN	=	у(	47	);			
Akt =	у(	48	);				
PIP3_Akt	=	у(	49	);			
PDK1 =	у(	50	);				
PIP3_Akt_PD	K1	=	у(	51	);		
pAkt =	у(	52	);				
PIP3_PDK1	=	у(	53	);			
PIP3_pAkt	=	у(	54	);			
PIP3_pAkt_PI	DK1	=	у(	55	);		
ppAkt =	у(	56	);				
PP2A =	у(	57	);				
ppAkt_PP2A	=	у(	58	);			
pAkt_PP2A	=	у(	59	);			
PP2Aoff	=	у(	60	);			
ppAkt_PP2Ao	ff	=	у(	61	);		
PIP2 = y(	62	);					
p1cl =	param	s(	1	,1);			
p2cl =	param	s(	2	,1);			
p3cl =	param	s(	3	,1);			
p4cl =	param	s(	4	,1);			
p1tr =	param	s(	5	,1);			

p2tr	=	params	(	6	,1);				
p3tr	=	params	(	7	,1);				
p4tr	=	params	(	8	,1);				
p5	=	params	(	9	,1);				
p13	=	params	(	10	,1);				
p6	=	params	(	11	,1);				
p7	=	params	(	12	,1);				
p8	=	params	(	13	,1);				
p9	=	params	(	14	,1);				
p10	=	params	(	15	,1);				
p11	=	params	(	16	,1);				
pdela	y1	=	params	5(	17	,1);			
pdela	y2	=	params	6(	18	,1);			
p12	=	params	(	19	,1);				
kf35	=	params	(	20	,1);				
kr35	=	params	(	21	,1);				
kf36	=	params	(	22	,1);				
kr36	=	params	(	23	,1);				
k_aRa	afRasGT	Р	=	paran	ns(	24	,1);		
kd_aF	RafRasG	TP	=	paran	ns(	25	,1);		
kd_Ra	afRasGT	P	=	paran	ns(	26	,1);		
kd_rR	afRasG	ΓP	=	paran	ns(	27	,1);		
k_dpF	Raf	=	params	6(	28	,1);			
kd_dp	oRaf	=	params	6(	29	,1);			
ked	=	params	(	30	,1);				
k_aM	EKRaf	=	params	6(	31	,1);			
kd_aN	<b>MEKRaf</b>	=	params	6(	32	,1);			
ked_N	<b>MEKRaf</b>	=	params	6(	33	,1);			
ked_N	MEKRaf2	2	=	paran	ns(	34	,1);		
k_dpN	ИЕК_рр	=	params	6(	35	,1);			
kd_dp	oMEK_pp	)	=	paran	ns(	36	,1);		
ked2	=	params	(	37	,1);				
k_dpN	ИЕК_р	=	params	6(	38	,1);			
kd_dp	MEK_p	=	params	6(	39	,1);			
k_aEF	RKMEK	=	params	6(	40	,1);			
kd_aE	ERKMEK	=	params	6(	41	,1);			

ked_ERKMEK	брр	=	param	s(	42	,1);
ked_ERKpME	Крр	=	param	s(	43	,1);
k_dpERKpp	=	params	S(	44	,1);	
kd_dpERKpp	=	params	S(	45	,1);	
ked3 =	param	s(	46	,1);		
k_dpERKp	=	params	S(	47	,1);	
kd_dpERKp	=	params	S(	48	,1);	
k_1PI3K	=	params	S(	49	,1);	
kd_1PI3K	=	params	S(	50	,1);	
k_aPIP2	=	params	S(	51	,1);	
kd_aPIP2	=	params	S(	52	,1);	
k_fPIP3	=	params	S(	53	,1);	
k_aPTEN	=	params	S(	54	,1);	
kd_aPTEN	=	params	S(	55	,1);	
k_fPIP2	=	params	S(	56	,1);	
k_aAkt =	param	s(	57	,1);		
kd_aAkt	=	params	S(	58	,1);	
k_aPDK1	=	params	S(	59	,1);	
kd_aPDK1	=	params	S(	60	,1);	
k_fAkt_p	=	params	S(	61	,1);	
k_fPIP3PDK1	=	params	S(	62	,1);	
k_aPP2A	=	params	S(	63	,1);	
kd_aPP2A	=	params	S(	64	,1);	
k_fAkt_pPP2A	١	=	param	s(	65	,1);
k_aPP2Aoff	=	params	S(	66	,1);	
kd_aPP2Aoff	=	params	s(	67	,1);	
k_fPP2A	=	params	S(	68	,1);	
D1	م1 ما * ا			J * II C		
RI = D2			א - אנםי ירעים א	/an120		, al * Deemalax
$R_2 = D_2$			)'''''''''''''''''''''''''''''''''''''	(gp130) hr * 11 G	/^2 - p4	. Roomplex ,
$R_{3} = D_{4}$	p111 1		or - pzi	(ap.120		, 1tr * Deemolox ,
R4 =	pou (		015/12 \\\2\ */4	(gp 130	η <sup>,</sup> 2 - μ τοι	+u RComplex;
	ha ((t	1 * 000	(1) (1 202000	+µ317		,

- pdelay1 \* SOCS3mRNA\_1 ; pdelay1 \* SOCS3mRNA\_2 ; p10 \* SOCS3mRNA ; R6 R7 R8 = = =

R9 p11 \* SOCS3mRNA = R10 = pdelav2 \* SOCS3 1 pdelav2 \* SOCS3 2 R11 = R12 = p12 \* SOCS3 : k aRafRasGTP\*Raf\*Ras GTP-kd aRafRasGTP\*Raf Ras GTP ; R13 = kd RafRasGTP\*Raf Ras GTP-kd rRafRasGTP\*aRaf\*aRas GTP; R14 = R15 = k\_dpRaf\*aRaf\*Ptase1-kd\_dpRaf\*aRaf\_Ptase1 R16 = ked\*aRaf Ptase1 R17 = k aMEKRaf\*MEK\*aRaf-kd aMEKRaf\*MEK aRaf ; R18 = ked MEKRaf\*MEK aRaf R19 = k aMEKRaf\*pMEK\*aRaf-kd aMEKRaf \*pMEK aRaf R20 = ked\_MEKRaf2\*pMEK\_aRaf ; R21 k\_dpMEK\_pp\*MEKpp\*Ptase2-kd\_dpMEK\_pp\*ppMEK\_Ptase2; = R22 = ked2\*ppMEK Ptase2; k dpMEK p\*pMEK\*Ptase2-kd dpMEK p\*pMEK Ptase2 ; R23 = ked2\*pMEK Ptase2 R24 = k aERKMEK\*MEKpp\*ERK - kd aERKMEK\*MEKpp ERK ; R25 = R26 = ked ERKMEKpp\*MEKpp ERK R27 k\_aERKMEK\*pERK\*MEKpp - kd\_aERKMEK\*pERK\_ppMEK = R28 = ked ERKpMEKpp\*pERK ppMEK k dpERKpp\*ERKpp\*Ptase3-kd dpERKpp\*ppERK Ptase3; R29 = R30 ked3\*ppERK Ptase3; = R31 k dpERKp\*pERK\*Ptase3-kd dpERKp\*pERK Ptase3 = R32 = ked3\*pERK Ptase3 R33 = k aPTEN\*PIP3\*PTEN-kd aPTEN\*PIP3 PTEN R34 k\_fPIP2\*PIP3\_PTEN; = R35 = k\_aAkt\*PIP3\*Akt-kd\_aAkt\*PIP3\_Akt; R36 k\_aPDK1\*PIP3\_Akt\*PDK1-kd\_aPDK1\*PIP3\_Akt\_PDK1 = k fAkt p\*PIP3 Akt PDK1 R37 = R38 = k fPIP3PDK1\*PIP3 PDK1 k\_aAkt\*PIP3\*pAkt- kd\_aAkt\*PIP3 pAkt R39 = k aPDK1 \*PIP3 pAkt\*PDK1-kd aPDK1 \*PIP3 pAkt PDK1; R40 = R41 k fAkt p\*PIP3 pAkt PDK1 ; = k aPP2A \*ppAkt\*PP2A-kd aPP2A \*ppAkt PP2A ; R42 = R43 = k fAkt pPP2A \* ppAkt PP2A; R44 = k aPP2A \*pAkt\*PP2A-kd aPP2A \*pAkt PP2A

R45	=	k_fAkt	_pPP2A	۸ *pAkt_PP2A ;
R46	=	k_aPP	2Aoff *p	pAkt*PP2Aoff-kd_aPP2Aoff *ppAkt_PP2Aoff ;
R47	=	k_fPP2	2A *ppÅ	kt_PP2Aoff ;
R48	=	p5 * R	complex	(1 +p13*SOCS3) ;</td
R49	=	p6*Rc	omplex_	_a ;
R50	=	p7*Rc	omplex_	_a *STAT3 - p8 * pSTAT3 ;
R51	=	kf35* I	Rcomple	ex_a * Ras_GDP - kr35* Rcomplex_a_Ras_GDP ;
R52	=	kf36 *	Rcompl	ex_a_Ras_GDP - kr36 * Rcomplex_a * Ras_GTP ;
R53	=	k_1PI3	3K*Rcor	nplex_a * PI3K - kd_1PI3K * Rcomplex_a_pPI3K ;
R54	=	k_aPII	2 * Rco	omplex_a_pPI3K * PIP2 - kd_aPIP2 * Rcomplex_a_pPI3K_PIP2;
R55	=	k_fPIP	3* Rcor	nplex_a_pPI3K_PIP2 ;
dsdt(	1	,1)	=	0 ;
dsdt(	2	,1)	=	-R1 ;
dsdt(	3	,1)	=	R1-2*R2 ;
dsdt(	4	,1)	=	-2*R2-2*R4 ;
dsdt(	5	,1)	=	R2+R4-R48+R49 ;
dsdt(	6	,1)	=	0 ;
dsdt(	7	,1)	=	R3-2*R4 ;
dsdt(	8	,1)	=	R48-R49 -R51+R52-R50 -R53 ;
dsdt(	9	,1)	=	R11-R12 ;
dsdt(	10	,1)	=	R51 -R52 ;
dsdt(	11	,1)	=	-R50 ;
dsdt(	12	,1)	=	R50 ;
dsdt(	13	,1)	=	R5-R6 ;
dsdt(	14	,1)	=	R6-R7 ;
dsdt(	15	,1)	=	R7-R8 ;
dsdt(	16	,1)	=	R9-R10 ;
dsdt(	17	,1)	=	R10-R11 ;

R53-R54+R55

-R51 ;

R54-R55

R52-R13

-R53 ; -R17+R24

-R25+R32

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dsdt(	25	,1)	=	R20-R21-R25+R26-R27+R28 ;
dsdt(	26	,1)	=	R28-R29 ;
dsdt(	27	,1)	=	R25-R26 ;
dsdt(	28	,1)	=	R26-R27+R30-R31 ;
dsdt(	29	,1)	=	R27-R28
dsdt(	30	.1)	=	R18-R19+R22-R23 ;
dsdt(	31	,1)	=	-R21+R22-R23+R24
dsdt(	32	,1)	=	-R29+R30-R31+R32
dsdt(	33	.1)	=	R21-R22 ;
dsdt(	34	.1)	=	R23-R24
dsdt(	35	.1)	=	R29-R30
dsdt(	36	,1) ,1)	=	R31-R32
dsdt(	37	.1)	=	R14 :
dsdt(	38	,1) ,1)	=	-R13+R16 ;
dsdt(	39	.1)	=	R13-R14
dsdt(	40	.1)	=	R14-R15-R17+R18-R19+R20 :
dsdt(	41	.1)	=	-R15+R16 ;
dsdt(	42	.1)	=	R15-R16
dsdt(	43	,1) ,1)	=	R17-R18
dsdt(	44	.1)	=	R19-R20
dsdt(	45	.1)	=	R55-R33-R35+R38-R39 :
dsdt(	46	,1)	=	-R33+R34 ;
dsdt(	47	,1)	=	R33-R34
dsdt(	48	.1)	=	-R35+R45
dsdt(	49	.1)	=	R35-R36
dsdt(	50	,1)	=	-R36+R38-R40 ;
dsdt(	51	,1)	=	R36-R37
dsdt(	52	.1)	=	R37-R39+R43-R44 ;
dsdt(	53	,1)	=	R37-R38+R41;
dsdt(	54	,1)	=	R39-R40
dsdt(	55	.1)	=	R40-R41
dsdt(	56	,1)	=	R41-R42-R46+R47 ;
dsdt(	57	.1)	=	-R42+R43-R44+R45+R47 ;
dsdt(	58	,1)	=	R42-R43 ;
dsdt(	59	,1)	=	R44-R45
dsdt(	60	,1)	=	-R46 ;
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dsdt(	61	,1)	=	R46-R47	;
dsdt(	62	,1)	=	-R54+R34	;

return