The Irish cystic fibrosis database

Siobhan M Cashman, Ana Patino, Marina Garcia Delgado, Loretta Byrne, Brian Denham, Miguel De Arce

Abstract

We have found records of 1014 Irish cystic fibrosis patients alive by December 1994, belonging to 883 families. Prevalence in the population is 1/3475 and incidence at birth 1/1461, with a gene frequency of $2 \cdot 6\%$. Twenty percent of the patients are aged over 20 years, but at present survival rate falls rapidly after that age. We have identified 85% of the mutations on the CFTR gene in a sample of 29% of the families (506 CF chromosomes). Mutation Δ F508 is found in 72% of Irish CF chromosomes, G551D in 6.9%, and R117H in 2%. These are the highest frequencies reported for the latter two mutations world wide. Another seven mutations are found in an additional 4% of CF families. We present new microsatellite haplotype data that could be useful for genetic counselling of CF families bearing some of the 15% of CF mutations still unidentified, and comment on possible uses of our database.

(7 Med Genet 1995;32:972-975)

The vital statistics and mutations found in the Irish cystic fibrosis (CF) patient population are not as well described as in other European countries. For instance, the only estimate of incidence for this population was obtained by O'Reilly et al¹ who, searching through paediatric hospital admissions for the year 1971, found a minimal figure of 1 affected for 1800 children under 1 year of age. However, Mulherin et al,² describing their adult clinic, found that 12% of their patients had been diagnosed after the age of 10 years. With regard to the prevalence of mutations on the CFTR gene, De Arce et al^3 reported the deletion Δ F508 in 76% of 88 Irish CF chromosomes, but this sample is small, and many more mutations causing the disease world wide have since been identified.⁴ The strong association between haplotypes for the intragenic microsatellite markers described by Morral et al⁵ and Zielinsky et al⁶ and specific mutations on the CFTR gene could prove particularly useful in diagnosis and in the identification of as yet unknown mutations on the CFTR gene,⁷⁸ and have been used as ethnic markers tracing the course of past migrations.910 Here we summarise initial observations on the incidence of CF and vital statistics of these patients in Ireland, and we report on the mutations found in a sample of 506 Irish CF chromosomes and their associated haplotypes.

Materials and methods THE DATABASE

The database (access by Microsoft for the PC) contains three large tables prepared to accept data under the headings Demography, Genetics, and Clinical data. The information on mutations and genetic markers was obtained in the authors' laboratories from a DNA bank built up over a period of four years with the cooperation of clinical colleagues. Other statistical and clinical data relevant to the Irish CF population were obtained as follows.

FREQUENCY OF CYSTIC FIBROSIS IN IRELAND AND VITAL STATISTICS

Four sources of data were consulted: (1) Department of Health statistics on the number of persons suffering from CF and availing of the "Long Term Illness Scheme", (2) records of the Irish CF Association, (3) current hospital charts of CF patients attending any of the nine CF clinics distributed throughout the country, having obtained the necessary permission from the Consultants and local Ethics Committees, and (4) medical publications reporting on previous surveys on the Irish CF population. Population data were obtained from the 1991 Irish Census.¹¹ Once a comprehensive list of patients alive by December 1994 had been obtained, yearly incidence at birth was calculated retrospectively for the period 1977-1986, thus correcting for cases diagnosed after 9 years of age.

FREOUENCY OF MUTATIONS ON THE CFTR GENE Mutation Δ F508 was screened for in an initial sample of 240 CF chromosomes by heteroduplex analysis of their PCR amplified exon 10^{12} A panel of at least 40 CF non- Δ F508 chromosomes were subsequently analysed by denaturing gradient gel electrophoresis13 of their PCR amplified products of each of the following exons: 4, 5, 6a, 7, 8, 9, 11, 12, 14a, 15, 16, 17b, 18, 20, 21 and 23. Conditions and primers have been described previously.1415 This was followed by direct sequencing of the amplified products with abnormal electrophoretic patterns using the dideoxy method of Sanger et al.¹⁶ After this initial work established the most common mutations, a further 266 CF chromosomes (133 CF patients) were screened for the six most common mutations using an enhanced ARMS test,¹⁷ kindly supplied by Cellmark Diagnostics (UK).

LINKED MARKERS AND MICROSATELLITES We also examined the linkage disequilibrium between CF mutations and three extragenic

Department of Genetics, Trinity College, Dublin 2, Ireland S M Cashman L Byrne M De Arce

Department of Genetics, University of Navarre, Pamplona, Spain A Patino M G Delgado

National Children's Hospital, Harcourt Street, Dublin 2, Ireland B Denham

Correspondence to: Dr De Arce.

Received 23 March 1995 Revised version accepted for publication 12 July 1995

Table 1 Age distribution of 1014 Irish CF patients alive by December 1994 compared to US

Age (y)	% Irish	% US*	
0–5	12.0	22.4	
6-10	22.7	22.1	
11-15	22.1	17.2	
16-20	22.7	13.5	
21–25	11.4	10.3	
26-30	5.9	7.2	
31–35	2.1	4.1	
36+	1.1	3.2	

* Data from Fitzsimons.²¹

restriction fragment length polymorphisms (RFLPs), namely KM-19/PstI,¹⁸ XV-2c/TaqI,¹⁹ and Mp6-d9/MspI.²⁰ Two hundred Irish CF chromosomes and 36 non-CF chromosomes were investigated for these markers. Haplotypes for the three intragenic microsatellite markers IVS8CA,⁵ IVS17bTA,⁶ and IVS17bCA⁶ were also investigated for a total of 109 normal, 150 CF Δ F508, and 92 CF non- Δ F508 chromosomes, using methods reported elsewhere.⁷¹⁰

Results

FREQUENCY OF CYSTIC FIBROSIS IN IRELAND AND VITAL STATISTICS

We found records of 1014 patients alive by December 1994, belonging to 883 families; 763 of these have one affected child, 109 have two affected children, and 11 have three affected. Population data from the 1991 Census allow us to estimate prevalence at all ages to be 1 in 3475. Average yearly incidence at birth for the 10 year period 1977–1986 was 1 in 1461, with a gene frequency of 2.6%. Age distribution for the whole patient population (table 1) shows that 20% are over 20 years of age. Five of these, four women and one man,

are married and have at least one child each. The genotype of three of the mothers was Δ F508/G551D, Δ F508/R117H, and Δ F508/ Δ F508 respectively. The CF father is a 29 year old of genotype Δ I507/R117H. Age at death was available for 98 patients who had died after 1986, showing a lifespan of at least 20 years for 23% patients, with no significant difference between the sexes. Age at diagnosis was available for 409 patients, showing that 64% were diagnosed within their first year of life, and 18% after their third birthday. These included seven patients of genotype Δ F508/R117H, for whom the mean age at diagnosis was 13.8years (SD 9.2 years). Two unrelated patients of genotype Δ F508/1717-1G-A were diagnosed at ages 12 and 29 years respectively, while another unrelated patient of this genotype was diagnosed at 1 month of age. The mean age at diagnosis for 59 homozygous $\Delta F508/\Delta F508$ patients was 2.3 years (SD 3.5 years).

FREQUENCY OF MUTATIONS ON THE CFTR GENE AND ASSOCIATED HAPLOTYPES

Table 2 shows the frequency of mutations found on 506 Irish CF chromosomes and their associated haplotypes. Microsatellite haplotypes found in 109 normal, 150 Δ F508, and 50 CF non- Δ F508 chromosomes bearing as yet unknown mutations are shown in table 3.

Discussion

FREQUENCY OF CF IN IRELAND AND VITAL STATISTICS

We have identified 1014 Irish CF patients on record by the end of December 1994. Care

Table 2 Haplotypes associated with the most common CF mutations in Ireland, as well as with Irish CF chromosomes with unidentified mutations and non-CF chromosomes

Mutation	Nm/Ncf*	%	X/K^{\dagger}	М‡	Microsatellite§	No chrs
ΔF508	367/506	72.5	Α	1	See table 3	2
			B	2		116
			B C	_		0
			Ď	2		2
G551D	35/506	6.9	в	2	16-7-17	20
R117H	10/506	2.0	С	1	16-30-13	5
			D B C C B B B D D C A B C	ī	16-31-13	ī
G542X	5/506	1.0	B	2	17-32-13	2
621+1G-T	4/506	0.8	B	2	21-7-17	2 2 2
			B	$\overline{2}$	21-31-13	$\overline{2}$
R560T	4/506	0.8	Đ	2	16-7-17	ī
			Đ	2	16-31-17	ī
1717–1G-A	3/506	0.6	Ē	ī	16-32-13	$\frac{1}{2}$
			Ă	2	17-32-13	2 1
N1303K	2/506	0.4	B	2 2	23-29-13	2
3659delC	2/506	0.4	ĉ	ī	16-35-13	2 2
ΔI507	2/506	0.4	ND	-	ND	-
R352Q	1/506	0.2	C	1	16-31-13	1
R553X	0/506	0.0	ND	-	ND	-
1078delT	0/506	0.0	ND		ND	
Total identified	435/506	85.6	112		ILE .	
Unidentified	71/506	14.4	Α	1	See table 3	3
			Ă	2	See tuble 5	3 5
			A B C	$\frac{1}{2}$		17
			č	ĩ		
			ă	2		5 8
Normal			D A	ĩ	See table 3	16
			Ă	2	See table 5	1
			A B C C D	2		3
			č	ĩ		13
			č	2		1
			ň	2 2		2

*Nm = number of chromosomes bearing the mutation. Ncf = number of total CF chromosomes screened (see text for methods). †A = allele 1 for KM-19 and allele 1 for XV-2c, B = alleles 1 and 2, C = alleles 2 and 1, D = alleles 2 and 2, respectively. ‡M refers to linked marker Mp6-dp, which has also two alleles, 1 and 2. § Triplets of figures represent numbers of dinucleotide repeats found at loci IVS8CA, IVS17bTA, and IVS17bCA respectively. || No of chromosomes haplotyped for all markers mentioned. ND = not done.

Table 3	Microsatellite	haplotypes	found in	Irish	normal	and	CF	chromosomes
---------	----------------	------------	----------	-------	--------	-----	----	-------------

Normal chromosomes		Non- $\Delta F508$ chromosomes‡			$\Delta F508$ chromosomes			
Haplotype*	No	%		No	%	Haplotype*	No	%
16-30-13	20	18.3	16-7-17	17	34	23-31-13	72	48
16-31-13	16	14.7	16-31-13	5	10	17-32-13	34	23
16-32-13	10	9.2	17-7-17	3	6	17-31-13	12	8
16-7-17	8	7.3	16-30-13	4	8	23-32-13	12	8
16-46-13	4	3.7	16-29-13	2	4	22-31-13	5	3
16-34-13	4	3.7	23-37-13	2	4	23-30-13	3	2 2
17-30-13	3	2.7	17-37-11	2	4	16-31-13	3	2
17-7-17	3	2.7	23-31-13	2	4	16-32-14	2	1.6
16-29-13	3	2.7	24-7-17	1	2	17-34-13	1	0.6
16-25-13	2	1.8	17-29-14	1	2	17-41-13	1	0.6
16-32-13	2	1.8	15-7-17	1	2	16-31-14	1	0.6
16-45-13	2	1.8	23-36-13	1	2	17-7-17	1	0.6
23-22-17	$\overline{2}$	1.8	17-32-13	3	6	21-7-17	1	0.6
16-35-13	2	1.8	16-7-13	2	4	23-31-17	1	0.6
16-51-13	2	1.8	16-32-13	1	2	18-32-13	1	0.6
Observed only	-		23-32-13	2	4			
once†	26		16-25-13	ī	2			
Total	109			50			150	

* See legend for table 2.

The following haplotypes were found only once in normal chromosomes: 16-36-13, 16-7-13, 16-50-13, 16-24-13, 16-33-13, 16-38-13, 16-49-13, 16-31-14, 16-29-17, 17-49-11, 17-50-11, 17-36-13, 17-29-14, 17-22-13, 17-71, 17-7-19, 18-7-13, 18-32-13, 18-35-13, 18-36-13, 21-32-13, 21-34-13, 23-30-13, 23-32-13, 23-33-13, and 24-39-13 (0.9% each). These do not include the mutations shown in table 2.

was taken to ensure that this was as close as possible to complete ascertainment by cross checking several sources. Average incidence at birth was estimated at 1/1461, which is comparable to the incidence reported for Brittany²² and among the highest in the world. In their one year survey of paediatric hospital records, O'Reilly *et al*¹ reported an incidence of 1/1800 children under 1 year of age, but this reflects only 64% of all cases, since we have found that 36% CF patients are diagnosed after that age. Twenty percent of Irish CF patients are aged 20 years or older, but survival rate still falls drastically after that age.

Table 1 shows the current age distribution of CF patients in Ireland, which is comparable to the distribution in the US in 1990,²¹ except for the relatively smaller proportion of patients aged 5 years or less in Ireland. This probably reflects the recent sudden decline in the number of Irish births, which dropped from a total of 74 080 in 1980 to 52 947 a decade later.¹¹ In the US, the proportion of adult patients had increased fourfold between 1969 and 1990.21 A similar increase cannot be documented in Ireland for lack of retrospective data, but the increasing age of the Irish CF population required the opening of the first Irish adult CF clinic in 1977,² now attended regularly by about 200 patients. The upward shift in age distribution reflects improved care, and perhaps also improved rates of diagnosis. Patients bearing "mild" mutations, such as R117H, are frequently diagnosed late compared to those homozygous for Δ F508. They have a milder course of the disease²³ and can be confidently classified as CF patients after genotyping, thus probably contributing to the increasing age of the patient population. Two unrelated compound heterozygotes Δ F508/1717-1G-A diagnosed at 12 and 29 years of age are unusual because this has been found in association with a severe phenotype in other patients.²³ Both of these patients carry the 1717-1G-A on a different haplotype to that of the third patient of this genotype diagnosed at 1 month (table 2); it is quite possible that this haplotype difference coincides with another alteration within one of their CF genes which could modify the effect of the genotype, as observed by Dork *et al*²⁴ in a patient bearing three mutations on the CFTR gene.

Infertility in female CF patients is considered to be greater than 10%.25 About 98% of males are infertile, mostly because of bilateral congenital absence of the vas deferens (CBAVD).26 The observation of genotype Δ I507/R117H in a CF patient showing a mild form of the disease and who has fathered a child is interesting. Mutation Δ I507 is assumed to have the same clinical effects as the more common $\Delta F508.^{27}$ Mutation R117H is usually found in mild disease, and has been reported in 10% of male compound heterozygotes expressing CBAVD (reviewed by Pignatti²⁸), most frequently in association with Δ F508. The genotype Δ F508/ R117H has also been found in an asymptomatic woman, and recent evidence suggests that the variable clinical manifestations found in compound heterozygotes bearing mutation R117H may be determined by "genetic background".29 The case presented here adds to this phenotypic variability, since he is both fertile and pancreatic sufficient. However, unilateral absence of the vas deferens has not been excluded, nor has "genetic background", as referred to by Kiesewetter et al,²⁹ been investigated in this patient as yet.

FREQUENCY OF MUTATIONS ON THE CFTR GENE AND ASSOCIATED HAPLOTYPES

Table 2 shows the most common mutations in Ireland. Mutation Δ F508 has been reported in >70% of CF chromosomes from all laboratories in the British Isles,⁴ except for Northern Ireland. Another two or three mutations are found in an additional 10% of CF chromosomes in all laboratories from the British Isles.⁴ Mutation 1078delT appears to be relatively frequent exclusively in Brittany,³⁰ but was absent among 40 Irish CF non- Δ F508 chromosomes, and is also very infrequent in England,⁴ suggesting that it might not be a marker common to Celtic populations as proposed by Audrezet *et al.*³⁰ Instead, the distribution of mutation G551D appears to follow more closely the area of long term Celtic settlements,³¹ and in view of the identical haplotype in all G551D chromosomes, we have suggested that all cases may be identical by descent.¹⁰ Mutation R117H, found initially in a US patient of Anglo-Irish ancestry,³² occurs with the highest frequency in Ireland, and with much lower frequency or not at all elsewhere.⁴

The haplotype associations shown in table 2 could be useful for counselling in some cases, for instance in families where DNA from the affected child is unobtainable,³³ or where a direct test for the less common mutations is not available. As has been observed in other populations,³⁴ there exists a strong association between the B (X/K:1/2) haplotype and the CF chromosome, even for those with uncharacterised mutations. This mutation occurs with low frequency in the sample of normal chromosomes. Microsatellite haplotypes have been studied in only 42 Irish chromosomes bearing the less common mutations (table 2), but the consistency of their association with certain mutations is already apparent. For instance, table 2 shows that mutations G542X, 621+1G-T, N1303K, 3659delC, R560T, and R117H are associated with the same haplotypes, or with haplotypes probably derived from these by slippage of the DNA polymerase, as has been seen in patients from other European populations.⁷⁻¹⁰ Since those haplotypes are relatively rare in normal chromosomes (table 3), the constancy of the association has been explained as suggesting identity by descent of all chromosomes bearing the same mutations.⁹ Haplotype 16-7-17 is found in CF non- Δ F508 chromosomes more frequently than would be expected from its frequency in normal chromosomes (p < 0.01, table 3), suggesting perhaps that another fairly common CF mutation may be associated with this haplotype in Ireland.

As the patient population benefits from longer and better quality of life, and the bases for new treatments are found, accessibility and concentration of clinical and genetic data are more necessary, and this would appear to justify the additional effort required to complete the database.

S M Cashman was funded by the Irish Cystic Fibrosis As-S M Cashman was funded by the Irish Cystic Fibrosis As-sociation. We are grateful to the following physicians: Professor M X Fitzgerald and Dr James Hayes from St Vincent's Hospital, Elm Park, Dublin 4, Professor Edward Tempany from Our Lady's Hospital, Crumlin, Dublin 12, Professor T Matthews from The Children's Hospital, Temple Street, Dublin 1, Dr Michael O'Mahoney from Limerick Regional Hospital, Dr Ger-ard Loftus from Galway Regional Hospital, Dr O'Kane from Mayo General Hospital, Dr John Gleeson from Sligo General Hospital, and all the staff at the CF clinic of the National Children's Hospital, Harcourt Street, Dublin 2. We are also grateful to all the patients and their families for their interest in our work.

- 1 O'Reilly D, Murphy J, McLaughlin J, et al. The prevalence O'Reilly D, Murphy J, McLaughin J, et al. The prevalence of coeliac disease and cystic fibrosis in Ireland, Scotland, England and Wales. Int J Epidemiol 1974;3:247-51.
 Mulherin D, Ward K, Coffey M, et al. Cystic fibrosis in adolescents and adults. Irish Med J 1991;84:48-51.
 De Arce M, Mulherin D, McWilliam P, et al. Frequency of deletion F508 among Irish cystic fibrosis patients. Hum Genet 1990;85:403-4.
 The Arcein Genetic Analysis Concentry. Bacutary Article Arcein Construction Concentry. Bacutary Concentry, Neurol. 51, 523-523.

- The Cysic Fibrosis Genetic Analysis Consortium. Population variation of common CF mutations. Hum Mutat 1994;4:167-77

- 5 Morral N, Gibau E, Zielinsky J, et al. Dinucleotide (CA/GT) repeat polymorphism in the cystic fibrosis transmembrane conductance regulator gene (CFTR). Hum Genet 1992; 88:356-8.
- 6 Zielinsky J, Marckiewicz D, Rininsland F, Rommens JM, Tsui LC. A cluster of highly polymorphic dinucleotide repeats in intron 17b of the cystic fibrosis transmembrane conductance regulator gene (CFTR). Am J Hum Genet 1991: 49:1256-62
- 7 Morral N, Nunes V, Casals T, et al. Microsatellite haplotypes for cystic fibrosis; mutation frameworks and evolutionary tracers. *Hum Molec Genet* 1993;2:1015-22.
- 8 Hughes D, Hill A, Redmond A, et al. Fluorescent multiplex microsatellites used to identify haplotype associations with 15 CFTR mutations in 124 Northern Irish CF families.
- 15 CFTR mutations in 124 Northern Irish CF families. Hum Genet 1995;95:462-4.
 9 Morral N, Bertranpetit J, Estivill X, et al. The origin of the major cystic fibrosis mutation (ΔF508) in European populations. Nature Genet 1994;7:165-75.
 10 Cashman SM, Patino A, Martinez A, et al. Identical intra-genic microsatellite haplotype found in cystic fibrosis chromosomes bearing mutation G551D in Irish, English, Scottish, Breton and Czech patients. Hum Hered 1995; 45:6-12 45:6-12
- 11 Central Statistics Office. Census of population of Ireland 1991. Stationery Office, Dublin, Ireland. 12 Kerem B, Zielinsky J, Markiewicz D, et al. Identification of
- Kerem B, Zielinsky J, Markiewicz D, et al. Identification of mutations in regions corresponding to the two putative nucleotide (ATP)-binding folds of the cyctic fibrosis gene. *Proc Natl Acad Sci USA* 1990;87:8447-51.
 Myers R, Maniatis T, Lerman L, et al. Detection and localisation of single base changes by denaturing gradient gel electrophoresis. *Methods Enzymol* 1987;155:501-27.
 Audrezet MO, Mercier B, Guere I, et al. Identification of 12 novel mutations in the CFTR gene. *Hum Molec Genet* 1993;2:51-4.
 Fanen P, Ghanem N, Vidaud M, et al. Molecular char-

- 1995,2.51-4.
 15 Fanen P, Ghanem N, Vidaud M, et al. Molecular characterisation of the cystic fibrosis gene: 16 novel mutations identified by analysis of the whole cystic fibrosis conductance transmembrane regulator (CFTR) coding region and the investment for the provided in the second secon
- and splice-site junctions. Genomics 1992;13:770-7.
 Sanger F, Nicklen S, Coulson A. DNA sequencing with chain terminating inhibition. Proc Natl Acad Sci USA 1977;74:5463-7.
 Termina P, Schwartz M, Packarten N, and David D. Schwartz M. Sci USA
- 17 Ferrie R, Schwartz M, Robertson N, et al. Developing, multiplexing and application of ARMS tests for common mutations in the CFTR gene. Am J Hum Genet 1992;51: 51 - 62
- 18 Feldman GL, Williamson R, Beaudet AL, et al. Prenatal
- Feldman GL, Williamson R, Beaudet AL, et al. Prenatal diagnosis of cystic fibrosis by DNA amplification for de-tection of KM-19 polymorphism. Lancet 1988;1:102.
 Rosembloom CL, Kerem B, Rommens JM, et al. DNA amplification for detection of the XV-2c polymorphism linked to cystic fibrosis. Nucleic Acids Res 1989;17:7117.
 Huth A, Estivill X, Grade K, et al. Polymerase chain reaction for the detection of the MP6-d9/Msp I RFLP, a marker closely linked to the cystic fibrosis mutation. Nucleic Acids Res 1980;17:7118
- closely linked to the cystic norosis mutation. *Ivacues Actas* Res 1989;17:7118.
 21 Fitzsimons SC. The changing epidemiology of cystic fibrosis. *J Pediat* 1993;12:1-9.
 22 Ferec C, Audrezet MP, Mercier B, et al. Detection of over 98% of cystic fibrosis mutations in a Celtic population. *Nature Genet* 1992;1:188-91.
 23 Cystic Fibrosis Genotype-Phenotype Consortium. Cor-relation between genotype and phenotyme in patients with
- Cystic Fibrosis Genotype-Phenotype Consortium. Correlation between genotype and phenotype in patients with cystic fibrosis. N Engl J Med 1993;329:1308-13.
 Dork T, Wulbrand U, Richter R, et al. Cystic fibrosis with three mutations in the cystic fibrosis transmembrane conductance regulator gene. Hum Genet 1991;87:441-6.
 Schwachman H, Kowalski M, Khew KT. Cystic fibrosis: a new outlook. Medicine (Baltimore) 1977;56:129.
 Kaplan E, Schwachman H, Perlmutter AD, et al. Reproductive failure in males with cystic fibrosis. N Engl J
- productive failure in males with cystic fibrosis. N Engl J Med 1968;279:65.
- 27 Kristidis P, Bozon D, Corev M, et al. Genetic determination of exocrine pancreatic function in cystic fibrosis. Am J Hum Genet 1992;50:1178-84.
- Pignati PF. Cystic Fibrosis. In: Humphries SE, Malcolm S, eds. From genotype to phenotype. Oxford: Bios Scientific Publishers, 1991:19–49.
 Kiesewetter S, Macek M, Davis C, et al. A mutation in

- Kleisewetter S, Macek M, Davis C, et al. A mutation in CFTR produces different phenotypes depending on chro-mosomal background. Nature Genet 1993;5:274-8.
 Audrezet MP, Mercier B, Guillermit H, et al. A mutation in exon 7 of the CFTR gene is common in the western part of France. J Med Genet 1992;29:679.
 Macek M Jr, Macek M, Serre JL, et al. Population study of CFTR gene mutations in Bohemia and Moravia: hy-pothesis on the historical spread of G551D and AF508 in Europe. Am J Hum Genet Suppl 1991;49:A474.
 Dean M, White M, Amos J, et al. Multiple mutations in highly conserved regions are found in mildly affected cystic fibrosis. Cell 1990;61:863-70.
 Beaudet AL, Feldman GL, Fernbach SD, et al. Linkage disequilibrium, cystic fibrosis and genetic counselling. Am J Hum Genet 1989;44:319-26.
 European Working Group on CF Genetics (EWGCFG). Gradient of distribution in Europe of the major CF muta-tion and of its associated haplotype. Hum Genet 1990;85: 436-41. 436 - 41