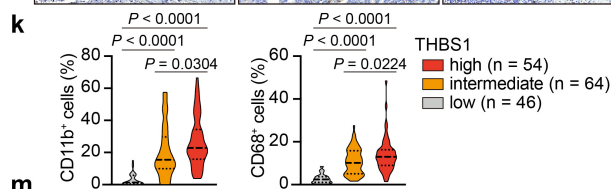
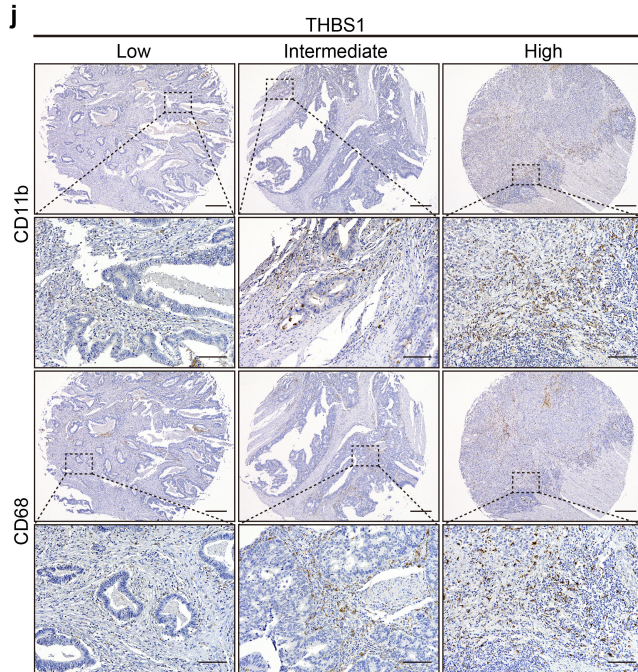
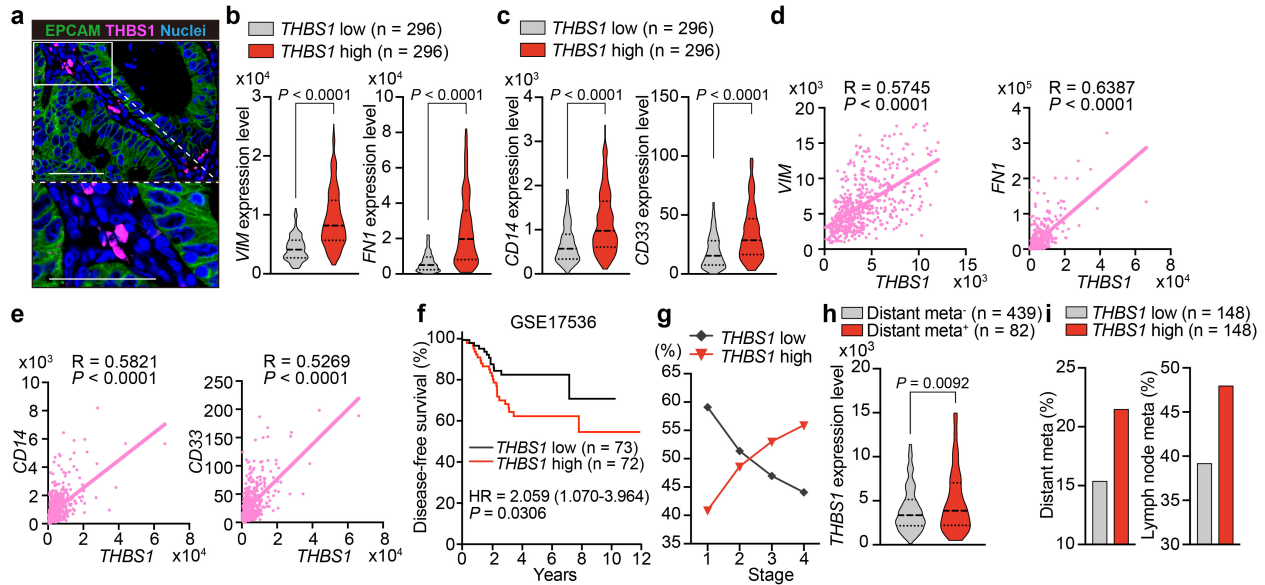


Supplementary Information

THBS1-producing tumor-infiltrating monocyte-like cells contribute to immunosuppression and metastasis in colorectal cancer

Mayuki Omatsu, Yuki Nakanishi, Kosuke Iwane, Naoki Aoyama, Angeles Duran, Yu Muta, Anxo Martinez-Ordoñez, Qixiu Han, Nobukazu Agatsuma, Kenta Mizukoshi, Munenori Kawai, Go Yamakawa, Mio Namikawa, Kensuke Hamada, Yuichi Fukunaga, Takahiro Utsumi, Makoto Sono, Tomonori Masuda, Akitaka Hata, Osamu Araki, Munemasa Nagao, Takaaki Yoshikawa, Satoshi Ogawa, Yukiko Hiramatsu, Motoyuki Tsuda, Takahisa Maruno, Toshiaki Kogame, Hiroaki Kasashima, Nobuyuki Kakiuchi, Masahiro M. Nakagawa, Kenji Kawada, Masakazu Yashiro, Kiyoshi Maeda, Yasuyuki Saito, Takashi Matozaki, Akihisa Fukuda, Kenji Kabashima, Kazutaka Obama, Seishi Ogawa, Nader Sheibani, Maria T. Diaz-Meco, Jorge Moscat, Hiroshi Seno

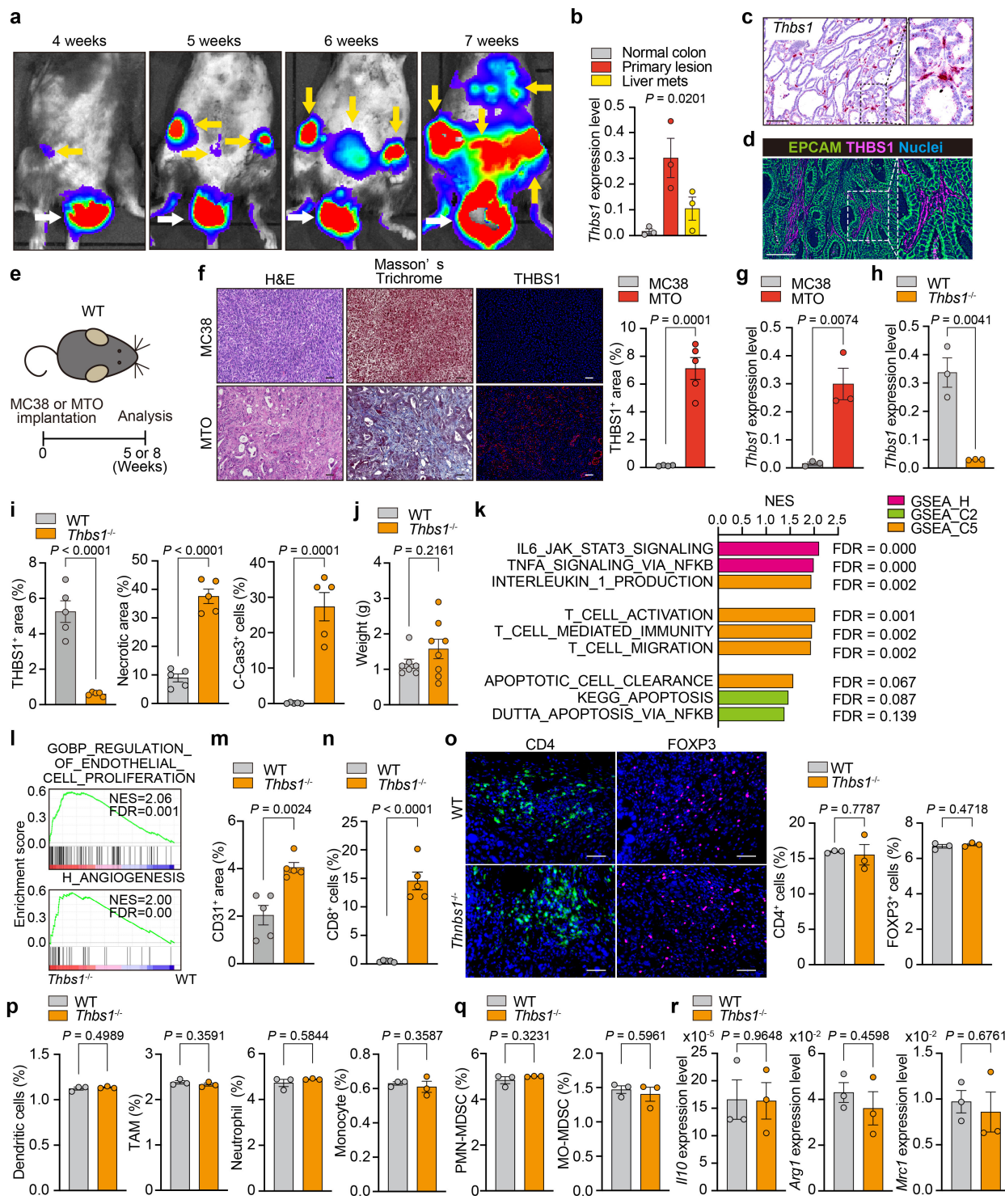


Variables	All patients (n = 331)	THBS1 high (n = 107)	THBS1 intermediate (n = 122)	THBS1 low (n = 102)	P value
Age, years	67.2 (11.2)	66.9 (11.9)	67.4 (10.5)	67.4 (11.2)	0.983
Age \geq 65	202 (60.8)	65 (60.7)	74 (60.7)	63 (61.8)	
Sex	169 (50.9)	43 (40.2)	62 (50.8)	63 (61.8)	0.008
Stage					0.002
1	3 (0.9)	3 (2.8)	0	0	
2	178 (53.6)	44 (41.1)	66 (54.1)	67 (65.7)	
3	151 (45.5)	60 (41.1)	56 (45.9)	35 (34.3)	
TNM classification					0.385
T					
T1	7 (2.1)	4 (3.7)	2 (1.6)	1 (0.98)	
T2	16 (4.8)	8 (7.5)	4 (3.3)	4 (3.9)	
T3	196 (59.0)	65 (60.7)	72 (59.0)	58 (56.9)	
T4	113 (34.0)	30 (28.0)	44 (36.1)	39 (38.2)	
TNM classification					0.035
N					
N0	181 (54.4)	47 (43.9)	66 (54.1)	67 (65.7)	
N1	107 (32.2)	41 (38.3)	44 (36.1)	22 (21.6)	
N2	37 (11.1)	15 (14.0)	11 (9.0)	11 (10.8)	
N3	7 (2.1)	4 (3.7)	1 (0.82)	2 (2.0)	
Primary location	100 (30.1)	36 (33.6)	37 (30.3)	27 (26.5)	0.528
Proximal					
Lymphatic invasion (n = 322)	232 (72.0)	82 (78.1)	78 (65.0)	72 (74.2)	0.078
Venous invasion	71 (21.4)	19 (17.8)	26 (21.3)	26 (25.5)	0.395
MSI-H (n = 301)	97 (29.2)	19 (19.6)	41 (36.0)	37 (41.6)	0.003
CMS4 (n = 118)	30 (25.4)	15 (36.6)	11 (22.0)	4 (14.8)	0.100
Cancer death	44 (13.3)	24 (22.4)	14 (11.5)	6 (5.9)	0.002
Recurrence	78 (23.5)	35 (32.7)	28 (23.0)	15 (14.7)	0.009
Mortality	70 (21.1)	30 (28.0)	28 (23.0)	12 (11.8)	0.013
Undifferentiated	24 (7.2)	13 (12.1)	8 (6.6)	3 (2.9)	0.035

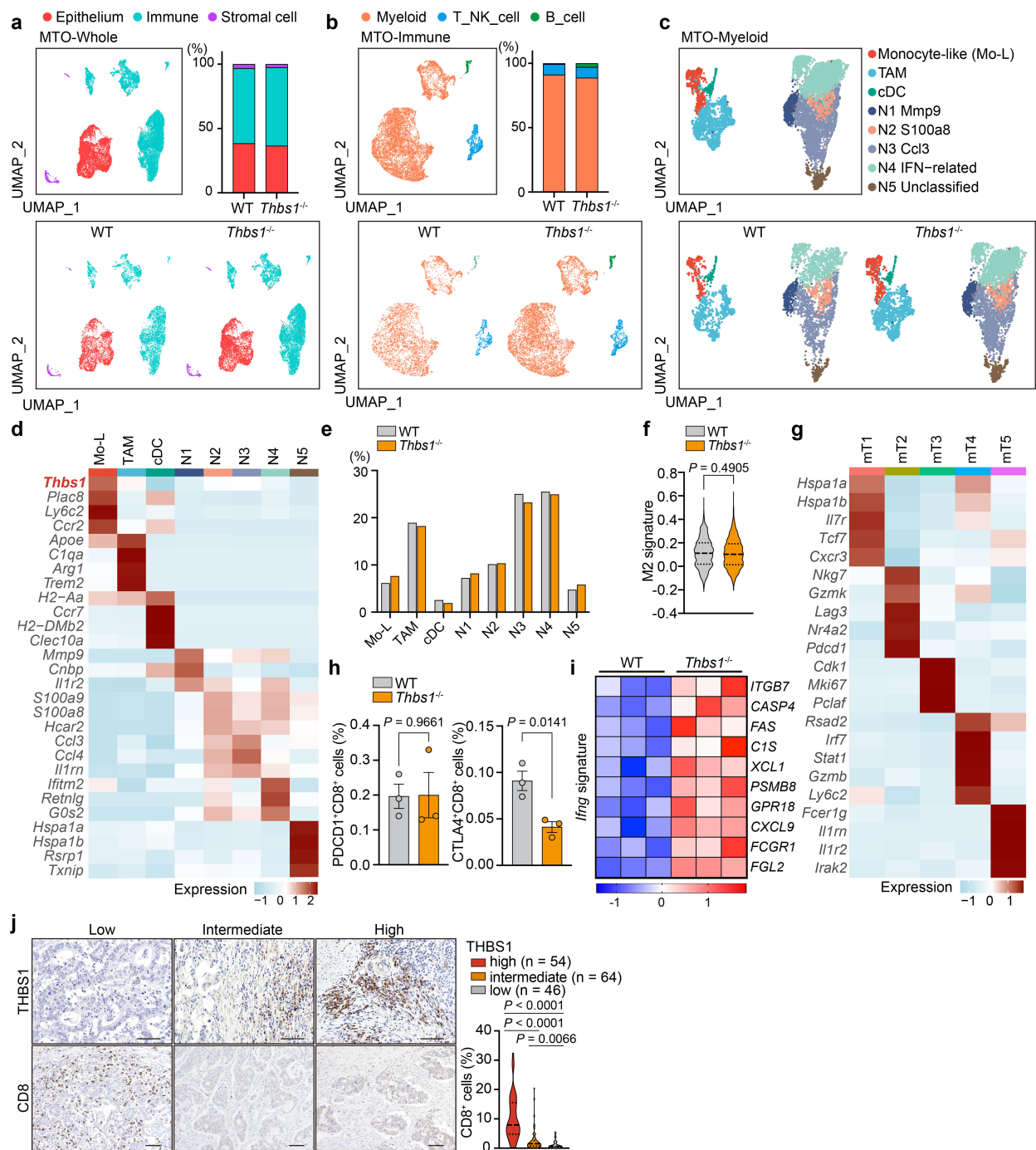
Values are mean (standard deviation) or number (%).

Variables	Recurrence		Lymph node met		Undifferentiated		MSS	
	OR (95%CI)	P value	OR (95%CI)	P value	OR (95%CI)	P value	OR (95%CI)	P value
THBS1		0.046		0.019		0.022		0.004
low	1 (reference)		1 (reference)		1 (reference)		1 (reference)	
intermediate	1.58 (0.76-3.29)		1.67 (0.95-2.96)		2.86 (0.69-11.8)		1.29 (0.72-2.30)	
high	2.48 (1.19-5.18)		2.32 (1.27-4.22)		5.72 (1.44-22.7)		2.94 (1.50-5.74)	

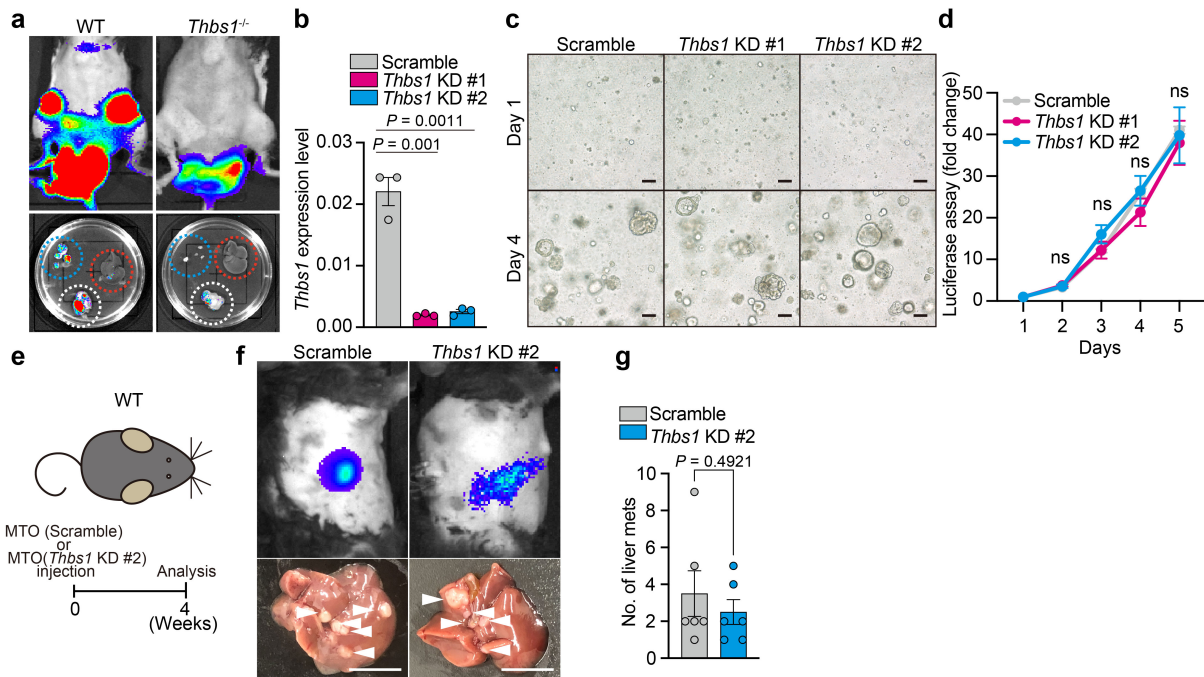
Supplementary Fig. 1 THBS1 expression positively correlated with mesenchyme- and myeloid-related genes and poor prognosis of CRC. **a**, Representative image of co-immunostaining for EPCAM and THBS1 in human CRC from three independent experiments with similar results. **b and c**, Transcript levels of mesenchyme- (**b**) and myeloid-related genes (**c**) in TCGA. **d and e**, Correlation of *THBS1* with indicated genes in TCGA (n = 592). **f**, Kaplan-Meier curve for disease-free survival in human CRC dataset (GSE17536). Hazard ratio with 95% confidence interval and *P* values, analyzed by Log-rank test, are shown. **g**, Proportion of stages 1 to 4 in *THBS1*-low (lower quartile) and *THBS1*-high CRC (upper quartile) in TCGA. **h**, Violin plot for *THBS1* expression in CRC with or without distant metastasis. **i**, Proportion of CRC with distant metastasis (left) or lymph node metastasis (right) in *THBS1*-low (lower quartile) and *THBS1*-high CRC (upper quartile) in TCGA. **j and k**, Immunostaining for CD11b and CD68 in CRC TMA samples (**j**) and proportion of positive cells (**k**). **l**, Basal characteristics and univariable analysis in CRC TMA samples using Pearson's chi-squared test. **m**, Summary of multivariable logistic regression analyses for indicated factors in column label. Detailed results of univariable and multivariable analyses for each factor are shown in Supplementary Tables 1–4. Scale bars, 25 μm (**a**), 200 μm (**j**, top), 50 μm (**j**, bottom). Mean \pm SEM. *P* values were calculated by two-tailed Mann–Whitney test in (**b,c**), Pearson's correlation analyses (**d,e**), and two-tailed, unpaired Student's *t* test in (**h,k**). Source data are provided as a Source Data file.



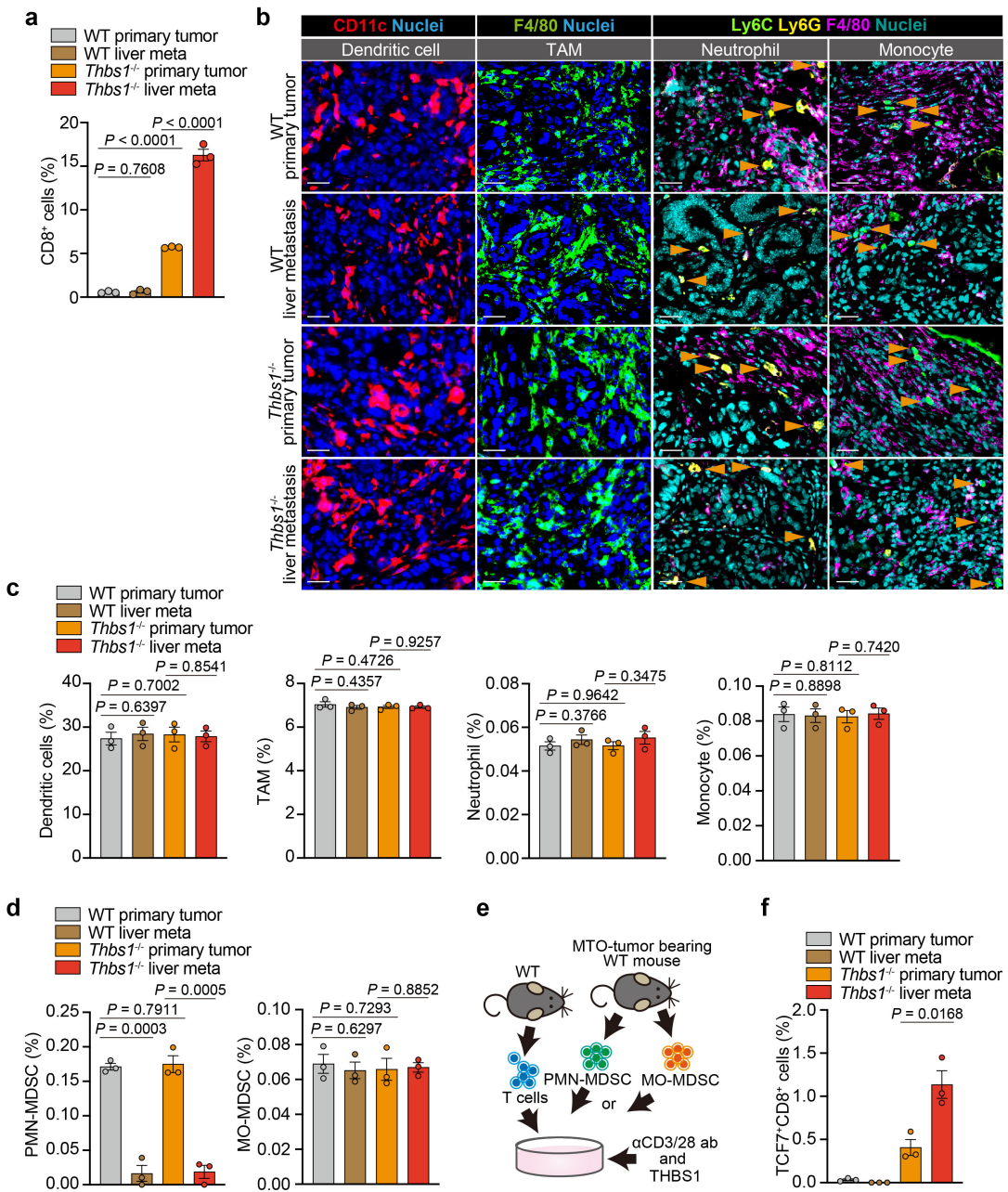
Supplementary Fig. 2 THBS1 is highly expressed in the stroma of mesenchymal tumor and suppresses inflammation, T cell activity, and tumor cell apoptosis. **a**, Bioluminescence imaging of orthotopic implantation of MTO to WT mouse rectum. Arrows: primary lesions (white), distant metastases (yellow). **b**, qRT-PCR analysis of *Thbs1* expression in orthotopic MTO tumor-bearing mice (n = 3). **c**, Representative image of RNAscope for *Thbs1* in the orthotopic MTO tumors in WT mice from two independent experiments with similar results. **d**, Representative image of co-immunostaining for EPCAM and THBS1 in orthotopic WT tumors from three independent experiments with similar results. **e**, Schematic representation of orthotopic implantation of MC38 or MTO to WT mouse rectum. **f**, Staining in the orthotopic MC38 (n = 4) and MTO (n = 5) tumors, and quantification of THBS1 staining. **g**, qRT-PCR analysis in the orthotopic MC38 or MTO tumors (n = 3). **h**, qRT-PCR analysis in the orthotopic MTO tumors in WT and *Thbs1*^{-/-} mice (n = 3). **i**, Quantification of Fig. 2**b** (n = 5). **j**, The weights of the orthotopic MTO tumors in WT (n = 7) and *Thbs1*^{-/-} (n = 8) mice. **k and l**, GSEA on RNAseq data of orthotopic MTO tumors in WT or *Thbs1*^{-/-} mice (n = 3). Normalized enrichment scores (NES) of indicated gene sets upregulated in *Thbs1*^{-/-} mice (**k**) and enrichment score curves (**l**) of GSEA. **m and n**, Quantification (n = 5) of Fig. 2**d** (**m**) and Fig. 2**f** (**n**). **o**, Immunostaining and quantification (n = 3) in the orthotopic MTO tumors in WT or *Thbs1*^{-/-} mice. **p and q**, FACS analyses of proportion of indicated cells in total cells in primary tumors in WT and *Thbs1*^{-/-} mice (n = 3). Dendritic cells, CD45⁺CD11c⁺; TAM, CD11b⁺F4/80⁺; neutrophils, CD45⁺CD11c⁻CD11b⁺Ly6C⁺Ly6G⁺F4/80⁻; monocytes, CD45⁺CD11c⁻CD11b⁺Ly6C⁺Ly6G⁻F4/80⁻; PMN-MDSC, CD45⁺CD11c⁻CD11b⁺Ly6C⁺Ly6G⁺; MO-MDSC, CD45⁺CD11c⁻CD11b⁺Ly6C⁺Ly6G⁻ cells. **r**, qRT-PCR analyses for indicated genes in the orthotopic MTO tumors in indicated mice (n = 3). Scale bars, 50 μm. Mean ± SEM. *P* values were calculated by one-way ANOVA in **b**, two-tailed, unpaired Student's *t* test in the rest. Source data are provided as a Source Data file.



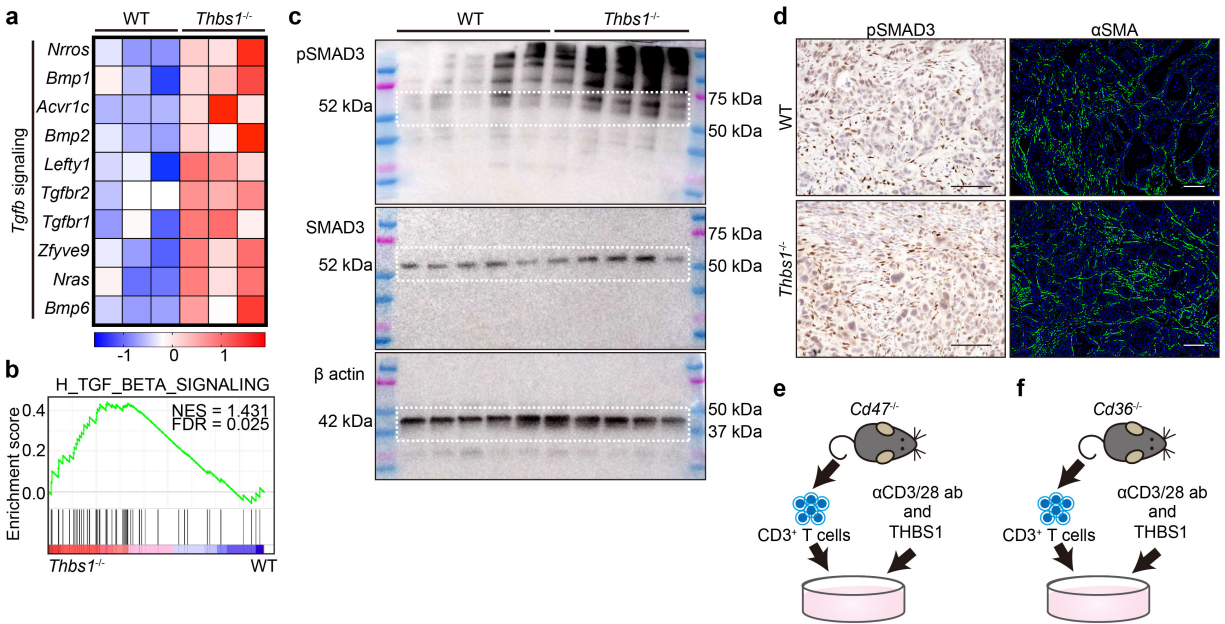
Supplementary Fig. 3 Immune landscapes of orthotopic MTO tumors in WT or *Thbs1*^{-/-} mice. **a**, UMAP plots for whole cells and proportion of each compartment in scRNA-seq of the orthotopic MTO tumors in WT and *Thbs1*^{-/-} mice (n = 2 tumors from two distinct mice per group, analyzed cell numbers are indicated in **Methods “Single-cell RNA sequencing”**). **b**, UMAP plots and proportion of re-clustered immune subsets in **(a)**. **c–e**, UMAP plots **(c)**, heatmap for representative genes **(d)**, and proportions **(e)** of re-clustered myeloid subsets in **(b)**. **f**, Violin plot for M2-related macrophage signature in re-clustered myeloid subsets of WT and *Thbs1*^{-/-} tumors in **(b)**. **g**, Heatmap for representative genes of each cluster of CD8 T subset in Fig. 2**g**. **h**, FACS analyses of proportion of indicated cells in total cells in the orthotopic MTO tumors in WT or *Thbs1*^{-/-} mice (n = 3). **i**, Heatmap of *Ifng* gene signature (Hallmark Collection of MSigDB) on the RNA-seq data from WT and *Thbs1*^{-/-} tumors (n = 3). **j**, Immunostaining in human CRC with low-, intermediate- or high-intensity of THBS1 in TMA samples. Scale bars, 50 μm. Mean ± SEM. *P* values were calculated by two-tailed, unpaired Student’s *t* test. Source data are provided as a Source Data file.



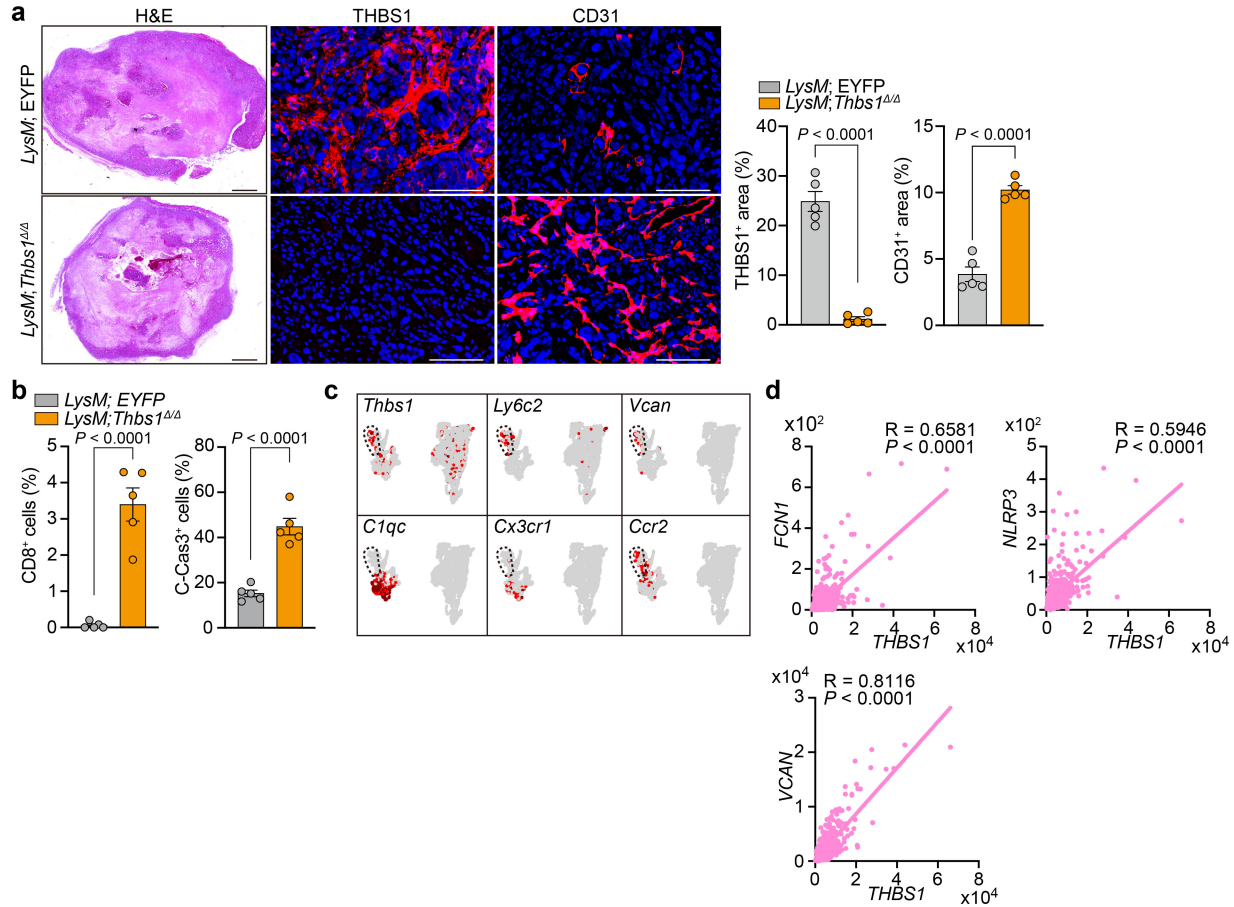
Supplementary Fig. 4 THBS1 loss in tumor epithelium does not affect progression of orthotopic MTO tumors. **a**, In vivo (top) and ex vivo (bottom) bioluminescence imaging of orthotopically MTO-inoculated WT or *Thbs1*^{-/-} mice. Dash lines denote primary rectal tumor (white), liver (red), or lymph nodes (blue). **b**, qRT-PCR analysis in control (scramble) or *Thbs1*-knockdown (*Thbs1* KD) MTO (n = 3). **c and d**, MTO growth assay. Representative images of organoids (**c**), and growth curve (n = 3), measured by luminescence intensity at indicated time (**d**, relative value to day 1). **e**, Schematic representation of splenic injection of *Thbs1*-knockdown MTO to WT mice (n = 6). **f and g**, Bioluminescence imaging (**f**, top) and macroscopic images (**f**, bottom) of liver, and numbers of liver metastases (**g**) of (**e**). White arrowheads denote liver metastases. Scale bars, 100 μ m (**c**), 1 cm (**f**). Mean \pm SEM. *P* values were calculated by two-tailed, unpaired Student's *t* test. Exact *P*-values are shown in the source data. Source data are provided as a Source Data file.



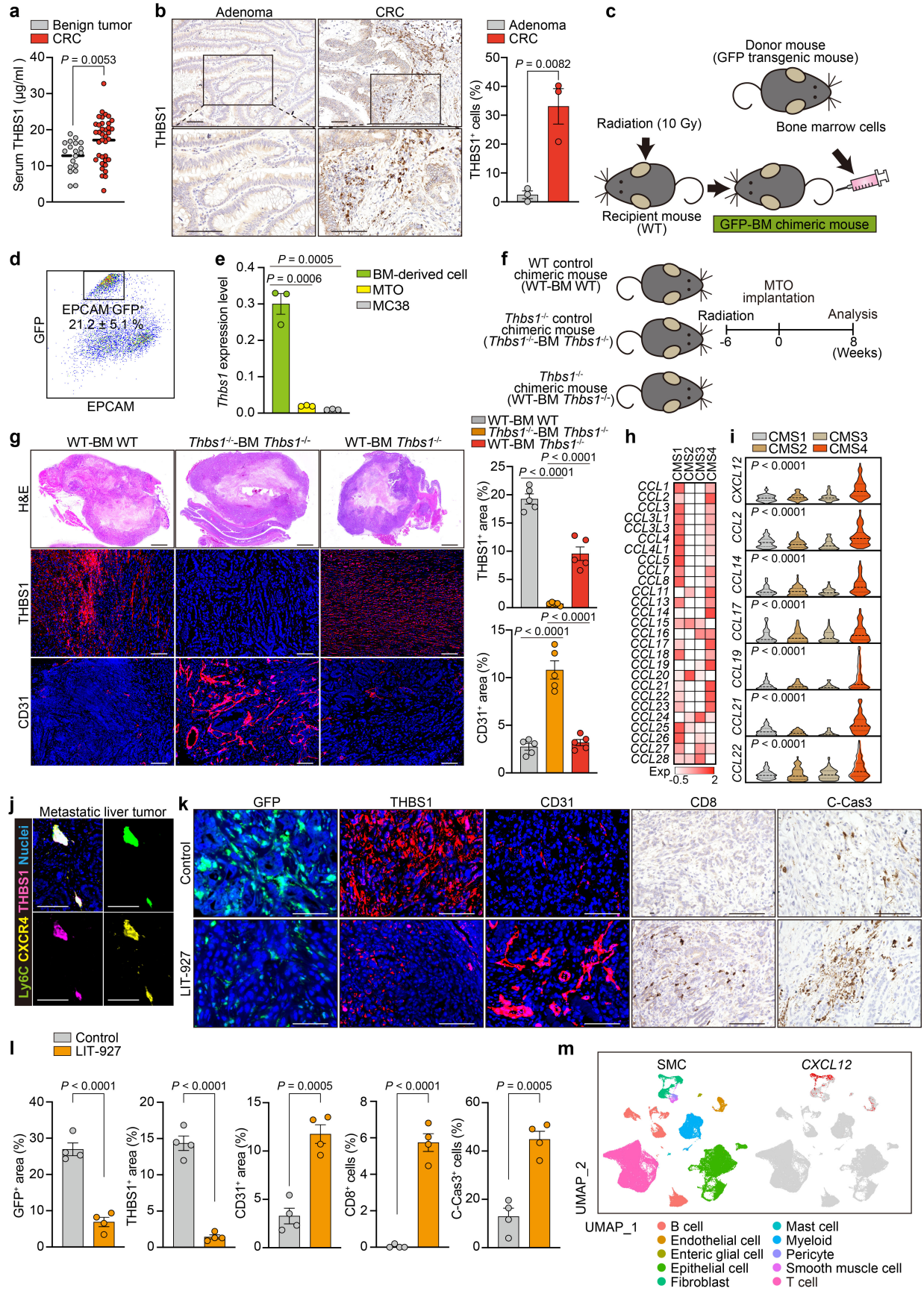
Supplementary Fig. 5 Impact of THBS1 loss on the immune TME of primary and metastatic lesions in orthotopic MTO tumors. a, Quantification of Fig. 3m (n = 3). **b and c**, Immunostaining for CD11c or F4/80 and co-immunostaining (**b**) for Ly6C, Ly6G and F4/80 in primary tumors and metastatic liver tumors and quantification (**c**, n = 3) in orthotopically MTO-inoculated WT or *Thbs1*^{-/-} mice. Orange arrows denote indicated cellular types. Scale bars, 50 μ m (**b**). **d**, Quantification of Fig. 3q (n = 3). PMN-MDSC, Ly6C⁺Ly6G⁺; MO-MDSC, Ly6C⁺Ly6G⁻ cells. **e**, Schematic representation of MDSC assay. **f**, Quantification of TCF7⁺CD8⁺ cells in Fig. 3s (n = 3). Mean \pm SEM. *P* values were calculated by two-tailed, unpaired Student's *t* test. Source data are provided as a Source Data file.



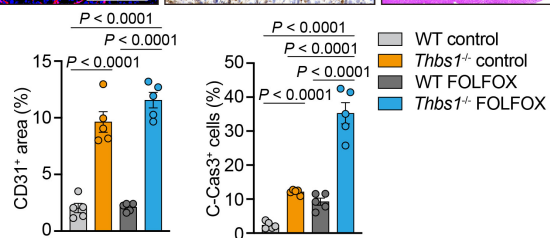
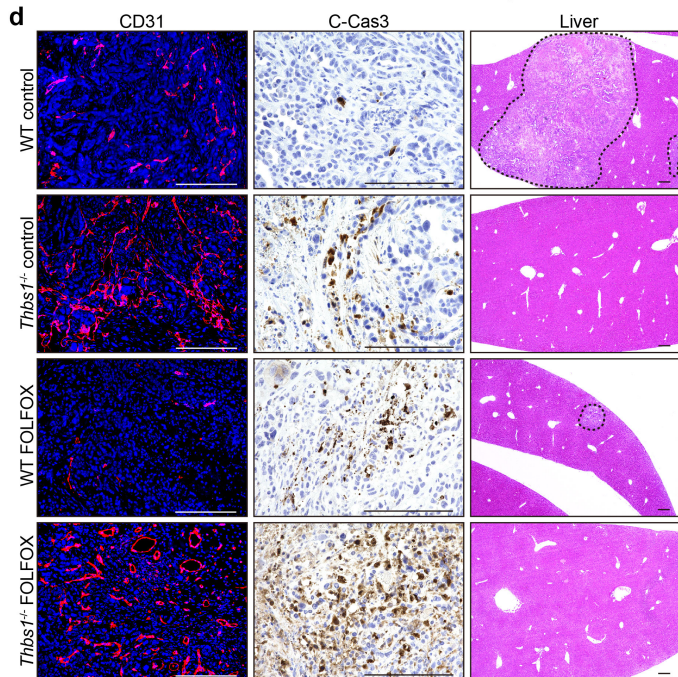
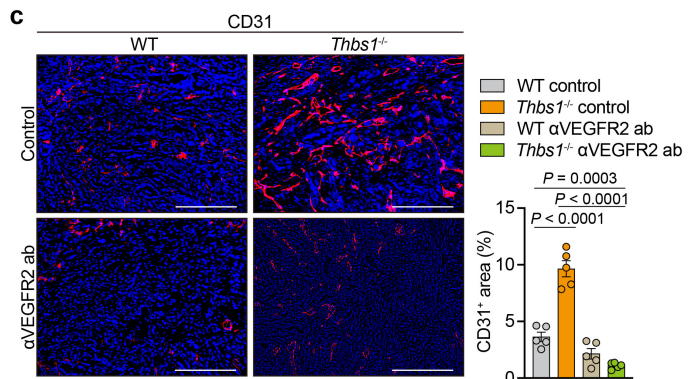
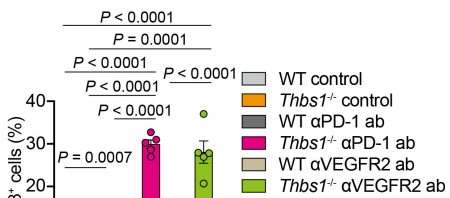
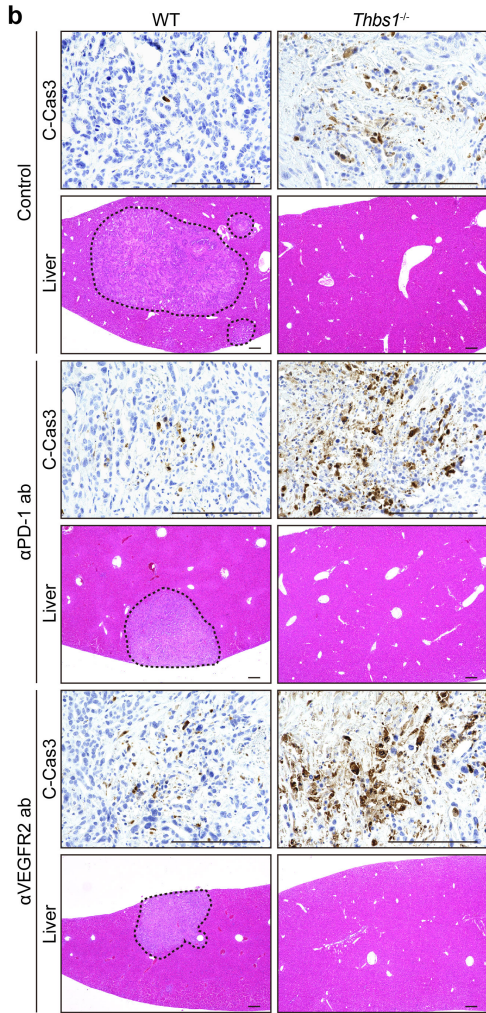
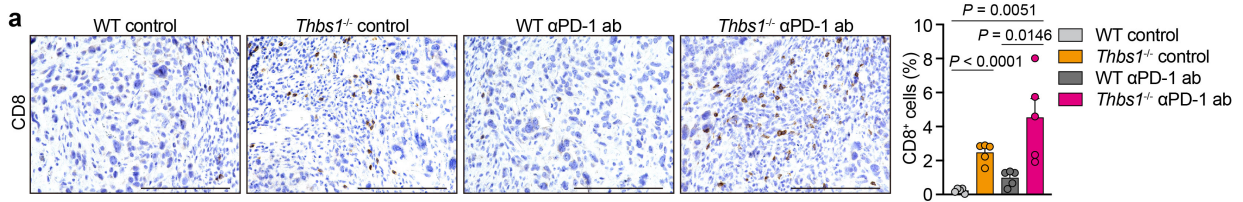
Supplementary Fig. 6 TGFβ signaling pathway in orthotopic MTO tumors in WT or *Thbs1*^{-/-} mice. **a** and **b**, Heatmap of representative genes (**a**) and GSEA (**b**) of TGFβ signaling pathway signature (Hallmark Collection of MSigDB) on the RNA-seq data from the orthotopic MTO tumors in WT or *Thbs1*^{-/-} mice (n = 3). **c**, Immunoblotting of phospho-SMAD3 (pSMAD3) and SMAD3 in the orthotopic MTO tumors in WT or *Thbs1*^{-/-} mice (n = 5). Dash lines denote predicted molecular weight for each protein. **d**, Representative immunostaining of pSMAD3 and αSMA in the orthotopic MTO tumors in WT or *Thbs1*^{-/-} mice from three independent experiments with similar results. Scale bars, 50 μm. **e** and **f**, Schematic representations of anti-CD3/CD28-mediated stimulation experiment of CD3⁺ T cells isolated from *Cd47*^{-/-} (**e**) and *Cd36*^{-/-} (**f**) mice. Source data are provided as a Source Data file.



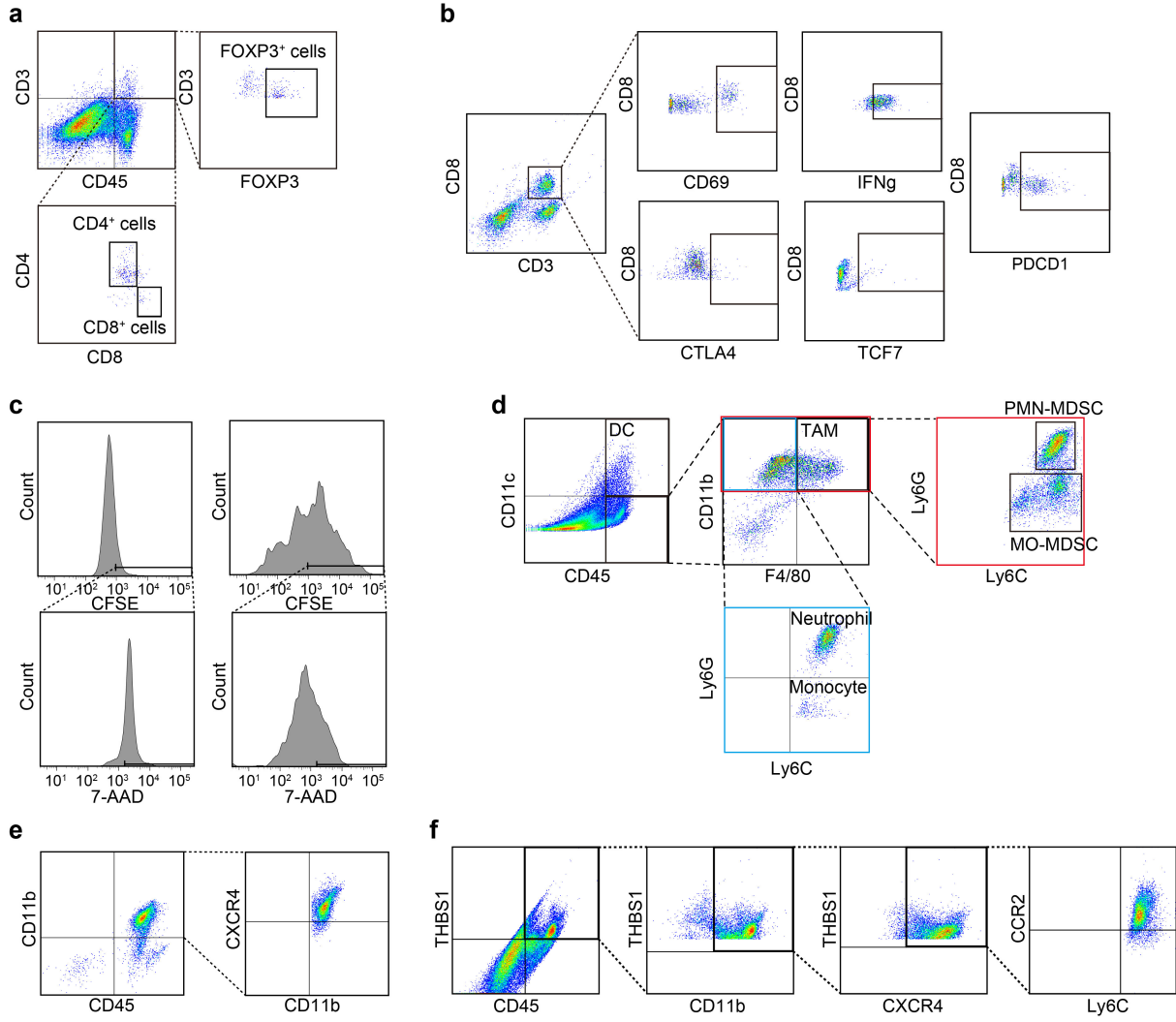
Supplementary Fig. 7 THBS1 is produced by monocyte/macrophage lineages in mesenchymal CRC. **a**, H&E and immunostaining in the orthotopic MTO tumors in *LysM;EYFP* or *LysM;Thbs1^{Δ/Δ}* mice and quantification (n = 5). Scale bars, 200 μ m (H&E), 50 μ m (the rest). **b**, Quantification of (Fig. 5g, n = 5). **c**, Dot plots of expression of selected genes in scRNA-seq data of the orthotopic MTO tumors in Supplementary Fig. 3c. Dash lines denote monocyte-like cluster. **d**, Correlation of *THBS1* with *FCN1*, *NLRP3*, or *VCAN* in TCGA (n = 592). Mean \pm SEM. *P* values were calculated by two-tailed, unpaired Student's *t* test (**a,b**) or Pearson's correlation analyses (**d**). Source data are provided as a Source Data file.



Supplementary Fig. 8 THBS1-expressing monocyte-like cells are recruited to primary tumor site via CXCL12/CXCR4 signaling. **a**, Serum THBS1 levels in patients with CRC (n = 37) or benign tumors (n: adenoma = 15, sessile serrated lesion = 4). **b**, Immunostaining of THBS1 in human adenoma and CRC and quantification (n = 3). Scale bars, 50 μ m. **c**, Schematic representation of generation of GFP-BM chimeric mice. **d**, Representative FACS analyses of orthotopic MTO tumors in GFP-BM chimeric mice (n = 3). **e**, qRT-PCR analysis for *Thbs1* in BM-derived (EPCMA⁺GFP⁺) cells in GFP-BM chimeric mice compared to MTO and MC38 cells (n = 3). **f**, Schematic representation of orthotopic implantation of MTO to the indicated BM-chimeric mice. **g**, H&E and immunostaining in the orthotopic MTO tumors of indicated BM-chimeric mice and quantification (n = 5). Scale bars, 200 μ m (H&E), 50 μ m (the rest). **h**, Heatmap for transcript expression of CC-chemokines in CRC from TCGA, stratified by CMS subtypes. **i**, Transcript levels of indicated genes in TCGA stratified by CMS classification (n: CMS1 = 85, CMS2 = 132, CMS3 = 78, CMS4 = 184). **j**, Representative co-immunostaining for Ly6C, CXCR4 and THBS1 in liver metastasis in orthotopically MTO-inoculated WT mice from three independent experiments with similar results. Scale bars 50 μ m. **k and l**, Immunostaining (**k**) and quantification (**l**; n = 5) in the orthotopic MTO tumors in GFP-BM chimeric mice with or without administration of LIT-927. Scale bars, 50 μ m. **m**, UMAP plot for whole cells showing cellular subsets and *CXCL12*-expressing cells of human CRC scRNA-seq data (SMC). Mean \pm SEM. *P* values were calculated by two-tailed Mann–Whitney test in (**a**), two-tailed unpaired Student's *t* test in (**b,e,g,l**), or one-way ANOVA in (**i**). Source data are provided as a Source Data file.



Supplementary Fig. 9 Loss of THBS1 partially improved response of aggressive CRC to current treatments **a**, Immunostaining for CD8 in the primary lesions in MTO-bearing WT or *Thbs1*^{-/-} mice with or without administration of anti-PD-1 antibody (α PD-1 ab), and quantification (n = 5). **b**, Immunostaining for C-Cas3 in the primary lesions and H&E staining of liver in MTO-bearing WT or *Thbs1*^{-/-} mice with or without administration of α PD-1 ab or anti-VEGFR2 antibody (α VEGFR2 ab), and quantification (n = 5). **c**, Immunostaining for CD31 in the primary lesions in MTO-bearing WT or *Thbs1*^{-/-} mice with or without administration of α VEGFR2 ab, and quantification (n = 5). **d**, Immunostaining for CD31 and C-Cas3 in the primary lesions and H&E staining of liver in MTO-bearing WT or *Thbs1*^{-/-} mice with or without administration of FOLFOX, and quantification (n = 5). Dash lines denote liver metastases. Mean \pm SEM. *P* values were calculated by two-tailed, unpaired Student's *t* test. Scale bars, 100 μ m. Source data are provided as a Source Data file.



Supplementary Fig. 10 FACS gating strategies a, Gating strategy for CD4⁺ T, CD8⁺ T, and FOXP⁺ T cells from orthotopic MTO tumors, related with Fig. 2e and 3n. T cells were collected from dissociated tumor as CD45⁺CD3⁺ cells. CD4⁺ T, CD8⁺ T, or FOXP3⁺ T cells were sorted from CD45⁺CD3⁺ cells. **b**, Gating strategies for active or dysfunctional CD8⁺ T cells, related with Fig. 2n, 3r, 4h, and 4l and Supplementary Fig. 3h. Active CD8⁺ T cells were collected as CD3⁺CD8⁺CD69⁺ or CD3⁺CD8⁺IFN γ ⁺ cells. Dysfunctional CD8⁺ T cells were collected as CD3⁺CD8⁺CTLA4⁺ or Cd3⁺CD8⁺PDCD1⁺ cells. Stem-like CD8⁺ T cells were collected as CD3⁺CD8⁺TCF7⁺ cells. **c**, Gating strategies for cytotoxicity assay related with Fig. 2q. Dead cells were measured with 7-AAD in MTO, pre-stained with CFSE. **d**, Gating strategies for myeloid cells in orthotopic MTO tumors, related with Fig. 3o and 3p and Supplementary Fig. 2p and 2q. Dendritic cells, TAMs, neutrophils, monocytes, PMN-MDSC, and MO-MDSC were sorted as CD45⁺CD11c⁺, CD45⁺CD11c⁻CD11b⁺F4/80⁺, CD45⁺CD11c⁻CD11b⁺F4/80⁻Ly6G⁺Ly6C⁺, CD45⁺CD11c⁻CD11b⁺F4/80⁻Ly6G⁻Ly6C⁺, CD45⁺CD11c⁻CD11b⁺Ly6G⁺Ly6C⁺, and CD45⁺CD11c⁻CD11b⁺Ly6G⁻Ly6C⁺, respectively. **e**, Strategies for sorting CD45⁺CD11b⁺CXCR4⁺ cells in BM or peripheral blood cells, related with Fig. 6a–c, 6o. **f**, Gating strategy for sorting THBS1⁺ cells in orthotopic MTO tumors, related with Fig. 6p.

Supplementary Table 1 Univariable and multivariable logistic regression analysis for recurrence

Variables	Univariable analysis, odds ratio (95%CI)	P value	Multivariable analysis, odds ratio (95%CI)	P value
THBS1		0.008		0.046
Low (reference)	1		1	
Intermediate	1.73 (0.87-3.45)		1.58 (0.76-3.29)	
High	2.82 (1.43-5.57)		2.48 (1.19-5.18)	
T		0.064		0.097
T1 (reference)	1		1	
T2	0.36 (0.04-3.26)		0.34 (0.03-3.40)	
T3	0.61 (0.11-3.24)		0.98 (0.17-5.84)	
T4	1.17 (0.22-6.31)		1.64 (0.27-9.86)	
N		< 0.0001		0.564
N0 (reference)	1		1	
N1	3.46 (1.91-6.27)		1 (1)	
N2	4.65 (2.11-10.2)		1.28 (0.56-2.91)	
N3	9.10 (1.91-43.3)		2.25 (0.45-11.3)	
Stage		<0.0001		0.0009
1 (reference)	1		1	
2	598053.8 (-)		442015.7	
3	2294164.7 (-)		1378820.7	
Histology type		0.030		0.649
Undifferentiated	2.51 (1.07-5.90)		1.26 (0.47-3.35)	

Univariable analysis was performed using Pearson's chi-squared test (left). Two-sided and unadjusted multivariable logistic regression analysis for the factors associated with recurrence was performed (right).

Supplementary Table 2 Univariable and multivariable logistic regression analysis for lymph node metastasis positive

Variables	Univariable analysis, odds ratio (95%CI)	P value	Multivariable analysis, odds ratio (95%CI)	P value
THBS1		0.007		0.019
Low (reference)	1		1	
Intermediate	1.62 (0.94-2.79)		1.67 (0.95-2.96)	
High	2.44 (1.40-4.28)		2.32 (1.27-4.22)	
T		< 0.0001		0 < 0.0001
T1 (reference)	1		1	
T2	1.17 (0.09-15.5)		1.23 (0.09-16.6)	
T3	0.09 (0.01-0.79)		0.10 (0.01-0.86)	
T4	0.20 (0.02-1.68)		0.21 (0.02-1.85)	
Histology type		0.010		0.056
Undifferentiated	3.14 (1.26-7.78)		2.45 (0.95-6.32)	

Univariable analysis was performed using Pearson's chi-squared test (left). Two-sided and unadjusted multivariable logistic regression analysis for the factors associated with lymph node metastasis was performed (right).

Supplementary Table 3 Univariable and multivariable logistic regression analysis for undifferentiated histology

Variables	Univariable analysis, odds ratio (95%CI)	P value	Multivariable analysis, odds ratio (95%CI)	P value
THBS1		0.032		0.022
Low (reference)	1		1	
Intermediate	2.32 (0.60-8.97)		2.86 (0.69-11.8)	
High	4.56 (1.26-16.5)		5.72 (1.44-22.7)	
T		0.069		0.058
T1 (reference)	1		1	
T2	266957.4 (-)		1678320.7 (-)	
T3	193759.4 (-)		1361536.4 (-)	
T4	566273.2 (-)		4479693.3 (-)	
N		0.001		0.003
N0 (reference)	1		1	
N1	1.47 (0.48-4.49)		1 (-)	
N2	7.94 (2.74-23.1)		6.58 (2.03-21.4)	
N3	9.89 (1.63-60.1)		7.29 (1.04-51.2)	
Stage		0.031		0.087
1 (reference)	1		1	
2	164885.4 (-)		1181578.8 (-)	
3	508015.9 (-)		1122911.6 (-)	

Univariable analysis was performed using Pearson's chi-squared test (left). Two-sided and unadjusted multivariable logistic regression analysis for the factors associated with undifferentiated histology was performed (right).

Supplementary Table 4 Univariable and multivariable logistic regression analysis for MSS status

Variables	Univariable analysis, odds ratio (95%CI)	P value	Multivariable analysis, odds ratio (95%CI)	P value
THBS1		0.003		0.004
Low (reference)	1		1	
Intermediate	1.27 (0.72-2.24)		1.29 (0.72-2.30)	
High	2.92 (1.52-5.56)		2.94 (1.50-5.74)	
T		0.020		0.082
T1 (reference)	1		1	
T2	1.42 x 10 ⁻⁶ (-)		6.16 x 10 ⁻⁷ (-)	
T3	5.75 x 10 ⁻⁷ (-)		2.58 x 10 ⁻⁷ (-)	
T4	3.75 x 10 ⁻⁷ (-)		1.85 x 10 ⁻⁷ (-)	
Primary tumor location		0.042		0.070
Proximal	0.59 (0.35-0.98)		0.61 (0.36-1.04)	

MSS: microsatellite stable.

Univariable analysis was performed using Pearson's chi-squared test (left). Two-sided and unadjusted multivariable logistic regression analysis for the factors associated with MSS was performed (right).

Supplementary Table 5 Mouse qRT-PCR primers

Gene	Forward	Reverse
<i>Thbs1</i>	GAAGCAACAAGTGGTGTCTCAGT	ACAGTCTATGTAGAGTTGAGCCC
<i>Ifng</i>	ATGAACGCTACACACTGCATC	CCATCCTTTTGCCAGTTCCTC
<i>Ctla4</i>	GTACCTCTGCAAGGTGGAAGCTC	CCAAAGGAGGAAGTCAGAATCCG
<i>Cd11b</i>	ATGGACGCTGATGGCAATACC	TCCCCATTCACGTCTCCCA
<i>Cxcl12</i>	TGCATCAGTGACGGTAAACCA	TTCTTCAGCCGTGCAACAATC
<i>Fcn1</i>	CACCATCCATCTTCCTGACTGC	ATAGGAGTCCCAGTCTCGGAAG
<i>Nlrp3</i>	ATTACCCGCCCGAGAAAGG	CATGAGTGTGGCTAGATCCAAG
<i>Ccl2</i>	TTAAAAACCTGGATCGGAACCAA	GCATTAGCTTCAGATTTACGGGT
<i>18s</i>	GTAACCCGTTGAACCCATT	CCATCCAATCGGTAGTAGCG
<i>IL10</i>	GCTCTTACTGACTGGCATGAG	CGCAGCTCTAGGAGCATGTG
<i>Vcan</i>	TTTTACCCGAGTTACCAGACTCA	GGAGTAGTTGTTACATCCGTTGC
<i>Arg1</i>	TGGCTTGCGAGACGTAGAC	GCTCAGGTGAATCGGCCTTTT
<i>Mrc1</i>	GCTGAATCCCAGAAATTCCGC	ATCACAGGCATACAGGGTGAC

Supplementary Table 6 Representative genes in indicated signatures

Signature	Genes
CMS4_UP ¹	<i>MSRB3, FERMT2, EFEMP2, SPOCK1, DDR2, TAGLN, CCDC80, GLI3, TNS1, STON1, PTRF, SFRP2, MGP, GAS1, BNC2, SLIT2, DPYSL3, AEBP1, PCDH7, MAP1B, CRYAB, PRRX1, FBN1, MXRA8, PTGIS, ZFPM2, MLLT11, MYL9</i>
CMS_DOWN ¹	<i>SEPT1, RMI1, ASF1B, STIL, CCNA2, HMGB2, CDC45, KIF18A, UNG, WHSC1, KIF18B, HK2, RBM47, DONSON, PLK4, CCDC134, EIF4E, RHPN2, FAM83F, FANCD2, HOOK1, CDCA2, EZH2, GMCL1, CENPA, ORC1, SPAG5, TMEM54</i>
Stem-like signature ²	<i>Tcf7, Xcl, Cxcr5, Ltb, Pdc4, Irf3, Slamf6, Cd200</i>
Dysfunction signature ³	<i>Ccl3, Cxcl13, Klrl3dl1, Klrl3dl2, Ifng, Cd7, Cd27, Akap5, Havcr2, Tnfrsf1b, Tnfrsf9, Lyst, Ptms, Tns3, Gbp2b, Entpd1, Pdc1, Phlda1, Csf1, Rdh10, Ctla4, Dgkh, Snap47, Itgae, Il2rb</i>
M2 macrophage-related signature ⁴	<i>Ccl4, Ccl13, Ccl20, Ccl22, Ccl276, Clec7a, Ctsa, Ctsb, Ctsc, Ctsd, Fn1, Il4ra, Irf4, Lyve1, Mmp9, Mmp14, Mmp19, Msr1, Tgfb1, Tgfb2, Tgfb3, Tnfsf8, Tbsf12, Vegfa, Vegfb, Vegfc</i>

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