

Supporting Information for

A peptide-binding domain shared with an Antarctic bacterium facilitates *Vibrio cholerae* human cell binding and intestinal colonization

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Supporting Information Text

Plasmid construction:

Plasmid pKEK2100 was created by PCR amplification of the R6K ori and oriT from pCVD442 (1) with primers R6K oriT F and R6K oriT R, and cloning via In Vivo Assembly (IVA) (2) into pKEK2094 PCR amplified with pUC118 ori F and pUC118 ori R. Δ PBD, Δ RIII_4, Δ IGL, Δ SBD, Δ UKD, and Δ 5D deletion constructs were synthesized as ds DNA with approximately 600 bp flanking the deleted domain coding sequence (IDT), which were as indicated in Fig. S1. The Δ 5D construct was PCR amplified with the Universal Δ Up and Universal Δ Dwn primers listed in Table S1, then digested with *NotI* and *ApaI* and ligated into pKEK229 digested similarly to form pKEK2054. The constructs Δ PBD, Δ RIII_4, Δ IGL, and Δ SBD were PCR amplified with the corresponding primers listed in Table S1 and cloned via IVA (2) into pKEK2100 PCR amplified with primers pUC118 MCS F and pUC118 MCS R. These fragments were then digested with *NotI* and *ApaI* and ligated into pKEK229 digested similarly to form pKEK2119 (Δ SBD), pKEK2120 (Δ IGL), pKEK2121 (Δ PBD), and pKEK2124 (Δ RIII_4). The Δ UKD construct was PCR amplified with Δ UKD F and Δ UKD R, and cloned via IVA (2) into pKEK2200 PCR amplified with primers pUC118 MCS F and pUC118 MCS R to form pKEK2157 (Δ UKD).

The deletions in *frhB* and *frhD* were constructed by Splicing by Overlap Extension (3) with the primers listed in Table S1, and the resulting deletion fragments were inserted into pKEK2100 PCR-amplified with primers pUC118 MCS F and pUC118 MCS R by IVA (2), resulting in pKEK2110 and pKEK2114. These plasmids were then digested with *NotI* and *ApaI*, and ligated into pKEK229, resulting in plasmids pKEK2122 (Δ *frhB*) and pKEK2123 (Δ *frhD*). The deletions in *lapD* and *lapG* in strains FY_VC_12120 and Fy_VC_11863 were PCR-amplified with the indicated primers and cloned by IVA (2) into pKEK2200 PCR-amplified with primers pUC118 MCS F and pUC118 MCS R to create pKEK2265 and 2266 respectively.

The Mp PBD was synthesized as a 620 bp ds DNA fragment (IDT), then PCR-amplified with primers MpIBP RIII-3 F and MpIBP RIII-3 R. This fragment was inserted via IVA (2) into pKEK2121 PCR-amplified with primers RIII-3VC>MP F and RIII-3VC>MP R, resulting in pKEK2335, which has the Vc PBD replaced with Mp PBD (*frhA*^{MpPBD}). pKEK2121 was also PCR-amplified with RIII-3VC>AbR F and RIII-3VC>AbR F, and a KanR fragment PCR-

amplified from SAD034 with primers ABD123 and ABD124 (4); these were assembled by IVA (2) resulting in pKEK2447, which replaces PBD with KanR.

***V. cholerae* strain construction:**

The *V. cholerae* mutants were generated through allelic exchange using the plasmids pKEK229 (5) or pKEK2200 (6) via *sacB* counterselection. The Vc strain expressing FrhA^{MpPBD} was created by first introducing *frhA::KanR* onto the Vc chromosome from pKEK2447, resulting in KKV3451. The FrhA^{MpPBD} in pKEK2335 was then introduced into the KKV3451 chromosome by replacing KanR, resulting in KKV3540.

Purification and Labelling of FrhA-PBD:

FrhA-PBD and its attendant split domain, which correspond to the red and orange domains in Figure 1B, respectively, spanning amino acids 1,155-1,460, were produced together as one recombinant protein. The codon-optimized DNA (GeneArt) for this 35-kDa construct was cloned between the *NdeI* and *XhoI* sites of the pET28a plasmid vector and transfected into *E. coli* BL-21 (DE3). After IPTG-induction of the *E. coli* culture, cells were harvested and lysed by sonication as previously described (7). FrhA-PBD was purified by affinity chromatography on nickel-NTA and size-exclusion chromatography on a HiLoad 16/60 Superdex-75 size-exclusion column (GE Healthcare) with a yield of 5 mg/L of culture.

FrhA-PBD in HEPES buffer (pH 8.5) was incubated in covered tubes for 2 h with fluorescein-5-isothiocyanate (FITC) or tetramethylrhodamine (TRITC) at a 20:1 molar ratio. The mixtures were then passed through a size-exclusion column equilibrated with 20 mM Tris-HCl (pH 9.0), 200 mM NaCl, 5 mM CaCl₂ followed by elution with the same buffer.

atgggaattcatgctttgttgtctttaacaaatctagcagcaaatacagctgctcgtgatt
M G I H A L L S L T N L A A N Q L L V I
gataaaaacggaaatatcgcgatcatcaacgcaggagaggcggttcctgaagggtgcgatt
D K N G N I A I I N A G E A V P E G A I
atcctcgacccgaatagtaacaatgatgcctgagcaggagccactgcccgtagcaca
I L D P N S N N L M P E Q E P L P V A Q
ctgggtgatgctgagggtaacgtccagccgatcaccgacgatatagagcagattttagcc
L V D A E G N V Q P I T D D I E Q I L A
gcattagaagaaggcgcagacccgactgcactagatgatctcgccccagcagcagggtggc
A L E E G A D P T A L D D L A **P A A G G**
cttcaagggtcatccatcacgggtagcgcctccattgagcgtgatggtgctgaaaccatc
L Q G S S I T G S A S I E R D G A E T I
gcgtaacacaatgtgatacatccggcttcgaagccattggcctatcaagaacgcaaagc
A S T Q F D T S G F E A I G L S R T Q S
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L S L L N L L Q A P T A P I T P I P P V
gatccagaagagcctgcctctcccattgtaattagcagtattacaggcgataacgcagcc
D P E E P A S P I V I S S I T G D N A A
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E G S N N T F S V S L S G T T A A E T T
attgtgctgacactggctggtgacactgacgaccaaaggtgtcgatttcaatggtacgtca
I V L T L A G D T A T K G V D F N G T S
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V I V V I N G V S Q T V P V N E D G T F
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Q V T V P T N T N S F S V Q V S T I D D
aatactacgaaggaaatgagaccttcaccctaagtgggtgcgggcacaatagcatagtc
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L S N A S T T A T T V T L T L A G G S A
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T A G T D F T S S E V T I T Y Q D G T T
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Q T V A V N G D G S F E V A I P A G D T
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G D A L I F T V K L S N V S S T S T S F
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D F L L Q D G T A T S D D Y G A A S F S
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L T I G G K E A I G T I V D N D N A P V
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S G G S S T N K V M E I E F N N G D K T
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L Y T D I H A Q A G R F Y E L D F D I A
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V V D N M G Y R G D F I K L S E I S T S
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L N D T D T S E T L S L K L K G M P E G

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 S T L A D S A F M K N D V I T D H D G V
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 L L Q S V Q S Q I Q P I T **D T V N L G S**
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G N D T V Y G G G G N L A A Y G G A G N
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D T L I G G D G N D A L R G G A D N D Y
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L S G G R G N D V L R G D S G N D V L I
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G G L G H D I L T G G S G E D L F K W V
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 aacctagcatctataattcagattaaagaagactag
 N L A S I I Q I K E D -

Fig. S1A. Amino acid sequence of FrhA *V. cholerae* O395 *frhA* gene (VC395_1738) and predicted amino acid sequence. The LapG cleavage site PAAG and the “RTX repeat” (COG2931) are in bold, as well as aa 884-1460, which were used to generate polyclonal α FrhA. The red highlighted nucleotides corresponding to aa 1155-1350 were deleted in Δ PBD, the grey highlighted nucleotides corresponding to aa 1351-1460 were deleted in Δ RIII_4, the yellow highlighted nucleotides corresponding to aa 1461-1659 were deleted in Δ IGL, the green highlighted nucleotides corresponding to aa 1677-1853 were deleted in Δ SBD, and the blue highlighted nucleotides corresponding to aa 1854-2016 were deleted in Δ UKD. The Δ 5D mutant has a deletion corresponding to aa 1155-2016.

CLUSTAL O(1.2.4) multiple sequence alignment

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MPPBD      eefevseiaaswvsythgesvttdgts--dlggvdndsakdqirwgnpaeskqsgygfi      58
VCPBD      DSFTVSGVVANWTSWSNGTNTVTFDGTNAPNGGGLDNDSGKDQIRWGQPASSYSSGYGFI    60
           :.* ** :.*.*.*:::* .*****. : **:****.*****:***.*****

MPPBD      dndsnlegrfdlnqdisvgtfthynypvysggaitsaemsvefsvldhlgvstpvtltvn    118
VCPBD      DNDSALNGEFALNQDIILGTFTHYNYPVYSSGGAITSASMDVAFSVTDAHGVLTPTLKLN    120
           **** *:*.* ***** :*****.*.* *** * ** *****.*

MPPBD      fdhnetpntndvnasrdivtvqnthvtferdgdiytvqivgfvfrevgnpdgevvtsiytne  178
VCPBD      FDHNETPNTNNPEASKDIIKVGNTNVTFENAGALYTLQVIGFRIPGT--NQIVTEIRTGE    178
           *****: :*:*:*. * **:****. * :*:*:*:*** *. .::*:.* *.*

MPPBD      naatsyelvvrvvvegdy 196
VCPBD      NATNSYELVVRVGPGEgy 196
           **:.****** *:*

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Fig. S1B. Amino acid alignment of FrhA-PBD and *Mpl*BP-PBD. These are the exact amino acids replaced in FrhA^{MpPBD}. Conserved residues are noted by asterisk, conserved residues in the ligand-binding site highlighted in red.

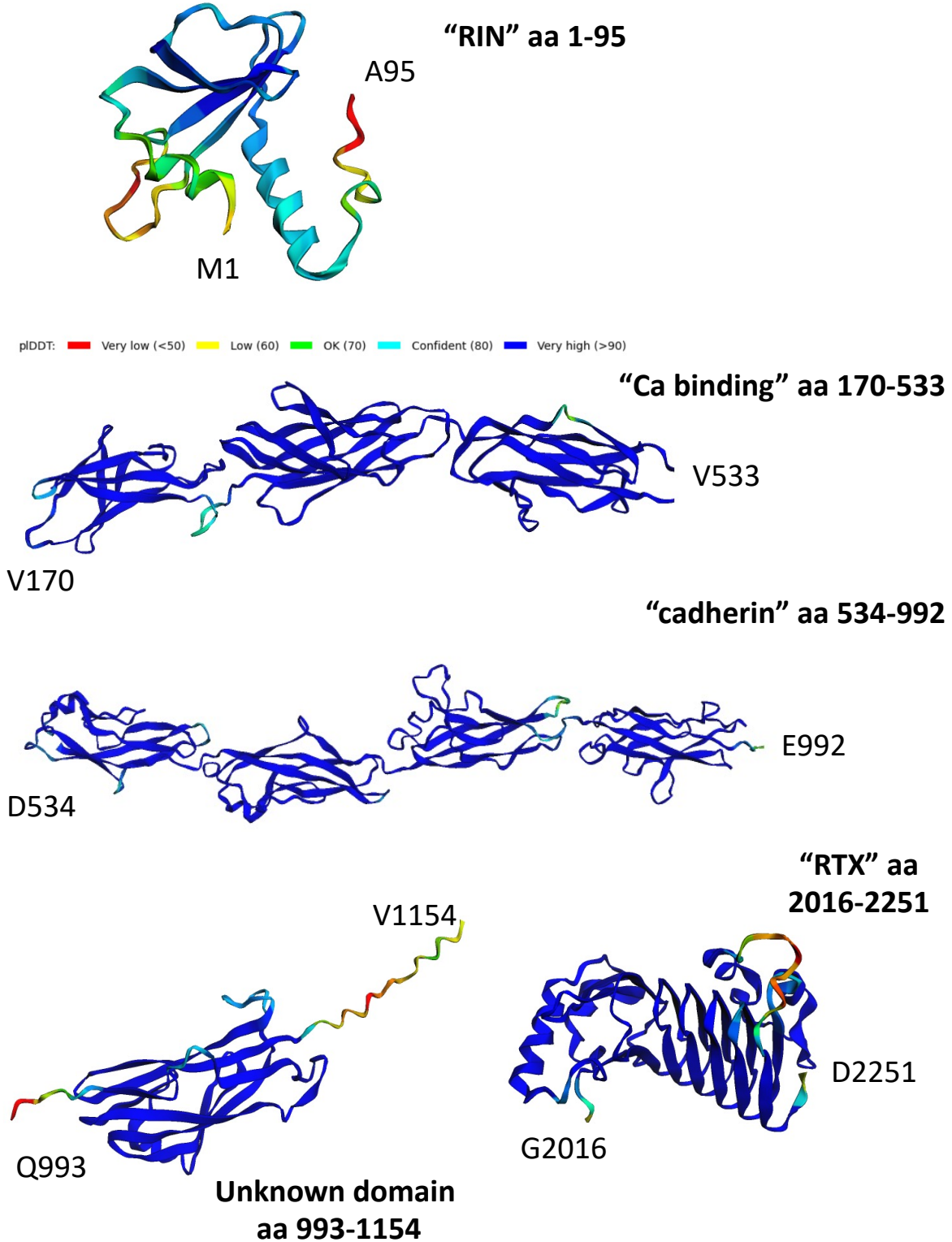
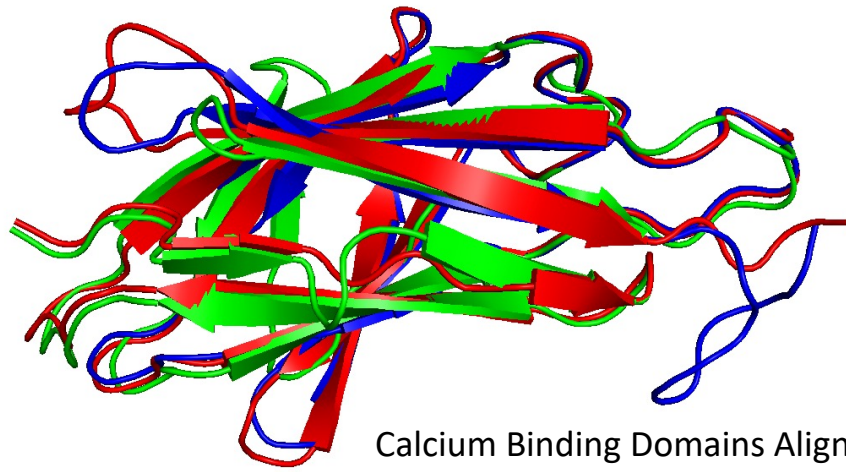
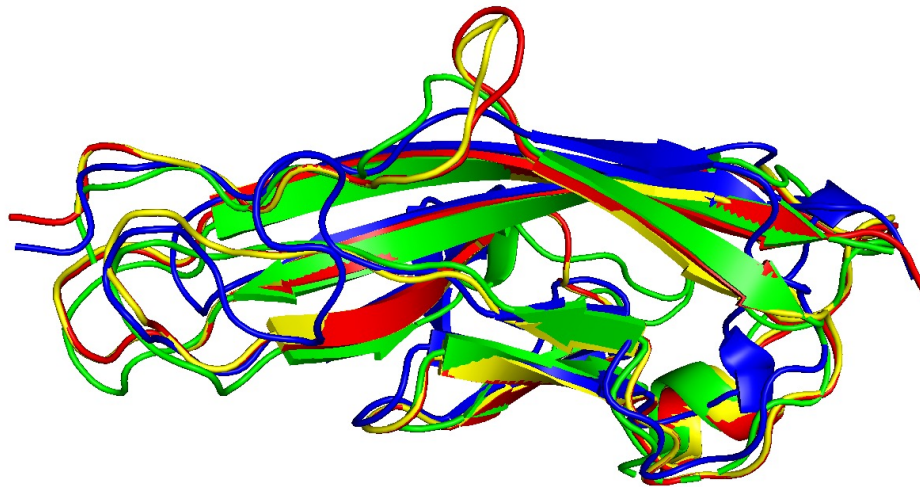


Fig. S2.A. AlphaFold predictions of various domains of FrhA



Calcium Binding Domains Alignment
R->G->B = N -> C Terminal appearance



Cadherin Domains Alignment
Y->R->G->B = N -> C Terminal appearance

Fig. S2.B. Structural alignment of predicted calcium binding domains and cadherin domains.

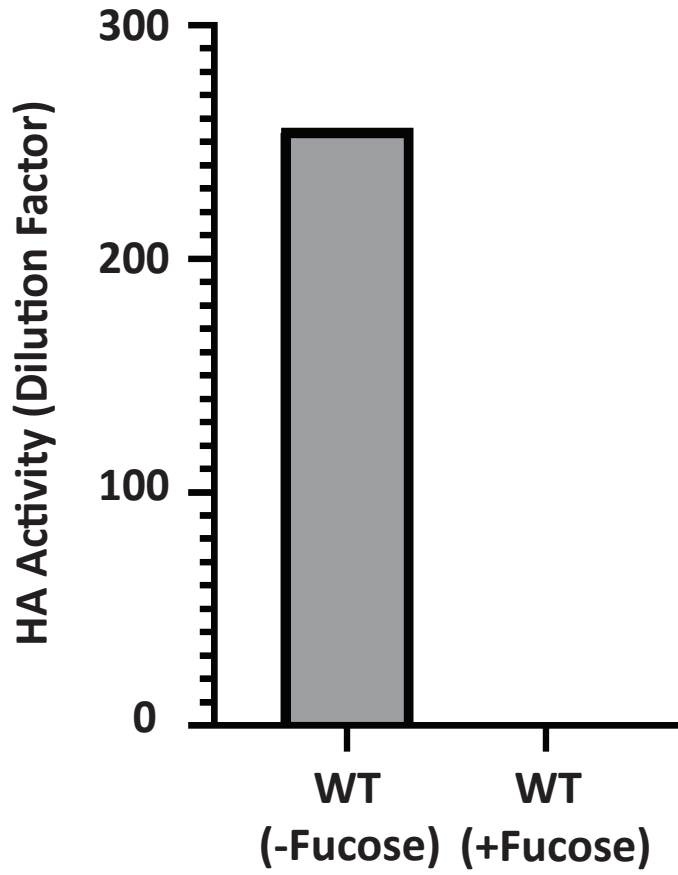


Fig. S3: Fucose inhibits *V. cholerae* HA activity. Hemagglutination of human O erythrocytes by *V. cholerae* strain KKV598 (WT) either without or with the addition of 100 mM fucose.

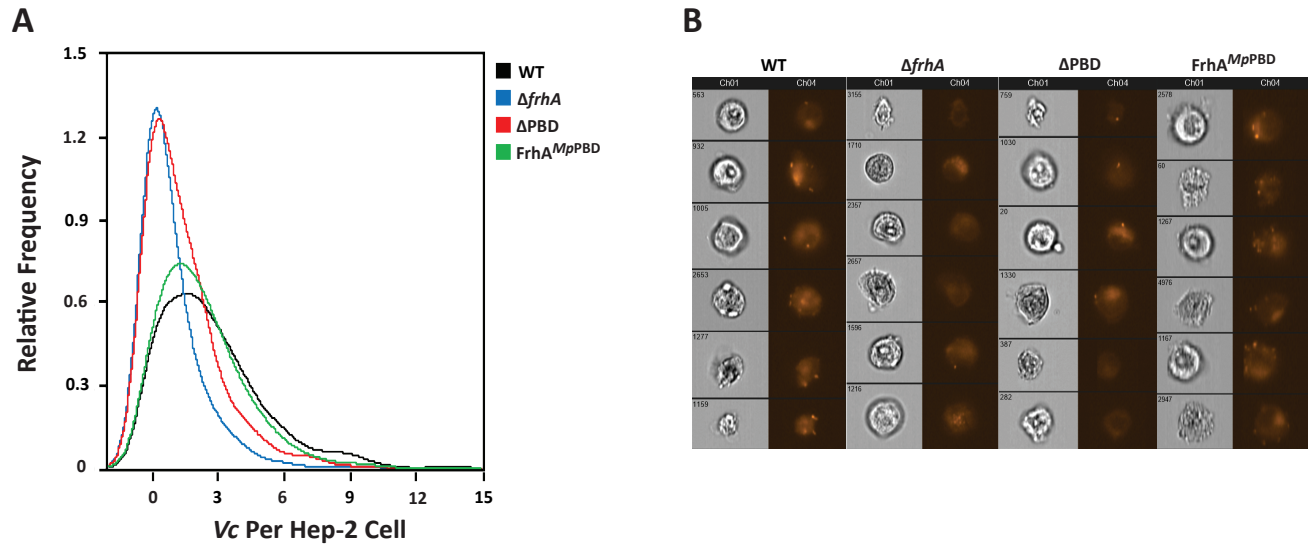


Fig. S4: *V. cholerae* binding to Hep-2 cells. **A.** Histogram of *V. cholerae* cells/Hep-2 cell as measured by Image Flow Cytometry; corresponds to Fig. 2A. **B.** Representative Hep-2 cells; corresponds to Fig. 2A and Fig. S4A.

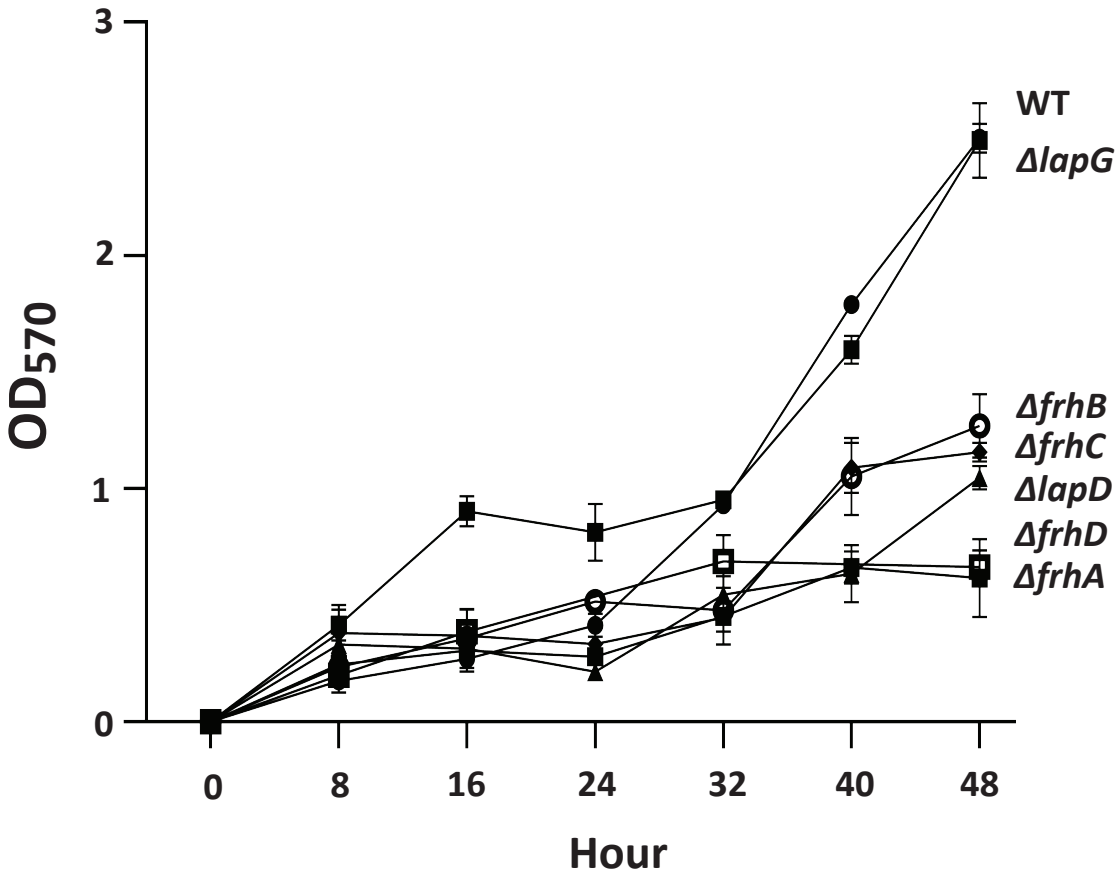


Fig. S5: Biofilm formation by *V. cholerae* *frhBCD* strains. *V. cholerae* strains KKV598 (WT), KKV3540 KKV3002 ($\Delta frhA$), KKV3032 ($\Delta frhB$), KKV2075 ($\Delta frhC$), KKV3033 ($\Delta frhD$), KKV2956 ($\Delta lapD$) and KKV2957 ($\Delta lapG$) were monitored for biofilm formation over 48 h, as described in Methods.

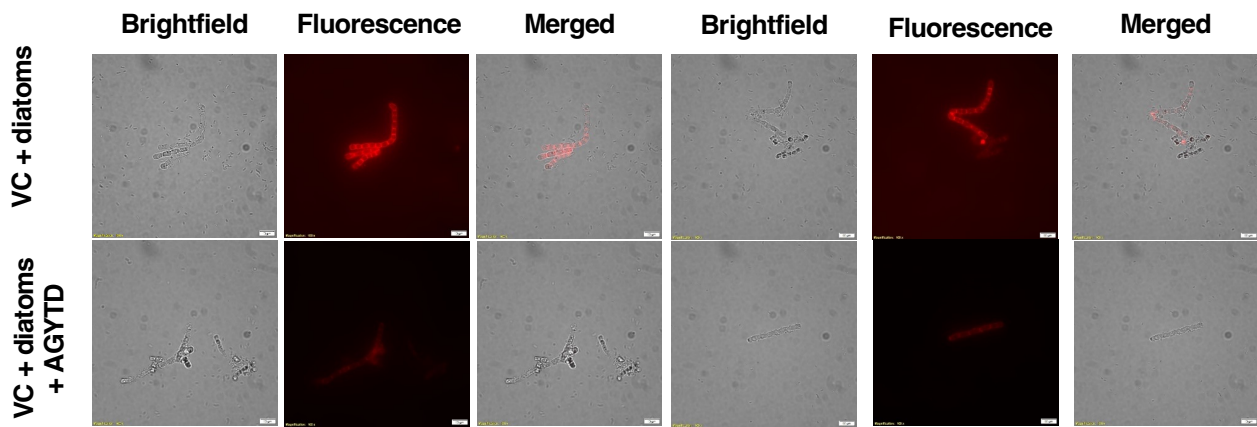


Fig. S6A: *V. cholerae* binds to diatoms via PBD: RFP-expressing *V. cholerae* were incubated with the diatom *E. spinifer* for 24 h, either without or with 500 mM AGYTD, and imaged by brightfield and fluorescence microscopy.

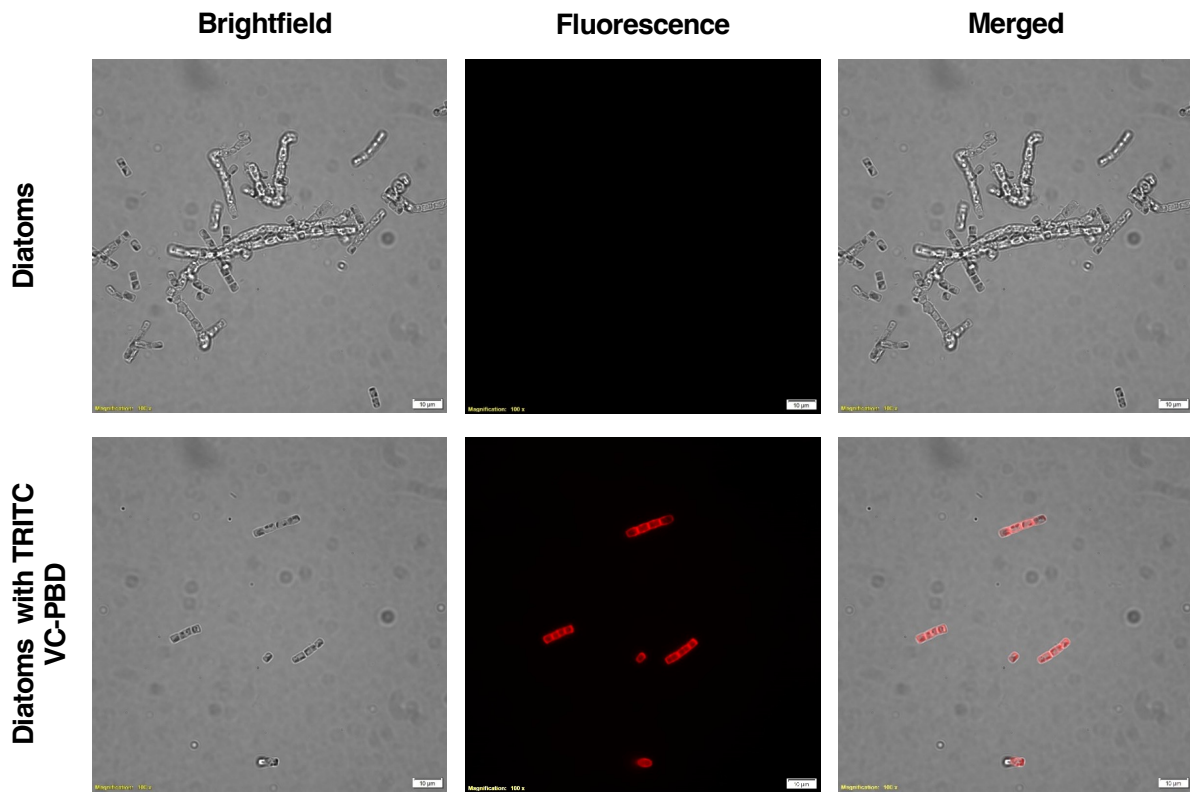


Fig S6B. *V. cholerae* PBD binds to diatoms: *E. spinifer* were incubated with TRITC-FrhA-PBD for 24 h and imaged by brightfield and fluorescence microscopy.

Table S1: *V. cholerae* binding to Hep-2 cells

Population	Count	Mean Bacteria per cell	Std. Dev.	Percent of Mean (WT)	Significance	Percent of Mean (Mp)	Significance
WT	491	2.668	2.343	1	NA	1.1789	***
Δ <i>frhA</i>	1556	0.8573	1.38	0.3213	*****	0.3788	-
Δ PBD	499	1.359	1.681	0.5093	*****	0.6005	-
WT +AGYTD	918	1.291	1.756	0.4838	*****	0.5704	-
WT +YTAGD	1319	2.146	2.14	0.8043	**	0.9482	-
FrhA ^{MpPBD}	1881	2.263	1.917	0.8482	**	1	NA
FrhA ^{MpPBD} +AGYTD	1915	1.193	1.371	0.4471	-	0.5271	*****
FrhA ^{MpPBD} +YTAGD	1655	2.056	2.049	0.7706	-	0.9085	**

Table S2: Oligonucleotides used in this study

primer	sequence
Universal Δ Up	GCGCACTAGTGCGGCCGC
Universal Δ Dwn	CGCGTCTAGATCAGGGCCC
R6K_oriT R	AAAAGGCCAGGAACCG TTTTGTCCGGTGTGGGGTTGAAG
R6K_oriT F	cactgagcgtcagacc CCATGTCAGCCGTTAAGTGTTC
pUC118 ori R	ggtctgacgctcagtggaacg
pUC118 ori F	CGGTTCTGGCCTTTTGCTG
ΔPBD F	CGAGCTCGGTACCCGG ACTGCGAAAGGTGCAGAGGC
ΔPBD R	CTTGCATGCCTGCAGG CCGTCGCATCATCAACAGGG
ΔSBD F	CGAGCTCGGTACCCGG CCGTGGTGGCCGGTAATGT
ΔSBD R	CTTGCATGCCTGCAGG CACACCATCATGATCAGTAATCACGTCAT
ΔRIII_4 F	CGAGCTCGGTACCCGG CTTGATGACCGTGGTGAAGTGA
ΔRIII_4 R	CTTGCATGCCTGCAGG TAGCCATTGATAGTCAGGTTCCGCA
ΔIGL F	CGAGCTCGGTACCCGG GGATGTTGCGTTCTCAGTGACAGA
ΔIGL R	CTTGCATGCCTGCAGG GGTATCGGTATCATTCAGTGAAGTGGAAA
ΔUKD F	CGAGCTCGGTACCCGG TACCGGAGCAACTGGA ACTCTGA
ΔUKD R	CTTGCATGCCTGCAGG AACCATCATTTTTCTCTACTTTGAATGACGATGA
ΔlapD FC F	CGAGCTCGGTACCCGG_TCCCGCATCAGAGCAAGAGC
ΔlapD FC R	CTTGCATGCCTGCAGG_TAATCAGCAGTGACGCTATCAACACATC
ΔlapG FC F	CGAGCTCGGTACCCGG_ACTGAATCGGCGGTCGGC

ΔlapG FC R	CTTGCATGCCTGCAGG_ACGGCAAACATACCAATCACTAAGATGC
frhB FC F	CGAGCTCGGTACCCGG_ATTGAACGTGTGATTCAACAGCTTGAAAAAG
frhB FC R	CTTGCATGCCTGCAGG AAACACAAAGAACTCTATAAGCAAGCTCAAGACA
frhB SOE F	ATGTAGTTTGGTGTCTTTAAATAGACGAAC
frhB SOE R	TTAAAGACACCAAACACTACATGCCTCTGACC
frhD FC F	CGAGCTCGGTACCCGG_TTTTCCGCCACACCTGTGATGT
frhD FC R	CTTGCATGCCTGCAGG_CTTTACGTTTCGACGCAGGTTTAATTTGT
frhD SOE F	ACAATTATTGGCGCTGAGGGAATAGCCCGC
frhD SOE R	CCCTCAGCGCCAATAATTGTTAAGGATATGCTATCTGCGGCG
RIII-3VC>MP F:	GAATTGCCATCCACATCAGGC
RIII-3VC>MP R:	AACAACAATGTCCACTTCACCAC
MpIBP RIII-3 F:	<u>AGTGGACATTGTTGTT</u> gaagagtgtgaagtgtctgagattgc
MpIBP RIII-3 R:	ATGTGGATGGCAATTC ataaccatccccctcaaccac
RIII-3VC>AbR R:	<u>GTCGACGGATCCCCGGAAT</u> AACAACAATGTCCACTTCACCAC
RIII-3VC>AbR F:	<u>GAAGCAGCTCCAGCCTACA</u> GAATTGCCATCCACATCAGGC
229 dPBD atbtc F	GAAGCAGCTCCAGCCTACAAACAACAATGTCCACTTCA
229 dPBD atbtc R	GTCGACGGATCCCCGGAATGAATTGCCATCCACATCAG
pUC118 MCS F	CCTGCAGGCATGCAAGCTT
pUC118 MCS R	CCGGGTACCGAGCTCGAATTC
ABD123	ATTCCGGGGATCCGTCGAC

ABD124	TGTAGGCTGGAGCTGCTTC
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Table S3: Plasmids used in this study

Plasmid	Description	reference
pFY_116	pGP704:: <i>Tn7-gfp</i>	(8)
pFY_117	pGP704:: <i>Tn7-rpf</i>	(8)
pFY_118	pUX-BF13	(8)
pKEK 229	Sucrose-counterselectable suicide vector: AmpR	(5)
pKEK 2054	Δ 5D in pKEK229	This study
pKEK2094	Cloning vector; pUC origin; AmpR	(6)
pKEK2100	pKEK2094 with R6K ori and oriT; AmpR	This Study
pKEK2106	Δ SBD in pKEK2100	This Study
pKEK 2107	Δ IGL in pKEK2100	This study
pKEK 2108	Δ PBD in pKEK2100	This study
pKEK 2109	Δ RIII_4 in pKEK2100	This study
pKEK2110	Δ <i>frhB</i> in pKEK2100	This study

pKEK2114	Δ <i>frhD</i> in pKEK2100	This study
pKEK2119	Δ SBD in pKEK229	This Study
pKEK2120	Δ IGL in pKEK229	This Study
pKEK2121	Δ PBD in pKEK229	This Study
pKEK2122	Δ <i>frhB</i> in pKEK229	This study
pKEK2123	Δ <i>frhD</i> in pKEK229	This study
pKEK2124	Δ RIII_4 in pKEK229	This Study
pKEK2157	Δ UKD in pKEK2200	This Study
pKEK 2200	Sucrose-counters selectable suicide vector; Cm ^R	(6)
pKEK 2265	Δ <i>lapD</i> in pKEK2200	This study
pKEK 2266	Δ <i>lapG</i> in pKEK2200	This study
pKEK 2270	<i>hapR</i> expression vector; Tc ^R	(9)

pKEK 2285	pBAD- <i>tfoXqstR</i> expression vector; AmpR	(9)
pKEK 2335	frhA ^{MPPBD} in pKEK 229	This Study
pKEK2447	frhA::KanR in pKEK229	This Study

Table S4: *V. cholerae* strains used in this study

Strain	Description	Reference/Source
O395	<i>V. cholerae</i> wildtype	(10)
FY_Vc_12114	$\Delta lacZ$; $\Delta frhA$	(11)
FY_VC_12120	$\Delta lacZ$; $\Delta lapD$	(6)
Fy_VC_11863	$\Delta lacZ$; $\Delta lapG$	(6)
SAD034	$\Delta VC1807::KanR$	(4)
KKV598	$\Delta lacZ$	(5)
KKV2075	$\Delta lacZ$; $\Delta frhC$	(12)
KKV2759	$\Delta lacZ$; $frhA^{\Delta 5D}$	This study
KKV2942	$\Delta lacZ$; $frhA^{\Delta PBBD}$	this study
KKV2956	$\Delta lacZ$, $\Delta lapD$	this study
KKV2957	$\Delta lacZ$, $\Delta lapG$	this study
KKV2985	$\Delta lacZ$; $frhA^{\Delta SBD}$	this study
KKV2986	$\Delta lacZ$; $frhA^{\Delta RIII_4}$	this study
KKV2987	$\Delta lacZ$; $frhA^{\Delta IGL}$	this study
KKV2988	$\Delta lacZ$; Tn7-rfp	This study
KKV3009	$\Delta lacZ$; $frhA^{\Delta UKD}$	this study
KKV3032	$\Delta lacZ$, $\Delta frhB$	this study
KKV3033	$\Delta lacZ$, $\Delta frhD$	this study
KKV3451	$\Delta lacZ$, $\Delta frhA::KanR$	This study
KKV3540	$\Delta lacZ$; $frhA^{MpPBBD}$	this study

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