### Figure S1



### Figure S1. MDS samples QC, integration, clustering, and cell-type assignment, related to Figure 1.

(A) Number of genes per cell (top) and number of UMIs per cell (bottom) in CD34+ sorted hematopoietic progenitors from samples MDS01-03 after QC filters, shown by each patient sample. (B) Number of genes per cell (top) and number of UMIs per cell (bottom) in CD34+ sorted hematopoietic progenitors from samples MDS04-06 after QC filters, shown by each patient sample. (C) UMAP of CD34+ sorted progenitor cells for each individual sample of MDS01-03 after integration using the Seurat package. (D) UMAP of CD34+ sorted progenitor cells for each individual sample of MDS04-06 after integration using the Seurat package. (E) Expression of lineage-specific genes from Velten et al.[S1] scored and projected onto the UMAP representation of cells from MDS01-03. (F) CD34 expression per progenitor cell-type of CD34- monocytes among CD34+ sorted hematopoietic progenitors. (G) Heatmap of top 10 differentially expressed genes for each progenitor subset for MDS01-03. (H) UMAPs comparing the graph-based clustering output when using the k-nearest neighbors (KNN) algorithm to perform clustering of cells with RNA data alone vs. when using the weighted nearest neighbors (WNN) algorithm that allows for the integration of both RNA and ADT data for clustering cells. The cell-type assignments determined from the KNN-RNA based clustering are projected onto the WNN-RNA+ADT clustering for comparison between the two methods. (I) UMAP of CD34+ sorted cells (n = 8,879 cells) from samples MDS04-06 with SF3B1 K700E mutations (n = 3), overlaid with cluster cell-type assignments. HSPC, hematopoietic stem progenitor cells; IMP, immature myeloid progenitors; MkP, megakaryocytic progenitors; MEP, megakaryocyticerythroid progenitors; EP, erythroid progenitors; NP, neutrophil progenitors; E/B/M, eosinophil/basophil/mast progenitor cells; T/B cell progenitors; Mono, monocyte; DC, dendritic cells; Pre-B, precursors B cells; Mono DC, monocyte/dendritic cell progenitors. (J) Heatmap of top 10 differentially expressed genes for each progenitor subset for MDS04-06. (K) Expression of lineage-specific genes from Velten et al. [S1] scored and projected onto the UMAP representation of cells from MDS04-06.

Figure S2



CD49f

r = 0.46 P = 0.36

FP

MEF

184

0.5 0.6 0.7 mean ADT expression

...

0.

0.

0.

EP

ME

MkF

1.25 1.50 1.75 mean ADT expression CD45 CD36 CD71 <sup>1.5</sup> r = 0.94 P = 0.0047 r = 0.98 P = 0.00075 1.5 r = 0.94 P = 0.0053 mean RNA expression EP 0 HSPO 1.0 0.2 ٩P NP 0.5 MAD 0.0 IMP HSPC •EP 1.6 0.4 0.8 1.2 mean ADT expression 3.0 3.5 4.0 4.5 mean ADT expression mean ADT expression

UMAP2

UMAP1

### Figure S2. MDS samples GoT, CITE-seq, and pseudotime analyses, related to Figure 2.

(A) SF3B1 K700E (7) and K666N (1) mutant cell fractions determined by GoT in single cells versus SF3B1 K700E and K666N mutation variant allele frequencies (VAF) determined in bulk sequencing of matched unsorted bone marrow mononuclear cells (MDS) or matched unsorted stem cell product (CH). (B) Fraction of cells in MDS samples by number of SF3B1 UMIs in standard 10x Genomics data without genotyping information, SF3B1 UMIs with K700E locus coverage in standard 10x data, SF3B1 UMIs with K700E locus coverage in GoT amplicon library, and SF3B1 UMIs with K700E locus coverage in ONT library. (C) UMAP of CD34+ sorted cells (n = 8,879 cells) from samples MDS04-06 with SF3B1 K700E mutations (left) and density plot of SF3B1<sup>mut</sup> vs. SF3B1<sup>wt</sup> cells (right). WT, cells with genotype data without SF3B1 mutation; MUT, cells with genotype data with SF3B1 mutation. (D) Normalized ratio of SF3B1<sup>mut</sup> cells in progenitor subsets with at least 300 genotyped cells. Bars show aggregate analysis of samples MDS01-03 with mean +/- s.e.m. of 100 downsampling iterations to 1 genotyping UMI per cell. Points represent the mean of n = 100 downsampling iteration per sample. (E) Comparison of the single-cell expression of markers captured in both CITE-seq (x-axes) and RNA-seq (y-axes) libraries. Correlation coefficient r calculated using Spearman's correlation. Cells are colored by each progenitor subset. (F) Comparison of the mean expression per progenitor subset of markers captured in both CITE-seq (x-axes) and RNA-seq (y-axes) libraries. Correlation coefficient r calculated using Spearman's Correlation. P-values derived from Student's tdistribution. (G) UMAP of progenitor cells from MDS01-03 samples overlaid with pseudotemporal ordering.

### Figure S3







### Figure S3. Long-read splicing and motif analysis, related to Figure 3.

(A) Long-read sequencing processing and splicing analysis pipeline. (B) Fraction of cryptic junctions detected in short-reads, long-reads or both when defining distinct cluster coverage thresholds. (C) Number of junctions per cell detected in 10x short-read compared to ONT full-length long-read (junction cluster coverage >= 1 read). (D) Number of detected cryptic 3' splice site junctions (in genes with ~1 read/cell in both short and long-read datasets and in clusters with coverage >= 600 reads) along increasing distances to the most 3' end junction of each transcript (see *inset*) in 10x short versus ONT full-length long-reads. Individual number of events as well as overlaps are itemized in the pie charts (bottom). (E) Comparison of the usage of various alternative 3' splice sites found in our MDS  $SF3B1^{mut}$  cells vs. a CD34+ sample with no SF3B1 mutation. (F) Bar plot of the number of cryptic 3' splice sites identified per gene in MDS. *Inset*: Gene example, *PFDN5*, with 2 unique cryptic 3' splice sites, showing the transcripts that have usage of either site. (G) Nucleotide enrichment (measured as log-odds ratio) across the 3' splice site region comparing cryptic vs. canonical sites in MDS02-03 samples.



# Figure S4. Comparison of splicing junctions detected with GoT-Splice vs. other tools, MDS samples, and cancer types, related to Figure 4.

(A) Left: Overlapping of all splicing junctions detected in GoT-Splice, FLAMES, and IsoQuant. Right: Comparison of splicing junctions supporting cryptic 3' ss between the two full-length isoform detection tools and GoT-Splice. (B) Estimated dPSI values from long-read tools FLAMES and IsoQuant compared to reported dPSI values measured in bulk data, like Figure 2g. (C) Bars showing the percentage of genes differentially spliced in SF3B1<sup>mut</sup> and SF3B1<sup>wt</sup> cells with BH-FDR adjusted P-value < 0.2 in each sample of the MDS cohort (MDS02(A/B)-03). (D) Differential splicing analysis between SF3B1<sup>mut</sup> and SF3B1<sup>wt</sup> cells across the aggregate of the MDS validation cohort (MDS04-06). Junctions with an absolute delta percent spliced-in (dPSI) > 2 and BH-FDR adjusted P-value < 0.2 were defined as differentially spliced. Bars (top) showing the percentage of genes differentially spliced in SF3B1<sup>mut</sup> and SF3B1<sup>wt</sup> cells of MDS validation cohort. (E) Bars showing the percentage of genes differentially spliced in *SF3B1<sup>mut</sup>* and *SF3B1<sup>wt</sup>* cells with BH-FDR adjusted *P*-value < 0.2 in each sample of the MDS validation cohort (MDS04-06). (F) Venn Diagram for the overlap of differentially spliced genes used more highly in SF3B1<sup>mut</sup> cells (P-values < 0.05, dPSI >0) from the bulk comparison of SF3B1<sup>mut</sup> vs. SF3B1<sup>wt</sup> cells in the MDS and MDS validation cohorts [i]. Increasing the read coverage threshold for the differentially spliced genes showed a more significant overlap between cohorts [ii]. P-values for the overlap from Fisher's exact test. (G) Differential isoform proportion in SF3B1<sup>mut</sup> vs SF3B1<sup>wt</sup> cells. Isoforms containing alternative/cryptic 3' ss overlapping with those detected in GoT-Splice are highlighted in red. *Top*: Quantification of significant (FDR adjusted *P*-value < 0.05) full-length isoforms detected containing alternative/cryptic 3'ss, split by direction of the logFC in proportion between MUT and WT cells. (H) Example of cryptic 3' ss in the gene ERGIC3 missed by IsoQuant and FLAMES but detected and quantified by GoT-Splice. (I) Venn Diagram showing the overlap of genes with cryptic 3' ss splicing events (dPSI > 0 and FDR adjusted P-value < 0.2) found in the MDS discovery cohort vs. those found in the bulk RNA sequencing of a set of published SF3B1<sup>mut</sup> CLL samples. (J) Box plot of the average expression levels found across all cells in the MDS discovery cohort for the genes cryptically mis-spliced in the MDS dataset only, in both the MDS and CLL datasets, and the CLL dataset only. P-values were computed using Wilcox rank sum test.

## Figure S5









# Figure S5. Application of GoT-Splice to other mutations and identification of cell-type specific splicing changes, related to Figure 5.

(A) Pie chart summarizing the distribution of different alternative splicing events detected after junction annotation in the DNMT3A<sup>mut</sup> CH sample. (B) Comparison of percent spliced-in (dPSI) values of alternative splicing events identified in DNMT3A<sup>mut</sup> vs. DNMT3A<sup>wt</sup> CH cells. Significant alternative splicing events are highlighted. (C) Sashimi plot of SRSF3 exon skipping event showing the expected increase in the PSI value in DNMT3A<sup>wt</sup> cells. (D) UMAP of progenitor cells from U2AF1<sup>S34F</sup> AML sample overlaid with genotyping data. WT, cells with genotype data without U2AF1 mutation; MUT, cells with genotype data with U2AF1 mutation. (E) Pie chart summarizing the distribution of different alternative splicing events detected after junction annotation in the U2AF1<sup>mut</sup> AML sample. (F) Comparison of percent spliced-in (dPSI) values of alternative splicing events identified in U2AF1<sup>mut</sup> vs. U2AF1<sup>wt</sup> cells. Significant alternative splicing events are highlighted. (G) Sashimi plot of KIN and DAP3 exon skipping events showing the expected increase in the exon skipping in U2AF1<sup>mut</sup> cells. (H) Heatmap of dPSI values between SF3B1<sup>mut</sup> and SF3B1<sup>wt</sup> cells for cryptic 3' splicing events identified in the main progenitor subsets across MDS validation samples. Rows (z-score normalized) correspond to cryptic 3' junctions found to be differentially spliced in at least one cell-type, with P-value <= 0.05 and dPSI >= 2. Columns correspond to cell-type. Genes with an SF3B1<sup>mut</sup> associated cryptic 3' splice site found in the MDS cohort are highlighted in red. (I) Pie chart showing the percent overlap of cryptically 3' spliced genes unique to MEPs and EPs in the primary MDS cohort that are also cryptically 3' spliced and unique to earlier progenitor cells (HSPCs and IMPs) in the MDS validation cohort (left) as well as the percent overlap with genes cryptically 3' spliced and unique to the MEPs and EPs in the MDS validation cohort (right). *P*-value for the overlap from Fisher's exact test. (J) Plots highlighting the differences in sequencing coverage along the transcript, between short- and long-read sequencing, for PPOX and UROD, demonstrating the ability for long-read sequencing to uncover splicing events than remain undetected in the 3' biased short-read sequencing.



HNRNPA1

#### Figure S6. CH cohort QC, integration, and GoT-Splice analyses, related to Figure 6.

(A) Number of genes per cell (top) and number of UMIs per cell (bottom) in CD34+ sorted hematopoietic progenitors from samples CH01-02 after QC filters, shown by each patient sample. (B) UMAP of CD34+ sorted progenitor cells for each individual sample of CH01-02 after integration using the Seurat package. (C) Expression of lineage-specific genes from Velten et al.[S1] scored and projected onto the UMAP representation of cells from CH01-02. (D) Heatmap of top 10 differentially expressed genes for each progenitor subset for CH01-02. (E) Fraction of cells in CH01-02 by number of SF3B1 UMIs in standard 10x Genomics data without genotyping information, SF3B1 UMIs with K666N (CH01) or K700E (CH02) locus coverage in standard 10x data, SF3B1 UMIs with K666N (CH01) or K700E (CH02) locus coverage in GoT amplicon library, and SF3B1 UMIs with K666N (CH01) or K700E (CH02) locus coverage in ONT library. (F) Per sample heatmap of relative expression of genes ordered by chromosome/chromosomal position following copy number variation analysis using the InferCNV package (see STAR Methods). Cells (y-axis) are stratified by SF3B1 genotype status. (G) Pseudotime in SF3B1<sup>mut</sup> vs. SF3B1<sup>wt</sup> cells per CH sample. P-value for comparison of means from Wilcoxon rank sum test. (H) Normalized ratio of SF3B1<sup>mut</sup> cells in HSPC and EP cells for CH01 and CH02. Bars show the mean of n = 100 downsampling iterations to 1 genotyping UMI per cell. (I) Comparison of the PSI values of identified cryptic junctions in WT cells only (gray) vs. MUT cells only (red) vs. all cells in pseudobulk in CH01-02. P-values for comparison of means from Wilcoxon rank sum test. (J) Comparison of the SF3B1<sup>mut</sup> cells PSI values for alternative splicing events that were found to be significantly differentially used (*P*-value < 0.05) between *SF3B1<sup>mut</sup>* and *SF3B1<sup>wt</sup>* in the primary CH cohort and that were also present in the CH validation sample. Spearman's rank correlation coefficient and P-value derived from Student's t-distribution are shown. (K) Sashimi plot examples of alternative splicing events in SERBP1 (top, cryptic 3' ss event) and HNRNPA1 (bottom, exon skipping/inclusion event) found to be significantly differentially spliced between SF3B1<sup>mut</sup> and SF3B1<sup>wt</sup> cells in the CH validation sample.

#### REFERENCES

<sup>[</sup>S1] Velten, L., Haas, S.F., Raffel, S., Blaszkiewicz, S., Islam, S., Hennig, B.P., Hirche, C., Lutz, C., Buss, E.C., Nowak, D., et al. (2017). Human haematopoietic stem cell lineage commitment is a continuous process. Nat Cell Biol 19, 271–281. 10.1038/ncb3493.