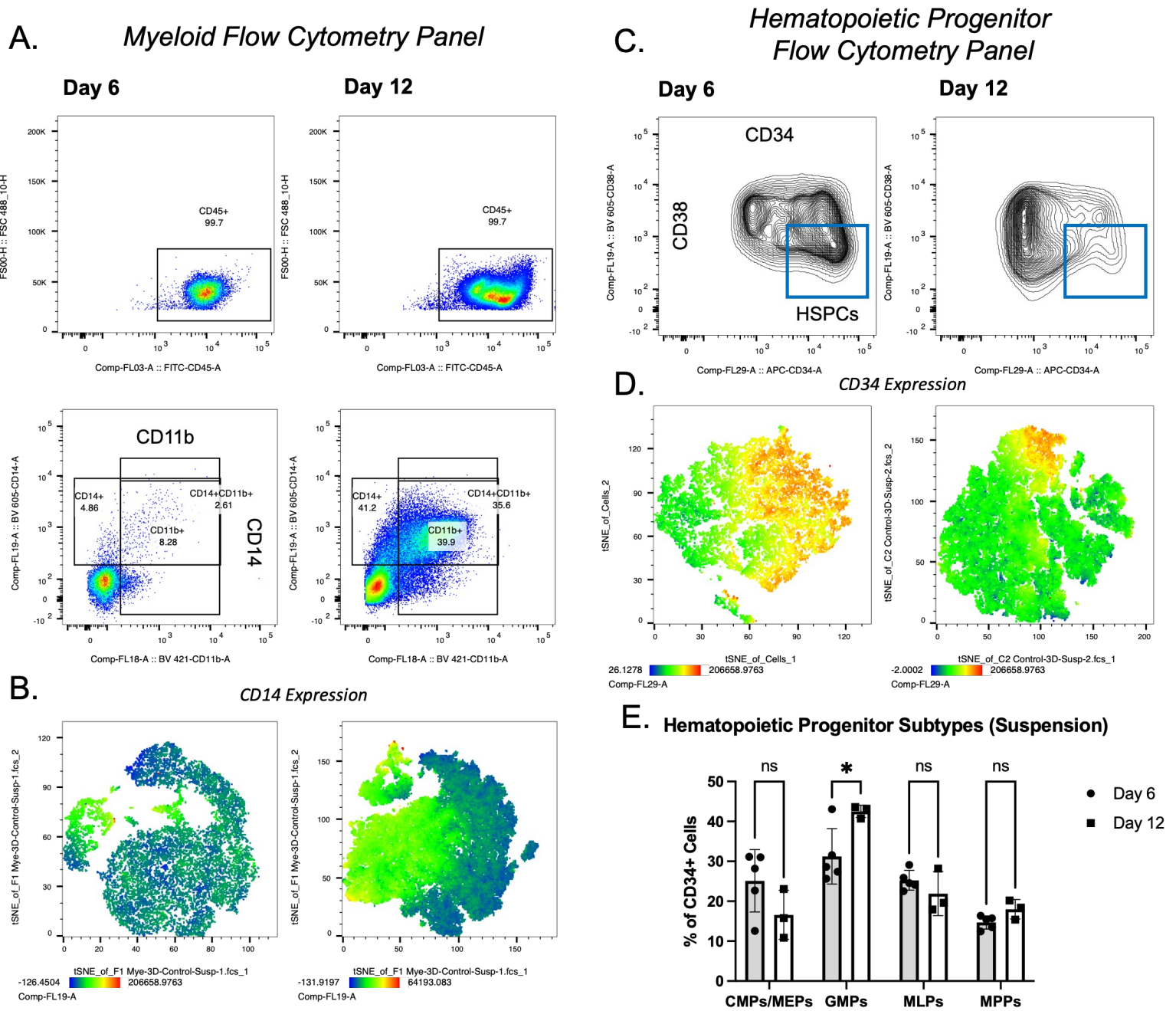


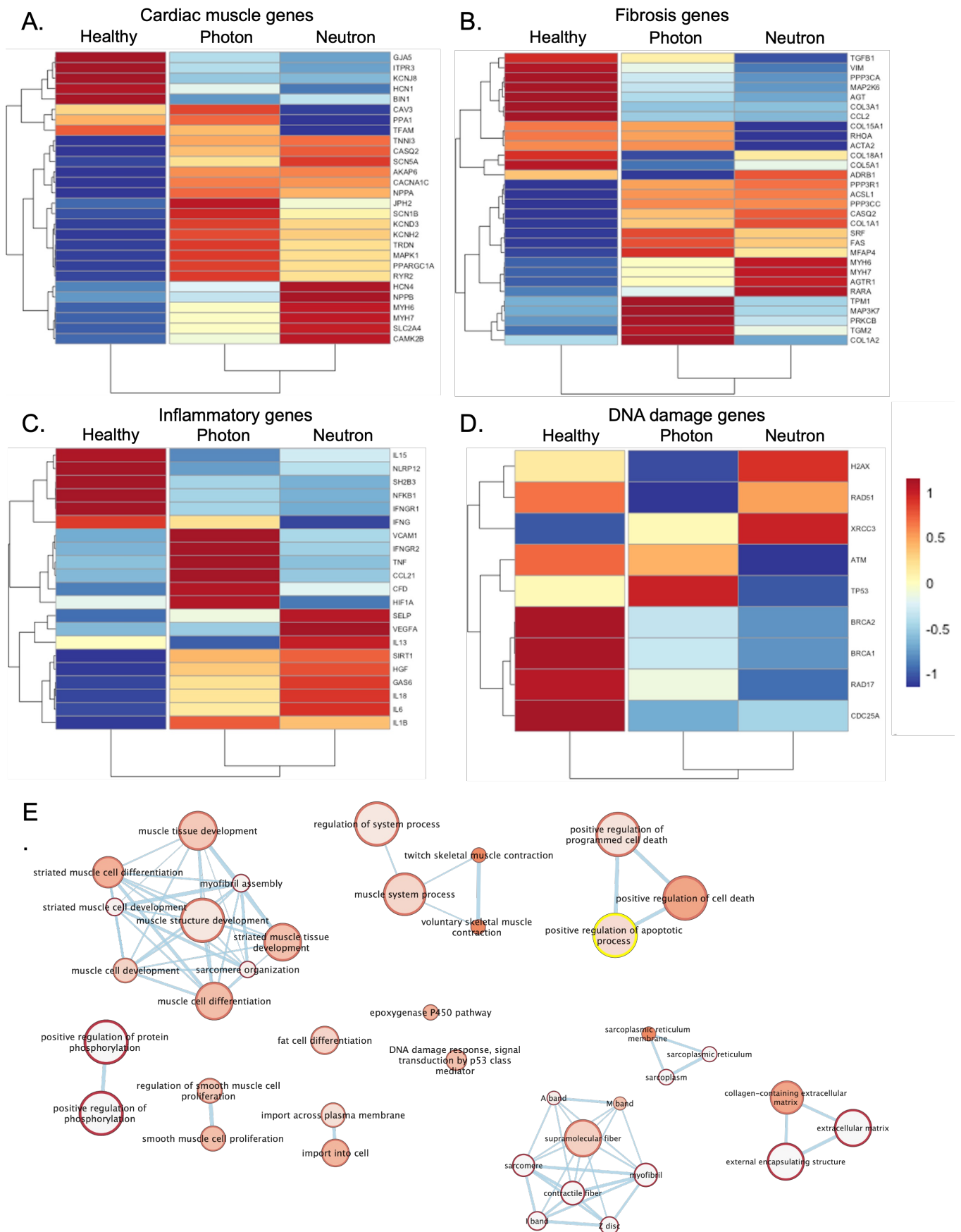
# Modeling and countering the effects of cosmic radiation using bioengineered human tissues

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## Supplemental Figures



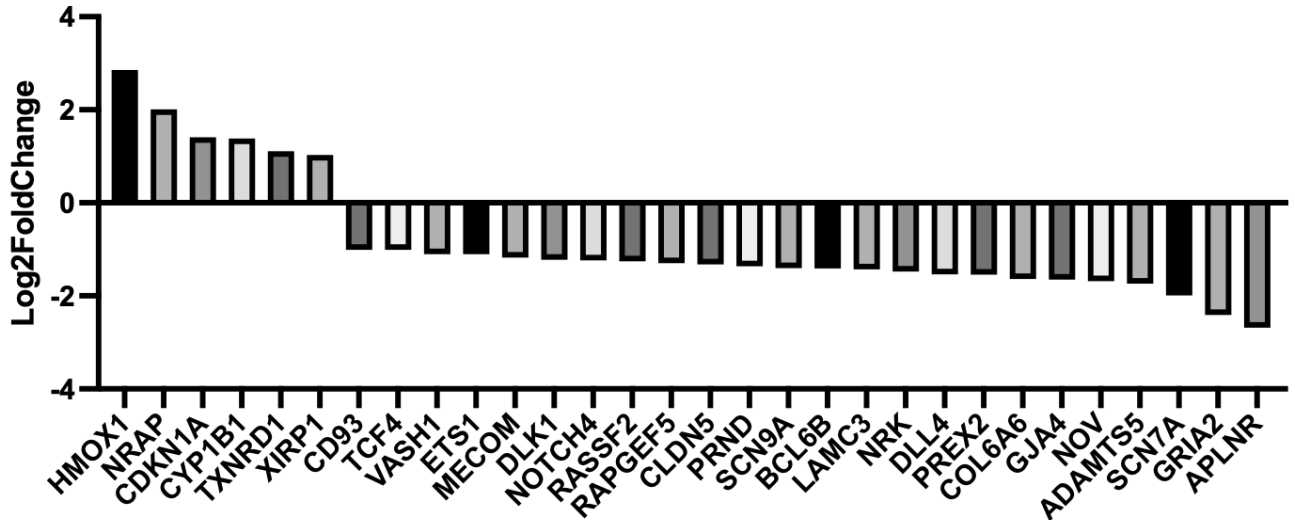
**Supplementary Figure 1. Sample flow cytometry gating for suspension cells in engineered bone marrow (eBM). Myeloid cell panel. (A-B) Shifting cell populations over time (A) and (B) dimensionality reduction for CD14). Hematopoietic cell panel. (C-D) Shifting cell populations over time (C) and (D) dimensionality reduction for CD34). (E) Subpopulations of hematopoietic progenitors described in Figure 2.**



**Supplementary Figure 2. Transcriptomic changes to engineered cardiac tissues (eCTs) in response to radiation (by key phenotypic genes). (A) Cardiac-specific genes (B) Fibrosis genes (c) Inflammatory genes (D) DNA damage/repair genes. (E) Network analysis using Cytoscape of differentially-expressed genes between neutron irradiated and control eCTs.**

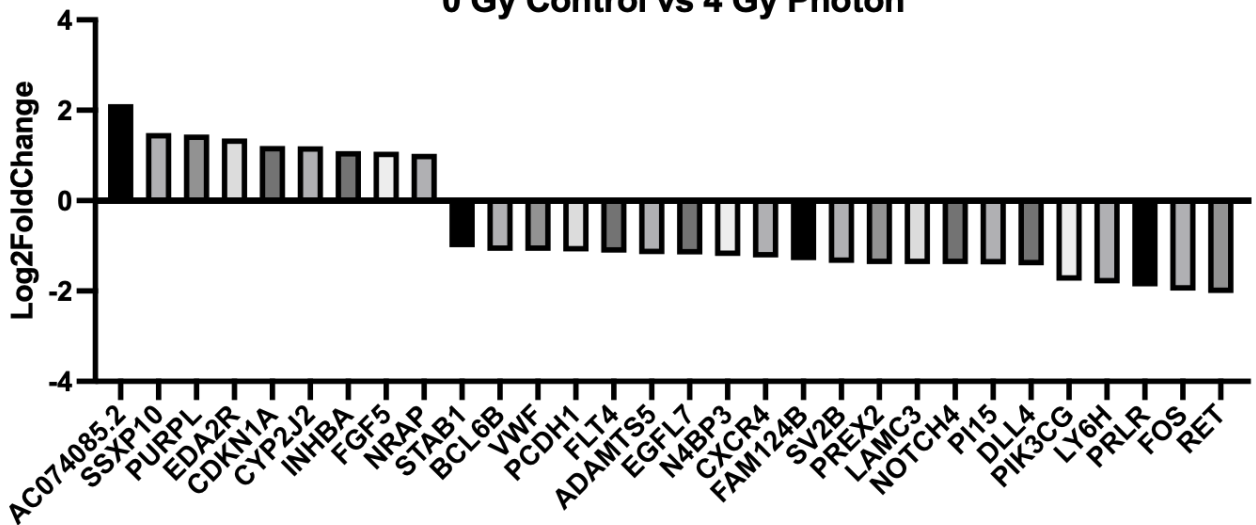
A.

Significant Differentially Expressed Genes - 0 Gy vs 1 Gy

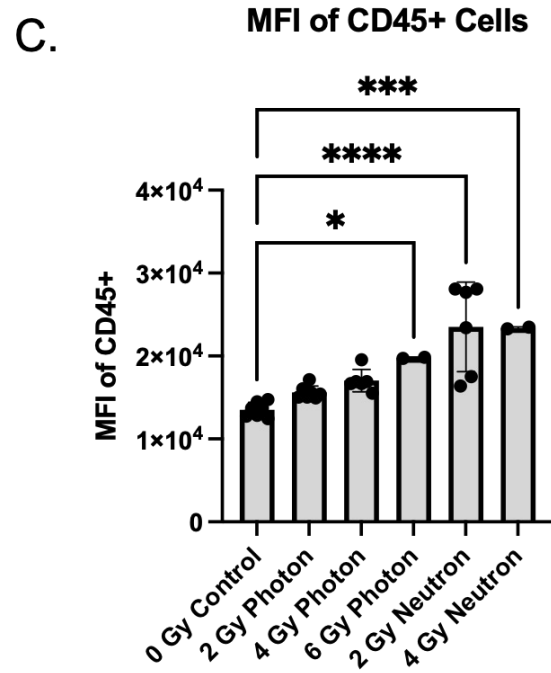
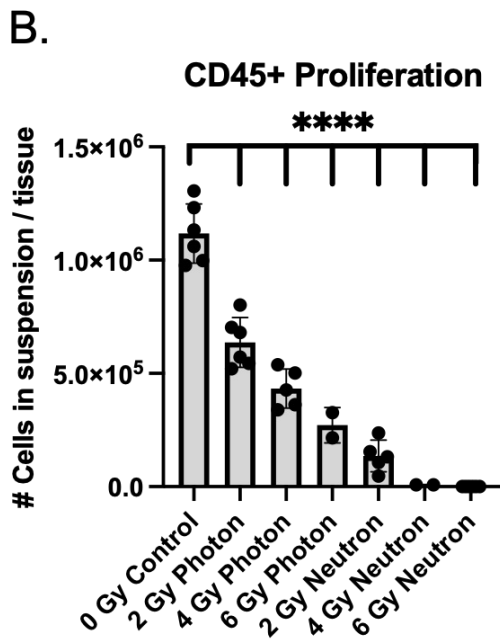
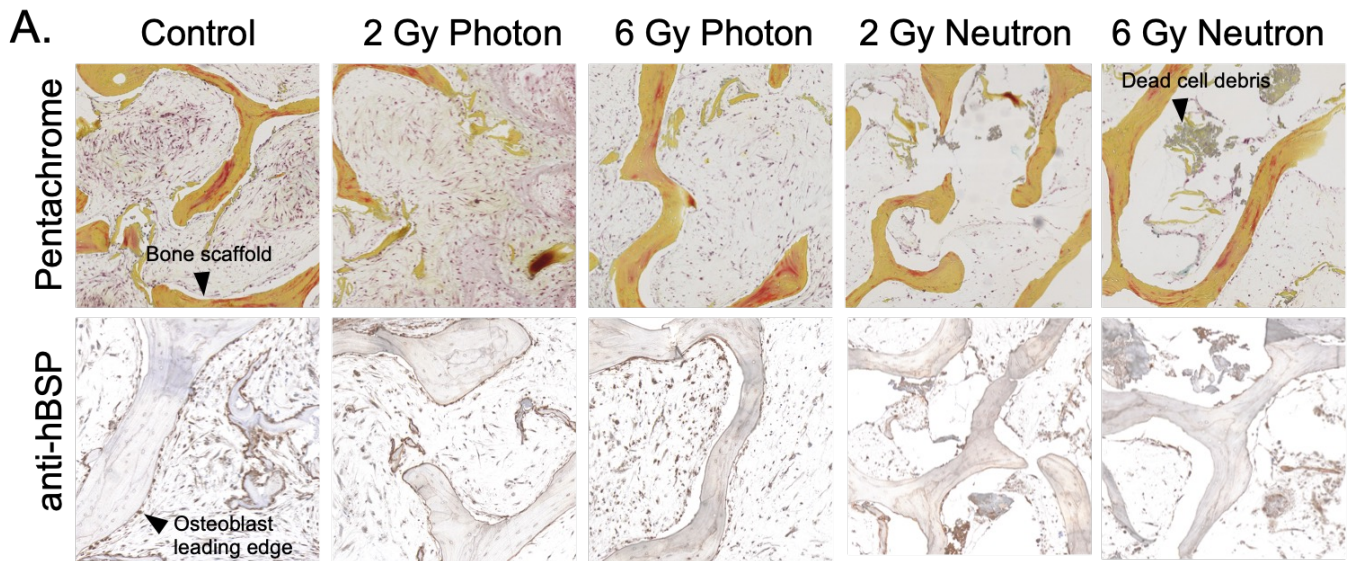


B.

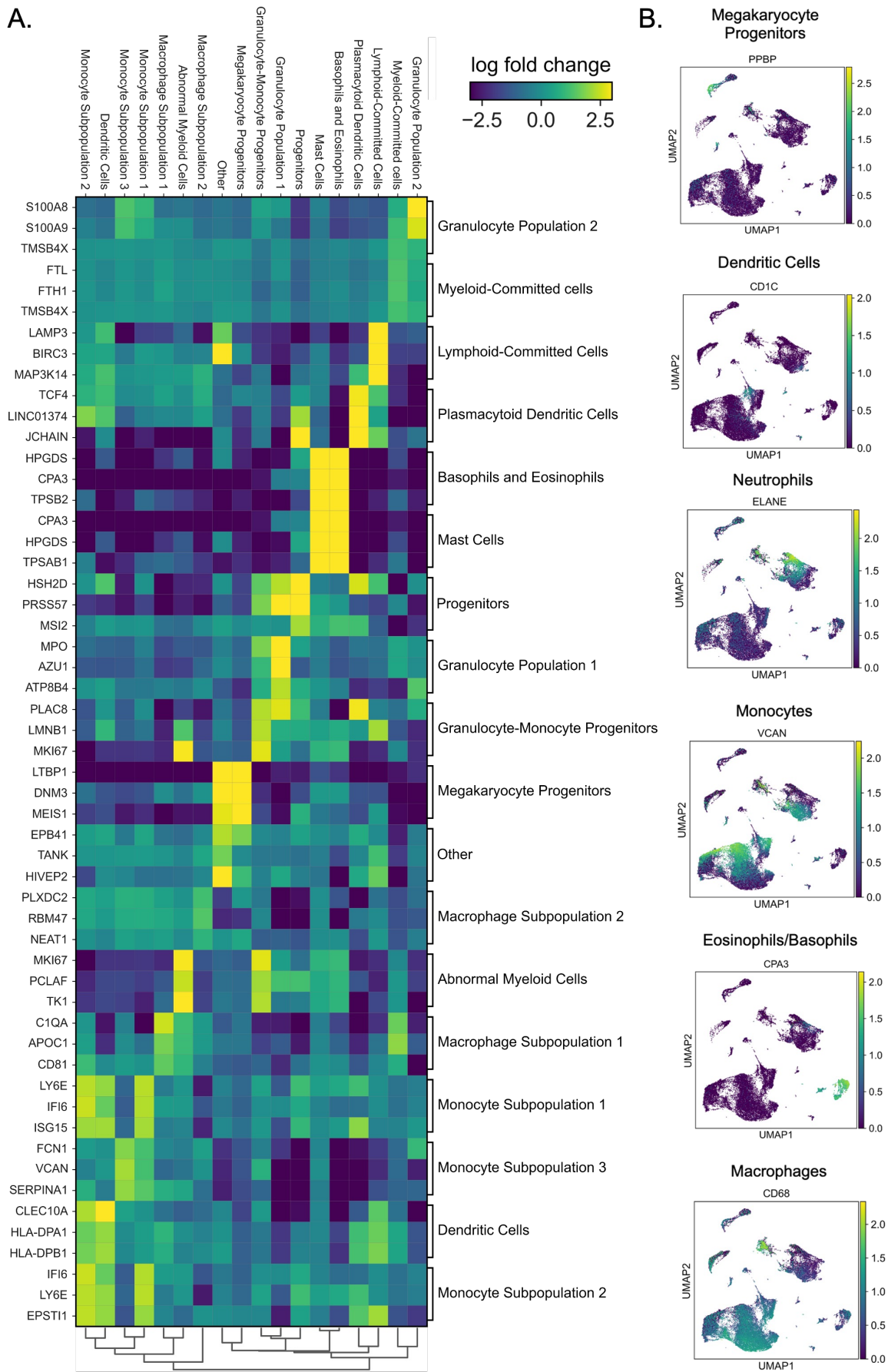
Significant Differentially Expressed Genes:  
0 Gy Control vs 4 Gy Photon



Supplementary Figure 3. Bulk gene expression changes in irradiated engineered cardiac tissues (eCTs) 3-weeks post-radiation. (A) Differentially expressed genes by fold change for (A) neutron and (B) photon radiation, compared to the control eCTs.

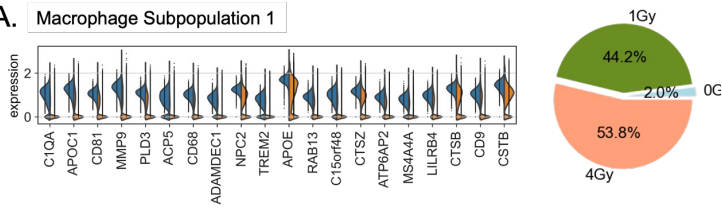


**Supplementary Figure 4. Dose-dependent effects of radiation on engineered bone marrow (eBM) tissues and hematopoietic cells 2 weeks after radiation exposure.** (A) Photon and neutron radiation decreased the intensity of pentachrome (top) and bone sialoprotein (BSP, bottom) staining in a dose-dependent manner. (B) Decreasing CD45+ cell production, and (C) increasing CD45+ cells median fluorescence intensity (MFI) with increasing radiation dose over two weeks. \*\*\*\*p<0.0001, \*\*\*p<0.001, \*\*p<0.01, \*p<0.05

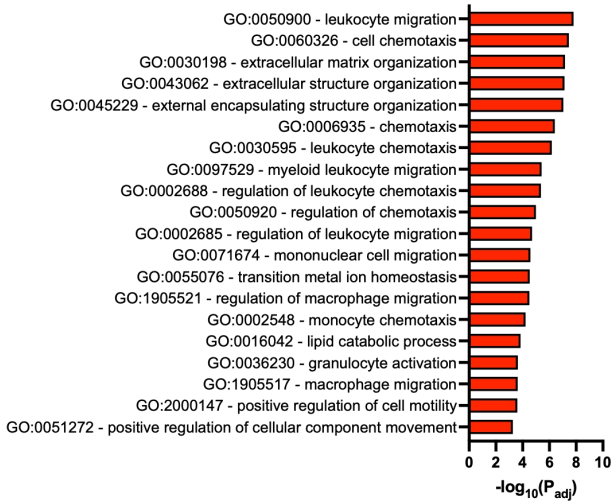


**Supplementary Figure 5. scRNAseq cell type classification of engineered bone marrow (eBM) blood cells. (A) Top genes for unique cell types. (B) Example marker genes for identifying known cell types.**

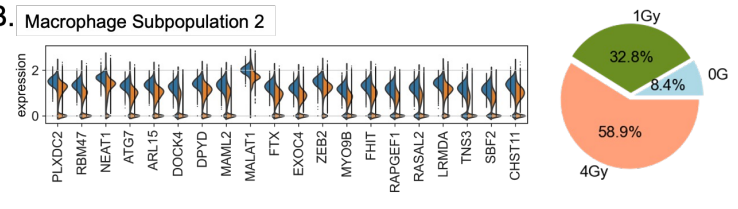
### A. Macrophage Subpopulation 1



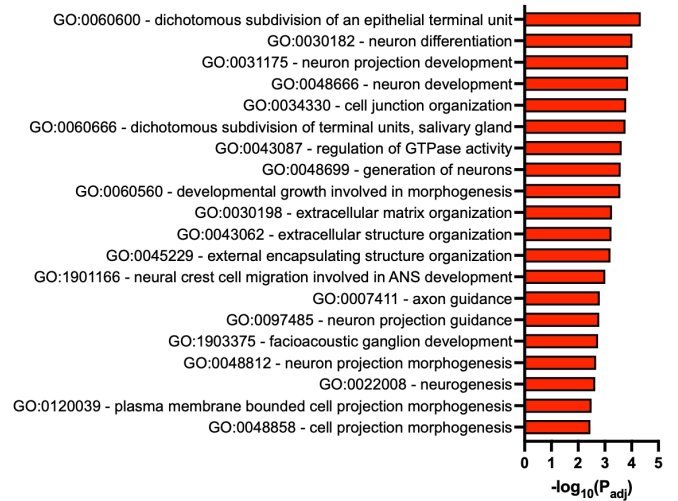
#### GO Biological Pathways



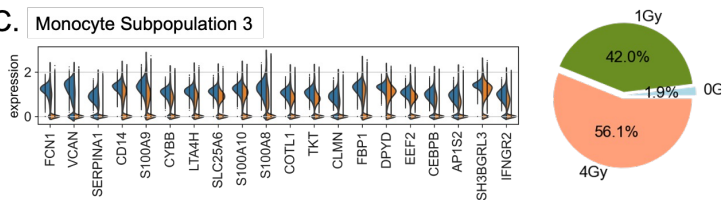
### B. Macrophage Subpopulation 2



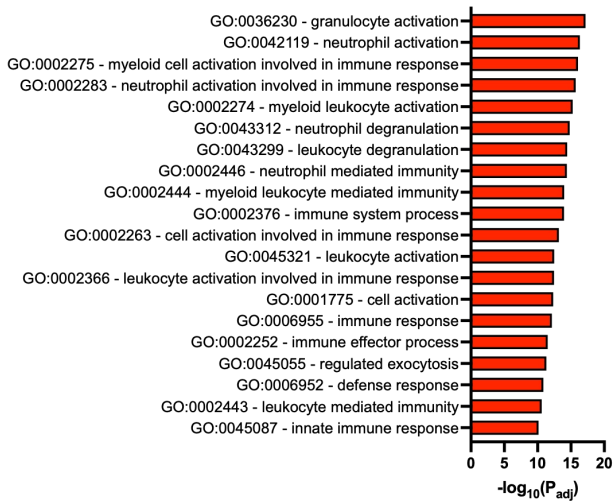
#### GO Biological Pathways



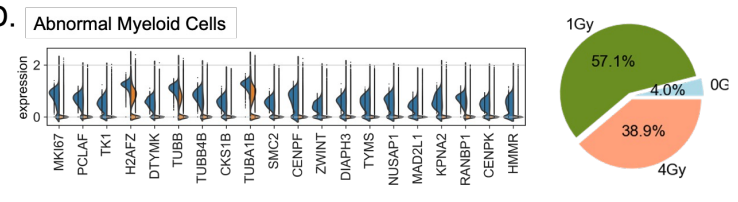
### C. Monocyte Subpopulation 3



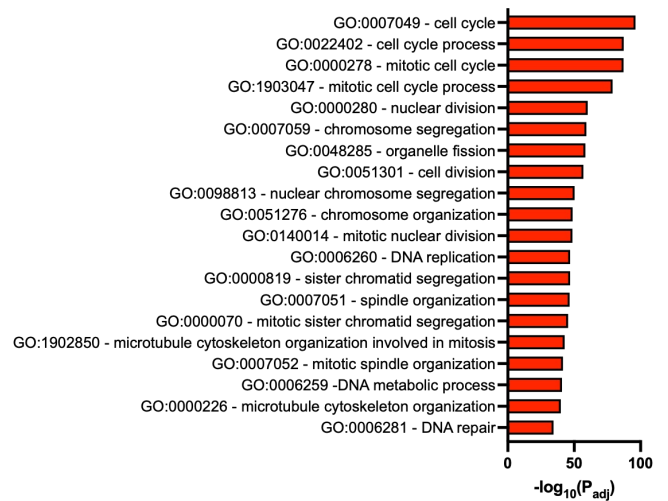
#### GO Biological Pathways



### D. Abnormal Myeloid Cells

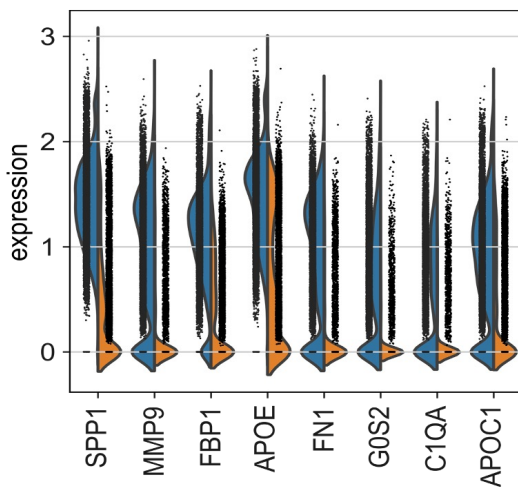


#### GO Biological Pathways

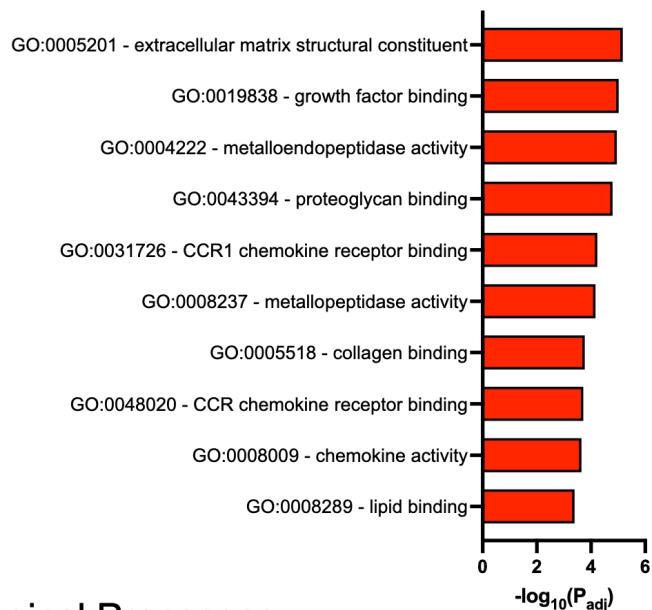


**Supplementary Figure 6. Identification of unique cell populations emerging in engineered bone marrow (eBM) 3-weeks post-irradiation.** Data correspond to single-cell RNA sequencing shown from eBMs in Figure 5. (A-D) Most significant differentially-expressed genes and pathways implicated in (A) Macrophage subpopulation 1, (B) Macrophage subpopulation 2, (C) Monocyte subpopulation 3, and (D) Abnormal myeloid cells.

**A.** Top significantly differentially-expressed genes in 1 Gy vs. Control

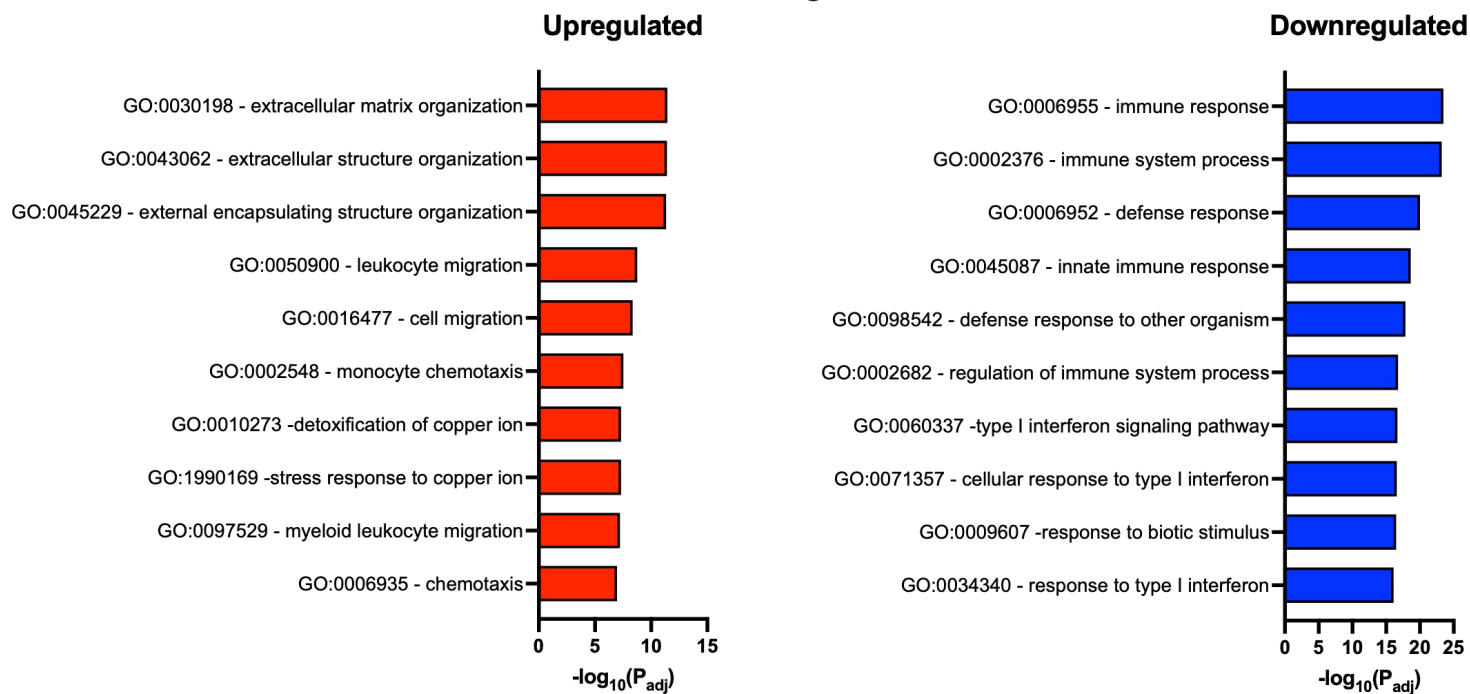


**B.** GO Molecular Functions Upregulated



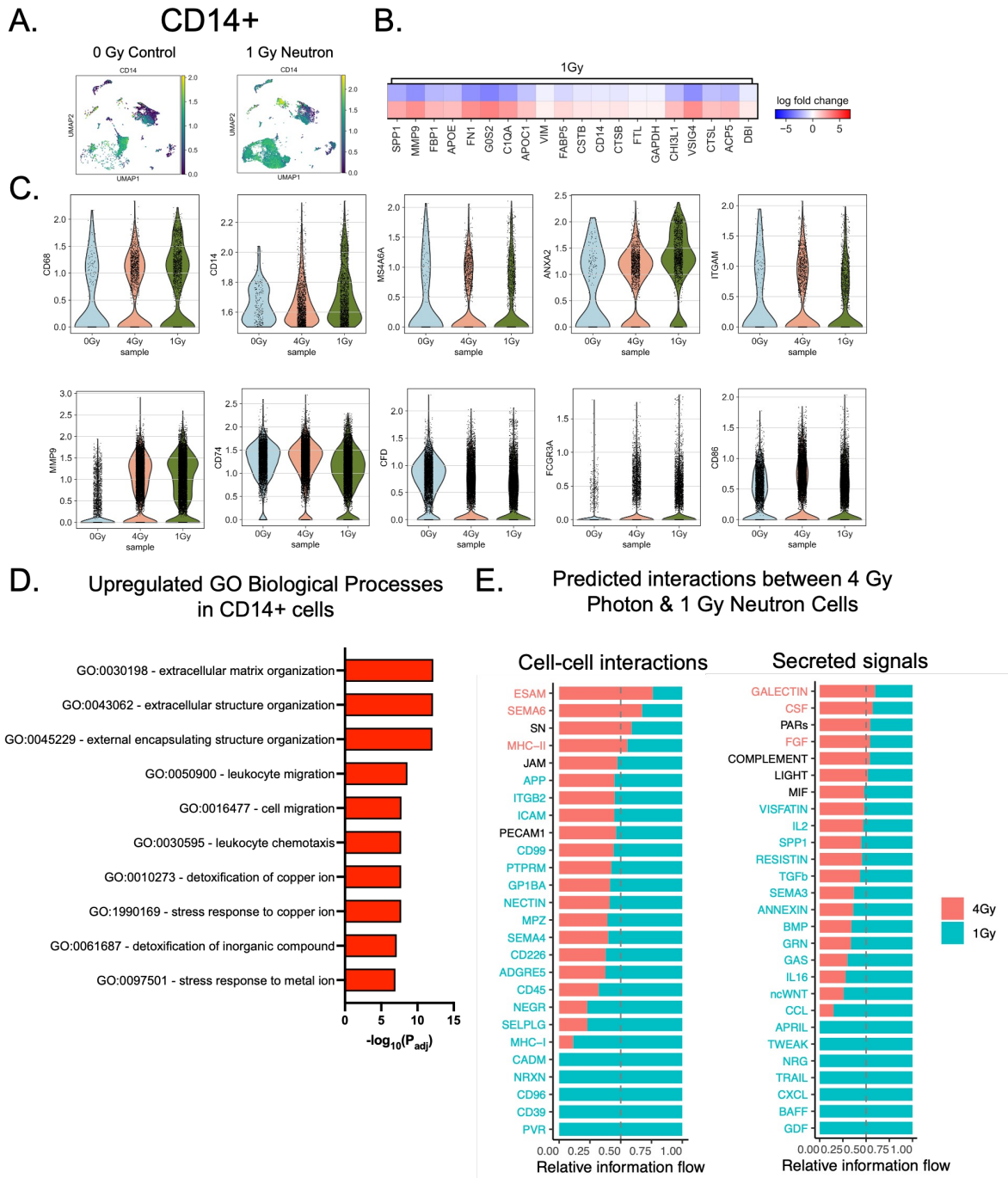
**C.**

GO Biological Processes



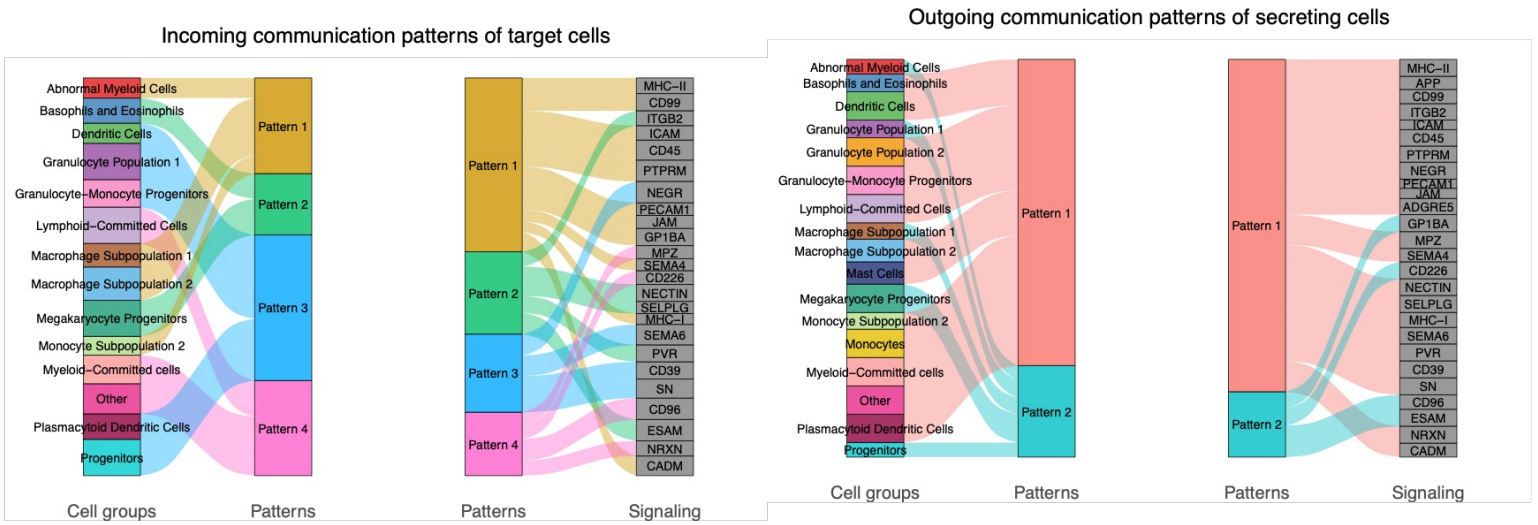
**Supplementary Figure 7. Total gene expression changes in all engineered bone marrow (eBM) cells by single cell RNA sequencing in Figure 5. (A) Top significant, differentially-expressed genes in 1 Gy cells as compared to the 0 Gy healthy control. (B) Pathway analysis of significantly upregulated genes in the neutron-irradiated samples in GO molecular functions. (C) upregulated/downregulated genes in the neutron-irradiated samples in GO biological processes.**



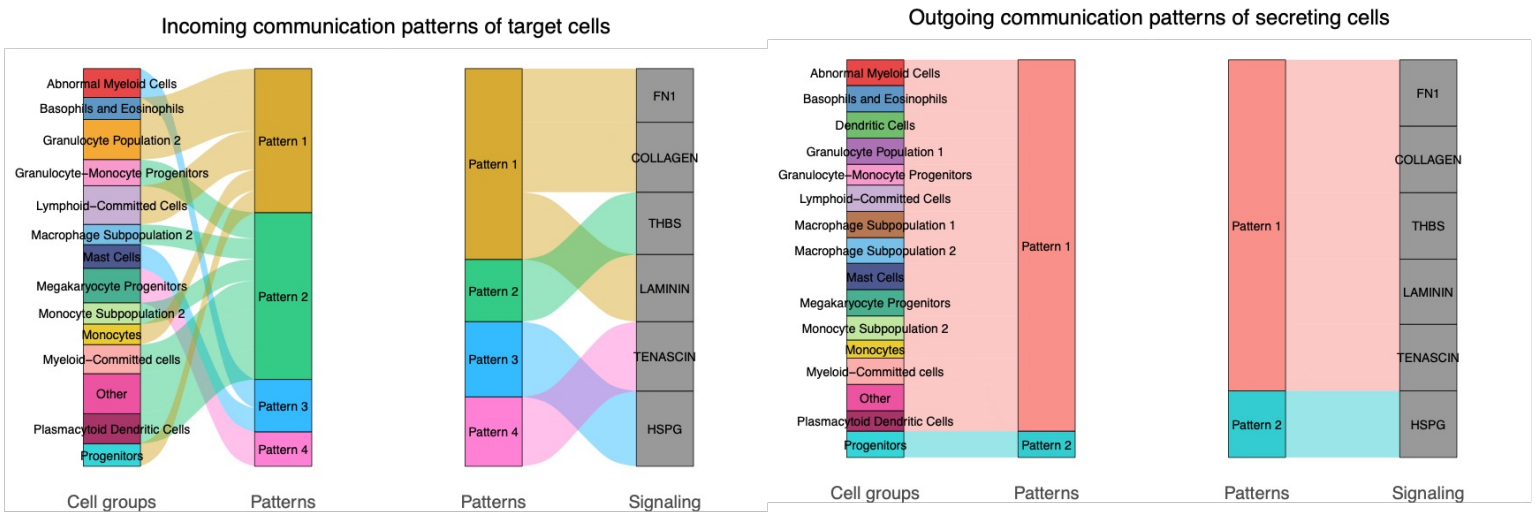


**Supplementary Figure 8. Differential expression of isolated CD14+ cells from engineered bone marrow (eBM) single-cell RNA sequencing in response to radiation. (A)** UMAP visualization of CD14+ myeloid cells. **(B)** Differential gene expression in neutron irradiated (1 Gy) tissues relative to controls. **(C)** Example violin plots of key myeloid genes. **(D)** GO pathway analysis of top biological processes. **(E)** Comparative analysis of cell-cell interactions and secreted signals in 4 Gy Photon versus 1 Gy Neutron eBM tissues.

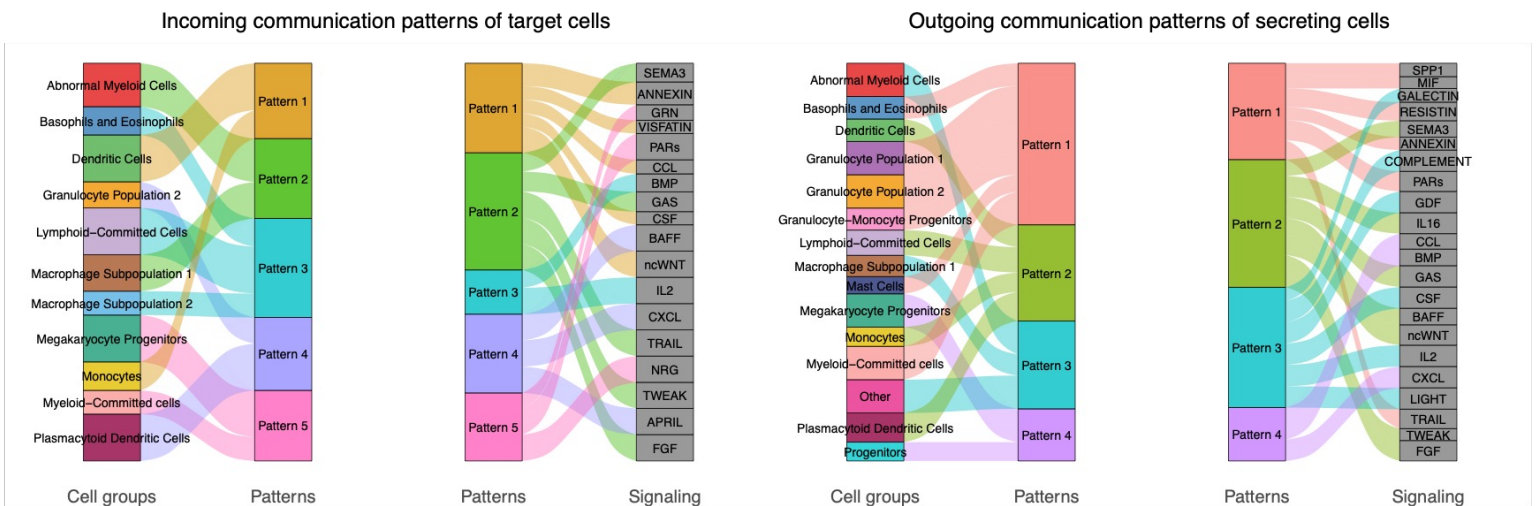
# A. 1 Gy: Inferred cell-cell interactions



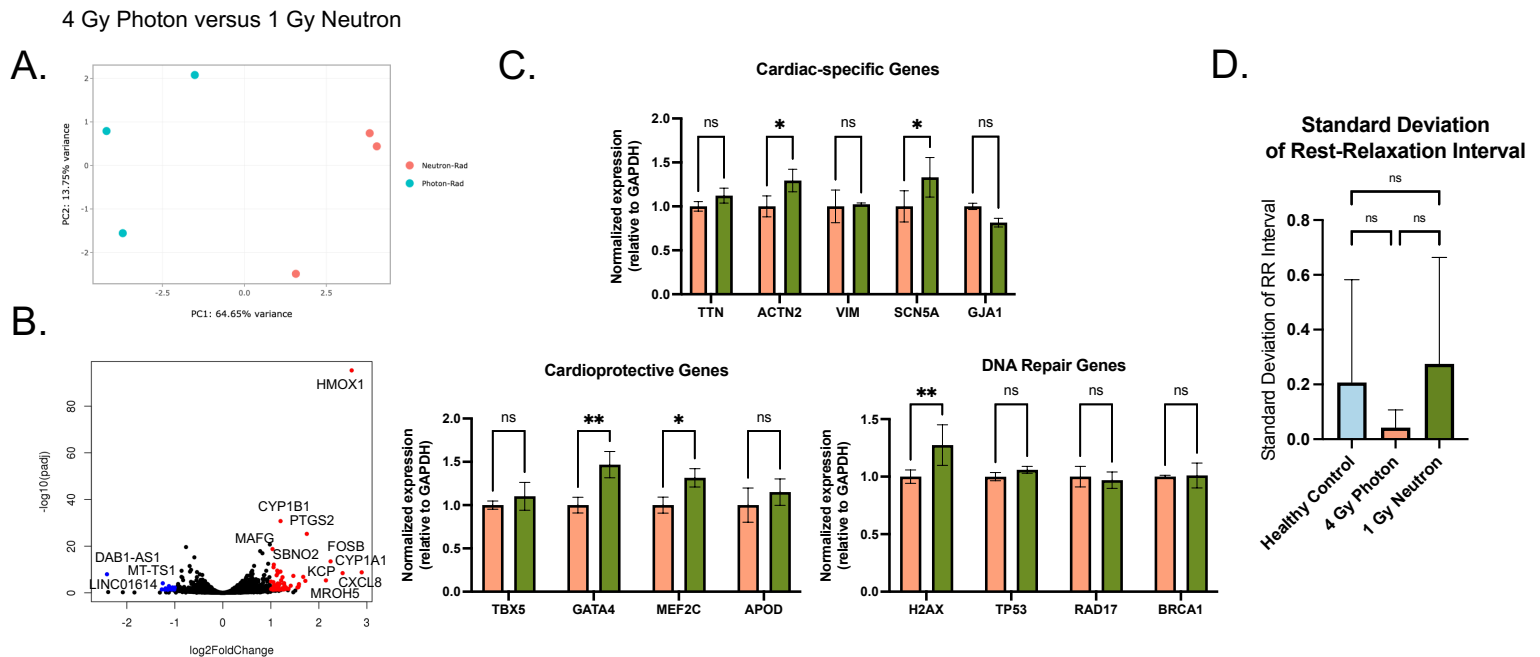
# B. 1 Gy: Inferred cell-ECM interactions



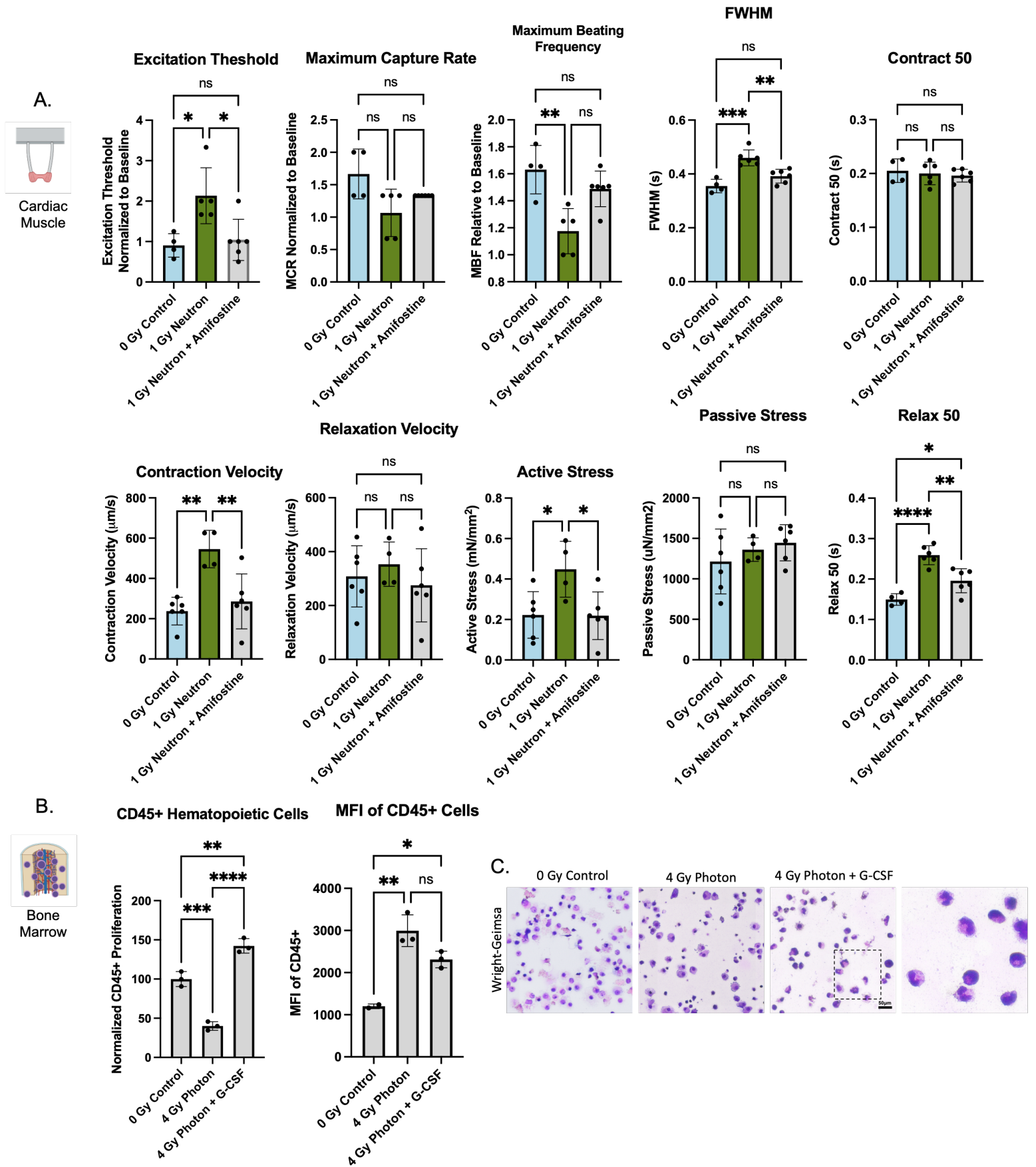
# C. 1 Gy: Inferred secreted signal interactions



**Supplementary Figure 9. Computationally-derived interactions within the 1Gy Neutron-irradiated engineered bone marrow (eBM) blood cells via CellChat. (A-C) Inferred incoming and outgoing (A) cell-cell, (B) cell-ECM, and (C) cell-secretome interactions.**



**Supplementary Figure 10. Differentially expressed changes associated with engineered cardiac tissues (eCT) in response to 4 Gy photon vs 1 Gy neutron radiation.** (A) Principle component analysis of eCT in response to photon and neutron radiation. (B) Differentially expressed genes between the two groups ( $p_{adj} < 0.05$ ,  $\log_2FC > 1$ ). (C) Individualized visualization of genes associated with cardiac function/maturation, cardioprotection, and DNA repair, shown with multiple, unpaired t-tests \* $p < 0.05$ ; \*\* $p < 0.01$ . (D) From calculating the standard deviation of the rest-relaxation interval during cardiac muscle tissue spontaneous beating, no differences were observed to indicate presences of arrhythmias ( $n = 2 - 4$  tissues).



**Supplementary Figure 11. Use of engineered human tissues for testing of therapeutic effects of radioprotective agents.** (A) Pre-treatment of engineered cardiac tissues (eCT) with radioprotective drug Amifostine prior to acute exposures prevented abnormal hypertrophic functionality of neutron-irradiated tissues. (B) G-CSF stimulation increases CD45+ cell production by (C) increasing neutrophil production *in vitro*.