

Cell Reports, Volume 42

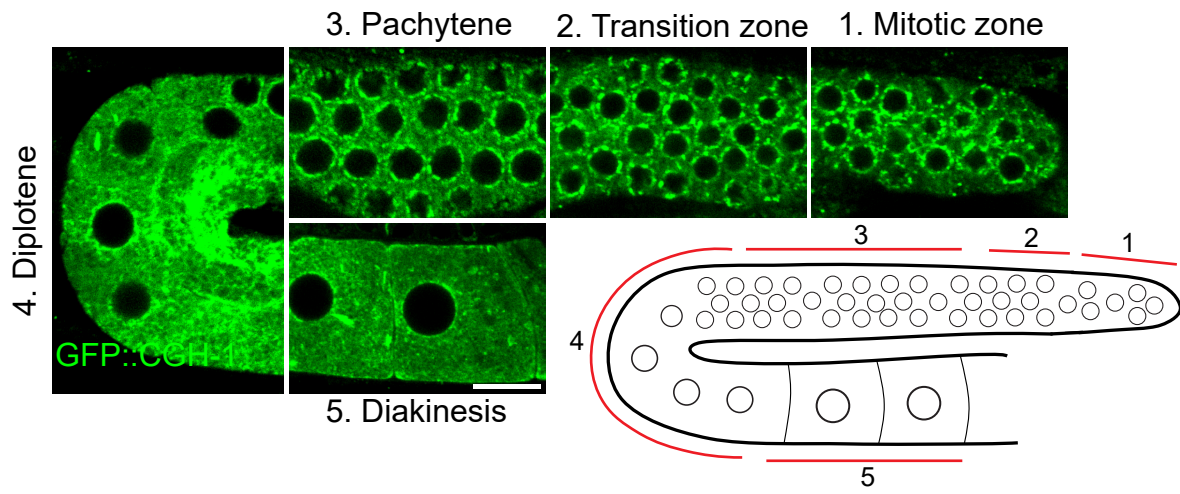
Supplemental information

**Condensate cooperativity underlies
transgenerational gene silencing**

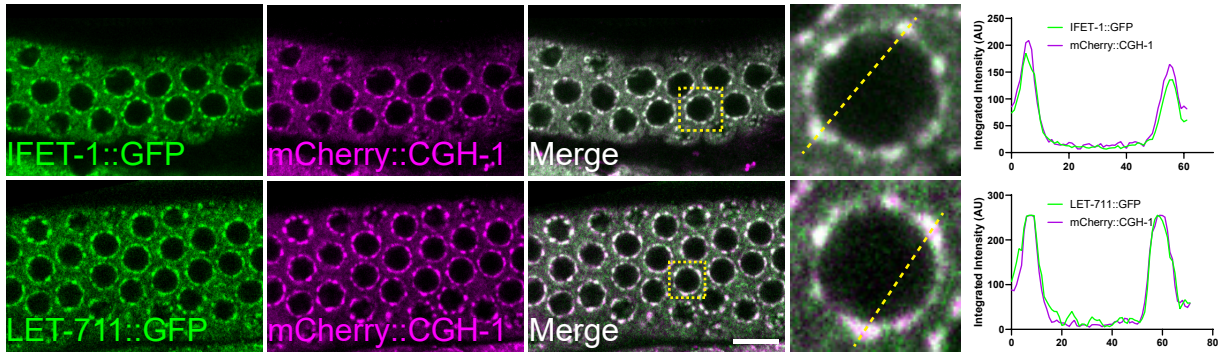
Zhenzhen Du, Kun Shi, Jordan S. Brown, Tao He, Wei-Sheng Wu, Ying Zhang, Heng-Chi Lee, and Donglei Zhang

Figure S1

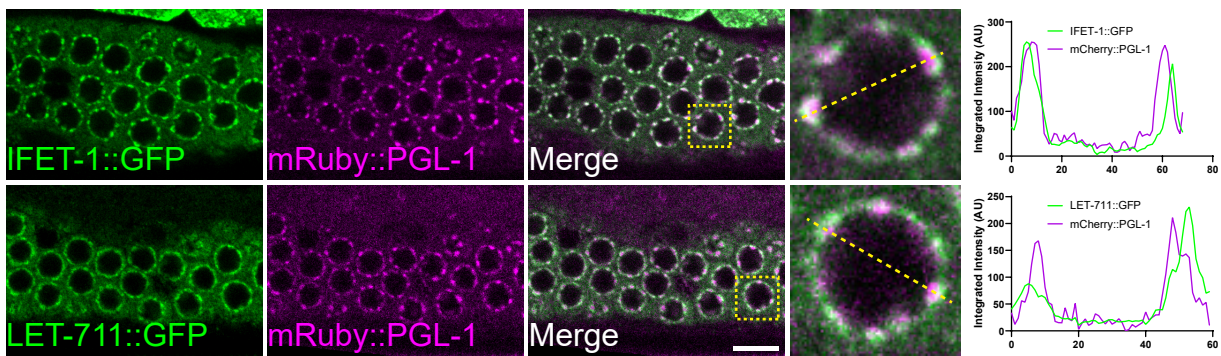
A



B



C



D

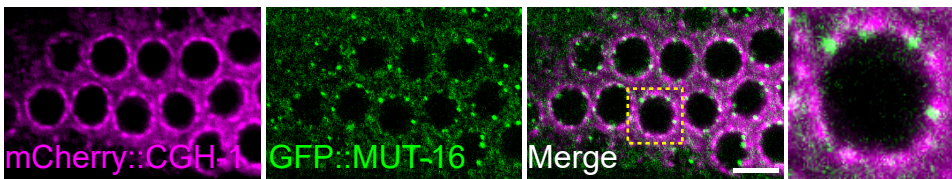
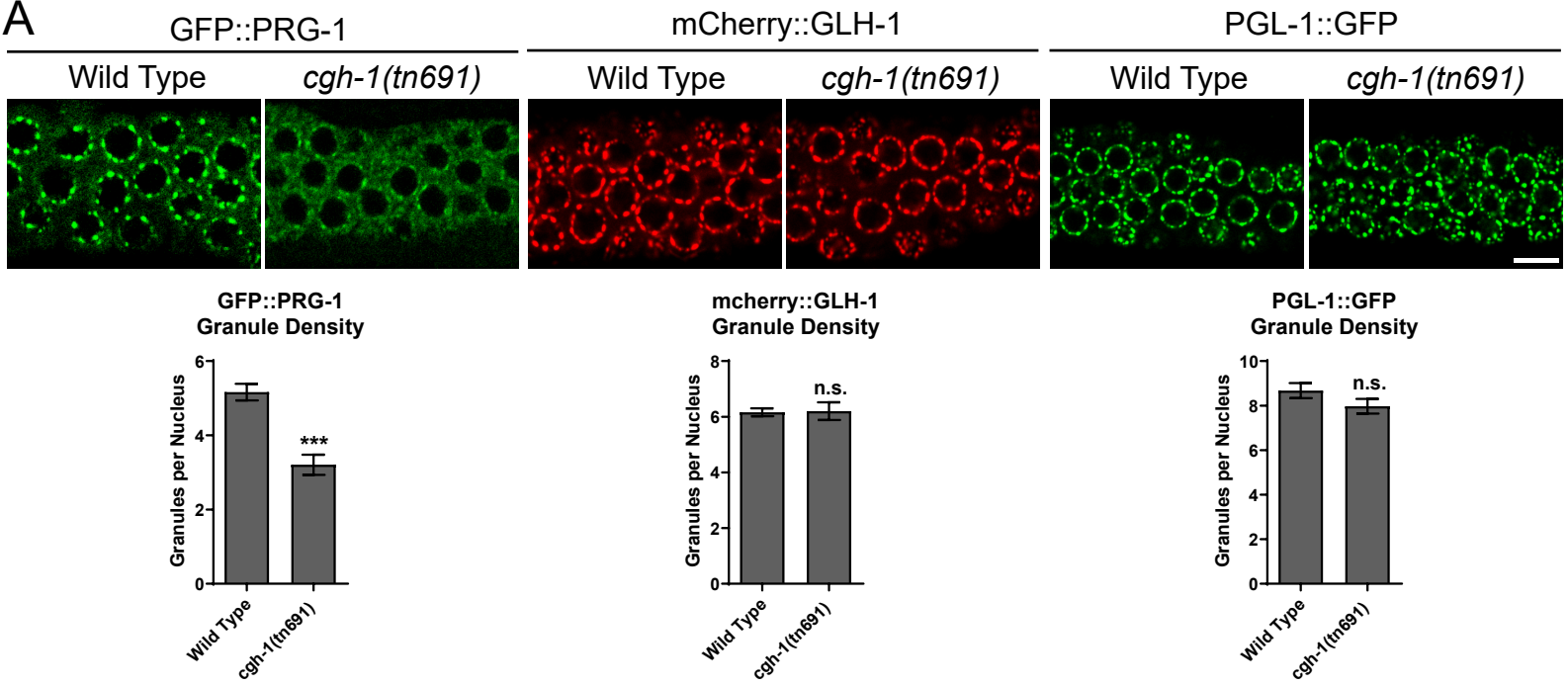


Figure S1. P bodies form perinuclear condensates on the cytoplasmic side of germ granules in the germline of *C. elegans*, related to Figure 1

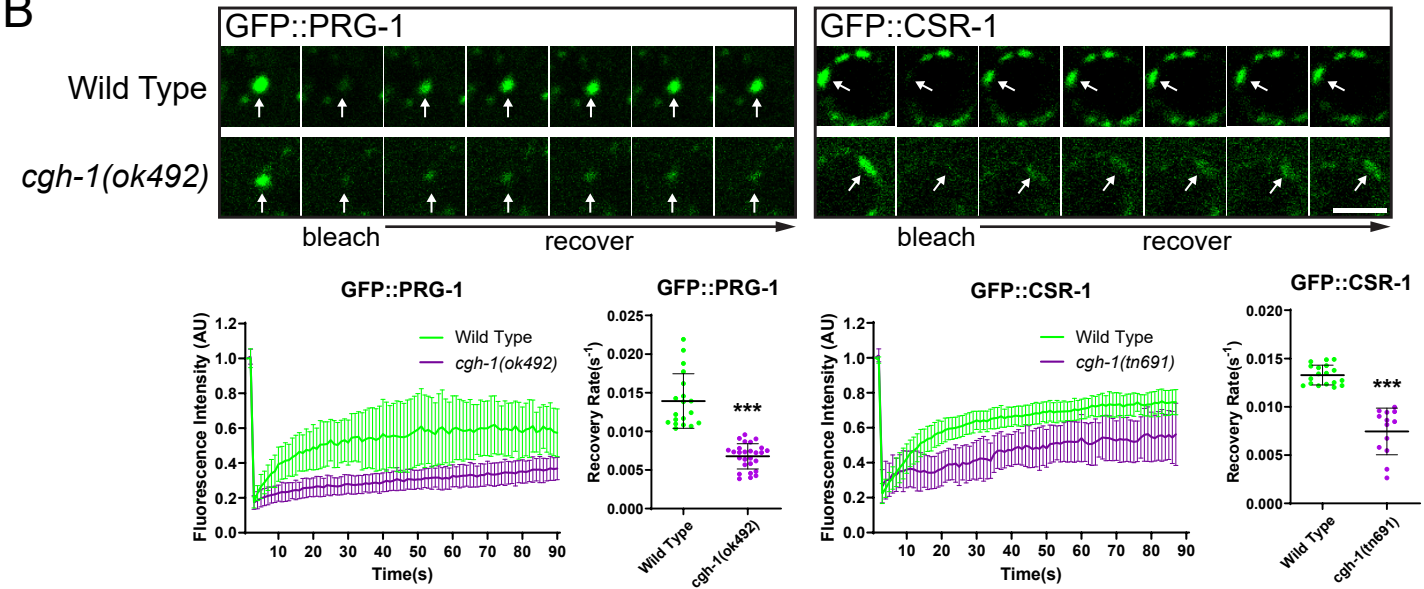
- (a) Fluorescent micrographs show the localization of P body marker CGH-1 in the indicated regions of the *C. elegans* germline. Bars: 10 micrometers. A cartoon shows the relative positions of distinct regions in the gonad of *C. elegans* (bottom right).
- (b) Fluorescent micrographs show the co-localization of two distinct P body markers in the pachytene region of adult germlines. Bars: 10 micrometers. The line in the merged image indicates the position of the line scan for measuring fluorescence intensity across single germline nuclei (right).
- (c) Fluorescent micrographs show the localization of the indicated P body and P granule markers in the pachytene region of adult germlines. Bars: 10 micrometers. The line in the merged image indicates the position of the line scan for measuring fluorescence intensity across single germline nuclei (right).
- (d) Fluorescent micrographs show the localization of the P body marker CGH-1 and Mutator foci marker MUT-16 in the pachytene region of the adult germline. Bars: 10 micrometers.

Figure S2

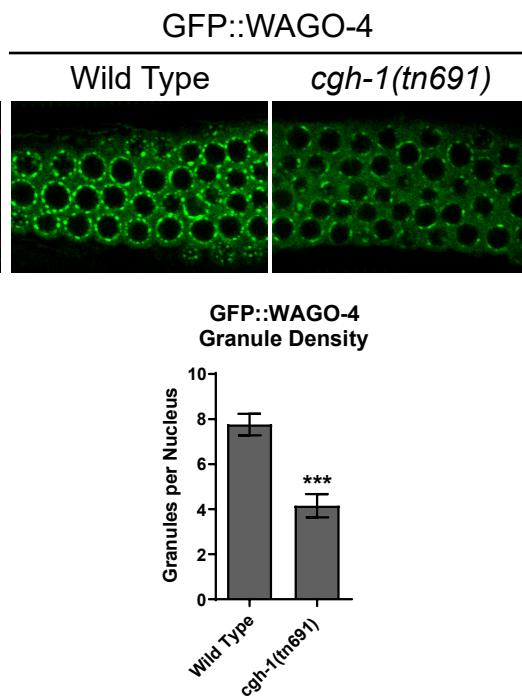
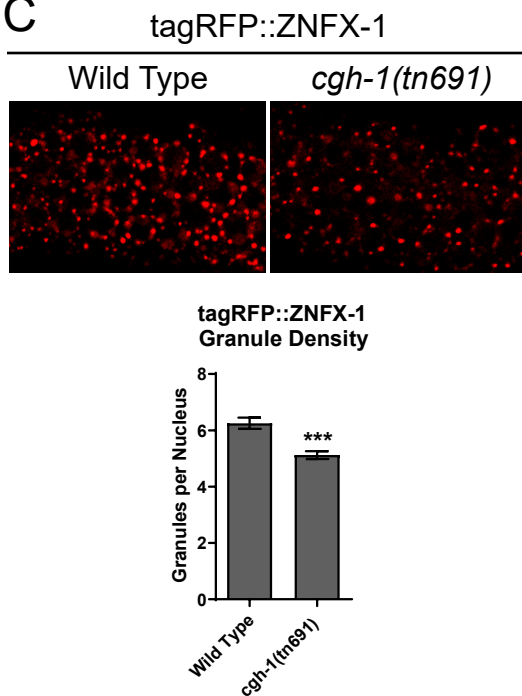
A



B



C



D

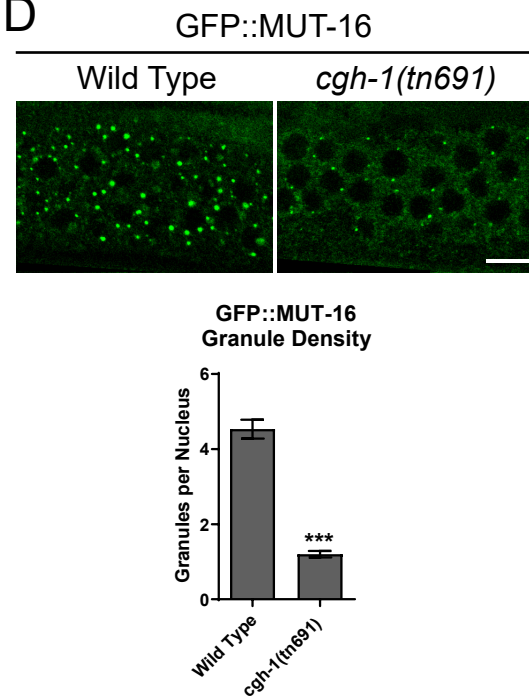


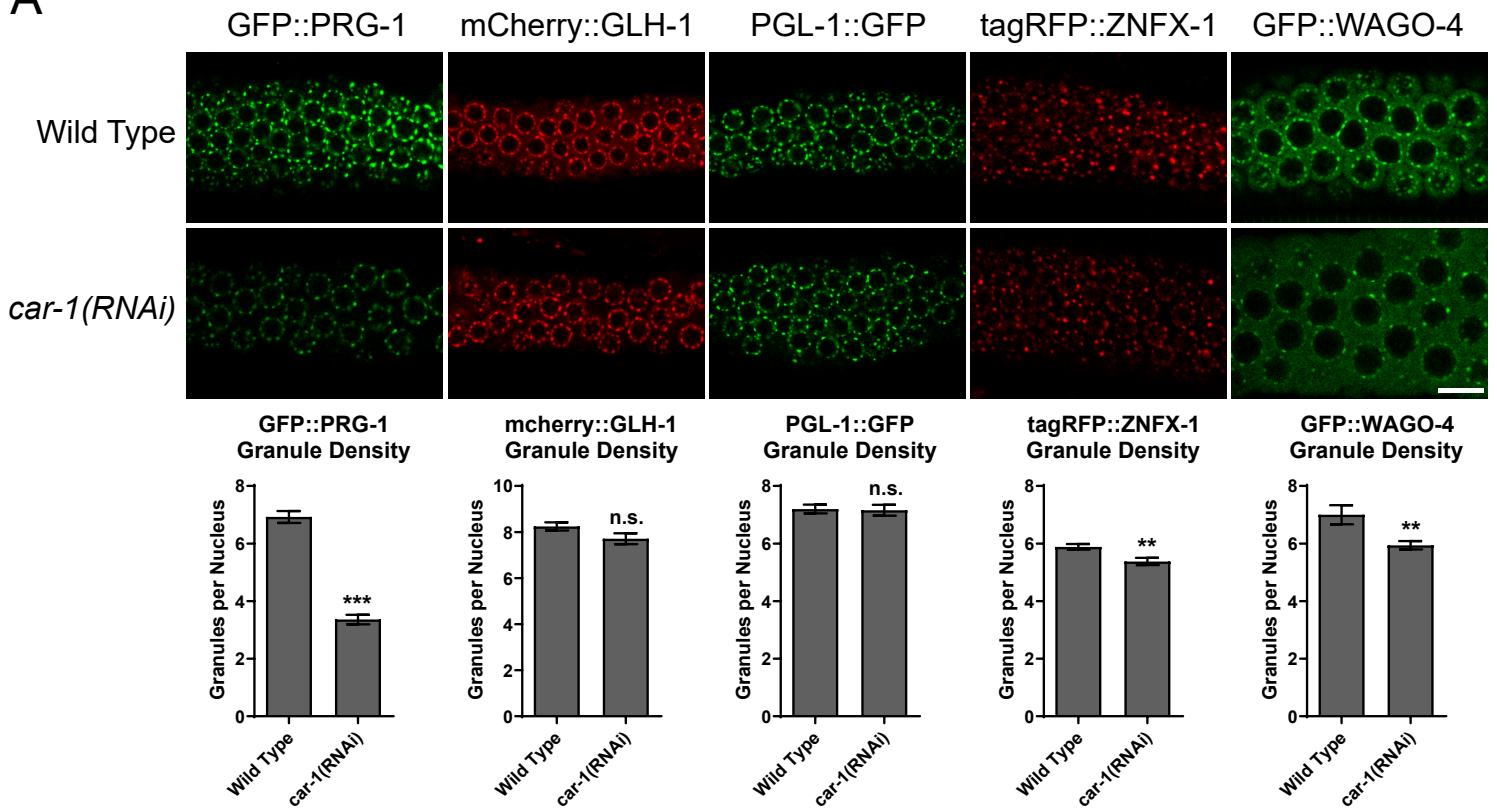
Figure S2. P body factor CGH-1 promotes the localization of small RNA pathway factors at germ granules, related to Figure 3

- (a) Fluorescent micrographs show the localization of indicated P granule factors in wild type or in *cgh-1(tn691)* mutant animals Bars: 10 micrometers (Top). Granule density (number of granule/nuclei/section) of indicated proteins in wild type or the indicated *cgh-1* mutants (bottom). Statistical analysis was performed using a one-tailed Student's t-test. Bars indicate the mean, errors bars indicate the standard deviation. Distributions represent data collected from 8-12 independent gonad images.
- (b) Fluorescence recovery after photobleaching (FRAP) analyses indicate that the dynamics of indicated P granule factors, including PRG-1 and CSR-1, are reduced in *cgh-1(ok492)* mutant animals. The arrows indicate the P granules that are photobleached. Bars: 10 micrometers (Top). The quantification of FRAP analyses show the average fluorescence signals of GFP::PRG-1 or GFP::CSR-1 at the indicated times (seconds) after photobleaching (bottom). >10 granules at different time points were calculated for each strain. Error bars indicate the standard deviations of fluorescence intensities or recovery ratios.
- (c-d) Fluorescent micrographs show the localization of indicated Z granule markers (left) Mutator foci marker MUT-16 (right) in the pachytene region of adult germlines in wild type or in *cgh-1(tn691)* mutant animals. Bars: 10

micrometers (top). Granule number (number of granule/nuclei/section) of indicated proteins in wild type or the *car-1* RNAi-treated animals (bottom). Statistical analysis was performed using a one-tailed Student's t-test. Bars indicate the mean, errors bars indicate the standard deviation. Distributions represent data collected from 8-12 independent gonad images.

Figure S3

A



B

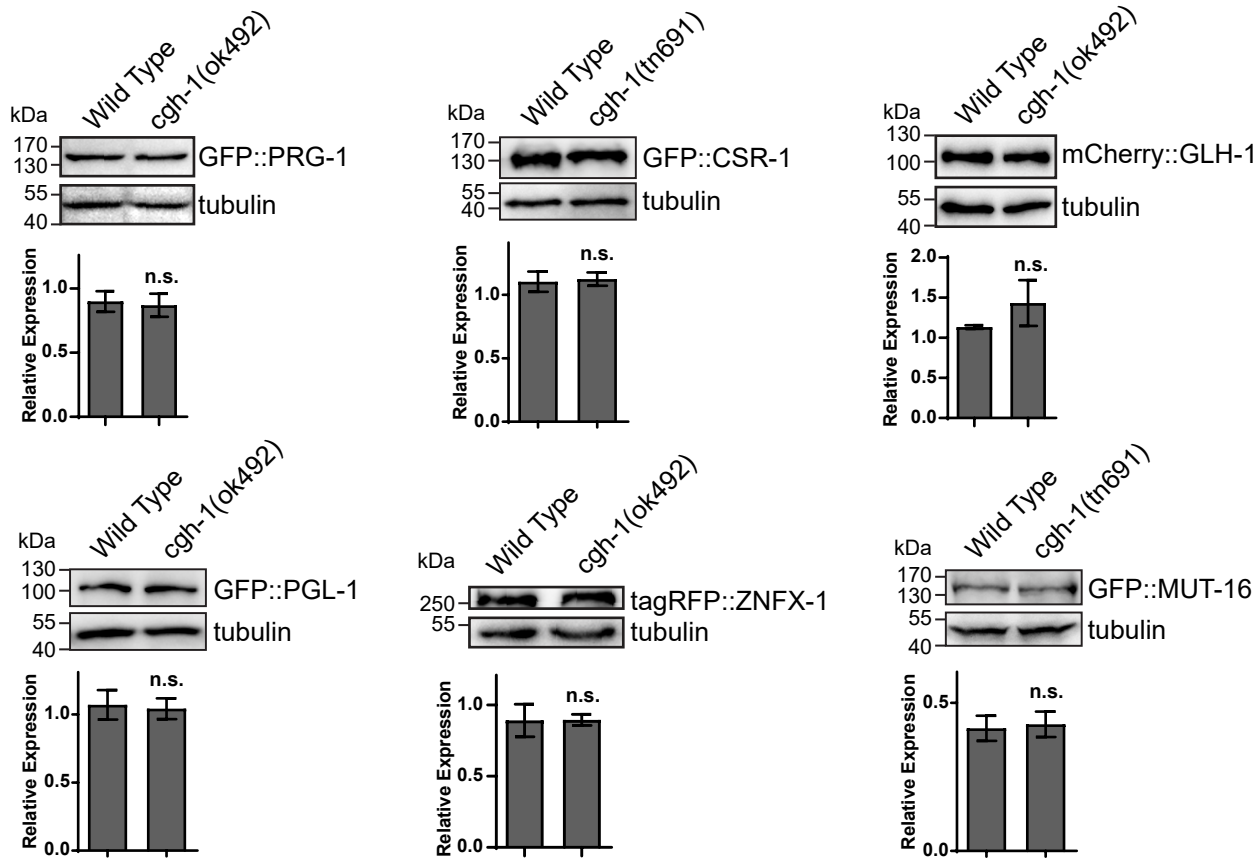
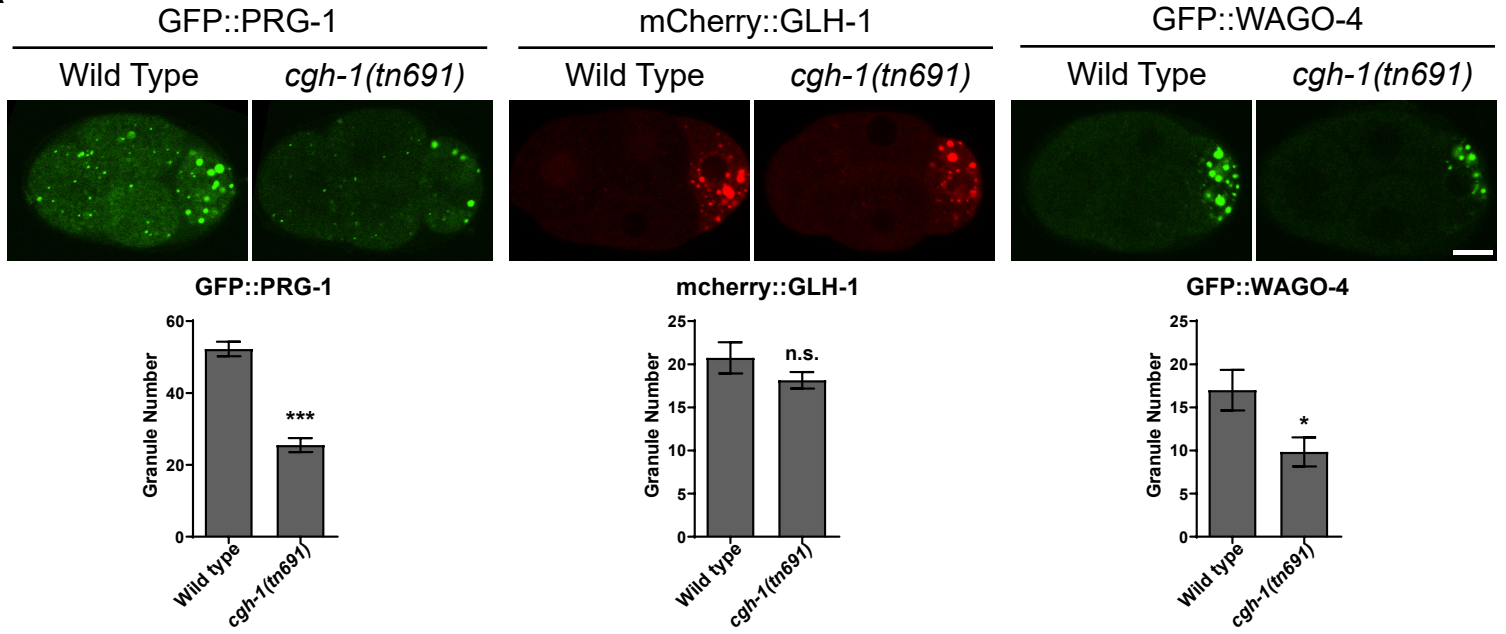


Figure S3. The localization and expression of small RNA pathway factors in P body mutants or RNAi-treated animals, related to Figure 3

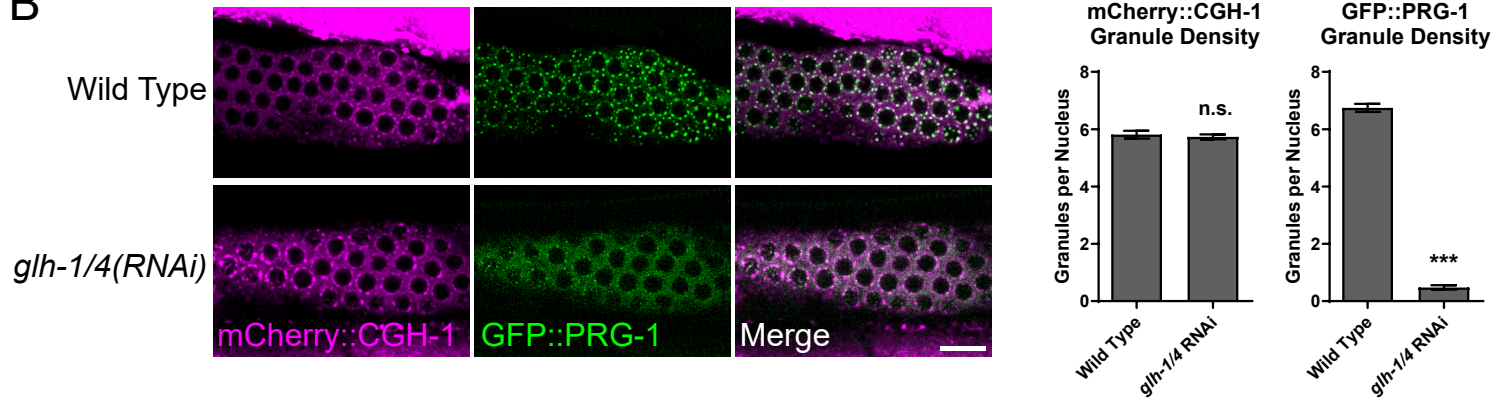
- (a) Fluorescent micrographs show the localization of indicated proteins in the pachytene region of adult germlines in wild type or in *car-1* RNAi-treated animals. Bars: 10 micrometers (top). Granule number (number of granule/nuclei/section) of indicated proteins in wild type or the *car-1* RNAi-treated animals (bottom). Statistical analysis was performed using a one-tailed Student's t-test. Bars indicate the mean, errors bars indicate the standard deviation. Distributions represent data collected from 8-12 independent gonad images.
- (b) Western blots show the levels of indicated proteins in wild type or in the indicated *cgh-1* mutants.

Figure S4

A



B



C

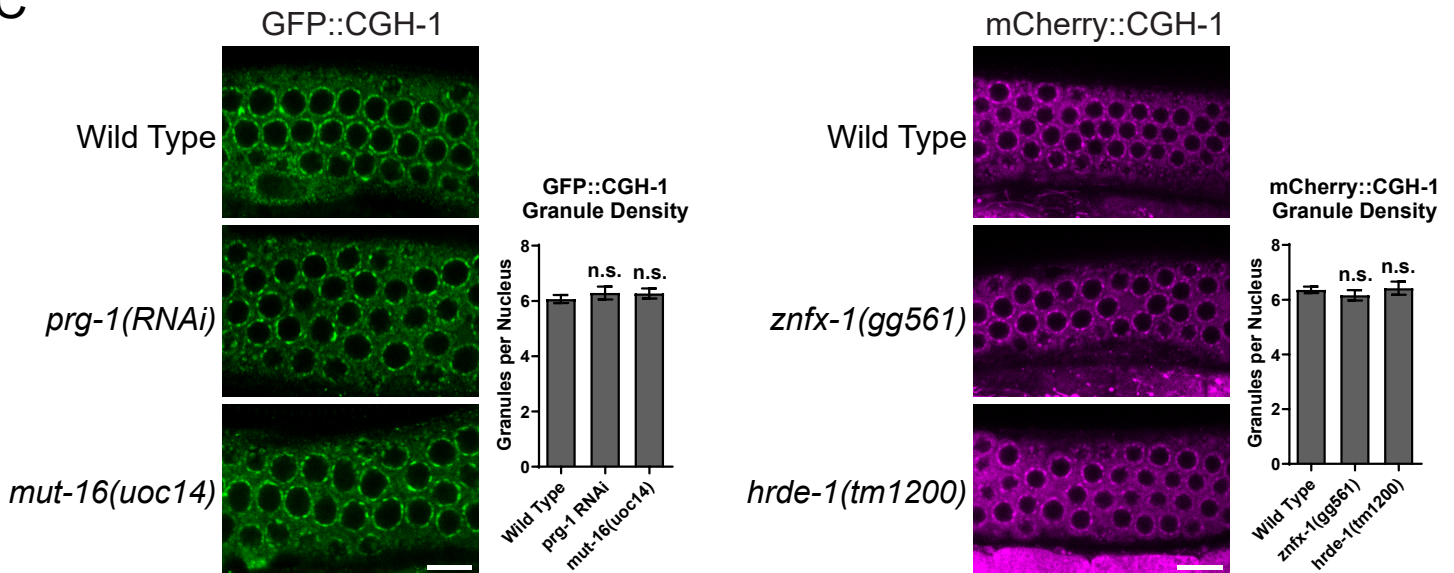


Figure S4. The localization of germ granule or P body factors in the indicated animals, related to Figure 3

- (a) Fluorescent micrographs show the localization of the indicated P granule factors in the 4-cell embryos wild type or *cgh-1(tn691)* mutant animals. Bars: 10 micrometers (Top). Granule number of indicated proteins in wild type embryos or the indicated *cgh-1* mutant embryos (bottom). Statistical analysis was performed using a one-tailed Student's t-test. Bars indicate the mean, errors bars indicate the standard deviation. Distributions represent data collected from 8-12 embryo images.
- (b) Fluorescent micrographs show the perinuclear localization of P granule factor PRG-1, but not P body factor CGH-1, is reduced in *glh-1* and *glh-4* double RNAi treated animals. Bars: 10 micrometers (left). Statistical analysis was performed using a one-tailed Student's t-test. Bars indicate the mean, errors bars indicate the standard deviation (right). Distributions represent data collected from 8-12 independent gonad images.
- (c) Fluorescent micrographs show the localization of P body marker GFP::CGH-1 in the pachytene region of adult germlines in wild type or in the indicated mutant or RNAi-treated animals. Bars: 10 micrometers. Statistical analysis was performed using a one-tailed Student's t-test. Bars indicate the mean, errors bars indicate the standard deviation. Distributions represent data collected from 8-12 independent gonad images.

Figure S5

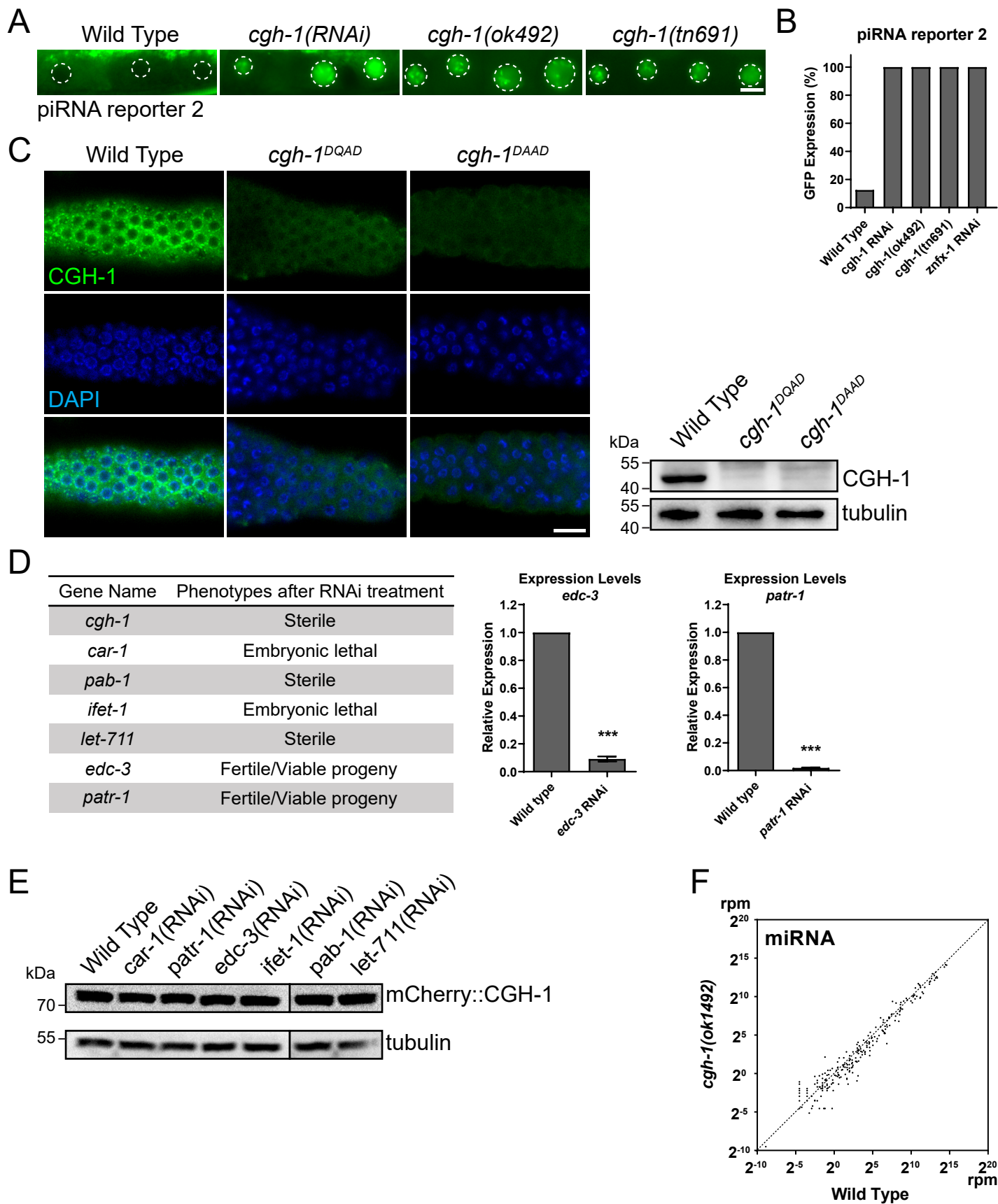


Figure S5. Characterization of the role of CGH-1 in piRNA-dependent gene silencing and the RNAi effects of knocking down P body factors, related to

Figure 4

- (a) GFP expression in the piRNA reporter #2 in the indicated strains. In this reporter, the expression of Histone H2B::GFP (nuclear) is silenced by a synthetic GFP-targeting piRNA in wild type animals. Dotted circles indicate the location of maturing oocyte nuclei. Bars: 10 micrometers.
- (b) Percentage of screened animals showing GFP expression in the piRNA reporter #2 in the indicated strains. We define the expression of GFP (on/off) by observing whether the indicated mutant or RNAi-treated animals exhibit visible nuclear GFP signals in their germline.
- (c) Fluorescent micrographs (left) and Western blots (right) showing the expression levels of CGH-1 wild type, CGH-1 DQAD, and CGH-1 DAAD. Bars: 10 micrometers.
- (d) The phenotypes (left) and qRT-PCR measurements of mRNA levels (right) of indicated RNAi knockdown of indicated P body factors. Statistical analysis was performed using a one-tailed Student's t-test. Bars indicate the mean, errors bars indicate the standard deviation. The qRT-PCR measurements present knockdown effects of three biological replates.
- (e) Western blots show the levels of CGH-1 in the indicated RNAi-treated animals.
- (f) A scatter plot showing the abundance of miRNAs in wild type worms compared to *cgh-1 (dz407)* mutant.

Figure S6

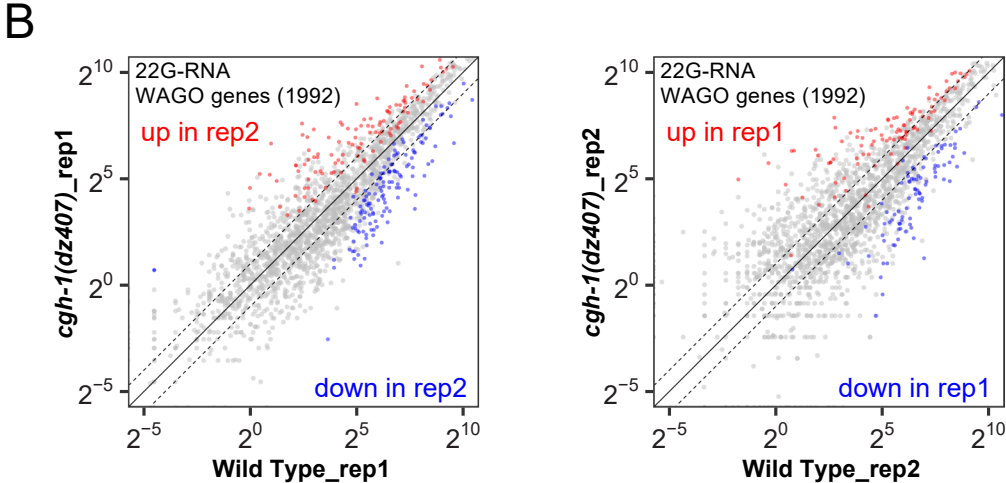
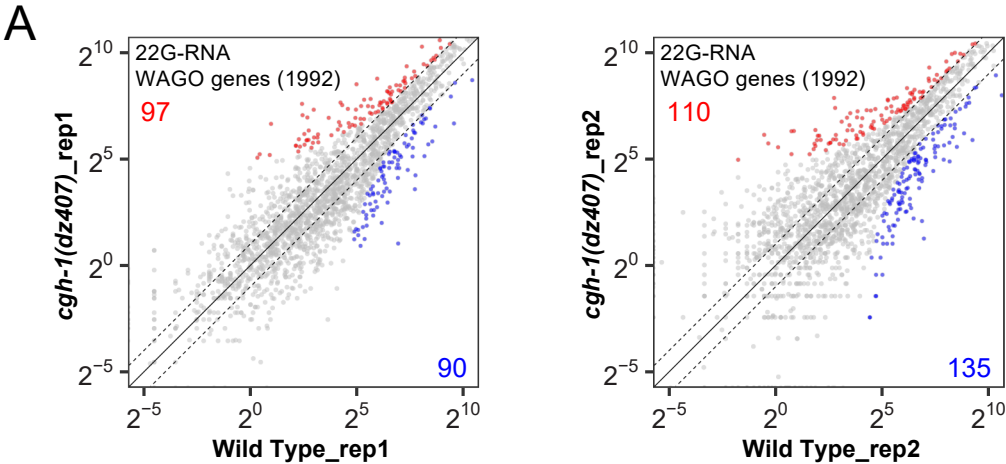
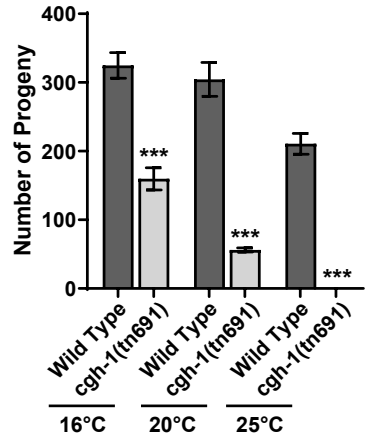


Figure S6. The comparison of changes in 22G-RNA levels in *cgh-1* mutants between two biological replicates, related to Figure 4.

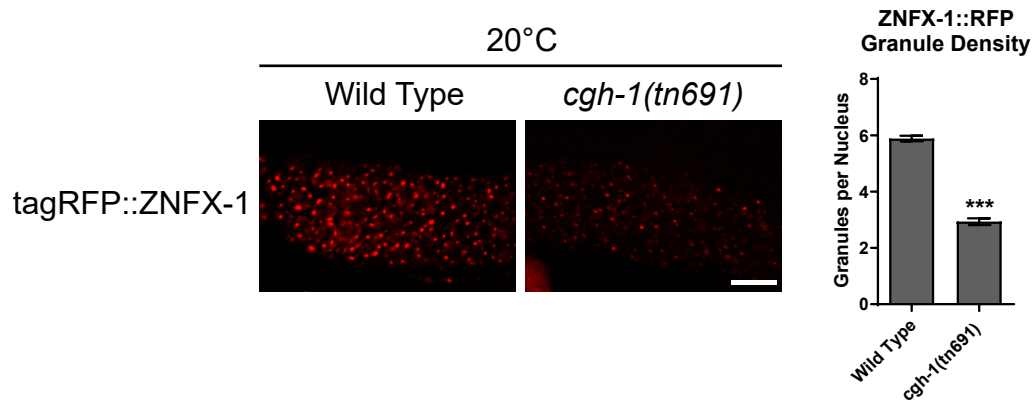
- (a) A scatter plot showing the abundance of 22G-RNAs mapped to WAGO target in wild-type worms compared to *cgh-1* mutant in two biological replicates. WAGO targets with significantly increased or decreased levels of 22G-RNA (adjusted $P < 0.05$, see STAR methods for details) and at least two-fold change are highlighted in red and blue dots, respectively.
- (b) A scatter plot showing the abundance of 22G-RNAs mapped to WAGO target in wild-type worms compared to *cgh-1* mutant in two biological replicates. The WAGO targets that exhibit significantly changes (adjusted $P < 0.05$, see STAR methods for details) and at least two-fold change in 22G-RNAs in one replicate (shown in Figure S6A) are highlighted in another replicate.

Figure S7

A



B



C

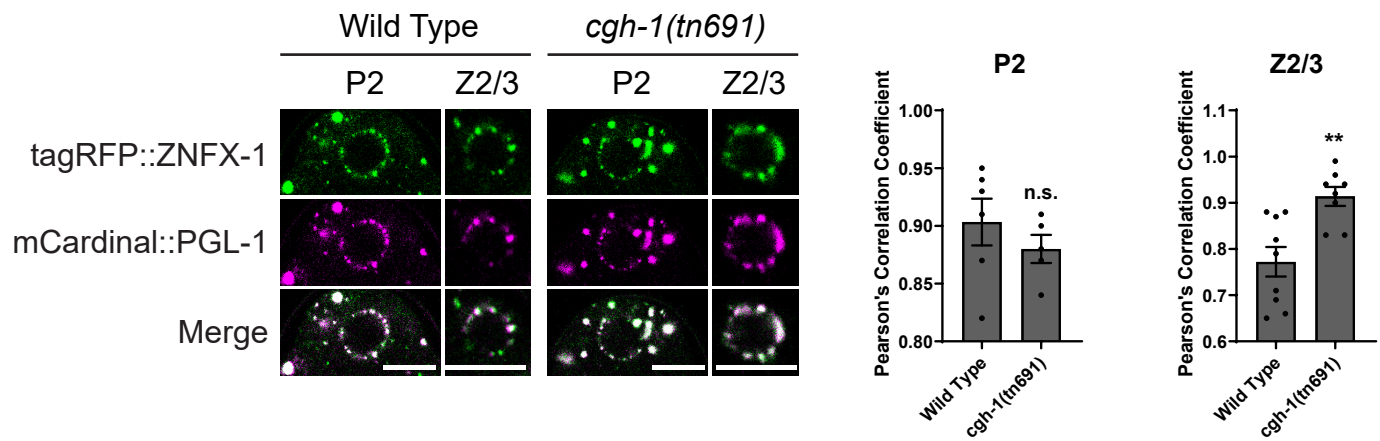


Figure S7. Characterization of temperature-sensitive *cgh-1 (tn691)* strain grown at permissive and non-permissive temperature, related to Figure 4, Figure 5, and Figure 6

- (a) The brood size of wild type or *cgh-1 (tn691)* mutant animals grown at the indicated temperature.
- (b) Fluorescent micrographs show the localization of indicated Z granule factor ZNFX-1 in the pachytene region of adult germlines in wild type or *cgh-1 (ok492)* mutant animals grown at 20 degree Celsius.
- (c) Fluorescent micrographs show the localization of P granule marker PGL-1 and Z granule marker ZNFX-1 in the P2 cells or Z2/Z3 cells of wild type or *cgh-1 (tn691)* mutant animals (left). Pearson's correlation coefficient of PGL-1 and ZNFX-1 signals (right).

