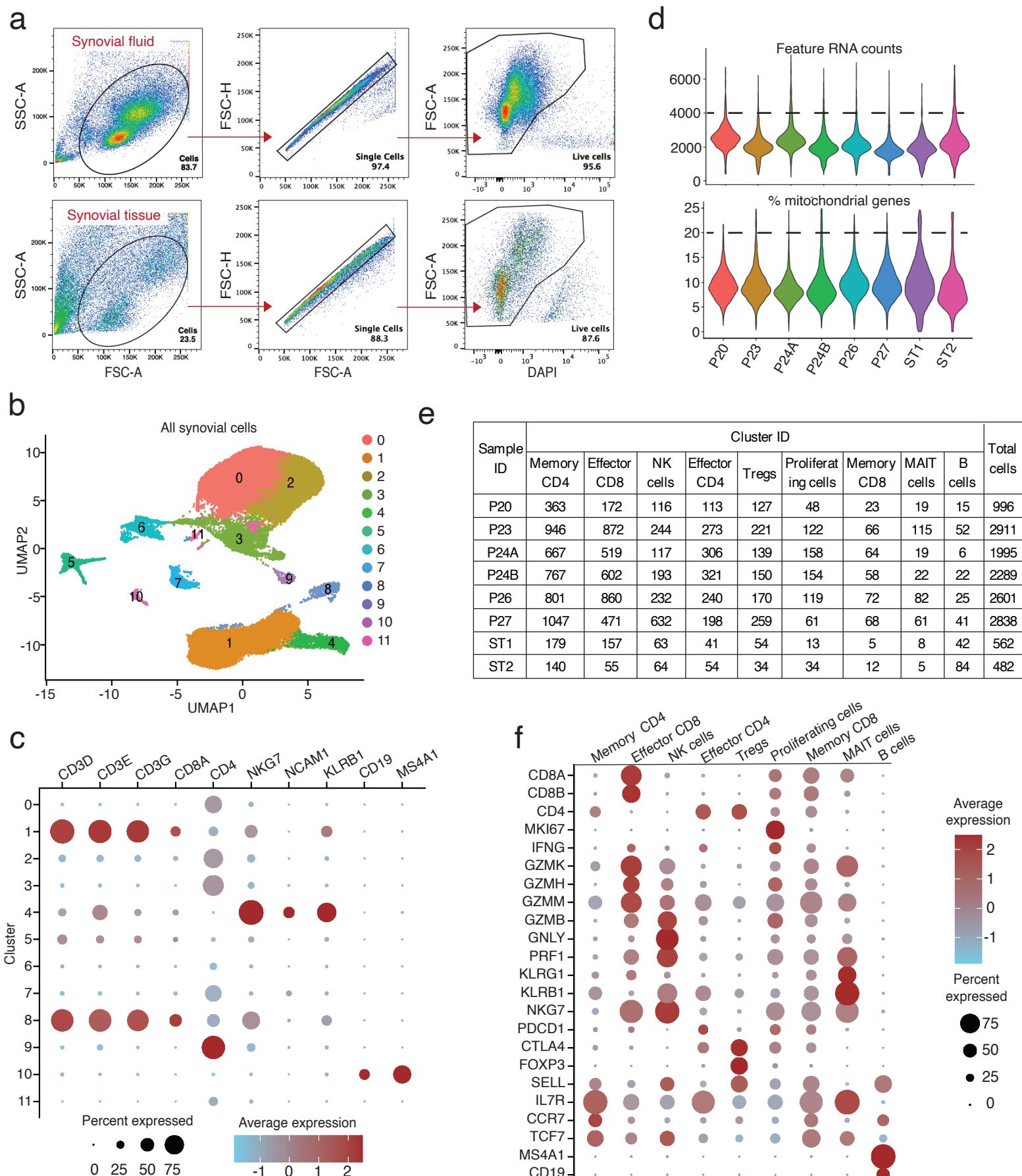


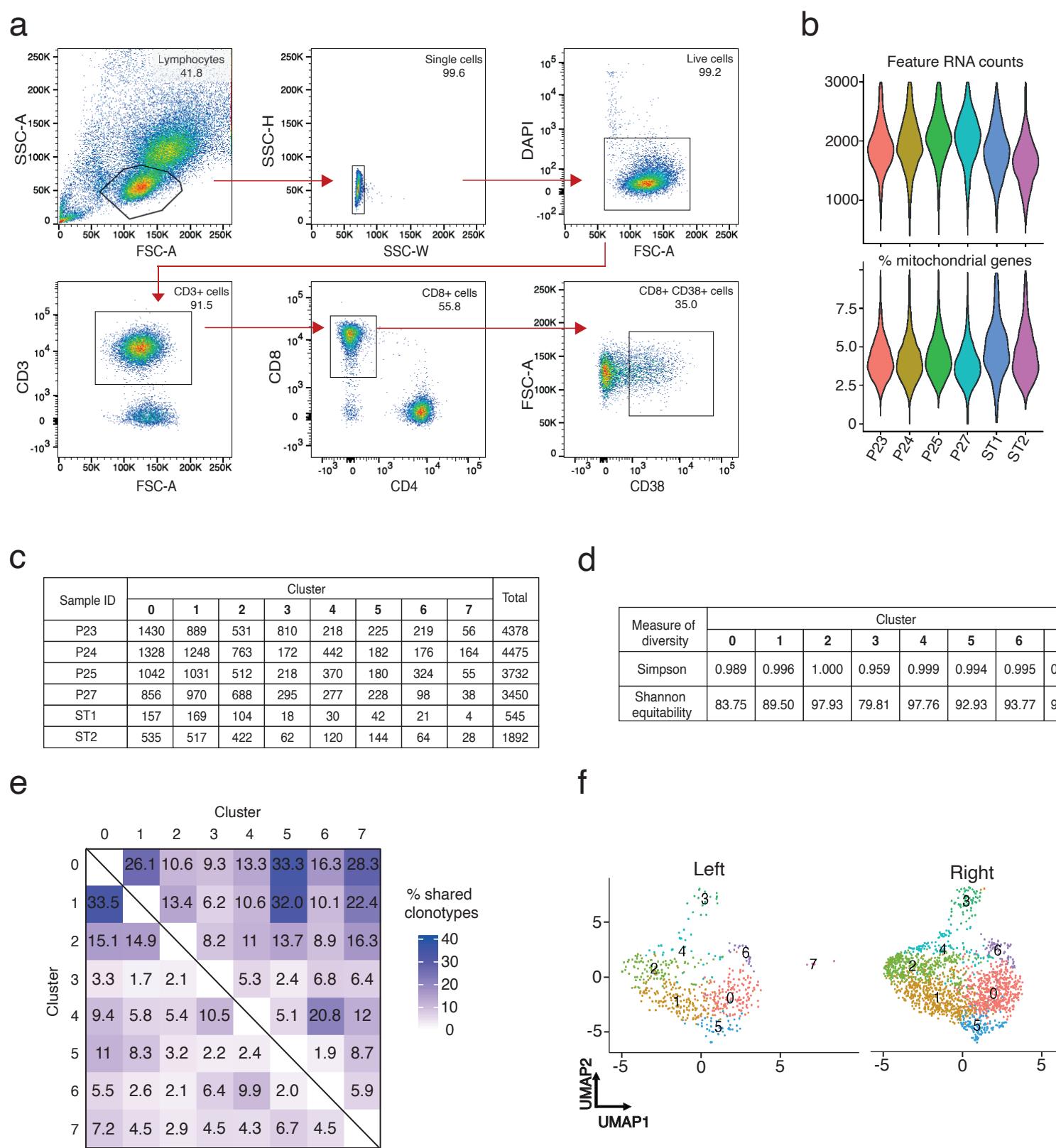
Supplementary figure 1. Multiplexed immunofluorescence of synovial tissue.

Multiplexed immunofluorescence images showing an example of a Ki67⁺PD-1⁺ synovial CD8 T cell (white arrow). CD3 staining is shown in red, CD8 in yellow, nuclear stain DAPI in blue, Ki67 in green and PD-1 in cyan.



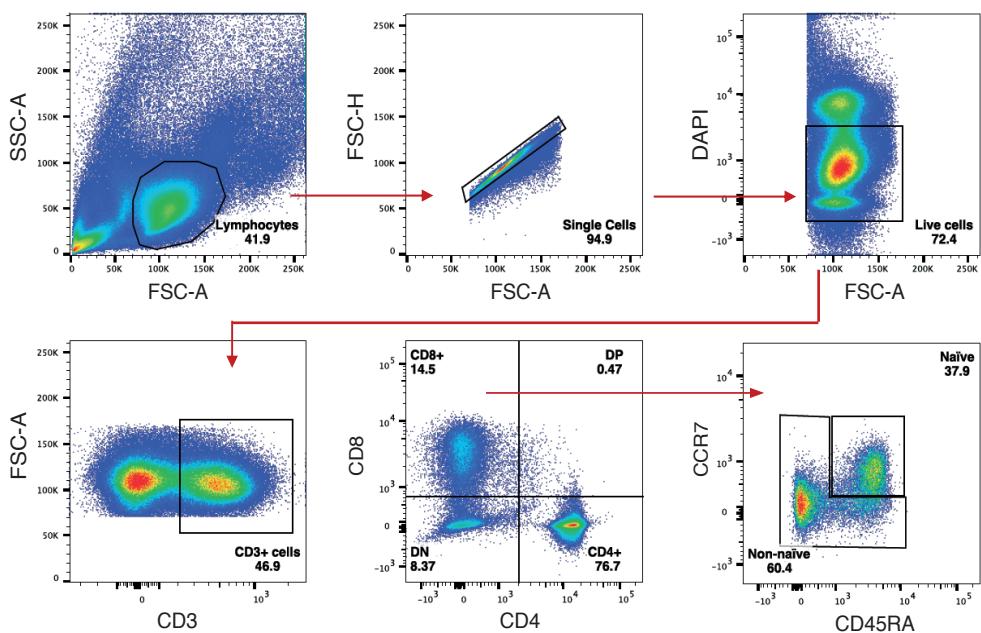
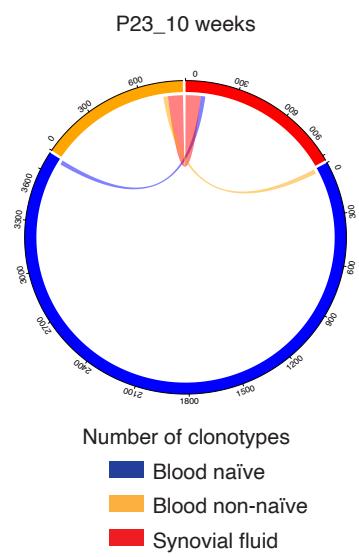
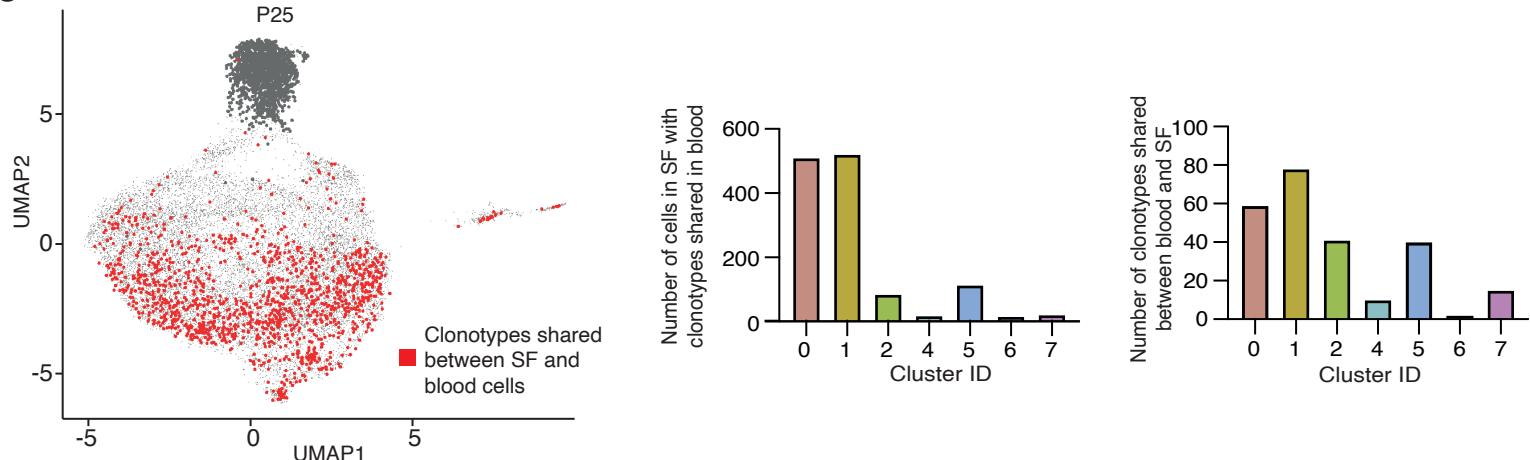
Supplementary figure 2. Single-cell gene expression analysis of live lymphocytes from ICI-arthritis synovium.

a) Gating scheme of isolation of live immune cells from ICI-arthritis synovium for scRNA-seq. **b)** UMAP visualization of all live cells from synovial samples that resolved into 12 different cell types. **c)** Dotplot visualization of differentially expressed genes that distinguish lymphocyte markers from other cell types. Color of the dot represents the average expression of the gene across cells in the cluster. Size of the dot represents the percentage of cells in the cluster with that gene detected. **d)** Distribution of all lymphocytes; cells with <4,000 total genes (feature RNA counts) and <20% mitochondrial genes were included in the analysis. **e)** Tabulation of QC filtered lymphocyte numbers contributing to nine distinct clusters from each individual patient. **f)** Dotplot visualization of differentially expressed genes that distinguish lymphocyte clusters.



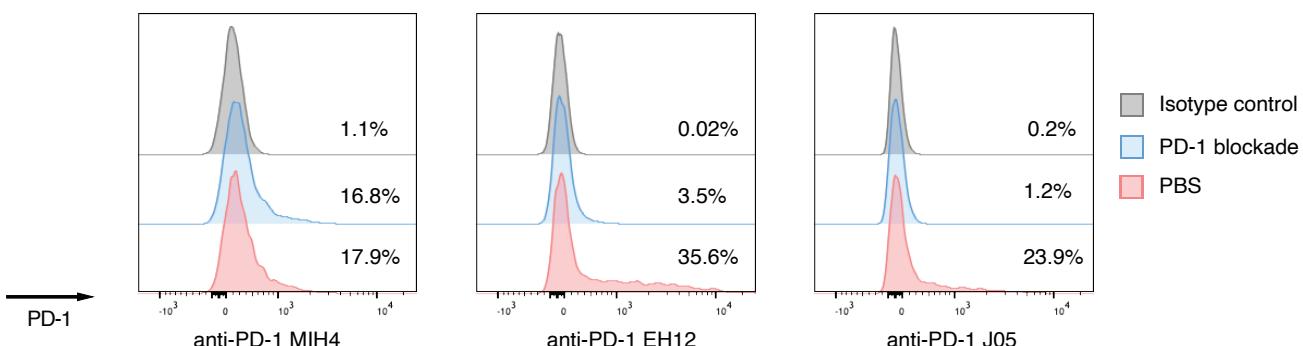
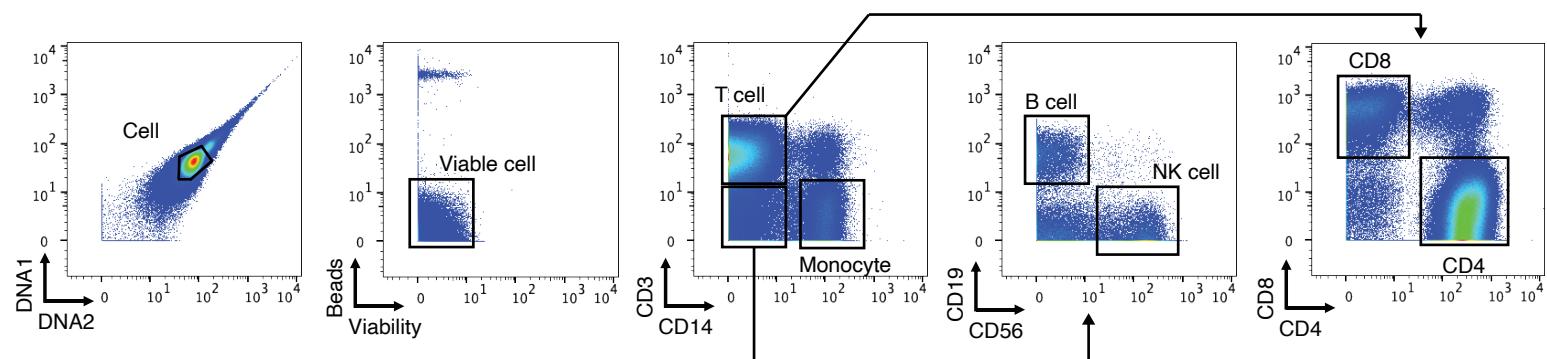
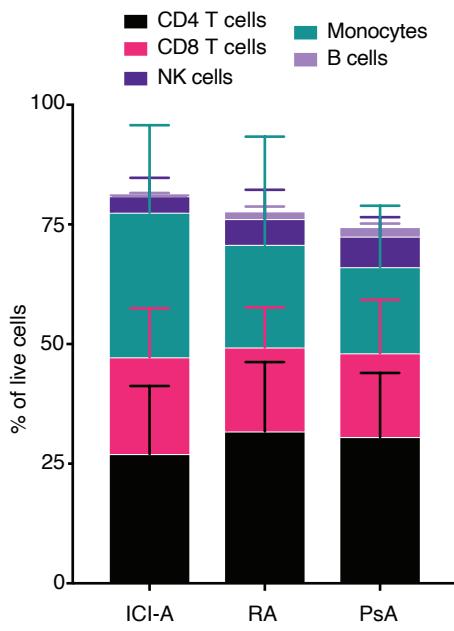
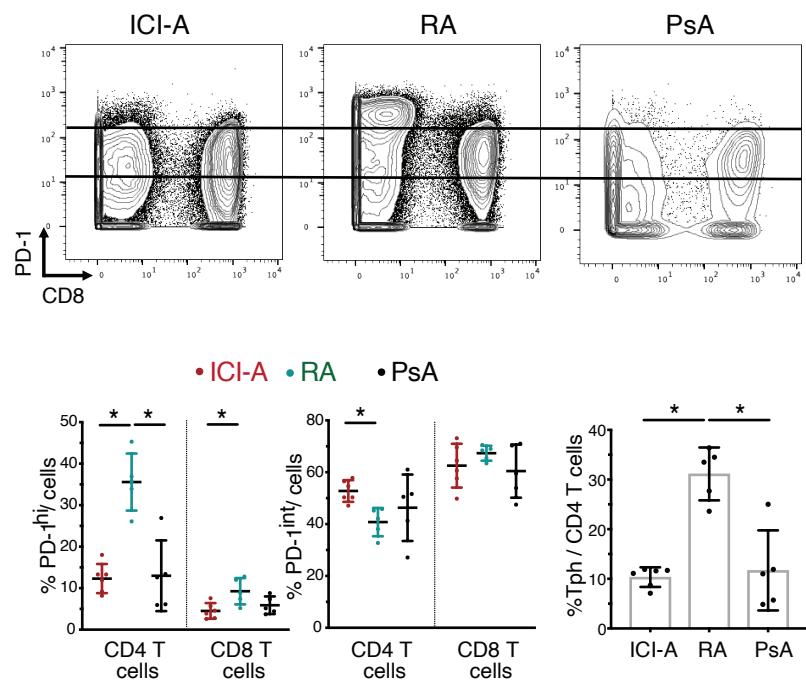
Supplementary figure 3. Single-cell analysis of CD8 T cells from ICI-arthritis synovial fluid and tissue.

a) Gating scheme of CD8 T cell isolation from ICI-arthritis synovium for scRNA-seq. **b)** Distribution of cells that passed quality control having <3,000 total genes (feature RNA counts) and <10% mitochondrial genes. **c)** Tabulation of QC filtered CD8 T cell numbers contributing to eight transcriptomically distinct clusters from each individual patient. **d)** Simpson and Shannon equitability indices calculated for the eight CD8 T cell clusters. **e)** The percentage of clonotypes shared between clusters. The row identity represents the denominator. **f)** Synovial tissue CD8 T cell scRNA-seq clusters from the left and right knee. Cluster IDs for figures (c) - (f): 0 - GZM+, 1 - Transitional, 2 - Tcm, 3 - MAIT, 4 - ZNF683+, 5 - KLRG1+, 6 - Dysfunctional, and 7 - Proliferating.

a**b****c**

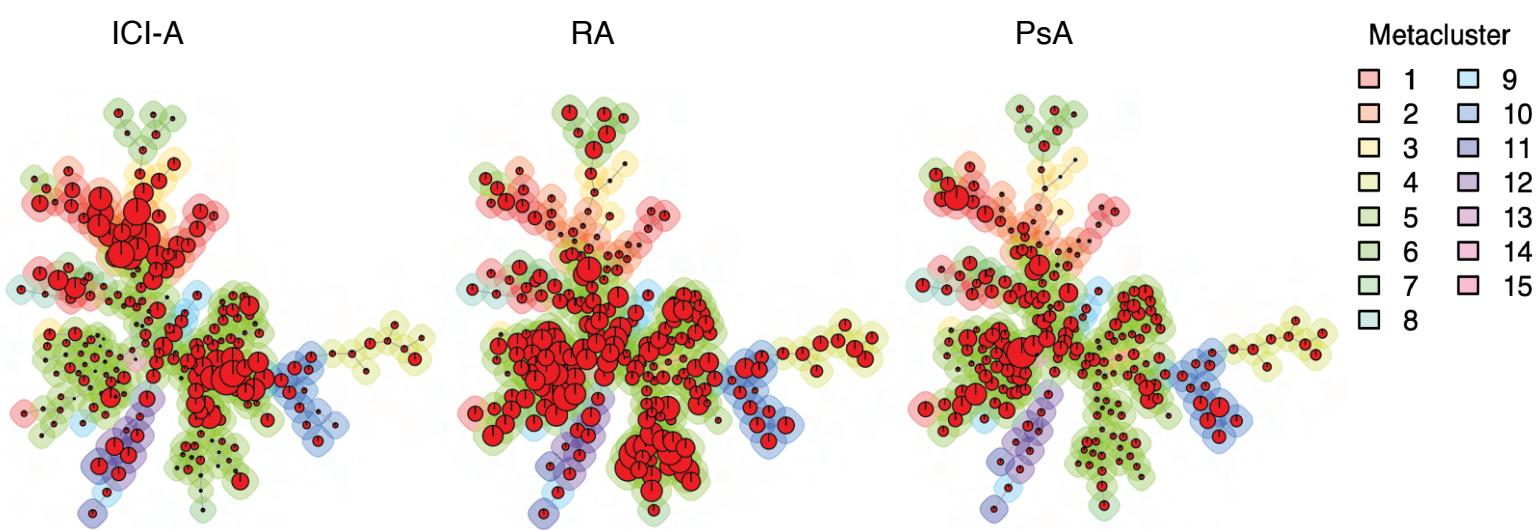
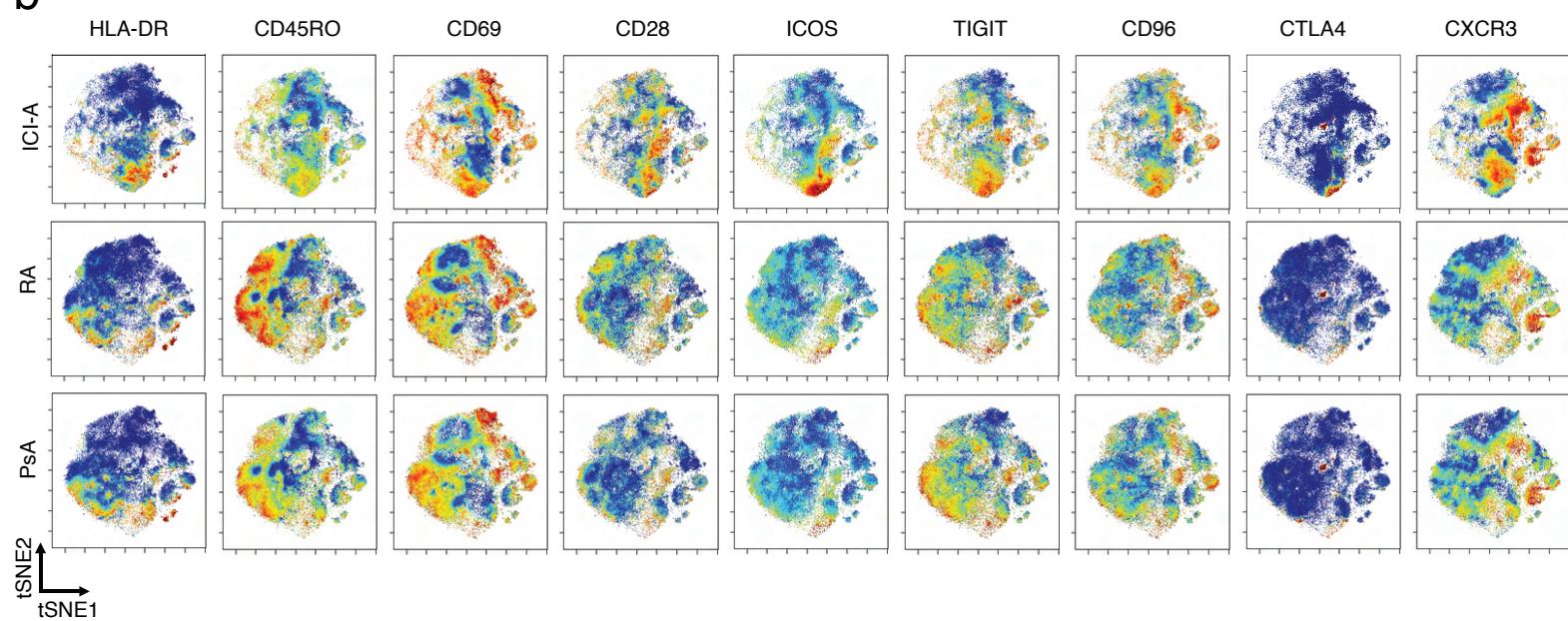
Supplementary figure 4. Single-cell analysis of CD8 T cells from peripheral blood of ICI-arthritis patients.

a) Gating scheme of naïve and non-naïve CD8 T cell isolation from ICI-arthritis blood for scRNA-seq and TCR repertoire analysis. **b)** Circle plot of clonotypes and shared clonotypes between synovial fluid CD8 cells and naïve and non-naïve blood CD8 cells after 10 weeks for P23. **c)** UMAP representation of clonotypes shared between paired synovial fluid CD8 cells and blood naïve and non-naïve CD8 cells in P25. MAIT cells were excluded in the analysis (dark gray dots). Bar graphs show the number of cells contained in the shared clonotypes and the number of shared clonotypes in each cluster. Cluster IDs for figure (c): 0 - GZM+, 1 - Transitional, 2 - Tcm, 3 - MAIT, 4 - ZNF683+, 5 - KLRG1+, 6 - Dysfunctional, and 7 - Proliferating .

a**b****c****d**

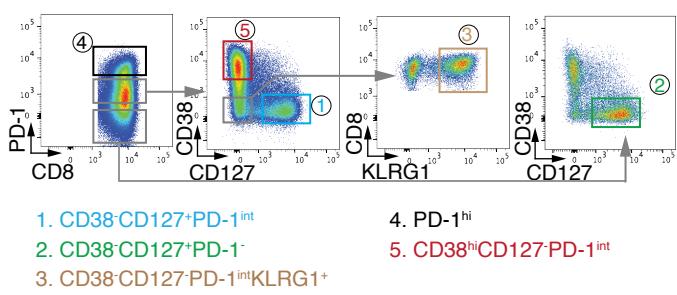
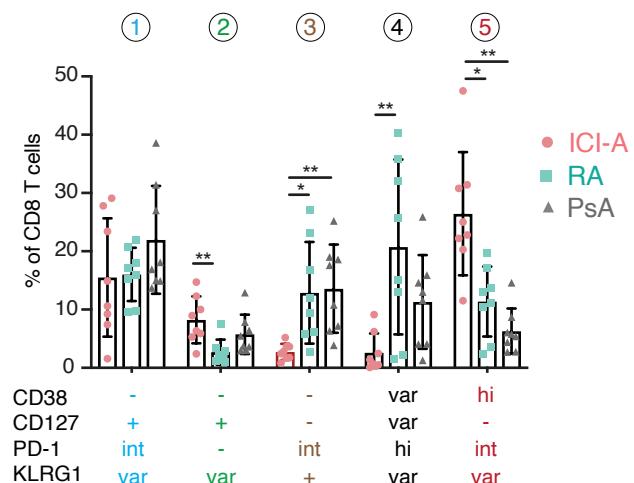
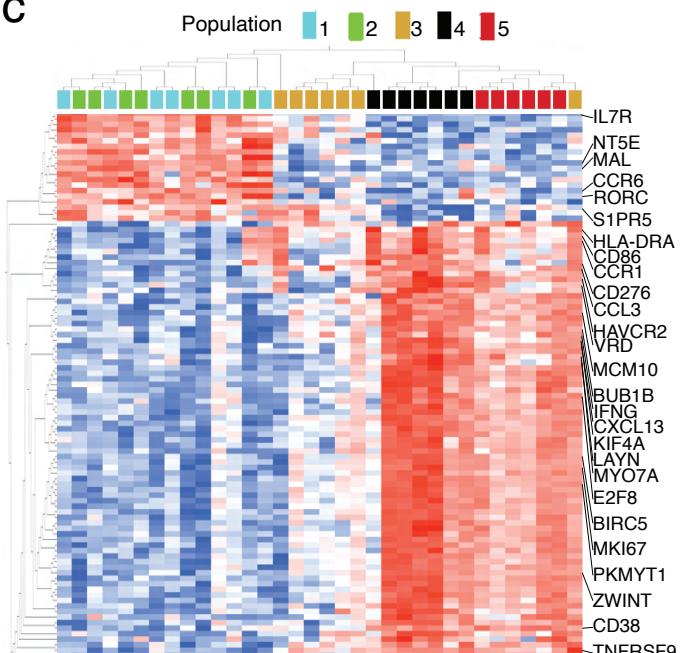
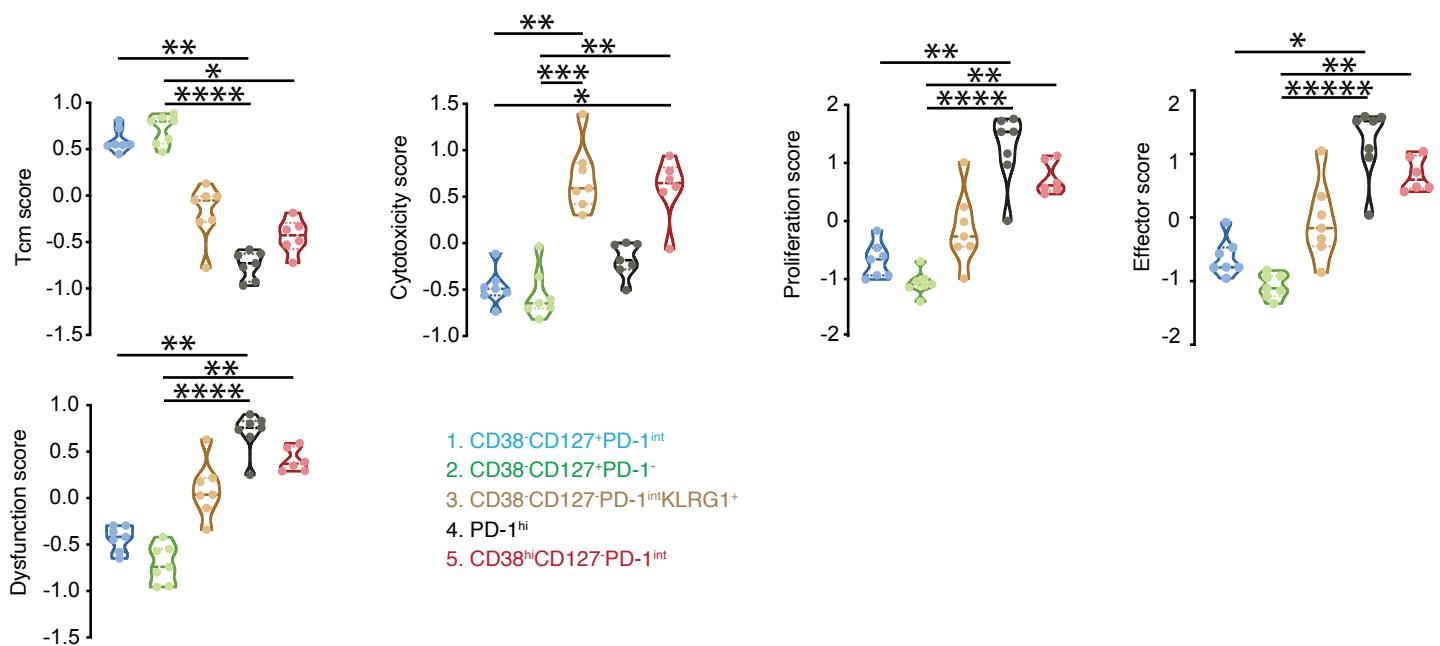
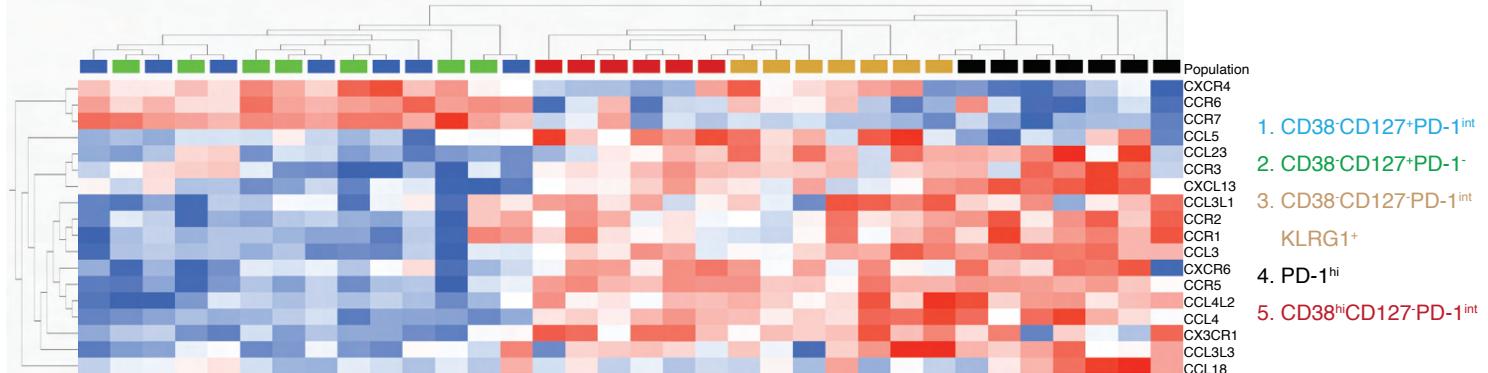
Supplementary figure 5. Basic analysis of mass cytometry of ICI-A, RA and PsA synovial fluid.

a) Flow cytometric detection of PD-1 on T cells using indicated detection antibody clones with or without prior incubation with pembrolizumab blockade *in vitro*. **b)** Example gating of mass cytometry data. **c)** Frequency of CD8 T cells, CD4 T cells, B cells, monocytes, and NK cells in ICI-A (n=6), RA (n=5) and PsA (n=5) synovial fluid. **d)** Example gating of high, intermediate and low PD-1 expression levels on T cells from ICI-A, RA and PsA synovial fluid detected by mass cytometry. Quantification of PD-1^{hi} cells, PD-1^{int} cells and PD-1^{hi}CXCR5⁻ CD4 Tph cells are shown. Mean ± SD shown. *p<0.05, **p<0.001 by Kruskal-Wallis test in (d).

a**b**

Supplementary figure 6. Unsupervised analysis of mass cytometry of ICI-A, RA and PsA synovial fluid.

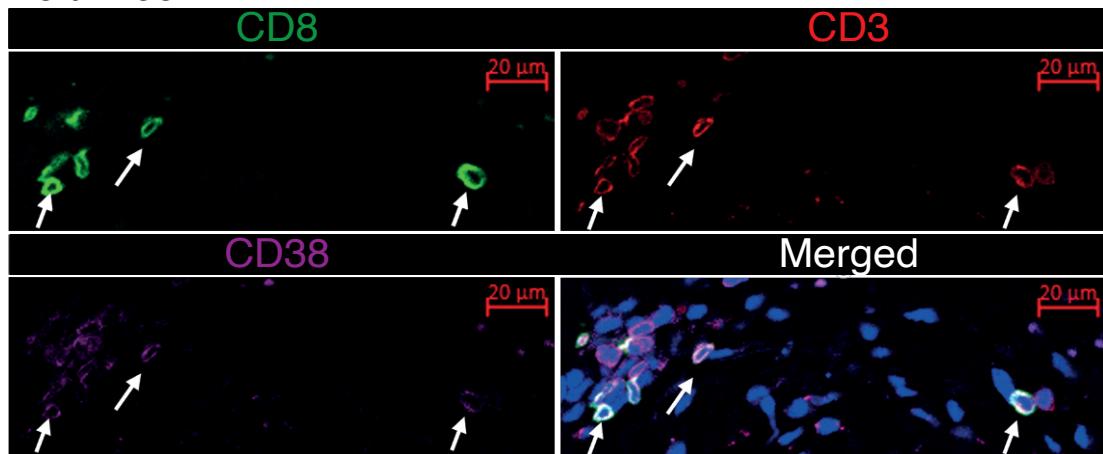
a) FlowSOM minimus spanning tree showing metaclusters on CD8 T cells from ICI-A, RA and PsA synovial fluid detected by mass cytometry. **b)** tSNE plots showing expression of indicated markers on CD8 T cells from ICI-A, RA and PsA synovial fluid detected by mass cytometry.

a**b****c****d****e**

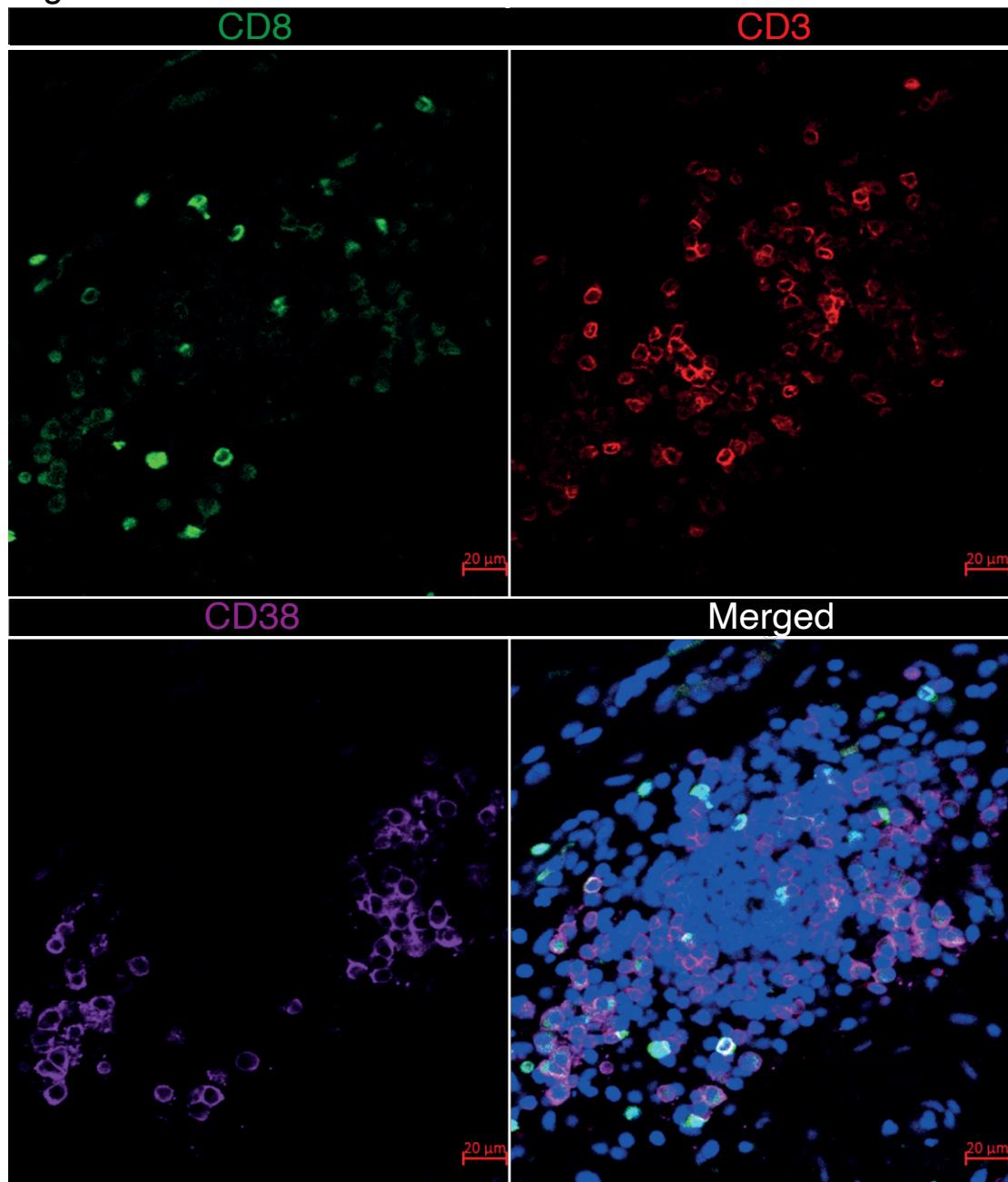
Supplementary figure 7. Transcriptional features of CD8 T cells in ICI-arthritis synovial fluid.

a) Sorting scheme to isolate five CD8 T cell populations with indicated surface phenotypes. **b)** Frequency of sorted CD8 T cell populations in ICI-arthritis (ICI-A), RA and PsA synovial fluid (n=8). **c)** Hierarchical clustering of CD8 T cell populations from ICI-A synovial fluid based on row-normalized mean expression of 100 top variable genes by RNA-seq. Colors indicate cell populations as in (a). **d)** Gene module scores of CD8 T cell populations as in (a) from ICI-A synovial fluid, calculated based on differentially expressed genes. **e)** Hierarchical clustering of CD8 T cell populations from ICI-A synovial fluid based on row-normalized mean expression of differentially expressed chemokines and chemokine receptors measured by RNA-seq. *p<0.05, **p<0.001, ***p<0.0001 by Kruskal-Wallis test in (b) and (d).

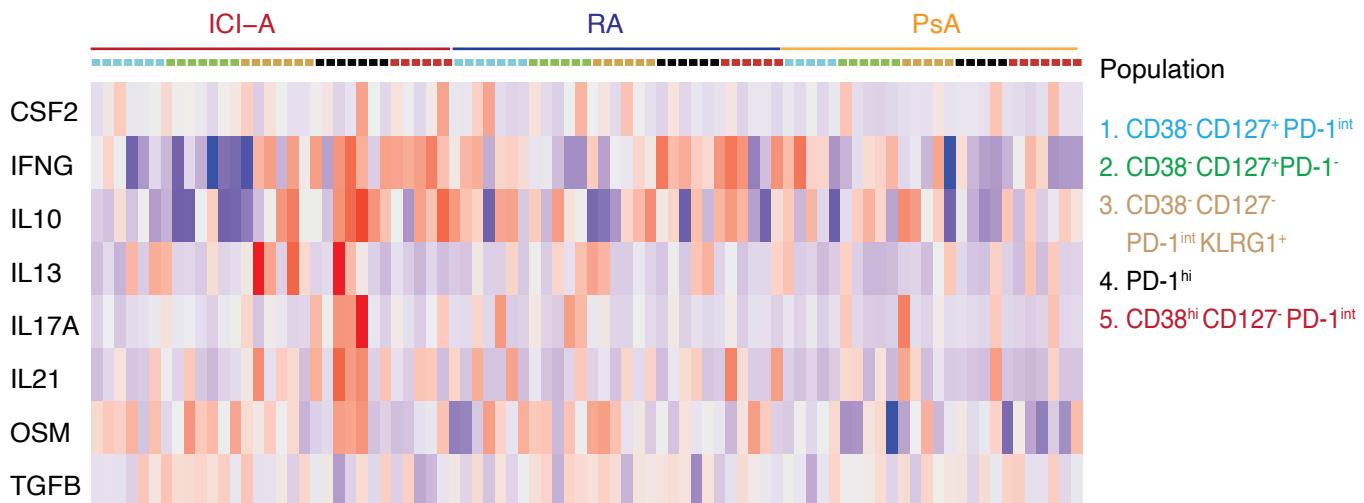
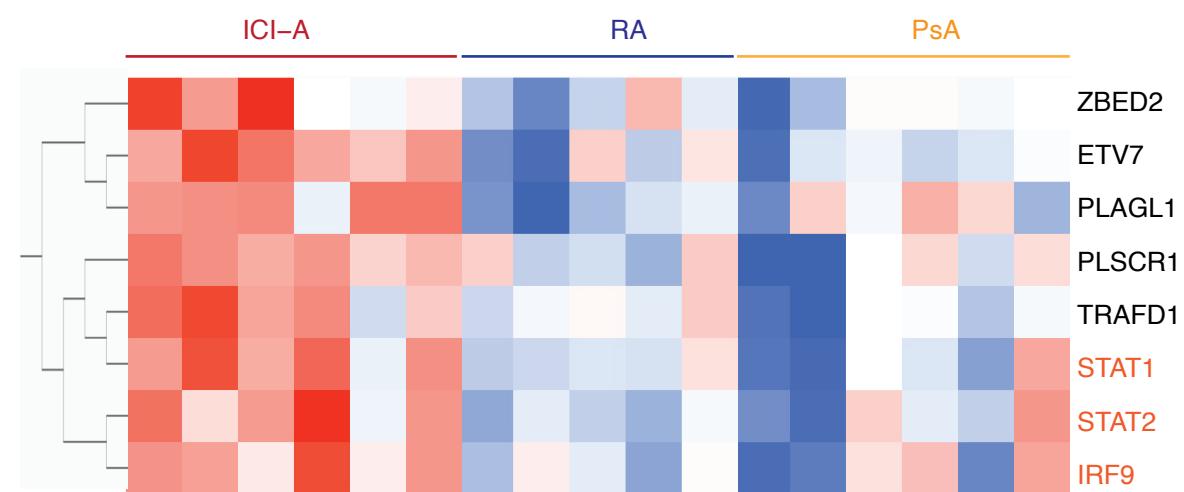
Left knee



Right knee

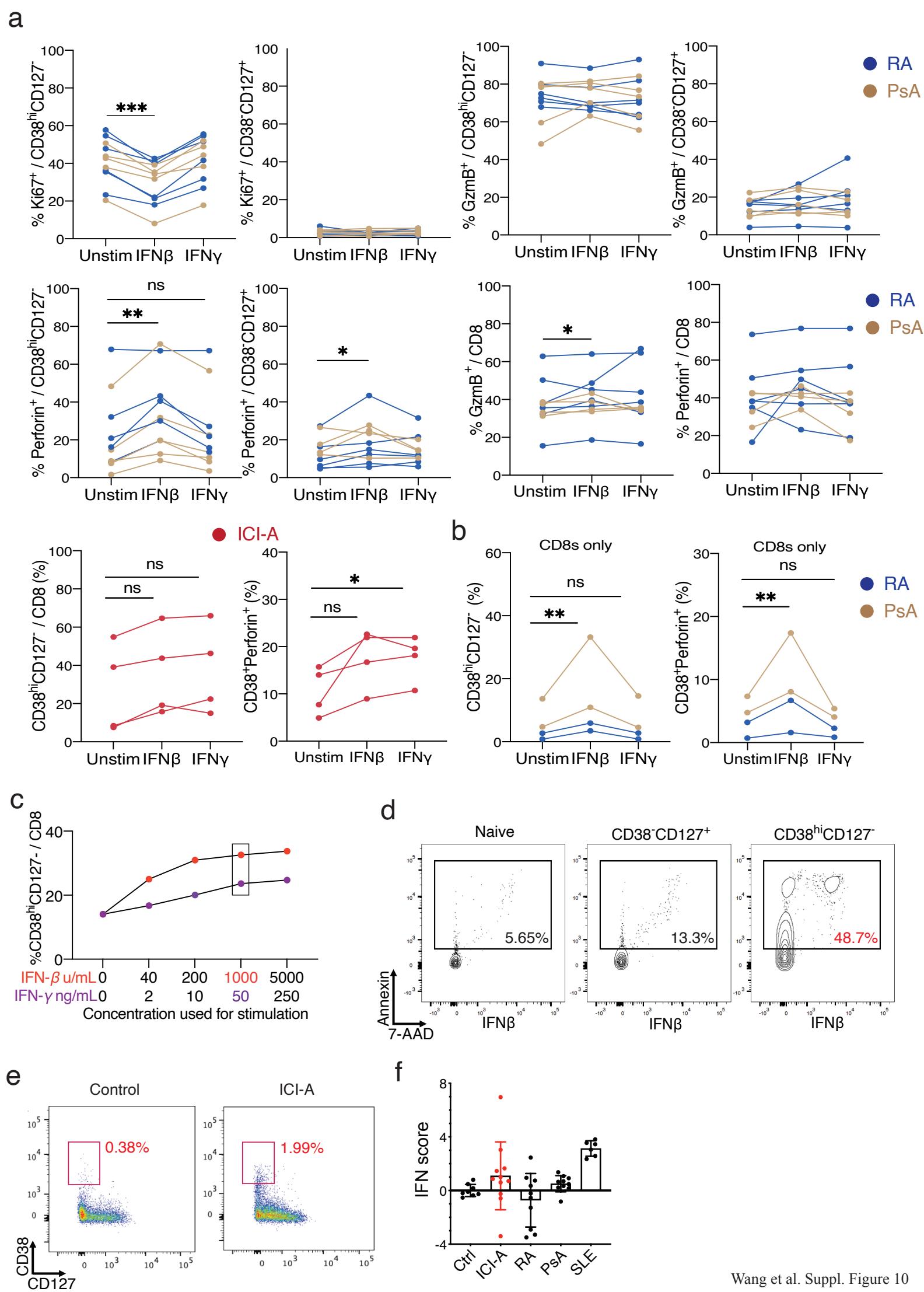


Supplementary figure 8. CD38 detection on CD8 T cells in synovium using multiplexed immunofluorescence.
Multiplexed immunofluorescence images showing CD38 protein detection on CD8 T cells (marked with white arrows) present in left and right knees of patient. CD8 staining is shown in green, CD3 in red and CD38 in magenta.

a**b**

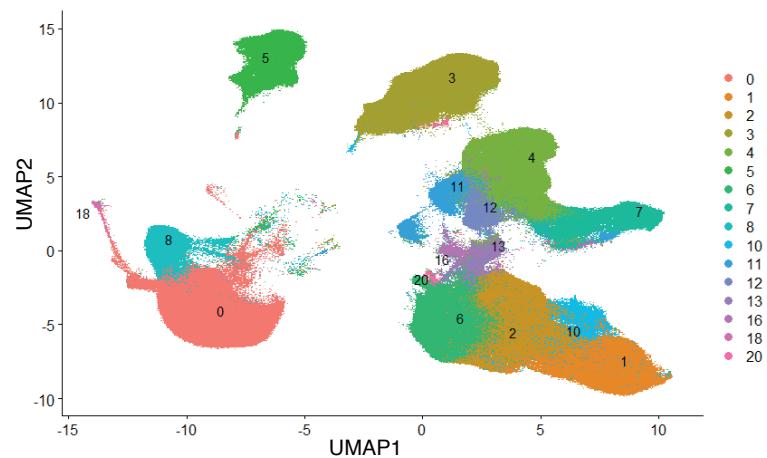
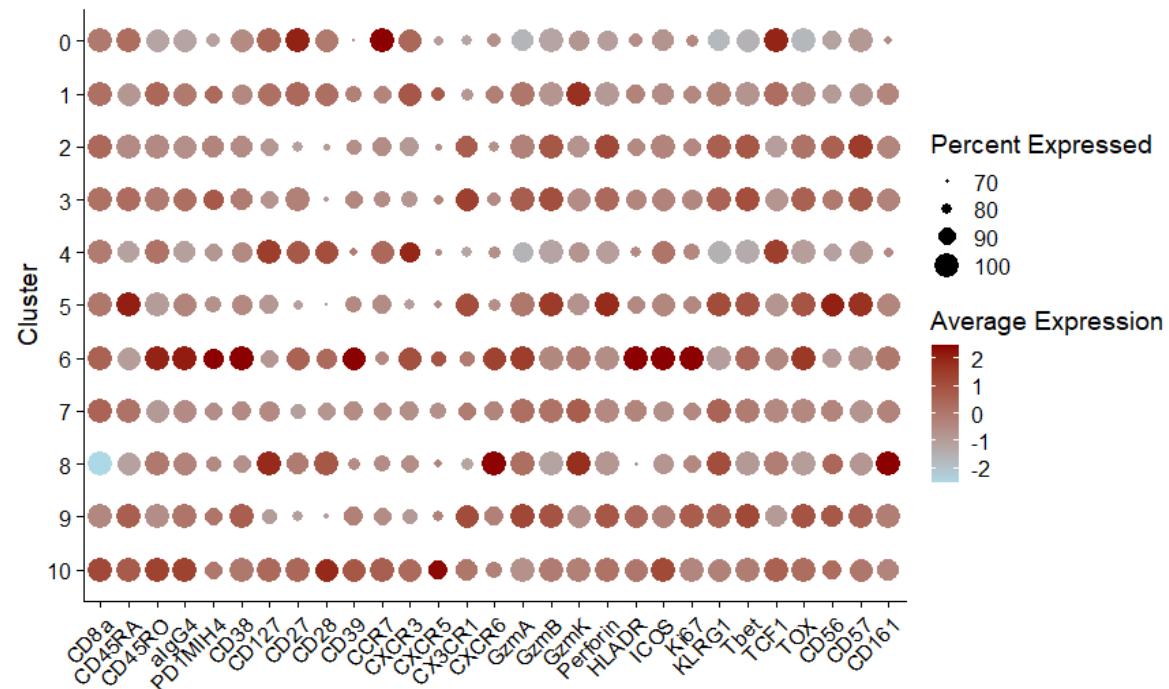
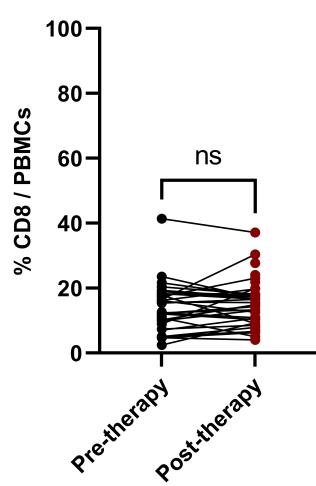
Supplementary figure 9. Distinct activation of IFN-inducible genes in ICI-arthritis.

a) Heatmap showing the expression of cytokine genes across the sorted populations ordered by diseases. **b)** Heatmap showing the expression of differentially transcription factor genes across the sorted populations ordered by diseases. Mean \pm SD shown.



Supplementary figure 10. Expression of intracellular markers with or without IFN treatment in SFMC and PBMC from various disease conditions.

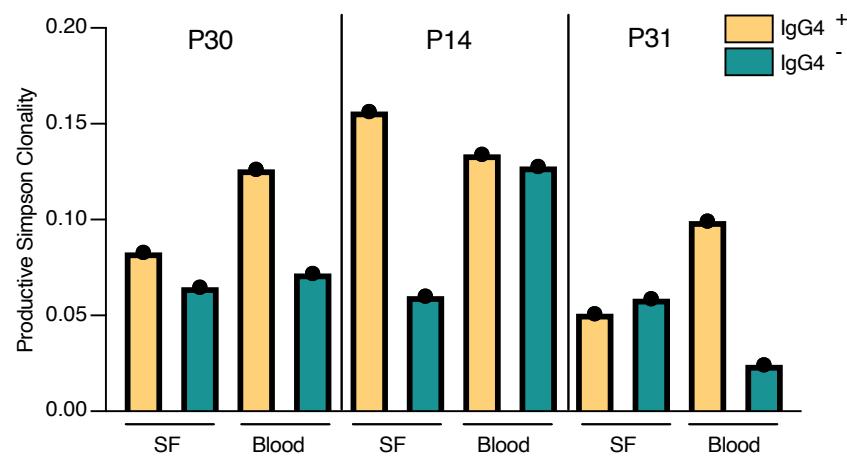
a) Frequency of Ki67⁺, granzyme B⁺, Perforin⁺ and cells in indicated CD8 T cell subsets from total SFMC (RA in blue, PsA in gold and ICI-A in red) cultured with IFN- β or IFN- γ for 72 hours. **b)** Frequency of CD38^{hi}CD127⁻ and CD38⁺Perforin⁺ cells in cultured CD8 T cells from SF (RA in blue, PsA in gold) with IFN- β or IFN- γ for 72 hours. Lines link the same patient sample under the different conditions in (a) and (b). **c)** Titration curve of cytokine concentrations for IFN stimulation experiments (IFN- β in red and IFN- γ in purple). Black box indicates concentration used in all experiments. **d)** Representative flow cytometry plots of Annexin versus 7-AAD gating on indicated PBMC-derived CD8 T cell subsets treated with IFN- β for cytotoxicity assays. See Fig. 4e. **e)** Representative flow cytometric plot of CD38 and CD127 expression on gated CD8 T cells from PBMC from healthy control and ICI-arthritis (ICI-A) patients. **f)** 8 gene-derived IFN score of PBMC from controls (n=8), ICI-A (n=11), RA (n=10), PsA (n=10), and SLE (n=6) patients. *p<0.05, **p<0.001, ***p<0.0001 by Kruskal-Wallis test in (a) and (b).

a**b****c**

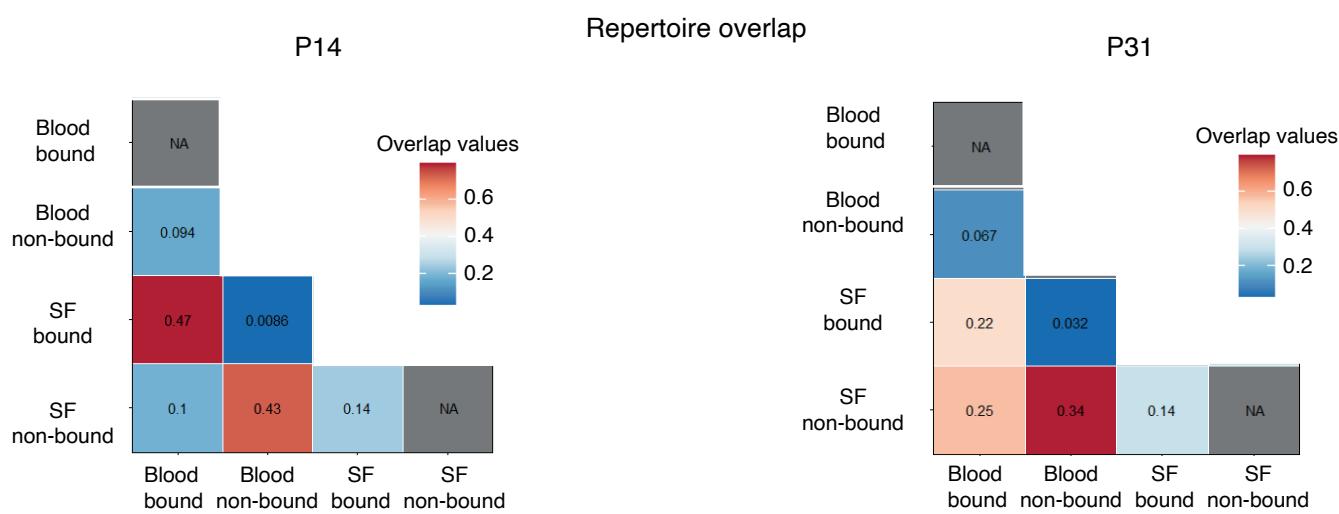
Supplementary figure 11. Features of CD8 T cells following ICI treatment.

a) UMAP visualization of total PBMCs analysed by CyTOF. **b)** Dotplot depicting expression of listed markers in CD8 T cells divided by the clusters visualized in Figure 5a. **c)** Frequency of CD8 T cells among total PBMCs pre and post initiation of therapy. Black lines connect paired patients before and 6-weeks post initiation of therapy. ns = not significant by Kruskal Wallis test.

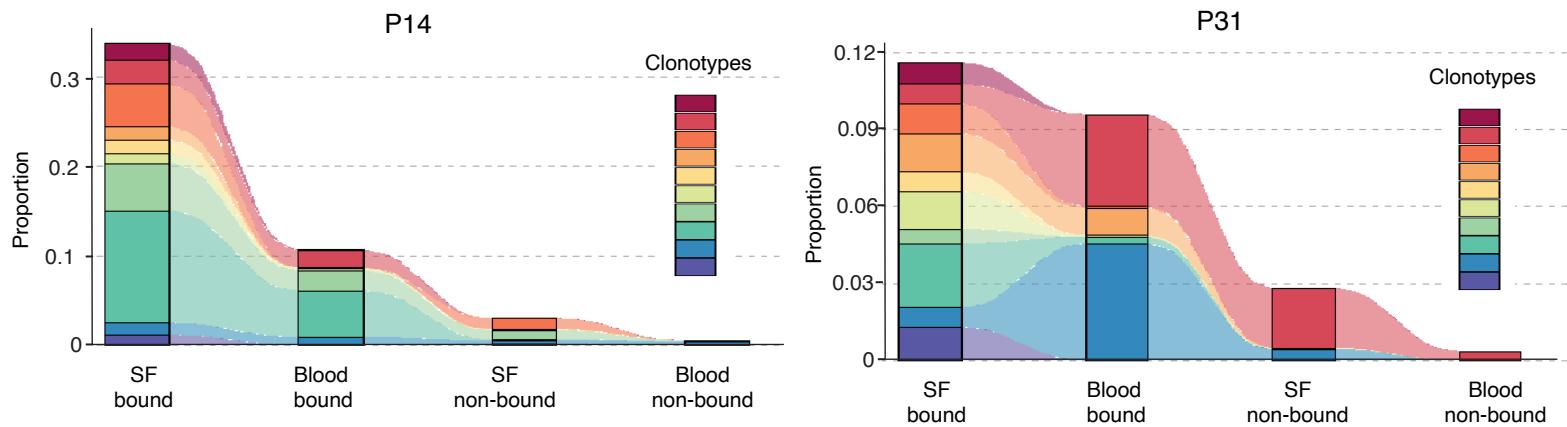
a



b



c



Supplementary figure 12. TCR repertoire overlap in drug-bound synovial fluid and blood CD8 T cells.

a) Productive Simpson clonality of all 3 patients in both blood and synovial fluid (SF). Values closer to zero are more polyclonal. b) Plot denoting the Morisita index value for each comparison demonstrating the repertoire overlap. Each plot depicts 1 patient each. See Fig. 5h. c) Alluvial plot demonstrating the location and overlap of the top 10 most abundant TCR clones in anti-PD-1 bound and not bound blood and synovial fluid. Each plot depicts 1 patient each.