Supplementary Materials for

Multimodal identification of rare potent effector CD8 T cells in solid tumors

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Materials and Methods

Mice: All mice were treated in accordance with the regulatory standards of the National Institutes of Health and American Association of Laboratory Animal Care and were approved by the UCSF Institution of Animal Care and Use Committee. Cd69-TFP-CreER^{T2} (denoted as Cd69-TFP) mice in the C57BL6/J background were custom-generated from Biocytogen Inc. and then maintained heterozygous (bred to C57BL6/J wild type mice) at the UCSF Animal Barrier facility under specific pathogen-free conditions. C57BL6/J (wild type; WT), C57BL6/J CD45.1 (B6.SJL-Ptprc^{a} Pepc^{b}/BoyJ), OT-I (C57BL/6-Tg(TcraTcrb)1100Mjb/J) mice were purchased for use from Jackson Laboratories and maintained in the same facility in the C57BL6/J background. For adoptive transfer experiments, CD45.1^{het}; OT-I^{het}; Cd69-TFP^{het} (denoted simply as CD45.1; OT1; Cd69-TFP) mice were used. Mice of either sex ranging in age from 6 to 14 weeks were used for experimentation. For experiments using the transgenic PyMTchOVA strain(*21*), mammary tumor-bearing females in the age range of 15 to 24 weeks were used. Adoptive transfer of T cells in these mice were done when mice developed at least 2 palpable tumors (> 25-30mm^2).

Mouse tumor digestion and flow cytometry: Tumors from mice were processed to generate single cell suspensions as described previously(*34*). Briefly, tumors were isolated and mechanically minced on ice using razor blades, followed by enzymatic digestion with 200 µg/mL DNAse (Sigma-Aldrich), 100U/mL Collagenase I (Worthington Biochemical) and 500U/mL Collagenase IV (Worthington Biochemical) for 30 min at 37°C while shaking. Digestion was quenched by adding excess 1X PBS, filtered through a 100µm mesh, spun down and red blood cells were removed by incubating with RBC lysis buffer (155 mM NH_4Cl, 12 mM $NaHCO_3$, 0.1 mM EDTA) at room temperature for 10 mins. The lysis was quenched with excess 1X PBS, spun down and resuspended in FACS buffer (2mM EDTA + 1% FCS in 1X PBS) to obtain single cell suspensions. Similarly, tumor draining lymph nodes (dLN) were isolated and mashed over 100µm filters in PBS to generate single cell suspensions.

For each sample, 2.5-3 million cells/sample were stained in a total of 50µL of antibody mixture for flow cytometry. Cells were washed with PBS prior to staining with Zombie NIR Fixable live/dead dye (1:500) (Biolegend) for 20 min at 4°C. Cells were washed in FACS buffer followed by surface staining for 30 min at 4°C with directly conjugated antibodies diluted in FACS buffer containing 1:100 anti-CD16/32 (Fc block; BioXCell) to block non-specific binding. Antibody dilutions ranged from 1:100-1:400, optimized separately. After surface staining, cells were washed again with FACS buffer. For intracellular staining, cells were fixed for 20 min at 4°C using the IC Fixation Buffer (BD Biosciences) and washed in permeabilization buffer from the FoxP3 Fix/Perm Kit (BD Biosciences). Antibodies against intracellular targets were diluted in permeabilization buffer containing 1:100 Fc Block and cells were incubated for 30 min at 4°C followed by another wash prior to readout on a BD LSRII or Fortessa Cytometer.

Processing and flow cytometry analysis of other mouse organs: To phenotype T cells under from lymphoid organs homeostasis, spleen and inguinal, mesenteric and brachial lymph nodes were isolated and mashed over 100µm filters washed with 1X PBS to generate single cell suspension of lymphocytes. For splenic suspensions, RBC lysis was performed as described above before staining for flow cytometry.

To profile thymocytes, thymus was isolated, cut into small pieces with a razor blade and minced by using gentleMACS dissociator (Miltenyi Biotec) in RPMI. Next, the mixture was spun down and resuspended in the digestion mixture described above and allowed to digest with shaking at 37°C for 20 mins, following which, the remaining tissue was either minced again using the gentleMACS dissociator and/or directly mashed over a 100µm filter in FACS buffer to generate a single cell suspension, ready to be processed for staining and flow cytometry.

Skin digestion was done as previously described(*35*). Briefly, mice are shaved and depilated prior to removal of dorsal skin. The skin was then rid of fat, minced with scissors and razor blade in the presence of 1 ml of digest media (2 mg/ml collagenase IV (Roche), 1 mg/ml hyaluronidase

(Worthington), 0.1 mg/ml DNase I (Roche) in RPMI-1640 (GIBCO). The minced skin was then moved to a 50 ml conical with 5 ml additional digest solution and incubated at 37°C for 45 min with shaking and intermittent vortexing before being washed and passed through a 70µm strainer prior to staining. TFP high vs. low gates were drawn by using a side-by-side WT control or using endogenous CD8 T cells in the context of adoptive transfer into a tumor-bearing mouse.

Tumor injections and adoptive transfer of CD8 T cells into tumors: The B78chOVA and MC38chOVA cancer cell lines, as previously described(*14, 34*), were generated by incorporating the same mcherry-OVA construct used to establish the PyMTchOVA spontaneous mouse line(*21*). For tumor injections, the corresponding cells were grown to near confluency (cultured in DMEM with 10% FCS (Benchmark) and 1% PSG (Gibco)) and harvested using 0.05% Trypsin-EDTA (Gibco) and washed 3x with PBS (Gibco). The number of cells to be injected per mouse was resuspended in PBS and mixed in a 1:1 ratio with Growth Factor Reduced Matrigel (Corning) to a final volume of 50µL per injection. The mixture was injected subcutaneously into the flanks of anesthetized and shaved mice. Tumors were allowed to grow for 14–21 days unless otherwise noted, before tumors and tumor-draining lymph nodes were harvested for analysis. CD8 T cells were isolated from CD45.1;OT-1;Cd69-TFP mice using the EasySep Negative Selection Kit (Stem Cell Bio), resuspended in 1X PBS at 10X concentration 100µL was injected into each tumor-bearing mice. For B78chOVA and PyMTchOVA tumors, 1 million and for MC38chOVA tumors, 200,000 CD8 T cells were injected retro-orbitally into each mouse either 5d (B78chOVA), 7d (MC38chOVA) post tumor injection or when mice had at least 2 palpable tumors (PyMTchOVA). Tumor measurements were done by measuring the longest dimension (length) and approximately perpendicular dimension (width) using digital calipers, rounded to one decimal place each.

Contralateral tumor injection and vaccination: 5 days post B78chOVA tumor injection, equal numbers (1 million) CD8 T cells from a CD45.1;OT-I;Cd69TFP and P14;Cd69TFP mice were injected retroorbitally into each mice. Next day, gp33-41 subcutaneous peptide (Anaspec)

vaccination was injected contralaterally to the tumor, with 50µg peptide + 50µL Common Freund's Adjuvant (CFA, Sigma) along with 50µL PBS for a total volume of 100µL. The vaccination site was identified by a white, hardened subcutaneous mass and isolated and processed similarly to the tumor for flow cytometry.

In vitro stimulation of naïve CD8 T cells: CD8 T cells were isolated from Cd69-TFP or WT mice as described above and plated in a 96 well round bottom plate (Corning) at 80,000 cells/well in T cell media-RPMI (Gibco) + 10% FCS (Benchmark) + Penicillin/Streptomycin + Glutamine (Gibco). TCR stimulation was induced by adding anti-CD3/CD28 Dynabeads (Applied Biosystems) at the concentration of 2µL per 80,000 cells (1:1 ratio of cells:beads), the plate was briefly spun down to bring cells and beads together before incubation at 37°C for varying lengths of time. 55µM β-mercaptoethanol (BME; Gibco) was added to the T cell media during stimulation. For repeated stimulation assays, 2 wells of each sample at every time point were pooled for mRNA isolation and qRT-PCR, while 2 other wells were used as duplicates for flow cytometry. After each cycle, beads taken off each well and replated for resting in T cell media containing 10 U/mL of Interleukin-2 (IL-2; Peprotech). To restart each stimulation cycle, cells from each biological replicate were pooled, counted and Dynabeads were added at the appropriate concentration for a 1:1 ratio and redistributed into wells for incubation.

Sorting and qPCR, resting or restimulation of homeostatic CD8 T cells: To sort sufficient CD8 T cells from homeostatic lymphoid organs, CD8 T cells were first isolated from spleens and inguinal, brachial, mesenteric lymph nodes Cd69-TFP or WT mice using the EasySep Negative selection kit. These cells were then sorted on TFP^{hi} (top 15%), TFP^{mid} (middle 30%) and TFP^{lo} (bottom 15%) from each mouse separately and rested in T cell media containing 10 U/mL Interleukin-7 in a 96 well round bottom plate and assayed at 0, 24 and 48h. Likewise, for qPCR analysis of populations high, mid and low for TFP, these populations were sorted into cold T cell media, pelleted and subjected to RNA extraction and qPCR with primers for Cd69 and 18s rRNA

as the reference gene. For the sort and restimulation experiment, Memory (CD44+CD62L+) TFP^{hi} cells and Effector (CD44+CD62L-) TFP^{lo} cells were sorted and incubated in T cell media + 55μM BME containing 1:1 anti CD3/CD28 Dynabeads in a 96 well round bottom plate with either 5μg/mL Actinomycin D (Sigma) in DMSO or DMSO alone (vehicle) for 3h, before profiling by flow cytometry. De novo CD69 surface expression was measured by the difference of CD69 MFI between the vehicle and Actinomycin D treated groups.

Restimulation and cytokine production of intratumoral CD8 T cells: OT-I T cells from B78chOVA tumors were sorted on a BD FACSAria Fusion or BD FACSAria2 (BD Biosciences) at d11-d13 post adoptive transfer of CD8 T cells from CD45.1; OT-I; Cd69-TFP mice, as described above. To prepare CD45-enriched fractions(*36*), tumors were digested as described above into single cell suspensions, centrifuged and resuspended in 30mL room temperature (RT) RPMI 1640. Then, 10mL Ficoll-Premium 1.084 (Cytiva) was carefully underlaid and the tubes centrifuged at 1025g for 20 mins at RT without braking. The resulting interface-localized cells were pipetted out, diluted in equal volume RPMI and centrifuged at 650g for 5 mins to collect the cells. This constituted a CD45-enriched fraction which was then processed for staining and FACS. The four CD69:TFP quadrants were sorted from each tumor sample (cells from 2-3 tumor samples were pooled for a single biological replicate) into serum-coated microcentrifuge tubes containing cold T cell media. These were subsequently plated in a 96-well V-bottom plate either in T cell media or T cell media containing PMA (50 ng/mL; Sigma-Aldrich), Ionomycin (500ng/mL; Invitrogen) + Brefeldin A (3μg/mL; Sigma-Aldrich) and BME (Gibco) for 3h, before cells were collected for surface and intracellular staining for cytokines and granzyme B.

Long-term ex vivo tumor slice overlay: Tumor slice overlay cultures were adapted, modified and extended from previous work(*22*). For tumor slice overlay cultures, B78chOVA tumors were injected bilateral subcutaneously into the flank of anesthetized and shaved mice. Tumors were allowed to grow for 11 – 13 days. 96 hours prior to tumor harvest and slicing, CD8 T cells were

677 isolated from CD45.1;OT-1;Cd69-TFP mice, as described above. Isolated CD8 T cells were
678 activated via 1:1 culture with Dynabeads Mouse T-Activator CD3/CD28 (Invitrogen) in T cell
679 media + BME in 96 well U-bottom plates for 48 hours. After activation, T cells were removed from
680 Dynabeads rested in T cell media with supplemented 10 U/mL IL-2 (PeproTech) for 48 hours
681 before use. For gating TFP high vs. low cells, CD8 T cells from CD45.1; OT-I mice were subjected
682 to similar pre-treatment and profiled by flow cytometry side-by-side along with the CD45.1;OT-
683 1;Cd69-TFP CD8 T cells at d0.

684 For slicing, tumors were harvested and stored in cold RPMI until use. Each well of a 24 well plate
685 was pre-filled with cold RPMI and stored on ice. Tumors were embedded in 1.5-2% agarose gel,
686 allowed to solidify, and sliced at a thickness of 350 – 400μm using a Compresstome VF310-0Z
687 Vibrating Microtome (Precisionary). Slices were immediately stored in pre-filled 24 well plate on
688 ice until use.

689 For the slice overlay, each well of a 24 well plate was pre-coated with 30μL of 1 part culture
690 medium:4 parts Matrigel and allowed to solidify at 37˚C. Tumor slices were removed from RPMI
691 and excess agarose was trimmed from slice edges (leaving a thin halo of agarose around slices
692 to use for handling). Slices were spread across solidified Matrigel bed in 24 well plates. Rested T
693 cells were stained with Violet Proliferation Dye 450 (BD Biosciences) diluted 1:1000 in PBS at 10
694 x 10^6 cells/mL for 15 minutes at 37˚C. Cells were washed 2x with PBS and resuspended in T cell
695 culture medium at 150 – 200 x 10^6 cells/mL. 5μL cell suspension (0.5 – 1 x 10^6 cells) was added
696 directly on top of each slice and incubated at 37˚C for 3 hours, with 5μL fresh media added to
697 each slice every 30 minutes to prevent slices from drying out. After incubation, 30μL non-diluted
698 Matrigel was added directly atop each slice and allowed to solidify at 37˚C. 2mL T cell culture
699 medium containing BME was added to each well. 1mL culture medium was removed and replaced
700 with fresh medium every 24 hours throughout the experiment.

Single cell RNA Sequencing and Analysis: Adoptively transferred CD45.1; OT-I; Cd69-TFP CD8 T cells were sorted from B78chOVA tumors d12 post transfer into four populations based on the CD69:TFP quadrants (Q1: TFP+/CD69-, Q2: TFP+/CD69+, Q3: TFP-/CD69-, and Q4: TFP-/CD69+). Sorted cells were separately labeled with lipid and cholesterol-modified oligonucleotides (LMO's) according to McGinnis et. al(*37*). Following 2 washes with PBS + 0.1% BSA, cells were pooled for encapsulation in one lane of a 10X 3' v3 kit with a target cell number of 18,000.

Following construction of the GEX library (according to manufacturer's instructions) and the LMO library(*37*), libraries were pooled at a 10:1 molar ratio for sequencing on the NovaSeq 6000. This resulted in 807M cDNA reads and 163M LMO reads. Transcript and LMO reads were counted using the CellRanger count function against the GRCm38 reference genome to generate feature barcode matrices. These matrices were loaded into Seurat and filtered to remove high mitochondrial % cells (> 15%) and cells with low nGene (< 200 genes). Cells were then demultiplexed using their LMO counts with cells having too few LMO nUMI or ambiguous identity (possible multiplets) filtered out using the demultiplex package(*37*). The resulting object had an average cDNA nUMI per cell of 7662 reads and average nGene per cell of 2115 genes and average LMO nUMI per cell of 1080 reads. The final object underwent scaling and then scoring for cell cycle signatures (S and G2M scores as computed using Seurat's built-in CellCycleScoring function. The object then underwent regression for cell cycle effects (S and G2M score as described in the Seurat vignette) and percent mitochondrial reads before PCA. K-Means clustering and UMAP dimensional reduction was then performed on the first 16 PC's.

Established subpopulations of exhausted T cells were marked by expression of canonical genes such as Stem-like or Progenitor (T_{EX}^{Prog}; *Tcf7, Ccr7, Jun*) (*8, 12*), Early Effector-like ($T_{EX}^{E.Eff}$; *Hsp90aa1, Hsp90ab1, Npm1*) (*38*), Late Effector or KLR-gene-expressing effector-like ($T_{EX}^{KLR.\ Eff}$; *Klrd1, Zeb2*) (*8*), Memory T_{EX}^{Mem}. (*Cxcr3, Ly6c2, Itgb7)* (*39*) and Interferon-Stimulated T_{EX}^{ISG} (*Cxcl10, Isg15, Ifit1*) (**Fig. S8A**). The Intermediate ($T_{EX}^{Int.}$) and Terminal ($T_{EX}^{Term.}$) subsets were

distinguished by exhaustion-related genes *Ctla4, Pdcd1, Tox* and those related to actin organization and TCR signaling such as *Tmsb4x, Coro1a, Actg1, Ccl5 and S100a6(8).* Additionally, two cell cycle gene-dominated clusters termed T_{EX}^{Cyc1} T_{EX}^{Cyc2} were identified (**Fig. 3B, Fig. S8A**).

Cytotoxic and exhaustion scores were generated by calculating the average expression of ensemble gene lists for each of the phenotypes—Exhaustion: *Ctla4, Pdcd1, Cd38, Entpd1, Tox*; Cytotoxic: *Prf1, Gzmc, Tnfrsf9, Ifng, Klrd1*.

qRT-PCR: At designated time points, CD8 T cells were isolated from the 96 well culture plates, or CD8 T cells were sorted into T cell media and centrifuged. The supernatant was aspirated out and the pellets stored at -80˚C until mRNA extraction using the RNEasy Micro Kit (Qiagen). Corresponding cDNA was synthesized from the mRNA samples using the cDNA amplification kit (Applied Biosystems). qPCR using pre-designed *Cd69* and *18s* probes (Invitrogen) with a TaqMan-based assay system (BioRad) or custom-made primers (iDT Technologies) for *Jun* (Fwd: 5′ ACGACCTTCTACGACGATGC 3′, Rev: 5′ CCAGGTTCAAGGTCATGCTC 3′)(*40*) , *Stat5a* (Fwd: 5’ CGCTGGACTCCATGCTTCTC 3’, Rev: 5’ GACGTGGGCTCCTTACACTGA 3’)(*41*) and *18s* (Fwd: 5’ CTTAGAGGGACAAGTGGCG 3’, Rev: 5’ ACGCTGAGCCAGTCAGTGTA 3’)(*42*) using the SsoFast assay system (BioRad) was used to quantify transcripts in a BioRad CFX94 machine.

Human tumor samples: All tumor samples were collected with patient consent after surgical resection under a UCSF IRB approved protocol (UCSF IRB# 20-31740), as described previously(*29*). In brief, freshly resected samples transported in ice-cold DPBS or Leibovitz’s L-15 medium before digestion and processing to generate a single-cell suspension. The following cancer indications were included in the cohort: Bladder cancer (BLAD), colorectal cancer (CRC), glioblastoma multiforme (GBM), gynecological cancers (GYN), hepatocellular cancers (HEP), head and neck cancer (HNSC), kidney cancer (KID), lung cancer (LUNG), melanoma (MEL),

pancreatic ductal adenocarcinoma (PDAC), pancreatic neuroendocrine tumors (PNET), sarcoma (SRC).

Transcriptomic analysis of human tumors: All tumor samples were collected under the UCSF Immunoprofiler project as described(*29*). Briefly, tumor samples were thoroughly minced with surgical scissors and transferred to GentleMACs C Tubes containing 800 U/ml Collagenase IV and 0.1 mg/ml DNase I in L-15/2% FCS per 0.3 g tissue. GentleMACs C Tubes were then installed onto the GentleMACs Octo Dissociator (Miltenyi Biotec) and incubated for 20 min (lymph node) or 35 min (tumor) according to the manufacturer's instructions. Samples were then quenched with 15 mL of sort buffer (PBS/2% FCS/2mM EDTA), filtered through 100µm filters and spun down. Red blood cell lysis was performed with 175 mM ammonium chloride, if needed. Freshly digested tumor samples were sorted by FACS into conventional T cell, Treg, Myeloid, tumor and in some cases, stromal compartments and bulk RNA-seq was performed on sorted cell fractions. mRNA was isolated from sorted fractions and libraries were prepared using Illumina Nextera XT DNA Library Prep kit. The libraries were sequenced using 100bp paired end sequencing on HiSeq4000. The sequencing reads we aligned to the Ensembl GRCh38.85 transcriptome build using STAR(*43*) and gene expression was computed using RSEM(*44*). Sequencing quality was evaluated by in-house the EHK score, where each sample was assigned a score of 0 through 10 based on the number of EHK genes that were expressed above a precalculated minimum threshold. The threshold was learned from our data by examining the expression distributions of EHK genes and validated using the corresponding distributions in TCGA. A score of 10 represented the highest quality data where 10 out of 10 EHK genes are expressed above the minimum threshold. The samples used for survival analysis and other gene expression analyses had an EHK score of greater than 7 to ensure data quality. Ensemble gene signatures scores

were calculated by converting the expression of each gene in the signature to a percentile rank among all genes and then determining the mean rank of all the genes in the signature.

Reanalysis of published datasets: Available, curated RNA-Seq data (*5, 7, 14*) on *Cd69* and upstream transcription factor expression was plotted directly without modification. A curated R object derived from Zheng et al.(*30*) generously shared by Dr. Miguel Reina-Campos, UCSD, was used for analysis in Fig. 5. Ensemble gene signatures were scored as mentioned above and plotted onto pre-existing UMAP dimensional reduction and already annotated cell clusters. While exhaustion and T^*_{EFF} genes were obtained from previously published work (*29*) and this study respectively, *TCF7, SELL, LEF1, CCR7, IL7R* genes were used for the Naïve score.

In vitro Killing Assay: MC38chOVA tumors with adoptively transferred Cd69-TFP-OT-I CD8 T cells in WT B6 mice were harvested at d8 post T cell transfer, digested as mentioned above, and sorted by CD69: TFP quadrants into cold T cell media. Sorted cells were centrifuged, resuspended in fresh, warm T cell media with BME and added onto MC38chOVA cells plated ~24h prior in flat-bottom 96 well plates. To each well containing 5000 MC38chOVA plated 24h prior, 5000 sorted T cells were added. As with the sort and restimulation experiments, each such collection of 5000 cells from a particular quadrant was pooled from 2-3 tumors and treated as a single biological replicate. Each experiment involved 7-8 tumors to obtain at least 3 biological replicates. Technical replicates were included and averaged wherever possible, i.e., at least 10,000 cells were sorted from a given quadrant and biological replicate. Percentage killing was obtained by measuring the fractional loss of live cells at 36h in no T cell vs. T cell added conditions relative to 0h. Live cell numbers from each condition was accurately measured by lightly detaching

cancer cells with trypsin and scoring against CountBright (ThermoFisher) absolute counting beads on a flow cytometer.

CITE-Seq analysis of human tumors: For CITE-Seq, post tumor digestion, cells were incubated with Human TruStain FcX Receptor Blocking Solution to prevent non-specific antibody binding before staining with Zombie Aqua Fixable Viability Dye and anti-human CD45 antibody in PBS/2%FCS/2mM EDTA/0.01% sodium azide and incubated for 25 min on ice in the dark. Live $CD45^+$ and CD45- cells were sorted on a BD FACSAria Fusion. $CD45^+$ and $CD45^-$ cells were pelleted and resuspended at 1x10^3 cells/ml in 0.04%BSA/PBS buffer before mixing in an 8:2 CD45+:CD45- ratio and loaded onto the Chromium Controller (10X Genomics) to generate 5′ v1.1 gel beads-in-emulsions (GEM). Pooled 8:2 CD45+:CD45- cells were resuspended in Cell Staining Buffer (BioLegend) and stained with a pool of 137 TotalSeq-C antibodies (Table) according to the manufacturer's protocol before loading onto the Chromium Controller (10X Genomics) for GEM generation. The cDNA libraries were generated using all or a subset of Chromium Next GEM Single Cell 5′ Library Kit for gene expression (GEX), Chromium Single Cell V(D)J Enrichment kit (10X Genomics) for T cell receptor (TCR), and Chromium Single Cell 5′ Feature Barcode Library kit for antibody derived tag (ADT) according to the manufacturer's instructions. The libraries were subsequently sequenced on a Novaseq S4 sequencer (Illumina) to generate fastqs with the following mean reads per cell: 42,000 (GEX), 34,000 (TCR), and 5,700 (ADT). For multimodal clustering and analysis, CLR normalization followed by weighted nearest neighbor (WNN) clustering was performed using the Seurat package in R. Naïve and Exhaustion scores were generated using the percentile rank method as mentioned above, but with protein (ADT) markers- Naïve : CD62L, CD45RA, IL7RA; Exhaustion: PD-1, CTLA-4, CD38, CD39.

Live 2-photon imaging of tumor slices and image analysis: Live imaging of tumor slices was performed on a custom-made 2-photon microscope as previously described(*1*). Briefly, 1 million CD2dsRed; OT-I; Cd69-TFP or control CD2dsRed; OT-I CD8 T cells were retro-orbitally injected into WT mice bearing MC38chOVA tumors injected 5-7d earlier and harvested 7-10d after T cell injection. Slices for imaging were generated as described above for the ex vivo slice culture assay. Slices were placed in a custom-made perfusion chamber and imaged under oxygenated and temperature-controlled perfusion of RPMI 1640, as described previously(*1*). Dual laser excitations at 825nm and 920nm were used to excite the requisite fluorophores. Image analysis was performed on Imaris (BitPlane) with custom-made plugins developed on Matlab (Mathworks) and Fiji. Surfaces were generated on CD8 T cells and in both CD2dsRed; OT-I and CD2dsRed; OT-I; Cd69-TFP bearing slices and the corresponding levels of the former in the 515-545nm range PMT were used to gate on TFP^{hi} vs. TFP^{lo} OT-Is.

Cell tracking was performed on Imaris and corresponding cell positions imported to Matlab for further analysis to fit the persistent random walk model (PRWM) to the cell trajectories(*45*) using the method of overlapping intervals (*46*). Briefly, the mean squared displacement (MSD) for a cell for given time interval t_i was obtained from the average of all squared displacements x_{ik} such that

$$\bar{x_i} = \frac{1}{n_i}\sum_{k=1}^{n_i} x_{ik} \quad (1)$$

and

$$n_i = N - i + 1 \quad (2)$$

where n_i is the number of overlapping time intervals of duration t_i and N the total number of time intervals for the experiment. Mathematically, the persistent random walk model can be written as

$$MSD(t) = n_d S^2 P[t - P\left(1 - e^{-\frac{t}{P}}\right)] \quad (3)$$

where S is the migration speed and P is the persistence time. The motility coefficient is given as

$$\mu = S^2 P \quad (4)$$

where n_d is the dimensionality of the random walk (in this case n_d=3). We fitted the PRWM in 3D to obtain estimates of speed, persistence time and motility of each cell track by non-linear regression.

Statistical Analysis: Statistical analysis was done in GraphPad Prism or in R. For testing null hypothesis between two groups, either Student's t tests and or the non-parametric Mann-Whitney U tests were used, depending on the number and distribution of data points. Likewise, for testing null hypotheses among 3 or more groups, ANOVA or non-parametric tests were performed, followed by post-hoc Holm-Sidak's test, correcting for multiple comparisons. Unless otherwise mentioned, data are representative of at least 2 independent experiments.

Supplementary Figures:

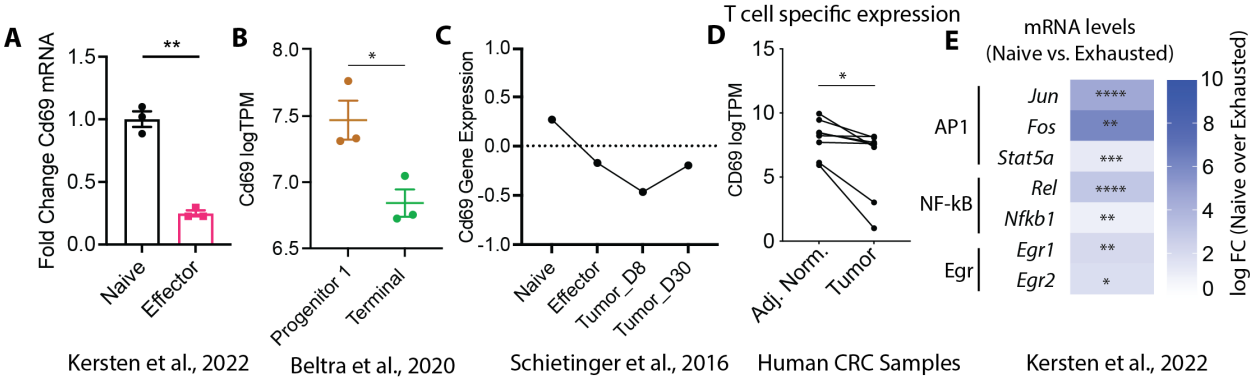


Fig. S1: **Resting Cd69 mRNA decreases with T cell differentiation towards exhaustion.** Cd69 mRNA expression in **(A)** Naïve vs. in vitro generated (stimulation with Dynabeads followed by rest in IL-2 containing media) effector CD8 T cells from published RNASeq data(*14*); **(B)** Progenitor 1 and Terminally exhausted T cell subsets from published data (*7*), **(C)** among Naïve, Effector, D8 tumor and D30 tumor infiltrating T cells from other published data (*5*), **(D)** in conventional T cells sorted from tumor and adjacent normal regions of human colorectal cancer patients; **(E)** depressed mRNA expression of factors associated with the Cd69 transcription in Naïve vs. Exhausted CD8 T cells from previous work (*14*) (symbols indicate FDR adjusted p-values). Plots show mean +/- SEM (A, B) p-values obtained by unpaired (A, B) and paired Student's t test (D).

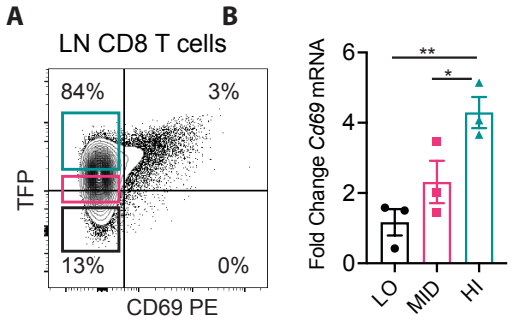


Fig. S2: Cd69-TFP reporter reads out Cd69 transcription. **(A)** TFP vs. CD69 in homeostatic lymph node (LN) CD8 T cells with percentage of cells in each quadrant and **(B)** corresponding *Cd69* mRNA by qPCR from color-coded sorted subpopulations.

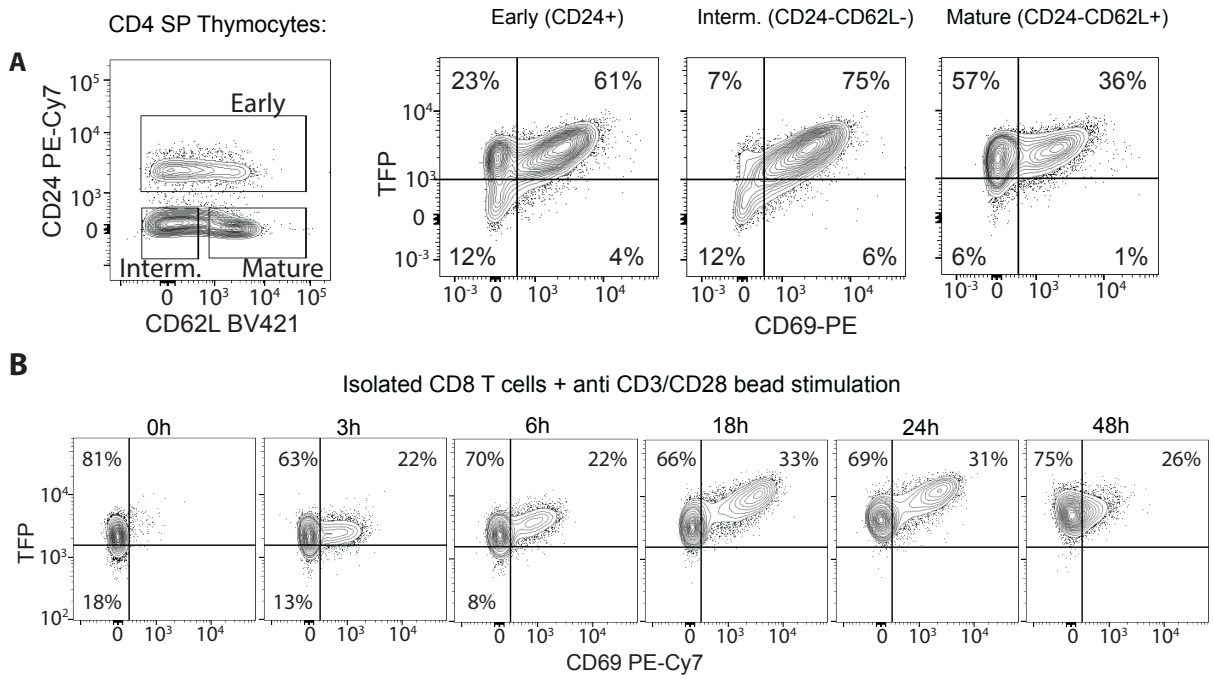


Fig. S3: TFP is upregulated along with CD69 in known contexts of TCR stimulation. (A) Representative flow cytometry plot of CD62L and CD24 expression in CD4+CD8- (CD4 single positive or SP) thymocytes to demarcate early, intermediate (interm.) and mature subsets with corresponding plots of TFP vs. CD69 in these subsets with varying degree of maturity during positive selection; **(B)** Representative flow cytometry plots of TFP vs. CD69 of isolated CD8 T cells from a naïve reporter mouse at different time points post stimulation with αCD3+αCD28 Dynabeads;

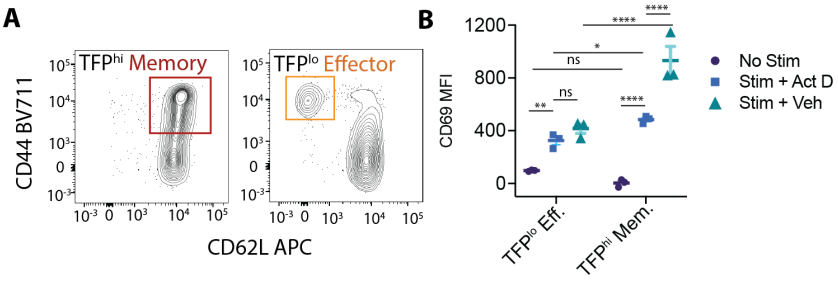
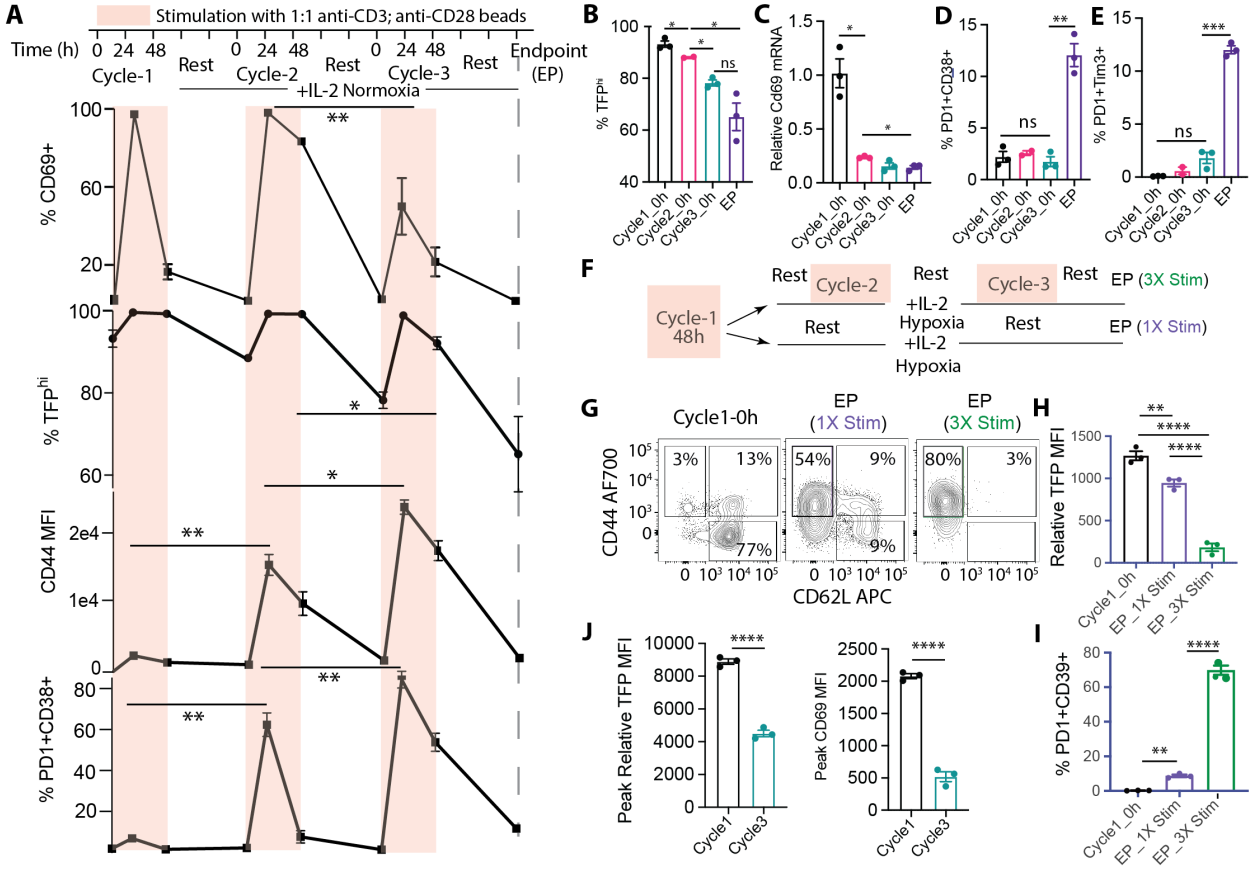


Fig. S4: Sensitivity to current stimulation is dependent on initial TFP level. (A) Typical phenotypic profile shown by flow cytometry plots of CD44 vs. CD62L from TFP^{lo} (bottom 20%) and TFP^{hi} (top 20%) splenic CD8 T cells, sorted TFP^{lo} Effector and TFP^{hi} Memory cells are shown by the corresponding color-coded gates matched with Fig. 1E; **(B)** CD69 MFI of the same sorted cells without stimulation (No Stim), 3h αCD3+αCD28 stimulation + DMSO (3h Stim + Vehicle) or 5μg/mL Actinomycin D (3h Stim + Act.D); (data representative of one out of at least 2 independent

901 experiments, each with 3 mice, bar graphs show mean +/- SEM, null hypothesis testing by
902 unpaired t test, adjusted for multiple comparisons).



903

904 **Fig. S5**: **Repeated TCR stimulation drives down TFP with acquisition of exhaustion markers**
905 **(A)** %CD69+, %TFP^{hi}, CD44 MFI, %PD1^{+}CD38^{+} of freshly isolated CD8 T cells through
906 successive cycles of 48h stimulation and 72h resting in ambient oxygen (normoxia) + IL-2;
907 **(B)** %TFP^{hi}, **(C)** *Cd69* mRNA by qPCR and **(D)** %PD1^{+}CD38^{+}, **(E)** %PD1^{+}Tim-3^{+} at the beginning
908 of cycles 1, 2, 3 and endpoint (EP); **(F)** experimental schematic showing 1X Stim vs. 3X Stim
909 conditions to parse the role of stimulation vs. IL-2 alone; **(G)** flow cytometry plots showing
910 representative CD44 vs. CD62L profiles of CD8 T cells at the timepoints and conditions indicated;
911 for the same experiment, **(H)** TFP (relative to WT control), **(I)** %PD1^{+}CD39^{+} of CD8 T cells at the
912 starting point (Cycle1_0h) and at endpoint (EP) either with 1X Stim followed by prolonged rest or
913 3X stim; **(J)** Peak Relative TFP and CD69 MFI between Cycle 1 and Cycle 3 in normoxia; (bar
914 graphs represent mean +/- SEM; null hypothesis testing by ANOVA followed by post-hoc Holm-
915 Šídák test; data representative of 2 independent experiments, each with 3 mice and technical
916 duplicates/biological replicate at every assay point).

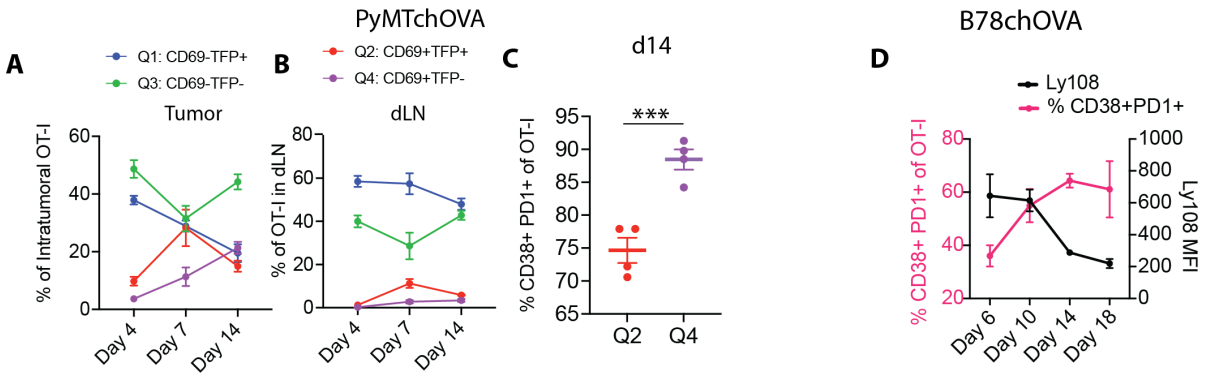


Fig. S6: Q4, as opposed to Q2 phenotype dominates terminally exhausted OT-Is in tumors

(A) TFP:CD69 quadrant distribution of OT-I T cells from a PyMTchOVA tumor or its corresponding **(B)** dLN at different time points post injection into tumor-bearing mice; **(C)** %$CD38^{+}PD1^{+}$ terminally exhausted cells among activated d14 intratumoral OT-Is belonging to TFP^{hi} Q2 and TFP^{lo} Q4 from PyMTchOVA tumors; **(D)** %CD38+PD1+ and Ly108 profiles over time (d6-d18) for all intratumoral OT-Is in B78chOVA. Null hypothesis testing by paired t test, bar graphs represent mean +/- SEM; data representative of 2 independent experiments, each with 2-3 mice for PyMTchOVA (each PyMTchOVA mice produced more than one tumor) and >=3 mice for B78chOVA per timepoint.

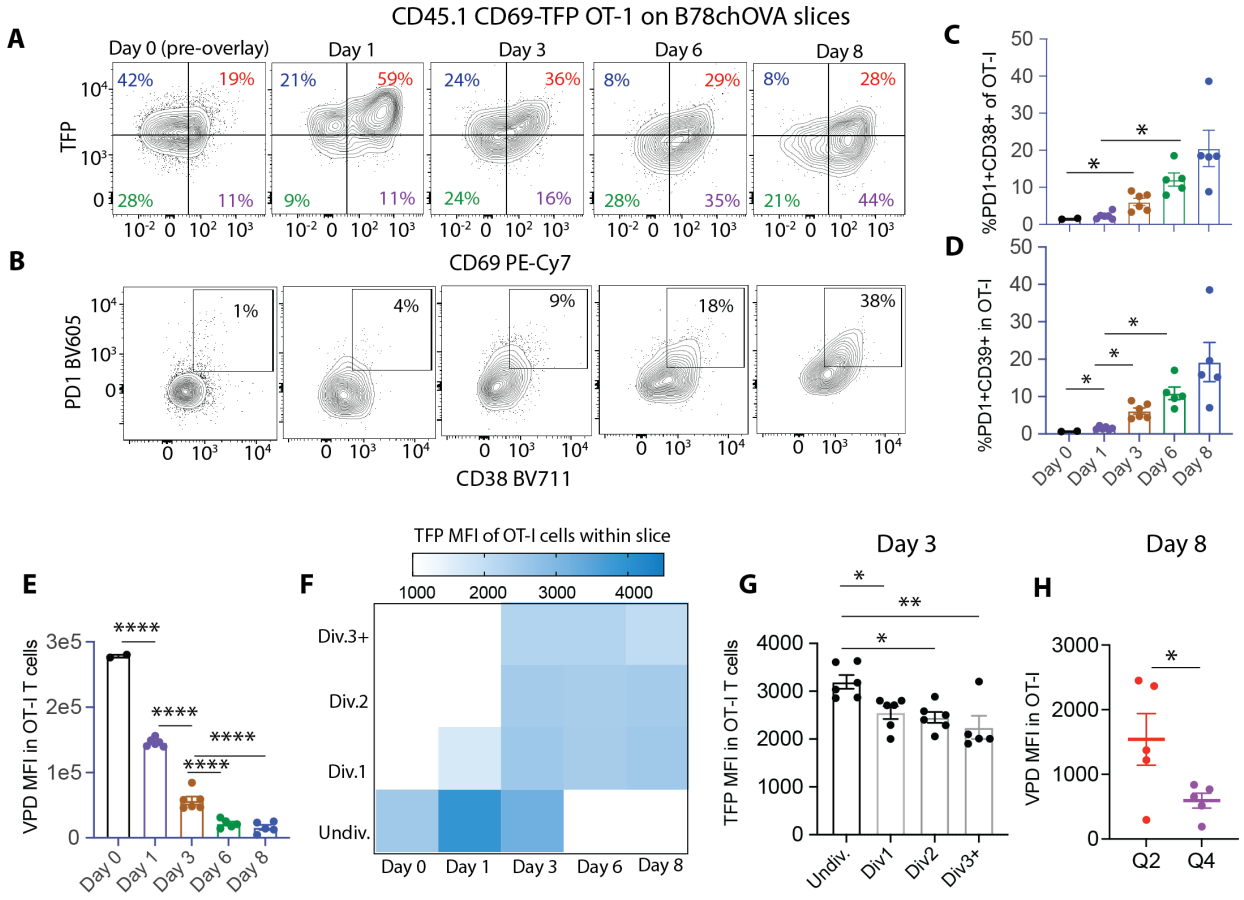
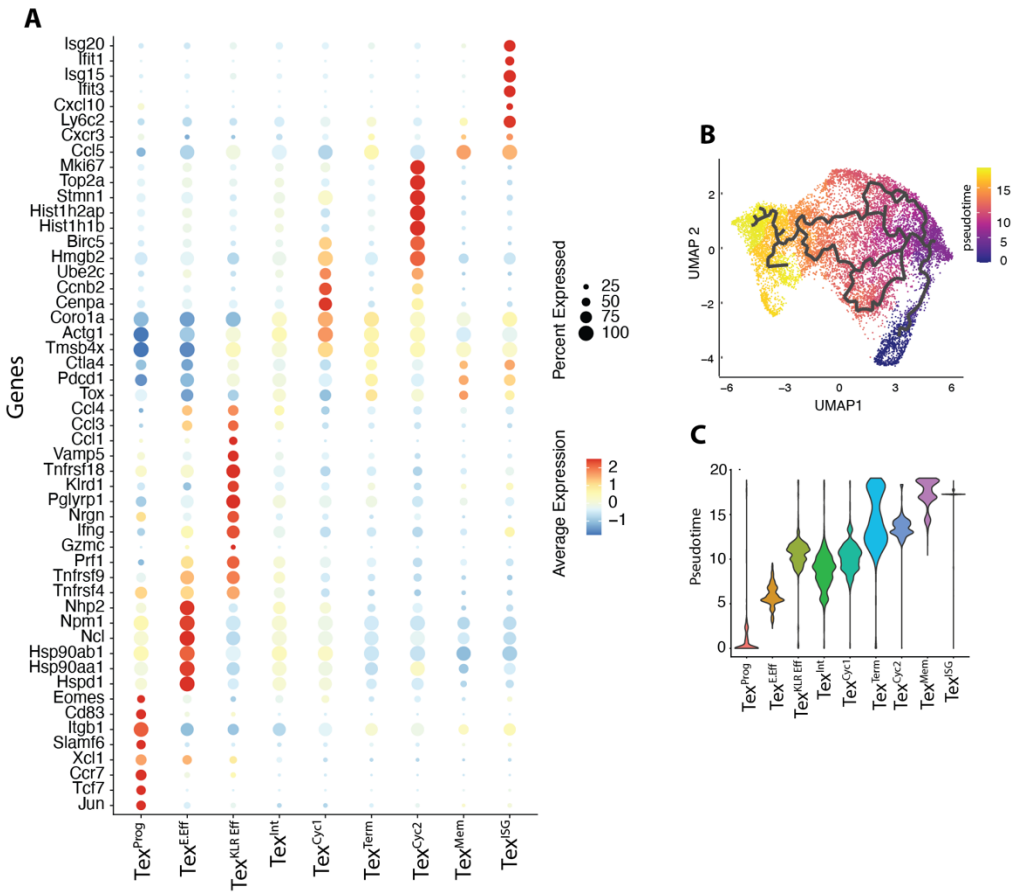


Fig. S7: Ex vivo tumor slice overlay culture mimics CD69:TFP dynamics in vivo (A) Representative flow cytometry plots of **(E)** TFP vs. CD69 and **(F)** PD1 vs. CD38 expression in slice-internal OT-I T cell from Day1-Day8, compared to Day0 (pre-overlay); **(B)** %PD1+CD38+, **(C)** %PD1+CD39+ and **(E)** Violet proliferation dye (VPD) MFI of Day 0 pre-overlay and slice-internal OT-I T cells at different time points after slice overlay (Day1-Day8); **(F)** heatmap of average TFP MFI of OT-I CD8 T cells at Day 0 pre-overlay and derived from slice culture from Day 1-Day 8 grouped by estimated number of divisions (>=3, 2, 1, or divided) and **(G)** corresponding bar graph showing this quantification for Day 3; (H) VPD MFI of slice-internal OT-Is from Q2 and Q4 at Day8; Bar graphs and line plots show mean +/- SEM, null hypothesis testing by ANOVA and post hoc Holm-Šídák test, or paired t test in H; data are representative of 2 independent experiments, each 5-6 slices/time point for each slice experiment and Day 0 pre-overlay samples in duplicate; TFP gated on WT controls CD8 T cells.



941

942 **Fig. S8: Gene expression based clustering of intratumoral OT-Is (A)** Dotplot representation
943 of differentially expressed and canonical T_{EX}-associated genes across the computationally
944 derived cell clusters from the scSeq of intratumoral Cd69-TFP:OT-I CD8 T cells at d12 post
945 injection into B78chOVA tumor-bearing mice; **(B)** UMAP representation of the scSeq data color-
946 coded by pseudotime derived from Monocle3 trajectory analysis; **(C)** Pseudotime spread of each
947 cluster.

948

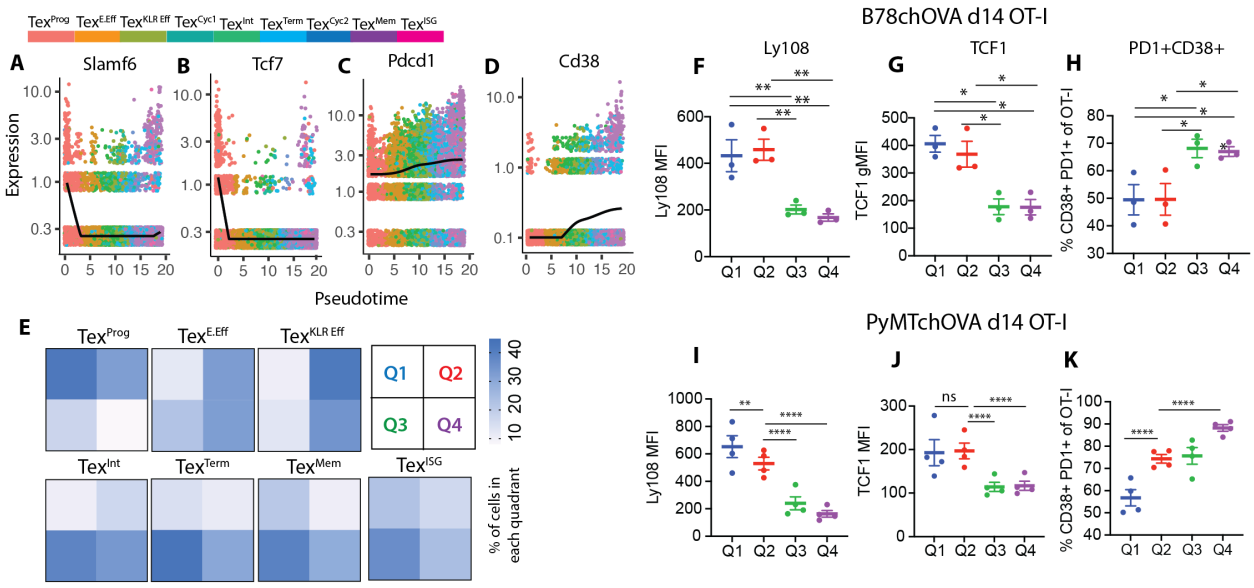
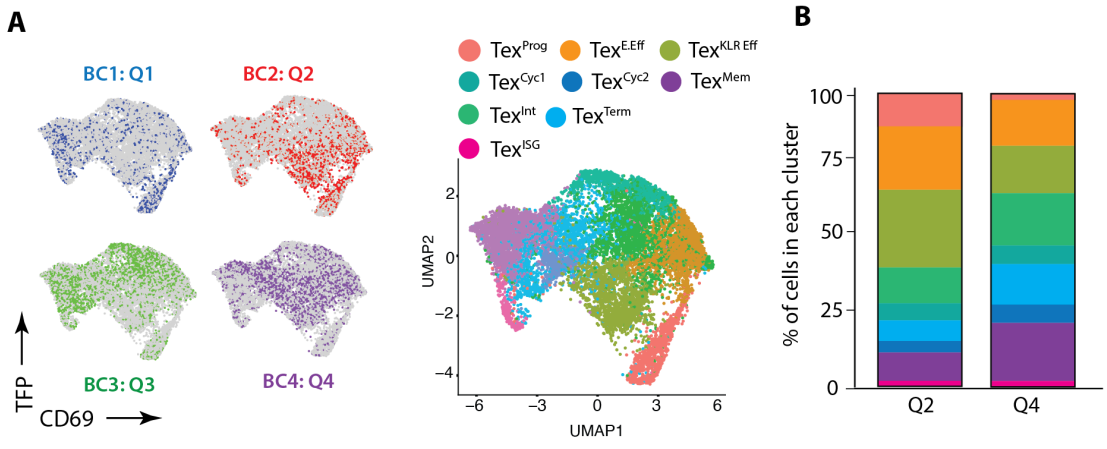
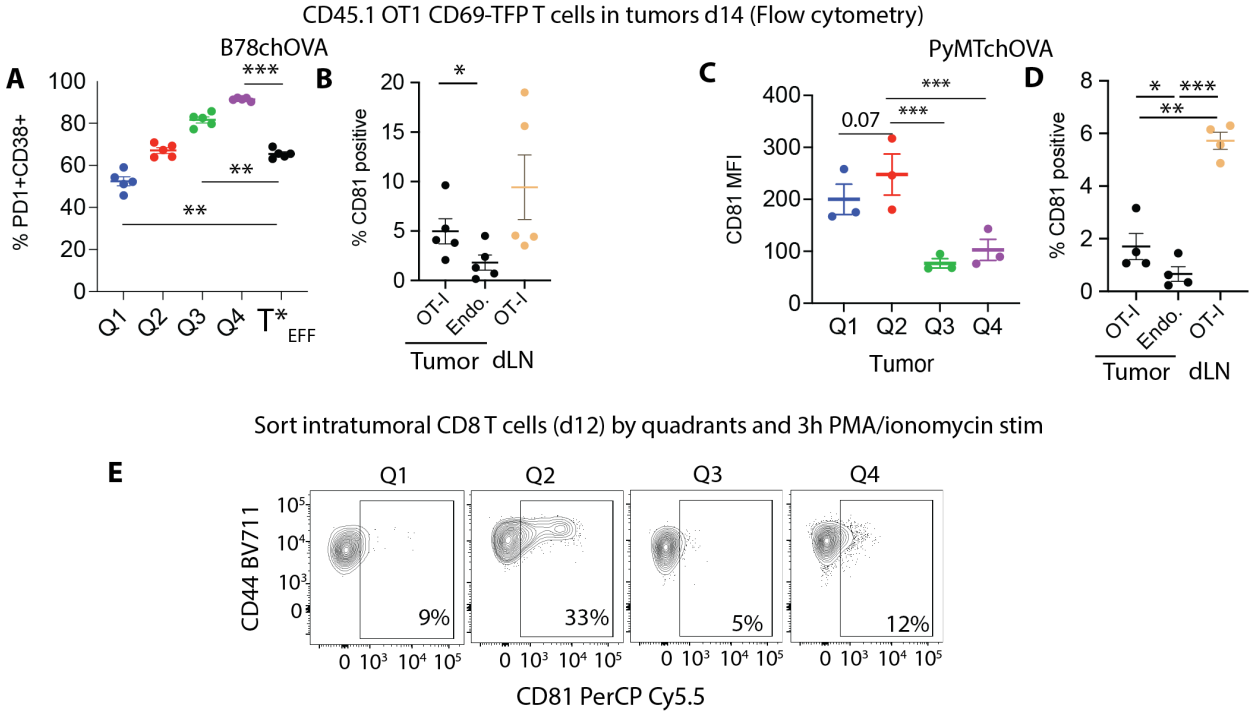


Fig. S9:Progenitor, intermediate and terminally exhausted CD8 T cells distribute distinctly among CD69:TFP quadrants (A-D) expression of select genes plotted against pseudotime and color-coded by clusters; best-fitting spline (degrees of freedom=5) to the gene expression pattern overlaid in black, random vertical jitter added to the plot for better visualization; **(E)** heatmaps of percentage of cells by CD69:TFP quadrants Q1-Q4; flow cytometric analysis of select markers from OT-I T cells isolated from **(F-H)** B78chOVA or **(I-K)** PyMTchOVA tumors 14 days post T cell injection grouped by quadrants; (data are mean +/- SEM, representative of >=2 independent experiments with 3-5 mice per experiment, null hypothesis testing by paired RM ANOVA with post-hoc paired t-tests).



961 **Fig. S10:Q2 is enriched in effectors but not devoid of terminally differentiated T cells. (A)**
962 Overlay of each CD69:TFP quadrant in the UMAP space with the corresponding clusters shown
963 side-by-side; **(B)** Stacked bar plot showing the distribution of cells in the computationally-defined
964 clusters among all Q2 and Q4 cells.



965

966 **Fig. S11: CD81 marks a rare subset of cells in Q2 (A)** %PD1+CD38+ of d14 intratumoral OT-I
967 T cells in B78chOVA tumors grouped by quadrants and with a subgating to show CD81+ Q2
968 (T^*_{EFF}) cells; **(B)** % CD81+ among d14 intratumoral OT-I, endogenous T cells and OT-I T cells in
969 the dLN of mice bearing B78chOVA tumors **(C)** CD81 expression in d14 intratumoral OT-I T cells
970 in PyMTchOVA tumors grouped by quadrants and **(D)** % CD81+ among d14 intratumoral OT-I,
971 endogenous T cells and OT-I T cells in the dLN of mice bearing PyMTchOVA tumors (data
972 representative of 2 independent experiments with 3-4 tumors per experiment; **(E)** CD81
973 expression among the quadrant-sorted populations; Bar graphs show mean +/- SEM; null
974 hypothesis testing by RM ANOVA and post hoc paired t test (A, C) and by ANOVA and post hoc
975 Holm-Šídák test (B, D).

976

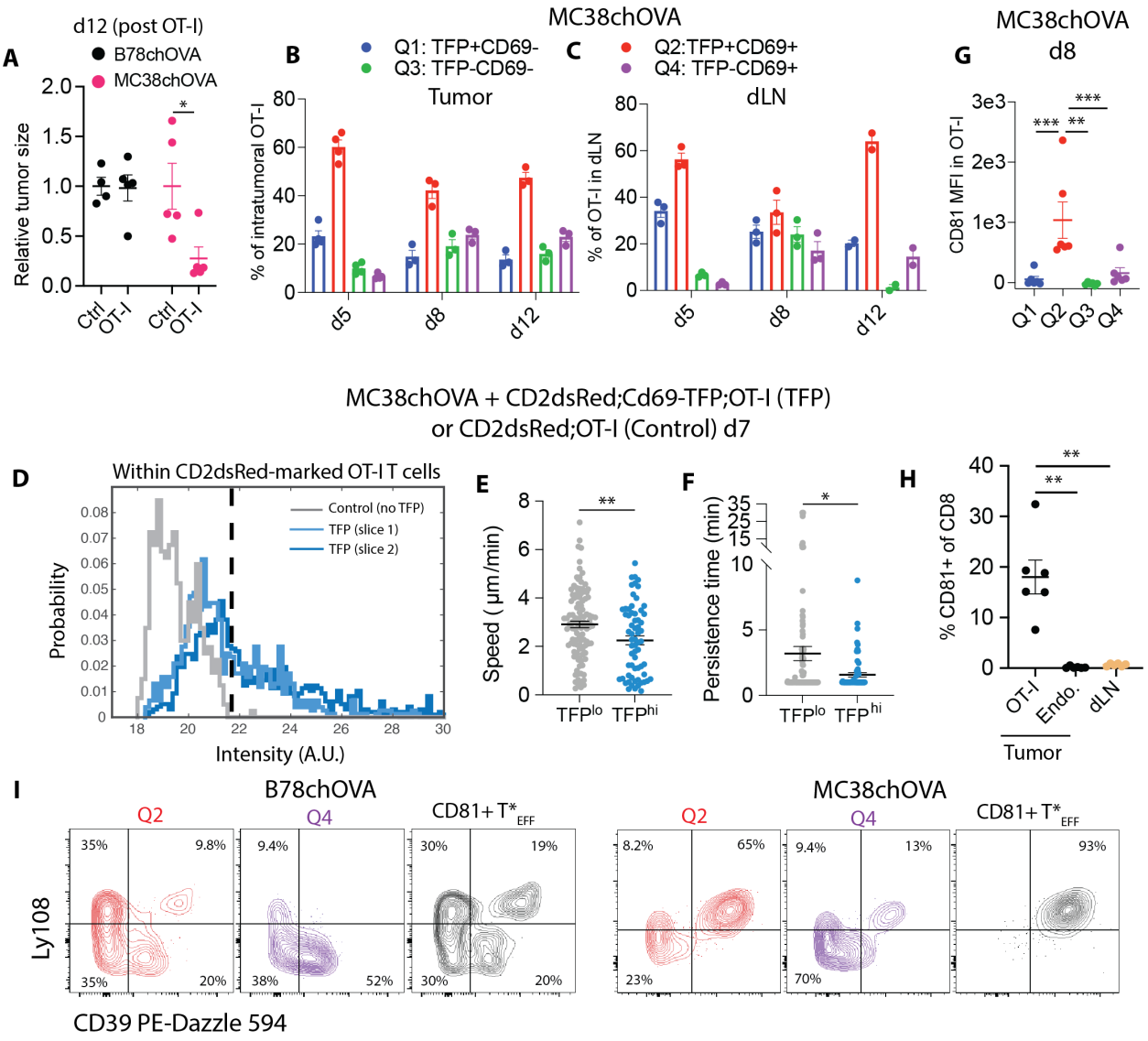


Fig. S12: **Features of Q2 OT-I T cells in MC38chOVA tumors. (A)** B78chOVA and MC38chOVA tumor sizes relative to the mean of the Ctrl group d12 post adoptive transfer of Cd69-TFP;CD45.1;OT-I T cells; in Bar graphs showing TFP:CD69 quadrant distribution among OT-I CD8 T cells in **(B)** tumors and **(C)** tdLN at d5, d8, d12 post T cell injection into MC38chOVA tumor-bearing mice corresponding dLNs (n=3-4 mice per group respectively); **(D)** Representative histograms of channel intensity within OT-I T cells in live tumor slices to find TFP^{hi} cells using CD2dsRed and CD2dsRed;Cd69-TFP OT-Is; **(E)** Speed and **(F)** Persistence of TFP^{hi} vs. TFP^{lo} intratumoral OT-Is d8 post adoptive transfer within live MC38chOVA tumor slices; **(G)** CD81 expression in d8 intratumoral OT-I T cells grouped by quadrants; **(H)** % CD81+ among d8 intratumoral OT-I, endogenous T cells and OT-I T cells in the dLN of mice bearing B78chOVA tumors; **(I)** Ly108 vs. CD39 expression profiles in d12 (B78chOVA) and d8 (MC38chOVA)

intratumoral OT-Is, separated by Q2, Q4 and CD81+ T*$_{EFF}$ subsets; bar graphs show mean +/- SEM, null hypothesis testing by unpaired t test (A), Mann-Whitney U test (E, F), paired RM ANOVA with post-hoc paired t-tests.

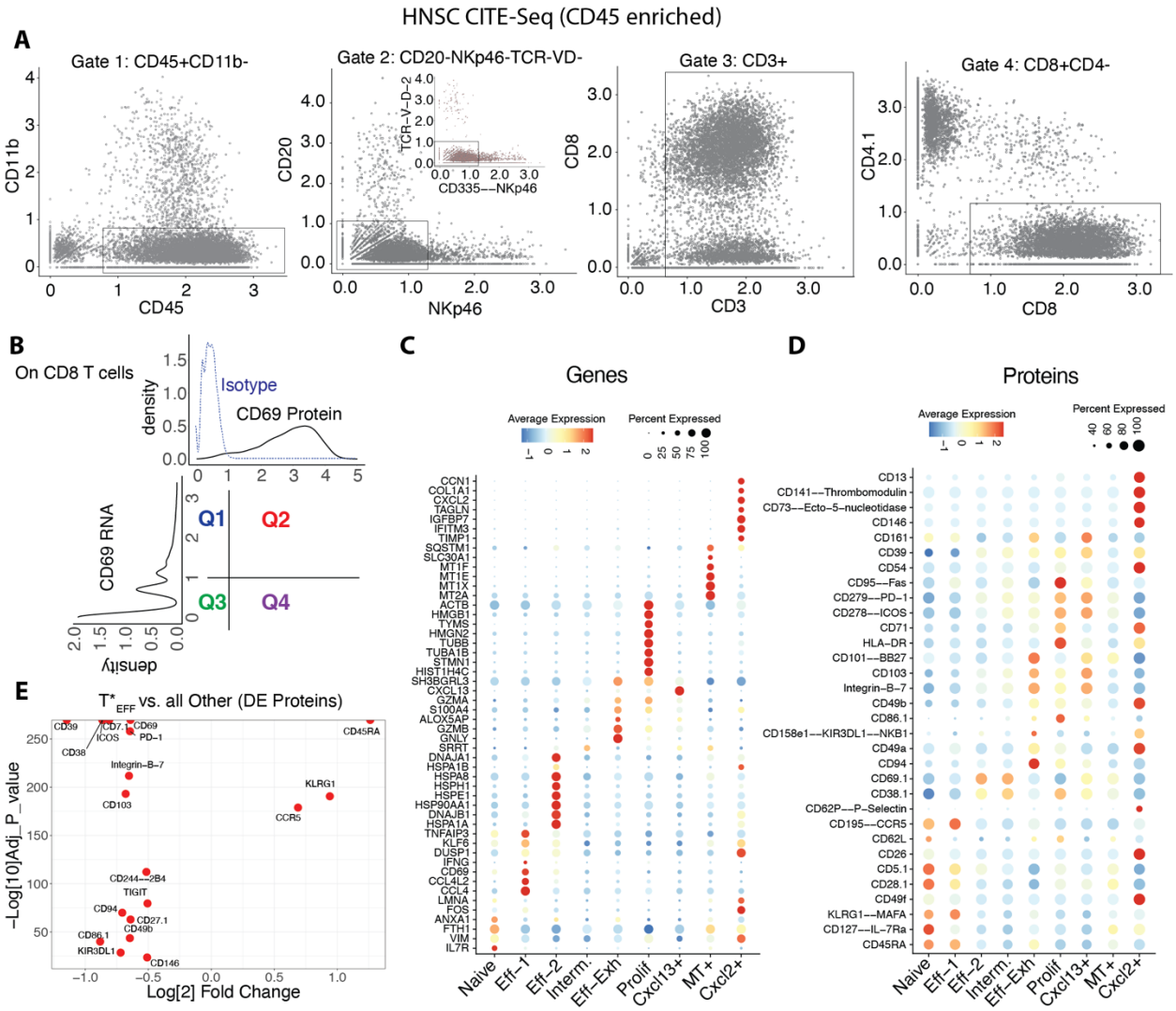


Fig. S13: **CITE-Seq of HNSC tumor sample allows mapping of quadrants onto cell phenotypes (A)** Gating scheme of CD45-enriched HNSC CITE-Seq data using protein markers to isolate a pure CD8 population; **(B)** Gating of the CD8 population into CD69 Protein: CD69 RNA quadrants; **(C)** DEGs and **(D)** DE Proteins for the computationally derived subsets obtained through multimodal clustering using both protein and RNA; **(E)** Volcano plot showing DE Proteins in the T*$_{EFF}$ (Q2 ∩ Eff-1) vs. all other CD8 T cells pre-filtered by a p-value <0.01 and average abs(log_2 fold change) >0.5;

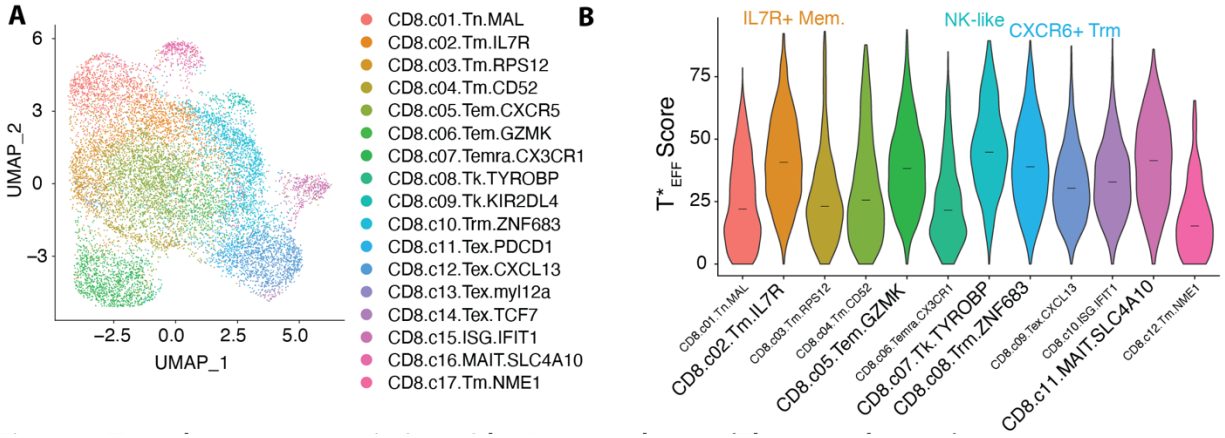


Fig. S14: **T*$_{EFF}$ phenotype association with CD8 metaclusters**. **(A)** UMAP representation of computationally-derived subsets among CD8 T cells in a pan-cancer T cell atlas(*30*) and **(B)** Violin plot showing the T*$_{EFF}$ signature score across those subsets – black line denotes median.