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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
X	A description of all covariates tested
\times	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
\boxtimes	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
\boxtimes	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
\boxtimes	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection

The SNP/Indel data of soyben accessions (Nat Genet 49, 773–779 (2017)) were used in this study.

Data analysis

The plant height, the fluorescent signal intensity and the relative fold change of western blot band were measured by Image J software. Confocal microscopy was performed using the LSM-700laser scanning confocal microscope(Zeiss). Haplotype network analysis was performed using PopART software. Phylogenetic analysis was based on UPGMA tree implemented in MEGA7 software. Quantitative data were subjected to statistical analysis performed with the GraphPad Prism 8.0.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

The Williams 82 soybean reference genome sequence was downloaded from the Phytozome v12 (https://phytozome.jgi.doe.gov/pz/

		tural population accessions were previously reported were deposited into the NCBI database under accession number nce Archive database in BIG Data Center under accession numbers PRJCA000205 and PRJCA001691.
Research invol	ving hur	man participants, their data, or biological material
	ut studies w	ith <u>human participants or human data</u> . See also policy information about <u>sex, gender (identity/presentation),</u>
Reporting on sex and	d gender	N/A
Reporting on race, et other socially relevar groupings		N/A
Population character	ristics	N/A
Recruitment		N/A
Ethics oversight		N/A
ote that full information	on the appro	val of the study protocol must also be provided in the manuscript.
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Clinical data

Dual use research of concern

Plants

Antibodies

Antibodies used Mouse monoclonal anti GFP (Proteintech, Wuhan, China; Cat No: 66002-1-lg;1:5000)

Mouse monoclonal anti MYC (Proteintech, Wuhan, China; Cat No: 60003-2-lg;1:5000) Mouse monoclonal anti Flag (Proteintech, Wuhan, China; Cat No:66008-4-lg;1:1000)

Rabbit ubiquitin polyclonal antibody (Proteintech, Wuhan, China; Cat No:10201-2-AP;1:1000)

Mouse monoclonal anti MBP (Proteintech, Wuhan, China; Cat No:66003-1-lg;1:5000)

Secondary antibody; HRP-conjugated Affinipure Goat Anti-Rabbit IgG(H+L) (Proteintech, Wuhan, China; Cat No:SA00001-2;1:5000) Secondary antibody; HRP-conjugated Affinipure Goat Anti-Mouse IgG(H+L)(Proteintech, Wuhan, China; Cat No:SA00001-1;1:5000)

Validation

All antibodies are commercially available and widely used in the scientific community. The following antibodies were validated by the supplier:

Mouse monoclonal anti GFP: https://www.ptgcn.com/products/eGFP-Antibody-66002-1-lg.htm Mouse monoclonal anti MYC: https://www.ptgcn.com/products/MYC-Antibody-60003-2-lg.htm Mouse monoclonal anti Flag:https://www.ptgcn.com/products/Flag-tag-Antibody-66008-4-lg.htm

Rabbit ubiquitin polyclonal antibody:https://www.ptgcn.com/products/ubiquitin-Antibody-10201-2-AP.htm

Mouse monoclonal anti MBP:https://www.ptgcn.com/products/MBP-Tag-Antibody-66003-1-lg.htm

HRP-conjugated Affinipure Goat Anti-Rabbit IgG(H+L):https://www.ptgcn.com/products/HRP-conjugated-Affinipure-Goat-Anti-

Rabbit-IgG-H-L-secondary-antibody.htm

 $HRP-conjugated\ Affinipure\ Goat\ Anti-Mouse\ IgG(H+L)\ \ \vdots\ https://www.ptgcn.com/products/HRP-conjugated-Affinipure-Goat-Anti-Mouse-IgG-H-L-secondary-antibody.htm$

Eukaryotic cell lines

Policy information about <u>cell lines and Sex and Gender in Research</u>

Cell line source(s) Y2H gold is a strain of yeast used in biological research for two hybrid screening.

Authentication The plasmid was transformed into the yeast strain Y2H gold using the lithium acetate method.

Mycoplasma contamination Y2H gold cell line was negative for mycoplasma contamination.

Commonly misidentified lines (See ICLAC register)

No cell lines used were listed in the database of commonly misidentified cell lines.