Supplemental Material

Safety and Preliminary Efficacy of ORBCEL-M Cell Therapy in Diabetic Kidney Disease: The Multicenter, Randomized, Placebo-controlled NEPHSTROM Trial

Norberto Perico, Giuseppe Remuzzi, Matthew D Griffin, Paul Cockwell, Alexander P Maxwell, Federica Casiraghi, Nadia Rubis, Tobia Peracchi, Alessandro Villa, Marta Todeschini, Fabiola Carrara, Bernadette A Magee, Piero L Ruggenenti, Stefano Rota, Laura Cappelletti, Veronica McInerney, Tomás P Griffin, Md Nahidul Islam, Martino Introna, Olga Pedrini, Josée Golay, Andrew A Finnerty, Jon Smythe, Willem E Fibbe, Stephen J Elliman, Timothy O'Brien MD PhD² for the NEPHSTROM Trial Consortium

Table of Contents

Supplemental Results. *Rationale for cohort 1 final analysis.*

Supplemental Table 1. *List of antibodies and the relative fluorochrome (as Format).*

Supplemental Figure 1. Correlations between percentages of Tregs or CD45RA⁻RO⁺ memory Tregs and serum concentrations of inflammatory mediators in the overall study cohort.

Supplemental Figure 2. Correlations between percentages of Tregs or CD45RA⁻RO⁺ memory Tregs and estimated glomerular filtration rate in the overall study cohort.

Supplemental Figure 3. Correlations between serum concentrations of inflammatory mediators and measured or estimated glomerular filtration rate in the overall study cohort.

NEPHSTROM Trial Consortium. Members, coordinating centers and contributions.

SUPPLEMENTAL RESULTS

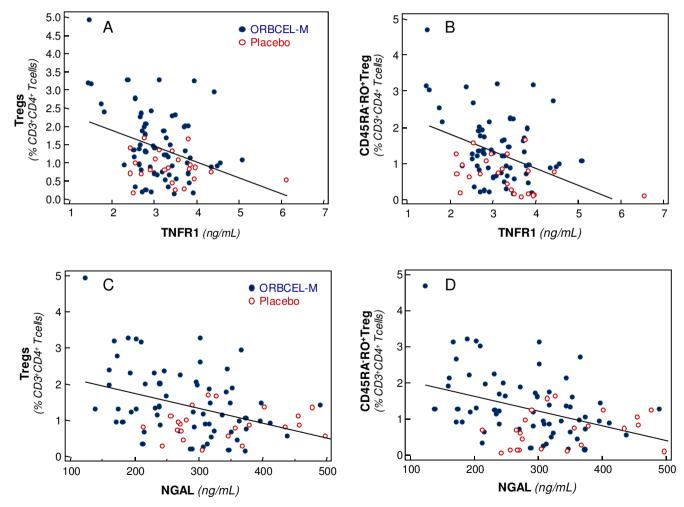
Rationale for cohort 1 final analysis

This report provides the final safety and preliminary efficacy analysis for the first cohort of 16 patients with progressive DKD who were randomized in double-blind fashion to receive an infusion of ORBCEL-M at the lowest planned dose (80 x 10⁶ cells) or a placebo infusion. The initial study protocol included two additional cohorts of 16 patients to be completed sequentially. These were to receive an intermediate dose (ORBCEL-M 160 x 10⁶ cells or placebo) and a high dose (ORBCEL-M 240 x 10⁶ cells or placebo) of the trial intervention. Once follow-up of the first dose cohort was completed, the Data Safety Monitoring Board (DSMB) reviewed the relevant data and provided the green light to activate the intermediate dose cohort. Following this, 13 of an intended 16 eligible patients were randomized, treated and are undergoing follow-up at the time of writing. However, due to COVID-19-related delays in enrolment to this second cohort, the approved shelf-life of remaining doses of cryopreserved investigational product for the cohort was exceeded and no further enrolment was possible. For this reason, the NEPHSTROM Sponsor and Trial Steering Committee made the decision to close the trial for further enrolment. As the 13 patients enrolled into the second cohort had completed their treatment and were continuing to be monitoring in double blind fashion, it was concluded that an unblinded analysis of data from the fully completed first cohort would yield important information regarding the safety and preliminary efficacy 80 x 10⁶ cell dose without biasing the subsequent analysis of data for the partially-enrolled second cohort once it is completed.

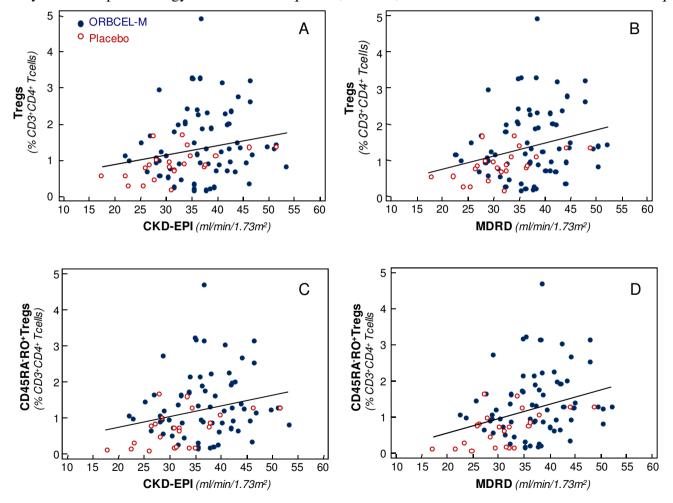
 $\textbf{SUPPLEMENTAL TABLE 1.} \ \textit{List of antibodies and the relative fluorochrome (as Format)}.$

1. Panel for CD4 ⁺ and CD8 ⁺ T cell, B cell, NK and monocyte subpopulation		
Antibody	Clone	Format
CD45	HI30	BUV395
CD3	HIT3a	BV510
CD19	HIB19	PECy7
CD27	M-T271	APCR700
IgD	IA6-2	FITC
IgM	G20-127	BV786
CD24	ML5	PE
CD38	HIT2	APC
CD16	B73.1	BB700
CD56	B159	BV650
CD11b	D12	BV711
CD14	M5E2	BUV737
CD64	10.1	PE-CF594
CX3CR1	2A9-1	BV421
HLA-DR	G46-6	BV605
CD33	WM-53	PE-Cy5
Additional reagents: Via-Probe or in alternative 7-AAD, Brillant Stain Buffer and Stain Buffer		
2. Panel for CD4 ⁺ Tregs		
Antibody	Clone	Format
CD45	HI30	BUV395
CD3	HIT3a	BV510
CD4	SK3	BB700
CD8	RPA-T8	BV711
CD45RA	HI100	PECy7
CD45RO	UHCL1	BV786
CD25	(MA251)	BV421
FOXP3	236A/E7	AF647
CD127	HIL-7R-M21	APC-R700
HLA-DR	G46-6	BV605
HELIOS	22F6	PE
CD197(CCR7)	150503	PE-CF594
CD62L	DREG-56	BV650
CD28	CD28.2	BUV737
CD95	DX2	PECy5
Additional reagents: Fixable Viability Stain 520, Transcription Factor Buffer Set, Brillant Stain Buffer,		
Stain Buffer		
3. Panel for Dendritic cells		
Antibodies	Clone	Format
CD45	HI30	BUV395
Lineage Cocktail 1(lin 1)	multiple	FITC
HLA-DR	G46-6	BV605
CD1c	F10/21A3	Alexa647
CD141	1A4	BB700
CD303	V24-785	BV421
CD123	9F5	PE
CD11c	B-ly6	BV711
CD33	WM-53	PE-Cy5
Additional reagents: Via-Probe or in alternative 7-AAD, Brillant Stain Buffer and Stain Buffer		

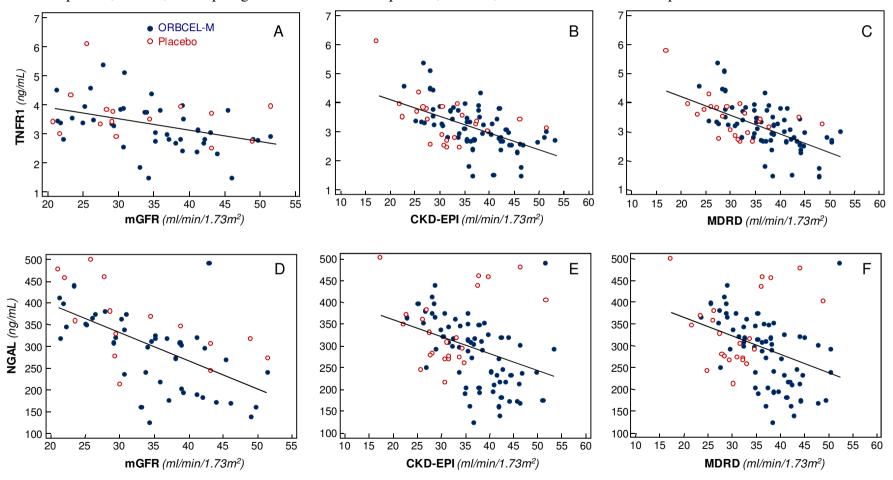
SUPPLEMENTAL FIGURE 1. Correlations between percentages of Tregs or CD45RA $^{-}$ RO $^{+}$ memory Tregs and serum concentrations of inflammatory mediators in the overall study cohort. Correlations between percentages of Tregs (A) or CD45RA $^{-}$ RO $^{+}$ memory Tregs (B) within peripheral blood CD3 $^{+}$ CD4 $^{+}$ T cells and serum TNFR1 concentrations in the overall study cohort (r = -0.3690 and -0.4024, respectively; both P<0.001). Correlations between percentages of Tregs (C) or CD45RA $^{-}$ RO $^{+}$ Tregs (D) within peripheral blood CD3 $^{+}$ CD4 $^{+}$ T cells and serum NGAL concentrations in the overall study cohort (r = -0.3879 and -0.3907, respectively; both P<0.001). TNFR1, tumor necrosis factor receptor 1; NGAL, neutrophil gelatinase-associated lipocalin.



SUPPLEMENTAL FIGURE 2. Correlations between percentages of Tregs or CD45RA-RO+ memory Tregs and estimated glomerular filtration rate in the overall study cohort. Correlations between percentages of Tregs within peripheral blood CD3+CD4+ T cells and glomerular filtration rate estimated with CKD-EPI (A) or MDRD (B) equations in the overall study cohort (r = 0.2235, P=0.035 and r = 0.2949, P<0.005, respectively). Correlations between percentages of CD45RA-RO+ memory Tregs within peripheral blood CD3+CD4+ T cells and glomerular filtration rate estimated with CKD-EPI (C) or MDRD (D) equations in the overall study cohort (r = 0.2578, P=0.015 and r = 0.3313, P<0.002, respectively). CKD-EPI, Chronic Kidney Disease Epidemiology Collaboration equation; MDRD, Modification of Diet in Renal Disease equation.



SUPPLEMENTAL FIGURE 3. Correlations between serum concentrations of inflammatory mediators and measured or estimated glomerular filtration rate in the overall study cohort. Correlations between serum TNFR1 concentrations and glomerular filtration rate measured by iohexol plasma clearance (A) or estimated by CKD-EPI (B) or MDRD (C) equations in the overall study cohort (r = -0.3929, *P*=0.002, r = -0.5493, *P*<0.001 and r = -0.5928, *P*<0.001, respectively). Correlations between serum NGAL concentrations and glomerular filtration rate measured by iohexol plasma clearance (D) or estimated by CKD-EPI (E) or MDRD (F) equations in the overall study cohort (r = -0.5932, *P*<0.001, r = -0.3486, *P*=0.001 and r = -0.3626, *P*=0.001, respectively). CKD-EPI, Chronic Kidney Disease Epidemiology Collaboration equation; MDRD, Modification of Diet in Renal Disease equation; NGAL, neutrophil gelatinase-associated lipocalin; TNFR1, tumor necrosis factor receptor 1.



NEPHSTROM Trial Consortium

<u>Coordinating Centre</u>: Mario Negri Institute for Pharmacological Research IRCCS, Clinical Research Centre for Rare Diseases Aldo e Cele Daccò, Ranica (Bergamo, Italy); Research Coordinator: Giuseppe Remuzzi (Bergamo, Italy), Trial coordinator: Norberto Perico, Nadia Rubis, Federica Casiraghi (Bergamo, Italy)

Centers including patients: Azienda Socio Sanitaria Territoriale Ospedale Papa Giovanni XXIII, Bergamo Italy (Giuseppe Remuzzi, Piero Ruggenenti, Stefano Rota, Laura Cappelletti, Valentina Portalupi, Eliana Gotti, Elena Perticucci, Grazia Natali, Alessandro Rambaldi, Giuseppe Gritti, Anna Maria Barbui, Silvia Ferrari, Silvia Mariani, Gianmaria Borleri); University of Galway Ireland (Matthew Griffin, Veronica McInerney, Tomás Griffin, Michelle Hennessy, Sarah Cormican, Nathan Devaney, Cassandra Phan, Amy Hanson, Sara Martyn, Joy Buckley, Sean Naughton, Julie Woods, Caroline Kelly, Amjad Hayat); Queen Elizabeth Hospital Birmingham, Birmingham UK (Paul Cockwell, Dimitrios Chanouzas, Lesley Fifer, Kulli Kuningas, Natalie Walmsley-Allen, Amisha Desai, Lucy Atchinson, Sinead White, Vijayan Suresh, Katie Kirkham, Fiona Evans)

<u>Data Monitoring Safety Board.</u> Dr Mark Little (Professor of Nephrology and Consultant Nephrologist, Trinity College Dublin / Tallaght and Beaumont Hospitals, Ireland). Prof. Piergiorgio Messa (Full professor of Nephrology – Università degli Studi di Milano, Italy). Christina Yap (PhD CStat, Professor of Clinical Trials Biostatistics, the Institute of Cancer Research, Sutton, UK).

Centralised activities: Regulatory activities, Monitoring and Pharmacovigilance: Laboratory of Pharmacovigilance and Monitoring for Clinical Investigations, Bergamo Italy (Nadia Rubis, Alessandro Villa, Olimpia Diadei, Paola Boccardo, Bergamo Italy with the collaboration of Lesley Fifer Birmingham UK and Veronica McInerney, Oleksii Noreiko, Galway Ireland); Database and Data Validation: Unit of Clinical Web Design and Development, Bergamo Italy (Davide Martinetti); Randomisation: Laboratory of Biostatistics, Bergamo Italy (Giovanni Antonio Giuliano); Data Analysis: Laboratory of Biostatistics, Bergamo Italy (Annalisa Perna, Tobia Peracchi); GFR measurement: Laboratory of Clinical Chemistry, Bergamo Italy (Daniela Cugini, Fabiola Carrara, Silvia Ferrari, Nadia Stucchi); anti-HLA measurement: N. I. Histocompatibility and Immunogenetics Laboratory, Belfast UK (A. Peter Maxwell, Bernadette Magee); Immune profile: Laboratory of Immunology of Organ Transplantation, Bergamo Italy (Federica Casiraghi, Marta Todeschini, Marilena Mister); Inflammatory Mediator Profile: Laboratory of M. Griffin, Galway Ireland (Matthew Griffin, Nahidul Islam, Tomás Griffin); Cell production and management: Orbsen Therapeutics Ltd, Galway Ireland (Stephen Elliman, Lisa O'Flynn, Yuka Shimizu, Galway Ireland), Leiden University Medical Center, Leiden Netherlands (Willem Fibbe, Helene Roelofs, Esther Steeneveld, Brigitte Wieles), Centro di Terapia Cellulare "Gilberto Lanzani" ASST-PG23 Bergamo, Italy (Martino Introna, Josée Golay, Olga Pedrini, Chiara Capelli), NHS Blood and Transplant, Liverpool UK (Jon Smythe, Eric Austen, Helen Murray, Vivien Hanson, Jenny Chan), Centre for Cell Manufacturing Ireland, Galway Ireland (Andrew Finnerty, Aoife Duffy, Miriam Holohan, Janusz Krawczyk, Matthew Duggan, Lauren Connolly), Galway Blood and Tissue Establishment, University Hospital Galway, Ireland (Amjad Hyatt, Janusz Krawczyk, Margaret Tarpey, Sean Naughton, Joy Buckley), Terumo BCT Europe NV, Zaventem Belgium (Layka Abbasi Asbagh, Brent Rice, Stefano Baila); EU H2020 project leader: University of Galway, Galway Ireland (Timothy O'Brien, Grace Davey, Michael Creane); EU H2020 project management: Pintail Ltd, Ireland (Ciaran Clissmann, Mark Sweetnam).