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APOBEC3B regulates R-loops and promotes transcription-associated mutagenesis in cancer

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Supplementary information

Supplementary Table 1. Supporting data sets including sequence information

- Layer 1 Proteomic data from A3B and control AP-MS experiments
- Layer 2 NTS and TS mutations in >16-fold overexpressed gene groups
- Layer 3 Sequences of oligonucleotides

Supplementary Note

To analyze the kinetics of R-loop resolution in the presence and absence of A3B, global R-loop distributions were first analyzed by DRIP-seq in WT MCF10A following 2 hrs PMA treatment compared to DMSO control treatment (workflow in **Extended Data Fig. 4a**, left). As anticipated, PMA caused changes in the overall R-loop landscape with 13,422 peaks increased, 16,432 peaks decreased, and 171,322 unchanged (Extended Data Fig. 4b-d). Many genes such as JUNB and FOS were induced strongly by PMA treatment and showed significant increases in DRIP signals (Extended Data Fig. 4e-f). Other genes such as NAXE and ARL4D showed decreases in DRIP signals (Extended Data Fig. 4g-h), whereas the majority such as GAPDH and GEMIN7 showed no changes in Rloop formation (Extended Data Fig. 4i-j). A parallel set of ChIP-seq experiments was done in MCF10A with Dox-inducible A3B-eGFP (Fig. 1c) to assess whether any of these R-loop categories might be bound by this deaminase (workflow in **Extended Data Fig.** 4a, right). An epitope-tagged protein was necessary because IP-grade antibodies have yet to be developed for endogenous A3B. Interestingly, A3B-eGFP appeared to bind preferentially to genomic DNA regions coincident with DRIP-seq peaks increased in PMA-

treated cells in comparison to vehicle control treated cells (ChIP-seq results superimposed over DRIP-seq results in **Extended Data Fig. 4b**). In contrast, A3B-eGFP ChIP-seq peaks were mainly unperturbed in genomic DNA regions in which R-loop levels decreased or remained unchanged upon PMA treatment (**Extended Data Fig. 4c-d**). Representative results were confirmed independently by ChIP-qPCR (**Extended Data Fig. 4k**). Moreover, quantification indicated that 43% of A3B ChIP peaks overlap with R-loop peaks induced by PMA (P = 0.0001, two-tailed binomial test; 54% overlap for ChIP peaks with >5-fold enrichment compared to uninduced Dox⁻ condition, P = 0.0008). These results indicate that A3B binds preferentially to genomic DNA regions with evidence for R-loop accumulation.

The PMA-inducibility of this system enabled an assessment of the kinetics of Rloop resolution in the presence and absence of A3B. WT and KO cells were treated with PMA or DMSO for 2 and 6 hrs and then analyzed by DRIP-seq (workflow in **Fig. 6a**). These timepoints were chosen for analysis because transcription induction and R-loop formation are rapid (2 hrs data above) and A3B protein levels are upregulated maximally by 6 hrs post-PMA treatment³⁰ (**Fig. 6b-c**). For example, *JUNB* and *DUSP1* have R-loop peaks that are strongly induced by PMA at 2 hrs and these return to near-control levels by 6 hrs (**Fig. 6d**). In contrast, R-loop peaks in the same genes remained significantly elevated and/or showed delayed resolution kinetics in KO cells after 6 hrs PMA treatment (**Fig. 6d**). PMA non-responsive control genes did not show R-loop induction or major differences in R-loop levels after 6 hrs PMA treatment (*e.g., GAPDH* and *HSPA8* in **Fig. 6e**). Moreover, independent IF confocal microscopy studies of the kinetics of R-loop resolution following PMA treatment of WT MCF10A cells showed that nucleoplasmic R-

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loop levels peak at 2 hrs and decline substantially by 6 hrs, whereas no decline (even a modest increase) was observed in *A3B* KO cells (**Fig. 6f-g**). As above, all DRIP-qPCR and nucleoplasmic R-loop signals were sensitive to RNase H treatment indicating specificity (**Fig. 6d-g**). Taken together with data from ChIP experiments, the results of these transcriptional activation experiments indicated that nuclear A3B is recruited to PMA-induced R-loops and contributes to their timely resolution.