

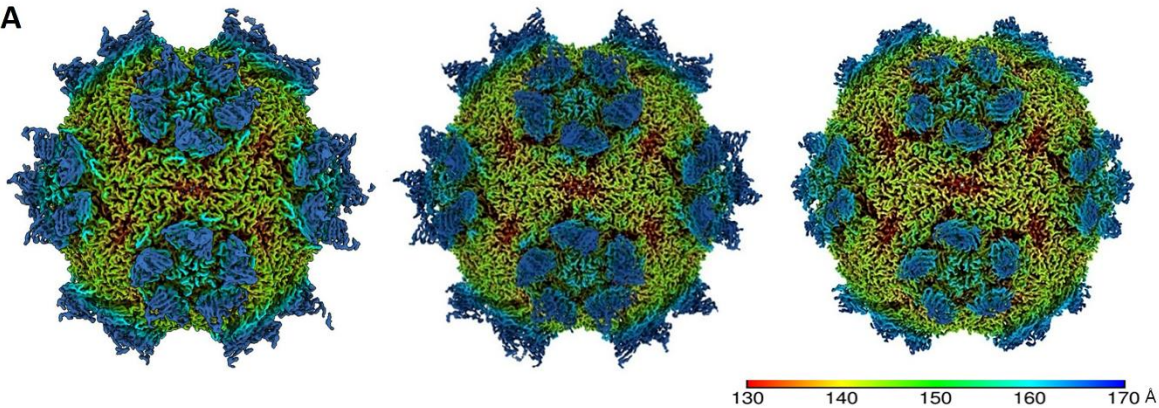
1
2 **Supplemental Figure 1: Motion Corrected Micrographs.** Example representative motion corrected
3 micrographs are shown. Fab is visible decorating the viral capsids. Central densities represents genome.
4
5

SIPV1

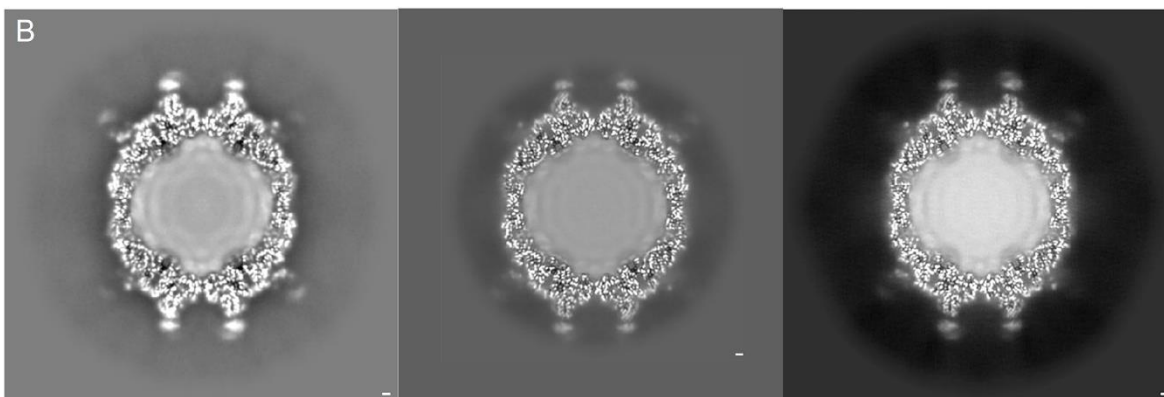
SIPV2

SIPV3

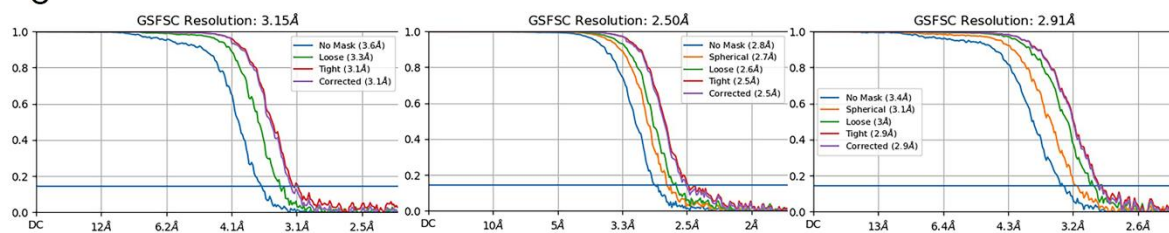
A



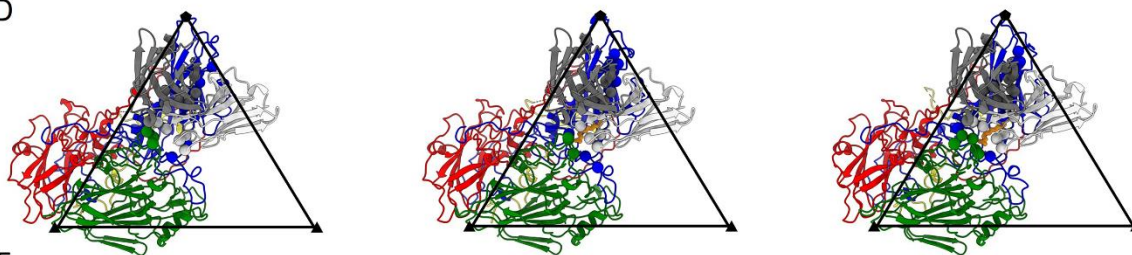
B



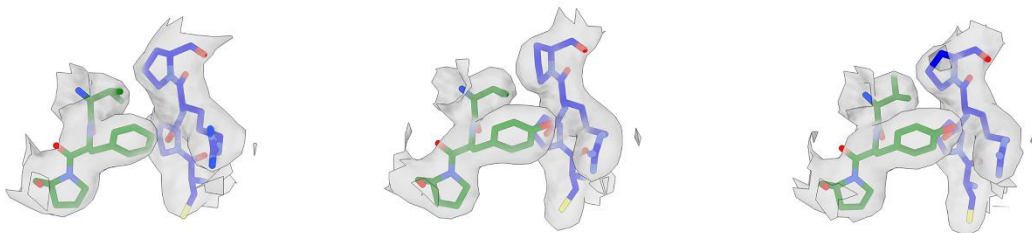
C



D

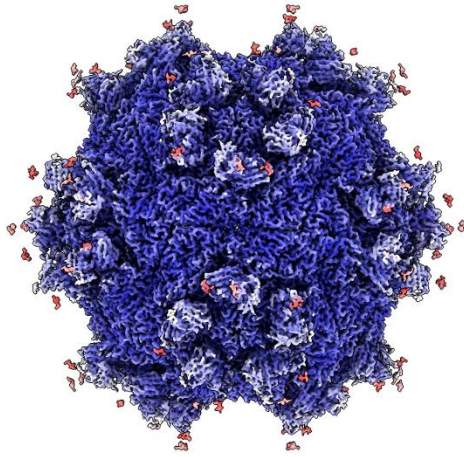


E

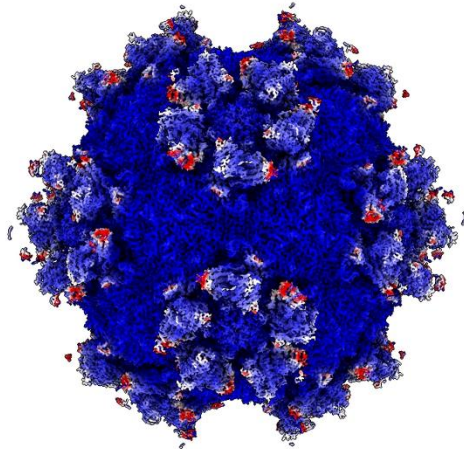


7 **Supplemental Figure 2: Inactivated Fab-virus maps: 9H2 Fab binds virus with same mode**
8 **of binding in all three serotypes (inactivated virus):** **A:** Surface rendered DeepEMhancer
9 sharpened maps are colored according to radius (color key) and show density corresponding to
10 the 9H2 Fab. **B:** The central section shows the Fab has similar magnitude of density as the capsid.
11 **C:** FSC curves indicate resolution ranging from 2.5 – 3.2 Å at the gold standard 0.143 cutoff. **D:**
12 Refined models of single virus protomer (VP1-4; blue, green, red, yellow) and bound 9H2 variable
13 domain (heavy and light chain; dark and light gray). SIPV1, 2 and 3 data were taken at
14 magnification 59000x, 75000x, and 59000x resulting in pixel sizes of 1.1, 0.89, and 1.1
15 respectively. **E:** For each complex, a representative area illustrating the quality of the model built
16 into the map is shown. The map area highlighted includes residues VP1 270-273 and VP2 192-
17 194. Coloration is VP1 (blue) and VP2 (green) with additional coloring by heteroatom.
18
19

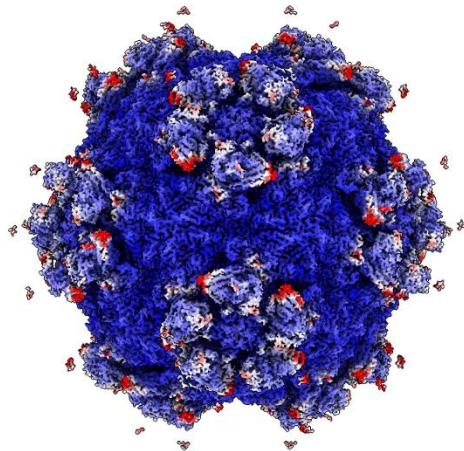
SIPV1



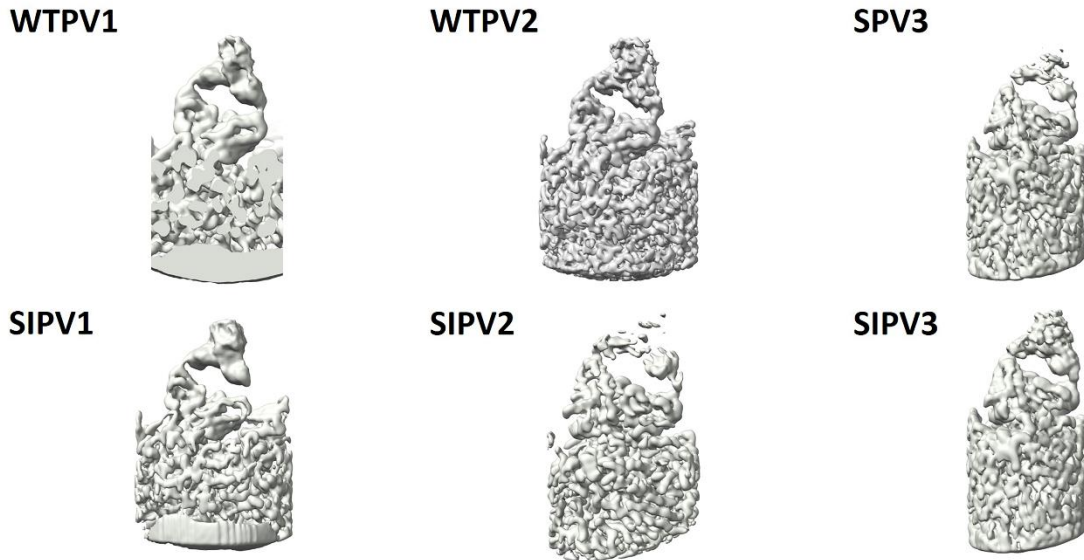
SIPV2



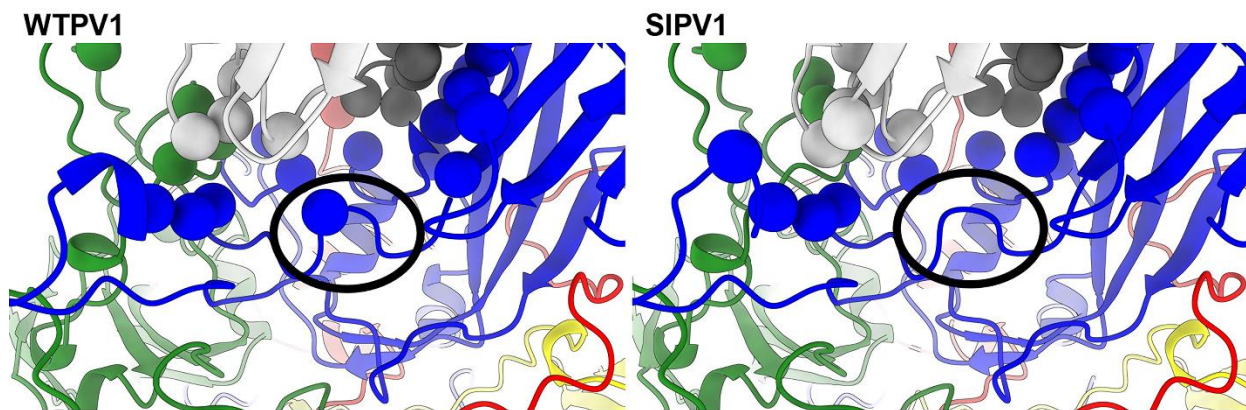
SIPV3



21 **Supplemental Figure 3: Local Resolution Maps:** Local resolution values were measured in
22 Angstroms and displayed with surface rendered for each inactive virus-Fab complex. The best
23 resolution is present in the capsid shell while the poorest is found in the hinge region of the Fab
24 fragments.
25
26



27
28 **Supplemental Figure 4: 9H2 Subparticles:** Examples of subparticle classes for each virus
29 complex with 9H2 are shown. Multiple subparticle classes were merged for the final maps which
30 were recombined to produce a symbreak full icosahedral map into which virus and 9H2 were
31 built (Methods).
32
33
34



35
36 **Supplemental Figure 5: Canyon Floor Loop:** Unlike the crystal structure 1HXS, between wild
37 type and inactivated Sabin, the structure and conformation of the loop (black circle) comprised
38 of VP1 residues 232-238 superimpose. Residues identified as contacts are displayed as
39 spheres matching their chain color (heavy and light chain; dark and light gray; blue,
40 green, red).
41
42

VP1 contacts 18-21

```

ISPV1 60 TVQTRHVQHRSRSESSIESFFARGACVAIIITVDNSASTKNKDKLFTVWKITYKDTVQLR 119
WTPV1 60 TVQTRHVQHRSRSESSIESFFARGACVTIMTVDNPASTTNKDKLFAVWKITYKDTVQLR 119
ISPV2 60 TVQTRHVIQRTRRSESTVESFFARGACVAIIIEVDNDAPTQRASRLFSVWKITYKDTVQLR 119
WTPV2 60 TVQTRHVIQRTRRSESTVESFFARGACVAIIIEVDNDAPTQRASKLFSVWKITYKDTVQLR 119
ISPV3 60 TVQTRHVQRRSRSESTIESFFARGACVAIIIEVDNEQPTTRAOKLFAMWRITYKDTVQLR 119
SPV3 60 TVQTRHVQRRSRSESTIESFFARGACVAIIIEVDNEQPTTRAOKLFAMWRITYKDTVQLR 119
*****:*:****:*****:*** *..:****:*****

ISPV1 120 RKLEFFTYSRFDMETFVVTANFTETNNGHALNQVYQIMYVPPGAPVPEKWDDYTWQTSS 179
WTPV1 120 RKLEFFTYSRFDMELTFVVTANFTETNNGHALNQVYQIMYVPPGAPVPEKWDDYTWQTSS 179
ISPV2 120 RKLEFFTYSRFDMETFVVTSNYIDANNHALNQVYQIMYIPPGAPIPGKWDDYTWQTSS 179
WTPV2 120 RKLEFFTYSRFDMETFVVTSNYIDANNHALNQVYQIMYIPPGAPIPGKWDDYTWQTSS 179
ISPV3 120 RKLEFFTYSRFDMETFVVTANFTNANNHALNQVYQIMYIPPGAPTPKSWDDYTWQTSS 179
SPV3 120 RKLEFFTYSRFDMETFVVTANFTNANNHALNQVYQIMYIPPGAPTPKSWDDYTWQTSS 179
*****:*****: :*****:***** *.:*****

ISPV1 180 NPSIFYTYGTAPARISVPYVGISNAYSHFYDGFSAKVPPLKQDS-AALGDSLGAASLNDFG 238
WTPV1 180 NPSIFYTYGTAPARISVPYVGISNAYSHFYDGFSAKVPPLKQDS-AALGDSLGAASLNDFG 238
ISPV2 180 NPSVFYTYGAPPARISVPYVGIANAYSHFYDGFSAKVPPLAGQA-STEGDSLGAASLNDFG 238
WTPV2 180 NPSVFYTYGAPPARISVPYVGIANAYSHFYDGFSAKVPPLAGQA-STEGDSLGAASLNDFG 238
SIPV3 180 NPSIFYTYGAAPARISVPYVGLANAYSHFYDGFSAKVPPLKTDANDQIGDSLGSAMTVDDFG 239
SPV3 180 NPSIFYTYGAAPARISVPYVGLANAYSHFYDGFSAKVPPLKTDANDQIGDSLGSAMTVDDFG 239
***:*****: *****:*****:*** : *****.* :*:***

ISPV1 239 ILAVRVVNDHNPTKVTSKIRVYLKPKHIRVWCPRPPRAVAYYGGVVDYKDGTLTPLSTKD 298
WTPV1 239 ILAVRVVNDHNPTKVTSKIRVYLKPKHIRVWCPRPPRAVAYYGGVVDYKDGTLTPLSTKD 298
ISPV2 239 SLAVRVVNDHNPTRLTSKIRVYMKPKHVRVWCPRPPRAVPYFGPGVDYKDG-LTLPPEKG 297
WTPV2 239 SLAVRVVNDHNPTKLTSLKIRVYMKPKHVRVWCPRPPRAVPYFGPGVDYKDG-LAPLPKG 297
ISPV3 240 VLAVRVVNDHNPTKVTSKVRIYMKPKHVRVWCPRPPRAVPYFGPGVDYRNN-LDPLSEKG 298
SPV3 240 VLAVRVVNDHNPTKVTSKVRIYMKPKHVRVWCPRPPRAVPYFGPGVDYRNN-LDPLSEKG 298
*****:***:*:****:*****:***** *:*****:.. * ** *.

```

VP2 contacts 3-6

```

ISPV1 121 ALGVFAVPEMCLAGDSNTTMMHTSYQANANPGEKGGTFTGTFTPDNNQTSPPARRFCVDYL 180
WTPV1 121 ALGVFAVPEMCLAGDSNTTMMHTSYQANANPGEKGGTFTGTFTPDNNQTSPPARRFCVDYL 180
ISPV2 121 ALGVFAVPEMCLAGDST-TMMHTKYENANANPGEKGGEFKGSFTLDTNATNPARNFPCVDYL 179
WTPV2 121 ALGVFAVPEMCLAGDST-TMMHTKYENANANPGEKGGEFKGSFTLDTNATNPARNFPCVDYL 179
ISPV3 121 ALGVFAIPEYCLAGDSDK-QRYTSYANANPGERGGKFKYSQFNKNNAVTSPPKREFCVDYL 179
SPV3 121 ALGVFAIPEYCLAGDSDK-QRYTSYANANPGERGGKFKYSQFNKNNAVTSPPKREFCVDYL 179
*****:*** ***** *.* *****:*** * . * . * ** *.******

```

VP3 contacts 1-2

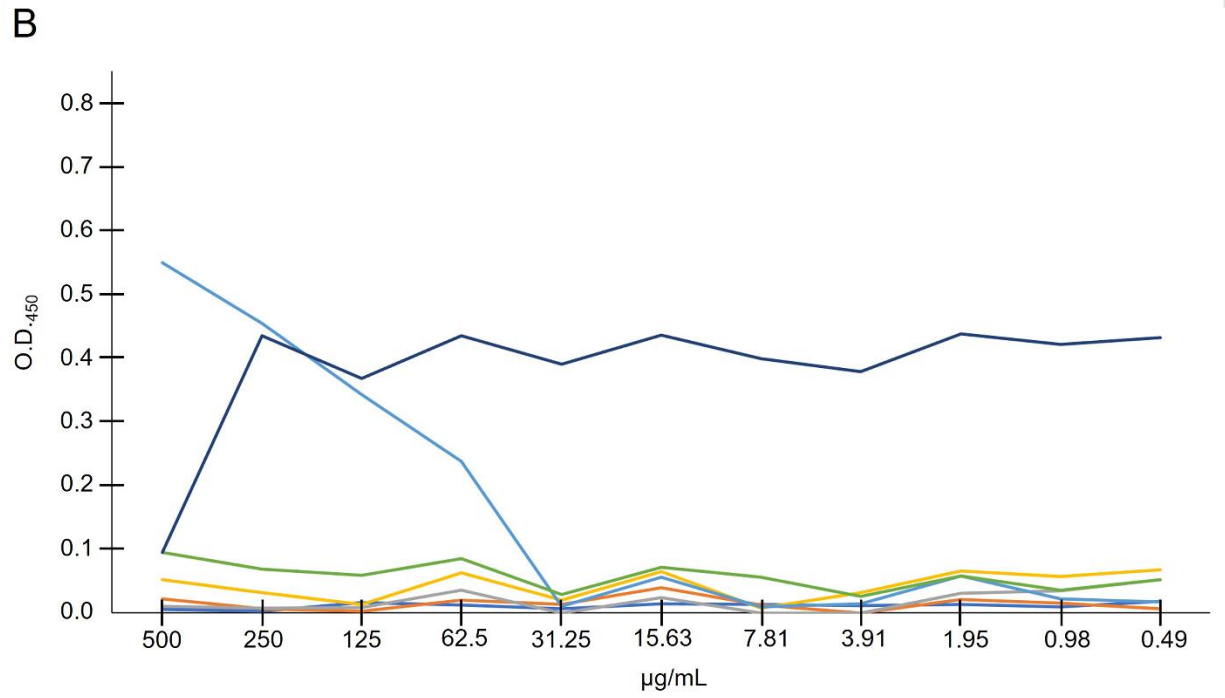
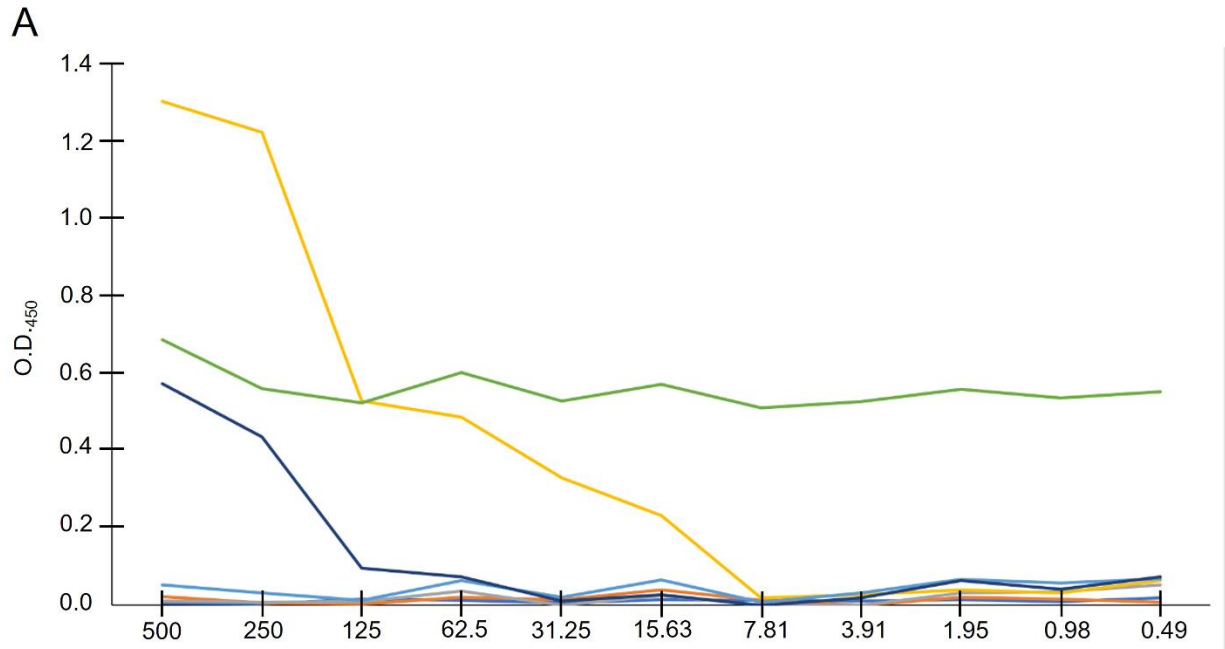
```

ISPV1 226 RDTTHIEQKA 235 3.15 Å
WTPV1 226 RDTTHIEQKA 235 3.13
ISPV2 226 RDTTHISQEA 235 2.50
WTPV2 226 RDTTHISQEA 235 2.73
ISPV3 226 RDTTHISQSA 235 2.91
SPV3 226 RDTTHISQSA 235 2.66

```

TOTAL 9H2 CONTACTS: 23-27

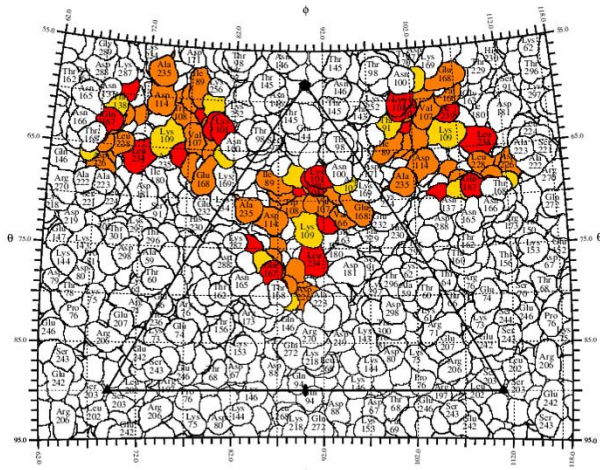
44 **Supplemental Figure 6: Sequence Alignment of Polioviruses:** 9H2 contacts are highlighted
45 in this Clustal Omega sequence alignment. Contact number is provided as low – high end
46 estimates for 9H2 footprint residue contacts. Variation likely resulted from differences in
47 resolution among maps and slight differences in chain and rotamer placement.
48
49



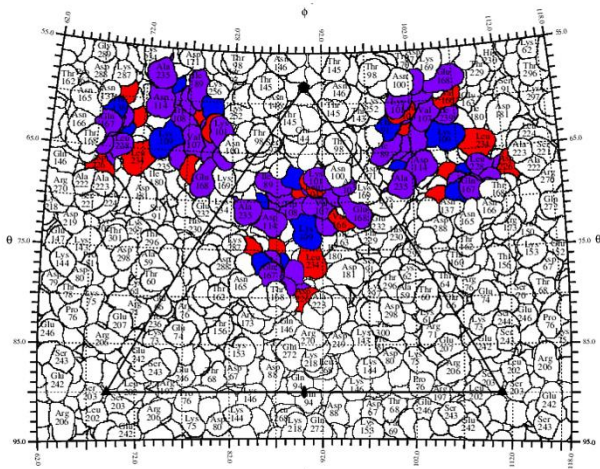
Controls	Experiment
— TBS-Tween20	— 9H2
— 9H2	(WTPV1+sPVR+9H2)
— sPVR	— sPVR
— 9H2 (WTPV1+9H2)	(WTPV1+9H2+sPVR)
— sPVR (WTPV1+sPVR)	

51 **Supplemental Figure 7: Binding Competition ELISA:** Binding of human monoclonal antibody
52 9H2 or recombinant soluble poliovirus receptor (sPVR) to wild type poliovirus type
53 1(Mahoney)(WTPV1) or WTPV1 complexed with 9H2 or sPVR. Binding was assessed by
54 enzyme-linked immunosorbent assay (ELISA) and reported optical density at 450nm. The results
55 are representative of three independent experiments. X axes are the concentration of 9H2 mAb
56 or sPVR in $\mu\text{g}/\text{mL}$. Y axes are optical density readings at 450 nm. Color key labels indicate the
57 specimen being serially diluted and controls. For both A and B the green line indicates the ability
58 of sPVR to compete off bound 9H2 by plotting the results of the sequential addition of virus, 9H2
59 held constant at 0.1 mg/mL across all dilution wells, followed by PVR at various dilutions. For both
60 A and B the blue line represents the ability for 9H2 to outcompete receptor; whereby the treatment
61 corresponds to the sequential addition of PV1, followed by the addition of a consistent amount
62 (0.1 mg/mL) of sPVR, and final addition of 9H2 at various dilutions. **A:** Human mAb-HRP
63 secondary antibody was used to report 9H2 bound to captured virus. **B:** Anti-sPVR secondary
64 mAb-HRP was used to report sPVR bound to captured virus. Application of 9H2 mAb at 0.1
65 mg/mL is capable of preventing subsequent sPVR (at 0.5 mg/mL) binding to the virus whereas
66 0.5 mg/mL 9H2 mAb can outcompete previously bound sPVR (at 0.1 mg/mL).
67
68

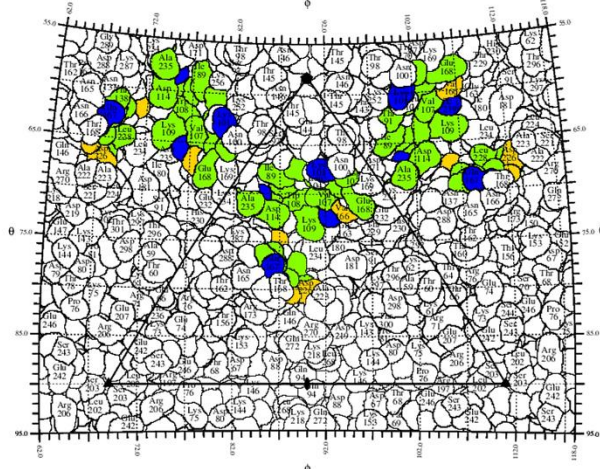
A



B



C



69

70 **Supplemental Figure 8: Live Footprint Comparisons: A-C:** 9H2 footprint comparisons
71 among live PVs showing differences between serotypes. PV1 (Red), PV2 (gold), PV3 (blue).
72 Overlaps shown; PV1 & PV2, orange; PV1 & PV3, purple; PV2 & PV3, green.

Supplemental Table 1: Data Collection, Processing, and Refinement Statistics

Data Collection and Processing	WTPV1	WTPV2	SPV3	SIPV1	SIPV2	SIPV3
magnification	59k	59k	120k	59k	75k	59k
voltage (kV)	300	300	200	300	300	300
e-/Å ²	40	40	50	40	40	40
defocus range (µm)	1-3	1-3	0.5-2	1-2.5	0.75-2.5	0.75-2.5
pixel size (Å)	1.1	1.1	1.2	1.1	0.86	1.1
symmetry imposed	ICOS	ICOS	ICOS	ICOS	ICOS	ICOS
initial micrographs	1592	2284	3737	2302	1921	734
final micrographs	1550	2185	2915	1883	1701	714
particles	79471	11599	84951	99462	111182	52115
map resolution (Å)	3.1	2.7	2.7	3.2	2.5	2.9
Refinement						
model resolution* d _{model} (Å)	3.3	2.9	2.8	3.3	2.6	3.0
model resolution** d _{FSC} model 0.143 (Å)	3.1	2.7	2.6	3.1	2.5	2.8
map sharpening B factor	176.3	109.3	135.5	181.3	129.5	145.5
non-hydrogen atoms	8318	8245	8343	8300	8310	8329
protein residues	1068	1059	1068	1064	1066	1066
ligands	N/A	PLM	PLM	N/A	PLM	PLM
protein B factors mean (Å ²)	100.83	58.74	57.55	91.93	54.20	73.54
ligand B factors (Å ²)	N/A	58.98	66.86	N/A	55.04	70.11
RMS deviation bond length (Å)	0.005	0.006	0.007	0.004	0.005	0.005
RMS deviation bond angle (°)	0.774	0.898	1.083	0.712	0.792	0.893
MolProbity score	0.95	1.20	1.11	1.00	1.13	1.31
Clashscore	0.67	1.78	0.67	1.47	1.77	1.52
Rotamer outliers (%)	0.11	1.55	1.74	1.41	0.44	1.74
Ramachandran favored (%)	96.49	97.41	96.67	98.00	96.77	96.48
Ramachandran allowed (%)	3.42	2.49	3.33	2.00	3.23	3.52
Ramachandran outliers (%)	0.09	0.10	0.00	0.00	0.00	0.00

74 Common statistics and parameters from the six 9H2 Fab-poliovirus complexes, with refinement
75 statistics generated from Phenix comprehensive validation. ICOS=icosahedral. PLM=palmitic
76 acid.

77
78 *Model resolution d_{model} is the resolution of the map calculated from the final model that
79 maximizes the correlation to the experimental map¹.

80 **Model resolution $d_{\text{FSC_model}}$ is the resolution cutoff at which the model and experimental map
81 Fourier coefficients are most similar¹.

82
83

84 Supplemental Table 2: Capsid Contacts

85

WTPV1

VP1	87 88 89 90 101 102 104 105 106 107 108 114 166 168 226 227 228 234 239 280 282
VP2	139 140 142 167
VP3	233 235

SIPV1

VP1	87 88 89 90 101 104 105 106 107 108 109 114 166 168 169 223 226 227 228 280 282
VP2	138 139 140 167
VP3	235

WTPV2

VP1	87 88 89 91 102 103 105 106 107 108 109 114 166 168 226 227 228 282
VP2	137 138 139 141 171
VP3	233 235

SIPV2

VP1	87 88 89 90 100 101 103 105 106 107 108 114 166 168 228 234 239 282
VP2	137 138 139 141
VP3	235

SPV3

VP1	87 88 89 91 101 102 103 105 106 107 108 109 114 115 168 228 229 240
VP2	137 138 139 140 141 166
VP3	233 235

SIPV3

VP1	87 88 89 91 100 101 102 103 105 106 107 108 114 168 224 228 229 240 281
VP2	137 140 141
VP3	235

86 Identified contact residue numbers in each 9H2-poliovirus complex for each virus chain.

87

88

89 **Supplemental Table 3: 9H2 Contacts**

90

WTPV1

Heavy chain	81 119 120 121 122 123 124 125 127 128 129 130
Light chain	45 46 50 73 74 114 115

WTPV2

Heavy chain	74 121 122 123 124 125 127 128 129 130 131
Light chain	45 46 50 74 114 115

SPV3

Heavy chain	74 81 120 121 122 123 124 125 127 128 129 130 131
Light chain	44 45 46 50 52 73 74 114 115

SIPV1

Heavy chain	81 119 120 121 122 123 124 125 127 128 129 130
Light chain	45 46 50 73 74 75 90 113 114 115

SIPV2

Heavy chain	46 120 121 123 124 125 127 128 129 130 131
Light chain	45 46 49 50 74 114 115

SIPV3

Heavy chain	46 120 121 122 123 124 125 127 128 129 130 131
Light chain	45 46 50 73 74 90 115

91 Identified contact residue numbers in each 9H2-poliovirus complex for each Fab chain.

92

93

94

Supplemental Table 4: 9H2 Fab Binding Constants

	KD (M)	Ka (1/Ms)	Ka error	Kd (1/s)	Kd error
SPV1	9.364 x 10 ⁻⁴	1.122 x 10 ³	8.470 x 10 ⁴	1.050	0.1795
SIPV2	2.097 x 10 ⁻⁶	2.301 x 10 ⁵	1.182 x 10 ⁵	0.4826	0.1374
SPV3	3.232 x 10 ⁻⁶	1.461 x 10 ⁵	6.838 x 10 ⁴	4.769 x 10 ⁻²	4.769 x 10 ⁻²

95 9H2 Fab binding constants as provided by analysis of biolayer interferometry data.

96

97

98

99

100

101

102

103
104
105
106
107
108
109

Supplemental References:

- 1) Afonine, P. V., Klaholz, B. P., Moriarty, N. W., Poon, B. K., Sobolev, O. V., Terwilliger, T. C., Adams, P. D., & Urzhumtsev, A. (2018). New tools for the analysis and validation of cryo-EM maps and atomic models. *Acta Crystallographica Section D Structural Biology*, 74(9), 814–840. <https://doi.org/10.1107/S2059798318009324>