

# An Evaluation of the Recycling in Measurements of Photorespiration

Received for publication August 4, 1986 and in revised form December 6, 1986

ALAIN GERBAUD\* AND MARCEL ANDRE

*Laboratoire de Chimie Biologique et de Photophysologie, INRA 78850 Thiverval-Grignon, France (A.G.), and Service de Radioagronomie, Centre d'Etudes Nucléaires de Cadarache, BP 1, 13108 Saint Paul lez Durance, France (A.G., M.A.)*

## ABSTRACT

All measurements of photorespiration and gross photosynthesis in leaves, whether using isotopes or not, are underestimated because of the recycling of O<sub>2</sub> or CO<sub>2</sub>. On the basis of a simple diffusion model, we propose a method for the calculation of the recycling and the corresponding underestimation of the measurements. This procedure can be applied when the stomatal resistance is known, and allows for a correction of certain results in the literature. It is found that measurements of the photorespiratory CO<sub>2</sub> release are usually underestimated by 20 to 100%, which sets the estimated rate of CO<sub>2</sub> photorespired at 30 to 50% of the net photosynthesis in C3 plants under normal conditions. In water stress studies, the correction of the photorespiration is still more important (1.5-3.3) because the stomata are closed more. Analysis of the diffusion of O<sub>2</sub> shows that its recycling is low and that the underestimation of photorespiration with <sup>18</sup>O<sub>2</sub> is negligible.

It is difficult to measure the gas exchanges of photorespiration because they are masked by opposite photosynthetic gas exchanges which occur at a higher rate. Various aspects of the problem have been reviewed by Jackson and Volk (10). One approach is to measure photorespiration by suppressing photosynthesis, either measuring the CO<sub>2</sub> release in CO<sub>2</sub>-free air or the temporary CO<sub>2</sub> outburst upon darkening. Another approach is to use isotopes, mainly <sup>18</sup>O<sub>2</sub> and <sup>14</sup>CO<sub>2</sub>, which allow one to distinguish between photosynthetic and photorespiratory gas fluxes.

Neither of these methods enables one to distinguish between photorespiration and the dark respiration which continues in light. The measured fluxes are always the sum of the two processes, so that the measurements should be corrected for dark respiration in so far as its rate in light is known.

Another drawback of these methods is that they overlook the recycling of gas fluxes. Recycling is the phenomenon whereby CO<sub>2</sub> released by photorespiration is taken up by photosynthesis instead of leaving the leaves, or whereby O<sub>2</sub> released by photosynthesis is taken up by photorespiration. The fluxes that are recycled are 'invisible' from outside the leaf and are therefore not measured. This results in underestimation of the measurements of photorespiration and gross (or true) photosynthesis. The occurrence of recycling has been known for years (18) but estimates of its magnitude vary widely from zero to 100% of the photorespiratory flux (7, 16, 18), so that it is usually disregarded.

The object of this paper is to show that recycling can be calculated—at least a minimum estimation of it—whenever the stomatal resistance is known. This is the case in many recent

studies where stomatal resistance is deduced from water vapor exchange and temperature measurements and is used for the calculation of the internal CO<sub>2</sub> concentration (3, 11).

A simple theory of gas exchanges, taking diffusion in account, is presented and applied to the correction of some measurements of gross photosynthesis and photorespiration found in the literature. The cases of <sup>18</sup>O<sub>2</sub> and <sup>14</sup>CO<sub>2</sub> practically are shown to be quite different, as there is little recycling of O<sub>2</sub>.

## THEORY

Most measurements of gas exchanges in plants refer to the following principles: the plant is enclosed in a chamber and its exchanges are calculated from the variation of gas concentration in the chamber. The plant air spaces are assumed to constitute a homogenous gas compartment, from which CO<sub>2</sub> is taken up by photosynthesis and to which it is supplied by photorespiration. The plant internal air space is separated from the chamber atmosphere by the epidermis; resistance to diffusion is determined by the sum of stomatal and boundary layer resistances.

Plant gas exchanges occur at two levels: between the cell organelles and the air spaces, and between the air spaces and the atmosphere. The problem is that the measurement of gas concentrations in the atmosphere allows the calculation of gas exchanges between plant and atmosphere, which are only the resultant of several physiological fluxes occurring inside the plant, in which we are interested. The supplementary information given by isotopes allows to trace back the internal gas flows, provided that they follow a simple model.

**Measurements with Carbon Isotopes.** Just as the measurement of <sup>12</sup>CO<sub>2</sub> concentration with an IR gas analyzer allows one to measure only the net CO<sub>2</sub> flux  $F$  (Fig. 1) from the chamber to the plant, the measurement of <sup>14</sup>CO<sub>2</sub> with an ionization chamber gives the net <sup>14</sup>CO<sub>2</sub> flux  $F^*$ , which is in this case identical to the gross <sup>14</sup>CO<sub>2</sub> flow, as long as there is no <sup>14</sup>CO<sub>2</sub> coming out of the plant. This condition imposes very fast measurements because the photorespiratory CO<sub>2</sub> becomes labeled after only 30 s of photosynthesis in the presence of <sup>14</sup>CO<sub>2</sub> (16). From the gross flow of <sup>14</sup>CO<sub>2</sub> into the plant, the total gross flow  $GPA$  of CO<sub>2</sub> into the plant can be deduced by a simple proportion. The value obtained differs from the true (or gross) photosynthesis by the amount of PR that is recycled (Fig. 1).

Usually the external concentration of the labeled CO<sub>2</sub>,  $Ce^*$ , is low compared to the unlabeled,  $Ce$ , and the fluxes of <sup>12</sup>CO<sub>2</sub> are dominant. Only the net fluxes of CO<sub>2</sub> into the plant,  $F^*$  and  $F$ , are measurable. A part of photorespiration contributes to the flow of CO<sub>2</sub> into the chloroplasts  $F'$  through recycling and another part is released out of the plant, although the net <sup>12</sup>CO<sub>2</sub> flux is usually into the leaf.

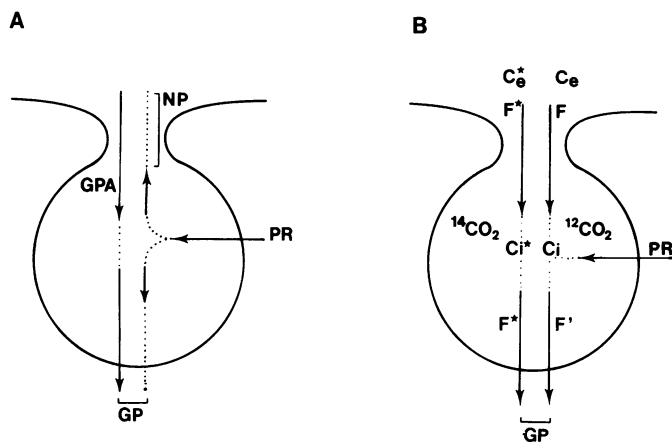


FIG. 1. Scheme of the photosynthetic  $\text{CO}_2$  exchanges inside a leaf. A, Physiologically defined fluxes; B, physical fluxes of the isotopes. The cavity figures all internal air spaces. PR, photorespiration; NP, net photosynthesis; GP, gross photosynthesis; \*, indicates labeled  $\text{CO}_2$ ; F, net  $\text{CO}_2$  flow into the leaf; F', gross (or true) flow of  $\text{CO}_2$ ;  $C_e$ ,  $C_i$ , external and internal  $\text{CO}_2$  concentrations. The size of the arrows is not proportional to the actual fluxes.

The law for gas diffusion can be written for each isotope

$$F^* = \frac{(C_e^* - C_i^*)}{R} \quad (1)$$

$$F = \frac{C_e - C_i}{R} \quad (2)$$

The two isotopes encounter the same resistance to carboxylation:

$$F'/F^* = C_i/C_i^* \quad (3)$$

and in the steady state there is no accumulation of  $^{12}\text{CO}_2$  in the air space:

$$F' = F + PR. \quad (4)$$

With these equations one can calculate the remaining parameters using values of F, F\*, C<sub>e</sub>, C<sub>e</sub>\*, and R as known variables:

$$C_i^* = C_e^* - F^* \cdot R \quad (5)$$

$$C_i = C_e - F \cdot R \quad (6)$$

$$F' = \frac{F^* \cdot C_i}{C_i^*} = F^* \frac{C_e - F \cdot R}{C_e^* - F^* \cdot R} \quad (7)$$

Gross photosynthesis can be obtained by the obvious relations:

$$GP = F^* + F' = F^* \left( 1 + \frac{C_e - F \cdot R}{C_e^* - F^* \cdot R} \right) \quad (8)$$

and  $PR = GP - NP$ . Let us use  $t_e$  and  $t_i$  as the labels for the external and internal  $\text{CO}_2$ , respectively:

$$t_e = \frac{C_e^*}{C_e^* + C_e}, \quad t_i = \frac{C_i^*}{C_i^* + C_i}$$

then

$$GP = \frac{F^*}{t_i} \quad (9)$$

**The Usual Approximation.** In many cases when R or  $t_i$  are not known, the above resolution is not possible. Authors approximate  $t_i$  by  $t_e$  and calculate approximations of GP and PR which we shall call GPA and PRA. The approximation of  $t_i$  by  $t_e$  is equivalent to the assumption that there is no recycling:  $t_i$  is less

than  $t_e$  because of the resistance R to the diffusion between the interior and exterior spaces. Actually there would be no recycling if there were no resistance.

Equation 9 is replaced by

$$GPA = \frac{F^*}{t_e}, \quad (10)$$

hence

$$PRA = \frac{F^*}{t_e} - NP. \quad (11)$$

These equations are well known. Our interest in calculating them with reference to the exact equations 8 and 9 is that we can then recalculate PR and GP as functions of PRA and GPA. The only supplementary information needed is the value of R.

**Calculation of the Corrected Values of Gross Photosynthesis and Photorespiration as Functions of the Approximated Values GPA and PRA.** Comparison of equations 9 and 10 shows that

$$GP = GPA \cdot t_e/t_i. \quad (12)$$

As the isotope concentration is very low ( $C_i^* \ll C_i$ ), the expression of  $t_i$  can be approximated by  $C_i^*/C_i$ . Let us replace  $C_i^*$  and  $C_i$  by their values from equations 5 and 6, then equation 12 can be written as:

$$\begin{aligned} GP &= GPA \cdot t_e \frac{C_i}{C_i^*} = GPA \cdot t_e \frac{C_e - F \cdot R}{C_e^* - F^* \cdot R} \\ &= GPA \frac{C_e - F \cdot R}{C_e^*/t_e - F^* \cdot R/t_e} \end{aligned}$$

or, approximating F by NP:

$$GP = GPA \frac{C_e - R \cdot NP}{C_e - R \cdot GPA}. \quad (13)$$

In a similar way we get

$$PR = PRA \frac{C_e}{C_e - R \cdot GPA}. \quad (14)$$

These equations show clearly that PRA and GPA would be equal to the real values if R was zero. We shall use equations 13 and 14 for recalculating corrected values of PR and GP from the values PRA and GPA found in the literature.

**Analysis of the Oxygen Exchanges.** Because of the analogy between the exchanges of  $\text{O}_2$  and  $\text{CO}_2$ , we can dispense with doing the same series of calculations again. It is enough to replace the  $\text{CO}_2$  fluxes by the corresponding  $\text{O}_2$  fluxes in the equations, keeping in mind that the equivalent of PR is  $GP_0$  and vice versa (Fig. 2), and NP is replaced by  $-NP_0$  because it is counted positively in the opposite direction.

Transforming equations 10 and 11 gives the values for  $\text{O}_2$  uptake and true photosynthesis obtained by neglecting the recycling:

$$PRA_0 = \frac{F_0^*}{t_{e0}} \quad (15)$$

$$GPA_0 = \frac{F_0^*}{t_{e0}} + NP_0 \quad (16)$$

and the true exchanges can be calculated by transforming equations 13 and 14:

$$GP_0 = GPA_0 \frac{C_{e0}}{C_{e0} - R \cdot PRA_0} \quad (17)$$

$$PR_0 = PRA_0 \frac{C_{e0} + R \cdot NP_0}{C_{e0} - R \cdot PRA_0} = PRA_0 \frac{C_{i0}}{C_{e0} - R \cdot PRA_0}. \quad (18)$$

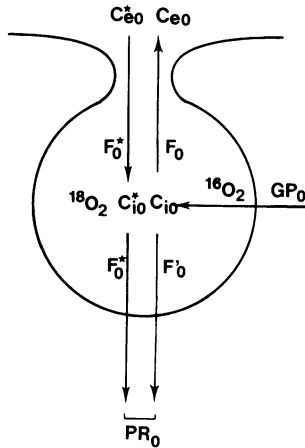


FIG. 2. Diagram of  $O_2$  gas exchanges (indexed 0) when labeled  $O_2$  is used, showing the equivalence with  $CO_2$  (Fig. 1B) by substituting  $PR_0$  for GP and  $GP_0$  for PR.

These equations will be used to compare the errors arising from the recycling of  $O_2$  or  $CO_2$ .

APPLICATION

**Comparison of the Errors Due to the Recycling of  $O_2$  and  $CO_2$ .** Although the formulas for  $O_2$  and  $CO_2$  are almost identical, they lead to different numerical results for the correction ratios  $PR/PRA$  and  $PR_0/PRA_0$ , respectively, equal to  $C_e/(C_e - R \cdot GPA)$  and  $C_{i0}/(C_{e0} - R \cdot PRA_0)$ . The main difference arises from the fact that the  $CO_2$  concentration is small and often of the same order of magnitude as the "gradient"  $R \cdot GPA$  (frequently around  $100 \mu l \cdot L^{-1}$ ), whereas the  $O_2$  concentration is much larger (20% is 606 times  $330 \mu l \cdot L^{-1}$ ) and the gradient term  $R \cdot PRA_0$  is negligible. In other words, the concentration of  $CO_2$  limits its diffusion, but this is not the case with  $O_2$ .

For example, if we assume that the internal  $CO_2$  concentration is  $230 \mu l \cdot L^{-1}$  for an external concentration of  $330 \mu l \cdot L^{-1}$ , the gradient  $R \cdot NP$  is  $100 \mu l \cdot L^{-1}$ . Taking the usual ratio of 1.2 for  $GPA/NP$ ,  $R \cdot GPA$  would be  $120 \mu l \cdot L^{-1}$ , so that the correction for photorespiration would be  $330/(330 - 120) = 1.5$ .

For  $O_2$ , if  $PRA_0$  is about the same as  $NP$  (2, 8) and if the resistance to diffusion is approximately the same as for  $CO_2$ , we get  $R \cdot PRA_0$  near  $100 \mu l \cdot L^{-1}$ , but  $C_e$  is equal to 20.6% or 206000  $\mu l \cdot L^{-1}$  hence  $C_{i0} = C_{e0} + R \cdot NP_0 = 206100 \mu l \cdot L^{-1}$  and  $C_{i0}/(C_{e0} - R \cdot PRA_0)$  will be very near one with an error of 5%. At worst.

The difference between  $O_2$  and  $CO_2$  fluxes can be illustrated if we represent the  $CO_2$  and  $O_2$  exchanges with unidirectional fluxes drawn on a realistic scale (Fig. 3). The unidirectional fluxes  $F_i$  and  $F_e$  follow the law of diffusion and are respectively equal to  $C_i/R$  and  $C_e/R$ . Their difference  $(C_i - C_e)/R$  is the net flux. If we consider that the ratio of recycling is the probability that a  $CO_2$  molecule coming from photorespiration is taken up in  $GP$  rather than in  $F_i$ , it is obvious that this probability is small if the unidirectional flux  $F_i$  is large. As  $F_i$  equals  $C_i/R$ , the probability of recycling is inversely related to the internal concentration. We can calculate that even in the less favorable case when the external  $O_2$  concentration is reduced (for example to 1%), the recycling of  $O_2$  will be negligible.

RESULTS AND DISCUSSION

We have applied the preceding calculations to the correction of measurements of gross photosynthesis and photorespiration found in the literature, where stomatal conductances were recorded. The data analyzed concern measurements with  $^{14}CO_2$

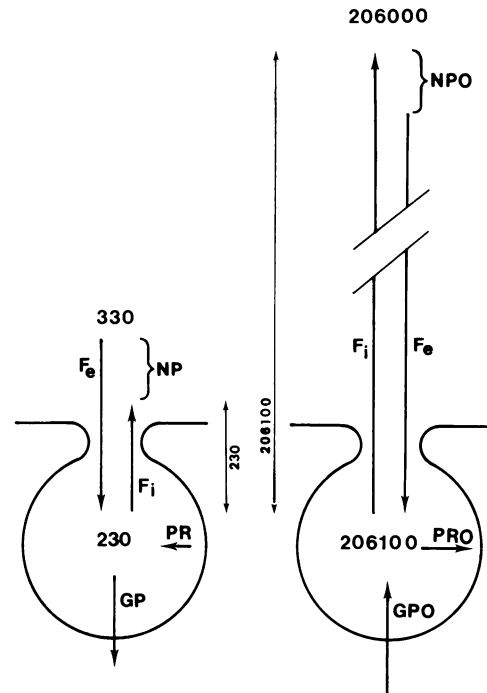


FIG. 3. Photosynthetic gas fluxes drawn at a realistic scale (for normal conditions and a C3 plant). Diffusional fluxes  $F_i$  and  $F_e$ , which are proportional to gas concentrations, are overwhelming in the case of  $O_2$ , compared to photosynthetic and photorespiratory fluxes. The consequence is that the recycling of  $O_2$ , proportional to  $FP_0/F_i$ , is negligible.

published by Ludwig and Canvin (17), Lawlor and Fock (14), Krampitz *et al.* (13), Fock *et al.* (5), and measurements of  $CO_2$  evolution in  $CO_2$  free air of Lawlor (15).

Measurements using  $^{14}CO_2$  are corrected on the basis of formulas 13 and 14:

$$PR = PRA \cdot C_e / (C_e - R \cdot GPA),$$

$$GP = GPA \cdot C_i / (C_e - R \cdot GPA)$$

where  $PRA$  and  $GPA$  are the underestimated measurements of photorespiration and gross photosynthesis;  $R$  is the stomatal + boundary layer resistance to  $CO_2$  diffusion, and  $C_e$  the outer concentration of  $CO_2$ .  $C_e$  is usually expressed in  $\mu l/L$ . For homogeneity of units,  $R \cdot GPA$  was multiplied by a conversion factor (1.4 when  $GPA$  is in  $mg CO_2 dm^{-2} h^{-1}$ ). Table I gives the results of the corrections. Analysis of the measurements of photorespiration in *Helianthus annuus* by Fock *et al.* (5) (not shown), gives similar results: the correction factor is around 1.6 independent of the level of  $CO_2$  or temperature.

We also applied similar reasoning to the correction of an example of measurement of photorespiration by the  $CO_2$  release in  $CO_2$ -free air. We took data from Lawlor (15) on wheat submitted to water stress. Since both the stomatal ( $c_s$ ) and mesophyll ( $c_m$ ) conductances were given, the calculation of the correction is straightforward, because in  $CO_2$  free air the ratio of recycled to released  $CO_2$  flux is  $c_m/c_{s+a}$  where  $c_{s+a}$  is the conductance of the stomata with the air layer. It is equal to  $c_s \cdot c_a / (c_s + c_a)$ .  $PRA$  is the flux of released  $CO_2$  and

$$PR = PRA (1 + c_a/c_{s+a}).$$

The results are shown in Table II.

The underestimation due to the recycling ranges from 1.15 to 3.3, which shows that recycling should rarely be considered as negligible. The accuracy of its calculation is mostly dependent on the accuracy of the diffusion resistance  $R$ , especially in the case of severe water stress, when  $R$  is large and the denominators

Table I. Correction of the Recycling in Measurements of Photorespiration with  $^{14}\text{CO}_2^a$   
Water stress experiments were made using PEG 4000 as osmoticum.

Type of Experiment	$R$	$NP$	$PRA$	$GPA$	$PR$	$GP$	$\text{CO}_2$	Correction Factor ( $PR/PRA$ )	
SUNFLOWER, $C_e$ , $\mu\text{L}$									
Variation of $\text{CO}_2$	50	0.68	1.2	5.8	7	6.7	7.9	49	1.15
Concentration	130	0.73	12	6.5	17.5	6.4	18.4	118	1.16
$700 \mu\text{E} \cdot \text{m}^{-2} \text{s}^{-1}$	250	0.78	26	6	32	7	33	222	1.16
Data from Ref. 17									
SUNFLOWER, $\Psi$ , bar	-6	2.24	11.6	5.0	16.9	6.1	17.7	294	1.22
Water stress	-9.5	5.8	9	3.2	11.9	4.9	13.9	257	1.5
$400 \mu\text{E} \cdot \text{m}^{-2} \text{s}^{-1}$	-13.5	6.8	6.5	3.2	8.3	4.3	10.8	268	1.35
Data from Ref. 14									
SUNFLOWER	-6	2.9	30.8	8	39.6	15.8	46.6	205	2
Water stress	-9	3.6	24.5	8	31.7	15.8	40.3	205	2
$1000 \mu\text{E} \cdot \text{m}^{-2} \text{s}^{-1}$	-13	6.8	12.7	7	19.8	16.7	29.4	210	2.4
	-15	12.9	6.4	6.3	12.6	20.8	27.2	216	3.3
	-18	51.8	1.3	4.7	6.3	?	?	210	?
Data from Ref. 13									
BEAN	-5.5	3.7	16.6	7.1	23.7	11.4	28	245	1.6
Water stress	-6.4	5.8	13.5	6.3	19.8	12.3	26.5	215	1.9
$1000 \mu\text{E} \cdot \text{m}^{-2} \text{s}^{-1}$	-8	7.1	9.2	5.5	14.7	10.1	19.3	240	1.8
Data from Ref 13	-9	12.5	6.5	4.7	11.2	11.6	18.1	216	2.5

<sup>a</sup>  $R$ , stomatal + boundary layer resistance  $\text{s} \cdot \text{cm}^{-1}$ ;  $NP$ , net photosynthesis. All gas exchanges are in  $\text{mg CO}_2 \cdot \text{dm}^{-2} \text{h}^{-1}$ ;  $PRA$ , apparent photorespiration ( $\text{CO}_2$  evolution calculated without correction of the recycling);  $GPA$ , apparent gross photosynthesis;  $PR$ , corrected photorespiratory  $\text{CO}_2$  evolution;  $GP$ , corrected gross photosynthesis;  $C_e$ , external  $\text{CO}_2$  concentration;  $C_i$ , internal  $\text{CO}_2$  concentration;  $\Psi$ , leaf water potential.

Table II. Correction of the Recycling in Measurements in  $\text{CO}_2$ -Free Air<sup>a</sup>

Symbols are as in Table I, except  $PIB$ , post-illumination burst;  $c_s$ , stomatal conductance;  $c_m$ , mesophyll conductance. Calculations are made with the hypothesis that the boundary layer resistance is  $ra = 0.6 \text{ s} \cdot \text{cm}^{-1}$ . The data are from wheat from Ref. 15.

Bar	$c_s$	$c_m$	$PIB$	$PRA$	$PR$	Corr	$NP$
-5	0.52	0.27	10.1	5.7	10	1.78	27.5
-8	0.29	0.23	8.6	4.6	9.2	2.0	21.2
-12	0.13	0.15	6.6	3.2	7.4	2.3	13
-16	0.065	0.072	4.5	1.7	3.7	2.2	4.3

in formulas 13 and 14 are small. Krampitz *et al.* (13) give standard deviations for conductance measurements. Although deviations are smaller at low water potentials where conductances are low (and resistances high), the relative deviations become very large, which causes a high relative uncertainty in the correction factors. Errors on  $R$  in other conditions would cause only moderate errors on the correction factors (2.5% at -6 bar in Krampitz' data for sunflower). Measurements made by Ludwig and Canvin (17) appear as the least underestimated (correction factor 1.15) because of the very low boundary layer + stomatal resistance of 0.78. Minimal values of stomatal resistance found in the literature range from 0.6 to 2.2 for young plants, but the most frequent values are between 2 and 4 (12), to which should be added the value of the boundary layer resistance; this value depends on experimental conditions and common ranges from  $1.8 \text{ s} \cdot \text{cm}^{-1}$  (14) to  $0.6 \text{ s} \cdot \text{cm}^{-1}$  (15). The range of variation of the diffusion resistances is considerable, but the recycling varies certainly much less because it is the product  $R \cdot GPA$  which is the variable factor in formula 14, and  $GPA$  is usually negatively correlated with  $R$ . Körner *et al.* (12) showed a linear relation between the photosynthetic capacity of well-watered leaves and their stomatal conductance.

Measurements of the rate of  $\text{CO}_2$  evolution in  $\text{CO}_2$ -free air suffer from similar underestimation values (Table II).

The literature offers few estimations of the recycling. Samish and Koller (18) describe a model and calculate corrections for

the recycling ranging from 1.3 to 2.3. However they could not make the full calculation of the correction factor because they do not dispose of photorespiration measurements. In a later paper (19), factors of 2 to 3 are given for measurements of photorespiration in soybean by  $\text{CO}_2$  evolution in  $\text{CO}_2$ -free air. The values are higher than ours due to higher stomatal resistances. D'Aoust and Canvin (4) have estimated, from the difference between the rate of  $\text{CO}_2$  evolution measured with  $^{14}\text{CO}_2$  and the post-illumination burst ( $PIB$ ) which is little subject to recycling, that the recycling would be around 30 to 40%, but this remained hypothetical, as the  $PIB$  itself can be underestimated for other reasons. It may be noticed that the  $PIB$  gives values that are probably quite near the true rate of photorespiration, (Table II).

The validity of the correction for recycling is dependent on the validity of the model for the diffusion of  $\text{O}_2$  and  $\text{CO}_2$  (Fig. 1). The main objection to that scheme is that it overlooks the direct recycling of dissolved gases inside the cells. This type of recycling should be added to recycling through the air spaces, which would make the underestimation of the measurements even greater. Although we do not know the internal diffusion resistances and concentrations, we can attempt to evaluate its magnitude compared with air-space recycling. The probability for a  $\text{CO}_2$  molecule released from a mitochondria of reaching directly a chloroplast, instead of going to the air space, is pro-

portional to the ratio

$$\frac{\text{'Diffusion to the air space' resistance}}{\text{'Diffusion to the chloroplast + carboxylation' resistance}}$$

The following argument suggests that this ratio would not be considerable. Although diffusion through liquid is much slower than through the gas phase, this is partly compensated for CO<sub>2</sub> by a high solubility (due to the formation of HCO<sub>3</sub><sup>-</sup>), and distances are short, so that the carboxylation resistance would be dominant. The same argument holds for O<sub>2</sub>: the fact that the internal concentration is nearly equilibrated with the external air means that the flux is limited by the photosynthetic capacity of the chloroplasts, not by the diffusion rate.

In spite of the number of untested assumptions involved, we shall consider that the direct recycling is small and that the error committed by neglecting it does not affect the overall validity of the calculations presented. In any case the corrections calculated are minimal ones.

### CONCLUSION

A primary consequence of the proposed correction is that it increases the ratio of photorespiration (as CO<sub>2</sub> evolution) to net photosynthesis. Whereas 20 to 30% of *NP* is a commonly cited figure (1, 5, 14, 17), corrected ratios would be in the range from 30 to 50% in nonstress conditions. These figures fit better with measurements of O<sub>2</sub> uptake made with <sup>18</sup>O<sub>2</sub> (9) and the accepted stoichiometry of the glycolate pathway. Tolbert (20) proposed a ratio of O<sub>2</sub> uptake to CO<sub>2</sub> evolution of three, and O<sub>2</sub> uptake would be 80 to 100% of *NP*. If this O<sub>2</sub> uptake is composed of about 15% of dark respiration, and 65 to 85% of photorespiratory O<sub>2</sub> uptake, then photorespiratory CO<sub>2</sub> evolution would be 20 to 30% of *PN*. Adding 15% for dark respiration, we come to a total CO<sub>2</sub> evolution of 35 to 45% of *PN* which fits well with our evaluation of photorespiration.

It is a problem that the CO<sub>2</sub> evolution does not seem to vary or may even increase with the CO<sub>2</sub> concentration (1, 5, 17). It has been hypothesized that this is due to decrease of the recycling with increasing CO<sub>2</sub> concentration (1), but Table I shows that this is not the case. This is also confirmed by the analysis of the results of Fock *et al.* (5) (data not shown), which shows that the correction factor is not affected by the CO<sub>2</sub> level. As it is known that O<sub>2</sub> uptake increases with decreasing CO<sub>2</sub> concentration (2, 8), photorespiratory CO<sub>2</sub> evolution should vary similarly, according to the model of the glycolate pathway. It follows that at high CO<sub>2</sub> level there is more CO<sub>2</sub> evolution than predicted by the model, but less at low CO<sub>2</sub> level. The last point could be explained by the occurrence of other O<sub>2</sub> uptake reactions, such as the Mehler reaction (6).

Although the correction changes the values taken for photorespiration appreciably, and to a lesser degree the values of gross photosynthesis, it does not change very much the qualitative aspects of the response to water stress. The decrease of *PR* with water stress in Lawlor's results is conserved, but the slight decrease of *PR* in the Krampitz *et al.* (13) data is changed to a slight increase. In all cases the increase in the ratio of *PR* to

photosynthesis cannot be due to the variation of the internal CO<sub>2</sub> concentration, as this is quite stable. How this ratio can be modified otherwise than through the CO<sub>2</sub> concentration is an open question. It can be envisioned that the dark respiration increases during stress, but it would require rates of dark respiration as high as 30% of normal net photosynthesis to explain the conservation of the CO<sub>2</sub> evolution when photosynthesis goes to zero. On the other hand, in water stressed plants, or whenever stomata are closed, the recycling could be considerable, causing an important underestimation of measurements of photorespiration; at the same time correction of the results becomes unfeasible because the uncertainty of values of high stomatal resistances is very large.

It appears that the problems of the measurement, as well as of the nature, of photorespiration are still far from solved.

### LITERATURE CITED

1. BRAVDO B, DT CANVIN 1974 Evaluation of the rate of O<sub>2</sub> production by photorespiration. In M Avron, ed, Proceedings of the third International Congress on Photosynthesis, Rehovot, Israel. Elsevier, Amsterdam
2. CANVIN DT, JA BERRY, MR BADGER, H FOCK, CB OSMOND 1980 Oxygen exchange in leaves in the light. *Plant Physiol* 66: 302-307
3. COWAN IR 1977 Stomatal behaviour and environment. *Adv Bot Res* 4: 117-228
4. D'AOUST AL, DT CANVIN 1974 Caractéristiques du <sup>14</sup>CO<sub>2</sub> dégagé à la lumière et à l'obscurité par des feuilles de haricot, de radis, de tabac et de tournesol pendant et après une photosynthèse en présence de <sup>14</sup>CO<sub>2</sub>. *Physiol Veg* 12: 545-560
5. FOCK H, K KLUG, DT CANVIN 1979 Effect of carbon dioxide and temperature on photosynthetic CO<sub>2</sub> evolution in sunflower leaves. *Planta* 145: 219-223
6. FURBANK RT, MR BADGER, CB OSMOND 1982 Photosynthetic oxygen exchange in isolated cells and chloroplasts of C3 plants. *Plant Physiol* 70: 927-931
7. GAUDILLERE JP 1981 CO<sub>2</sub> evolution outside leaves during photosynthesis. In G Akoyunoglou, ed, Photosynthesis. 4. Regulation of Carbon Metabolism. Balaban International Science Service, Philadelphia
8. GERBAUD A, M ANDRE 1979 Photosynthesis and photorespiration in whole plants of wheat. *Plant Physiol* 64: 735-738
9. GERBAUD A, M ANDRE 1980 Effect of CO<sub>2</sub>, O<sub>2</sub> and light on photosynthesis and photorespiration in wheat. *Plant Physiol* 66: 1032-1036
10. JACKSON WA, RJ VOLK 1970 Photorespiration. *Annu Rev Plant Physiol* 21: 385-432
11. JARVIS JP 1971 The estimation of resistances to carbon dioxide transfer. In Z Sestak, J Catsky, PG Jarvis, eds, *Plant Photosynthetic Production. Manual of Methods*. Dr W Junk NV, The Hague, pp 566-631
12. KÖRNER CH, JA SCHEEL, A BAUER 1979 Maximum leaf diffusive conductance in vascular plants. *Photosynthetica* 13: 45-82
13. KRAMPITZ MJ, K KLUG, HP FOCK 1984 Rates of photosynthetic CO<sub>2</sub> uptake, photorespiratory CO<sub>2</sub> evolution and dark respiration in water-stressed sunflower and bean leaves. *Photosynthetica* 18: 322-328
14. LAWLOR DW, H FOCK 1975 Photosynthesis and photorespiratory CO<sub>2</sub> evolution of water-stressed sunflower leaves. *Planta* 126: 247-258
15. LAWLOR DW 1976 Water stress induced changes in photosynthesis, photorespiration, respiration and CO<sub>2</sub> compensation concentration of wheat. *Photosynthetica* 10: 378-387
16. LUDWIG JL, DT CANVIN 1971 An open gas-exchange system for the simultaneous measurement of the CO<sub>2</sub> and <sup>14</sup>CO<sub>2</sub> fluxes from leaves. *Can J Bot* 49: 1299-1313
17. LUDWIG LJ, DT CANVIN 1971 The rate of photorespiration during photosynthesis and the relationship of the substrate of light respiration to the products of photosynthesis in sunflower leaves. *Plant Physiol* 48: 712-719
18. SAMISH YB, D KOLLER 1968 Photorespiration in green plants during photosynthesis estimated by use of isotopic carbon. *Plant Physiol* 43: 1129-1132
19. SAMISH YB, JE PALLAS JR, GM DORNHOFF, RM SHIBLES 1972 A reevaluation of soybean leaf photorespiration. *Plant Physiol* 50: 28-30
20. TOLBERT NE 1980 Photorespiration. In DD Davies, ed, *The Biochemistry of Plants*, Vol 2. Academic Press, New York