

**Supplemental information**

**Cell-type-specific regulation of APOE and CLU  
levels in human neurons  
by the Alzheimer's disease risk gene SORL1**

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# Supplemental Information

## Supplemental Figure and Table Legends

### Supplemental Figure legends

**Supp. Figure 1. Overview of iPSC differentiation protocols.** Schematic of (A) iN, (B) iMGL, (C) iA, and (D) iEC differentiation protocol used in this study. For differentiation of iNs (neurons) and iAs (astrocytes), we used lentiviral transduction of cell type specific transcription factors to directly differentiate iPSCs into respective cell types. For differentiation of iMGLs (microglia) and iECs, non-viral differentiation methods were used. (E) Frequency distribution of the standard deviation of the log<sub>2</sub> RNA expression level of all genes detected by RNAseq across cell type. (F) Shown are heatmaps of differentially expressed genes (DEGs) for a subset of pathways enriched in iNs, iAs, and/or iMGLs. Also shown are protein-level results for each pathway generated using TMT-MS. See also Supplemental Table S1 for full data set.

**Supp. Figure 2. Expression of LOAD GWAS genes in SORL1 WT/KO iNs, iAs, iMGLs.** (A) Heatmap of RNA expression levels of LOAD GWAS genes in iNs, iAs, and iMGLs. (B) Table of known LOAD GWAS genes differentially expressed in SORL1 WT/KO neurons, astrocytes, or microglia. (C-O) RNA levels (TPM) of LOAD GWAS genes in 5 different cell types (Only genes that are differentially expressed with SORL1 null (at least in one cell type) are graphed).

**Supp. Figure 3. Generation of SORL1 KO iPSCs using CRISPR/Cas9.** (A) Table showing the iPSC line source information. (B) Table with the information on the gRNAs used to target SORL1, and the edited sequence results for each clone used in this paper. (C) Sequence chromatogram of Line 2 WT and line 2 KO lines. (D-E) Table showing all the potential off-target loci identified by IDT CRISPR-Cas9 guide RNA design checker for (D) Cell line 1 and (E) Cell line 2. All potential off-target loci were sequenced and confirmed to be wild type for each of the lines. \* For line#2, WT line showed a trend towards higher chromosome 20 dosage. Because this may indicate a low level of chromosome 20 duplication, we examined the RNAseq data from the resultant neurons. Examining all DEGs in our dataset in the line 2 comparison, 8.0% of genes detected were differentially expressed based on a cutoff of  $q < 0.05$ . Within chromosome 20, only 6.5% of the genes detected were DEGs with the same cutoff, and only 24 of these were upregulated in wild type compared to SORL1 KO. These results suggest that wild type neurons in line 2 do not appear to express higher levels of chromosome 20 genes compared to their paired KO neurons.

**Supp. Figure 4. SORL1 KO neurons show increased A $\beta$  levels without altering APP levels.** (A) Representative western blot of CLU protein expression in SORL1 WT and KO iNs. (B-C) Quantification of (B) CLUa chain/GAPDH and (C) CLU Precursor/GAPDH protein expression. (D-E) Representative western blot of APP protein expression in SORL1 WT and KO iNs in cell line 1 and 2. (F-J) Quantification of APP/GAPDH in iNs. (G,K) A $\beta$ 38 levels, (H,L) A $\beta$ 40 levels, and (I,M) A $\beta$  42/40 ratio in media generated from SORL1 WT and KO iNs in cell lines 1 and 2. Data show mean +/- SEM from three independent differentiations, for each differentiation, 3 wells were used for each group. Values are normalized to WT for each pair. Unpaired student's t test (two tailed).

**Supp. Figure 5. Loss of APOE does not affect SORL1 levels and p-tau levels in iNs.** (A) Representative western blot of an isogenic set of iNs with APOE  $\epsilon$ 3/ $\epsilon$ 3, APOE  $\epsilon$ 4/ $\epsilon$ 4, and APOE KO genotypes. (B) Quantification of SORL1/GAPDH and (C) p-tau/tau. (D) Representative western blot of iNs that received APOE shRNA KD transduction. (E) Quantification of secreted APOE in the media, (F) SORL1/GAPDH, and (G) p-tau/tau. Data show mean +/- SEM from three independent differentiations, for each differentiation, 2-3 wells were used for each group. Values are normalized to APOE  $\epsilon$ 3/ $\epsilon$ 3 iNs or iNs with empty virus. One-way ANOVA with Tukey's multiple comparisons test.

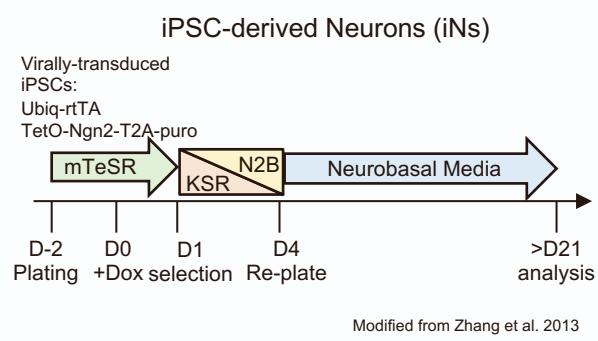
**Supp. Figure 6. Interrogation of candidate pathways associated with SORL1 loss-of-function.** (A) Total A $\beta$  levels after 72hr treatment of d17 iNs with 10uM or 20uM R33. Elevated A $\beta$  levels in SORL1 KO iNs are partially rescued with 20 uM R33. (B) A $\beta$ 42/40 is not altered with R33 treatment. (C) Total A $\beta$  levels in media of iNs after 72hr treatment with 100mM trehalose treatment. Elevated A $\beta$  levels in SORL1 KO iNs are not rescued with trehalose. (D) A $\beta$ 42/40 is not altered with trehalose treatment. Data show mean +/- SEM from three differentiations, n=3 per differentiation for each line. Values are normalized to WT iNs treated with vehicle. One-way ANOVA with Tukey's multiple comparisons test.

**Supp. Figure 7. SORL1 and APOE expression in iNs and iAs based on SORL1 and APOE genotype.** (A,D) Western blot of iN d21 protein lysates and (G) iA d21 protein lysates from ROSMAP cohort. (B,C,E,F) SORL1 and APOE protein expression normalized to GAPDH categorized based on diagnosis. Data show mean +/- SEM. One-way ANOVA with Tukey's multiple comparisons test. (H,J) SORL1 protein levels and (I,K) APOE protein levels in iNs and iAs western blots from the ROSMAP cohort categorized based on APOE genotype. (L,N) SORL1 protein levels and (M,O) APOE protein levels in iNs and iAs western blots from the ROSMAP cohort categorized based on SORL1 5' haplotype. Data show mean +/- SEM. One-way ANOVA with Tukey's multiple comparisons test.

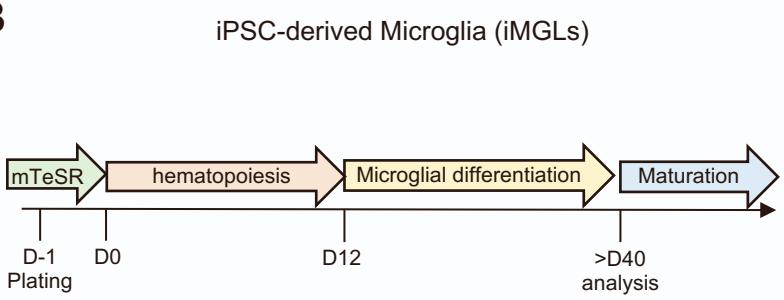
**Supp. Figure 8. Association of SORL1, APOE, and CLU RNA levels in ROSMAP post-mortem brain tissue.** (A,B) RNAseq from medial frontal cortex brain lysates of ROSMAP cohort. Shown are Pearson correlations between (A) SORL1 and APOE RNA levels ( $r=0.03$ ,  $p=0.28$ ) and (B) SORL1 and CLU RNA levels ( $r=0.47$ ,  $p=0.0001$ ).

# Supplemental Figure 1

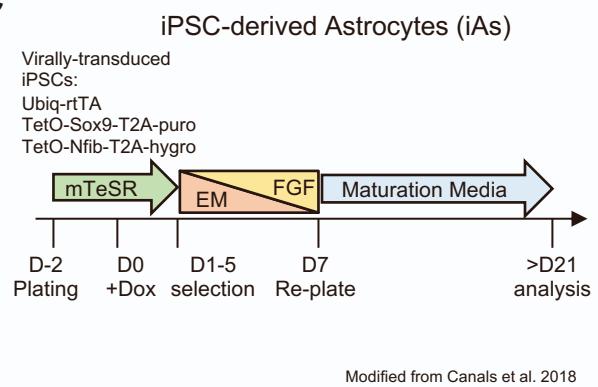
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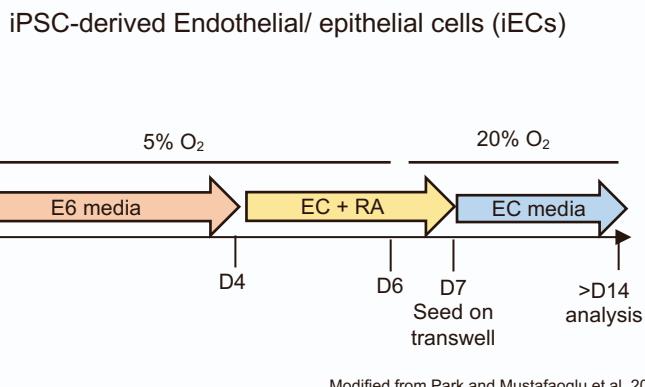
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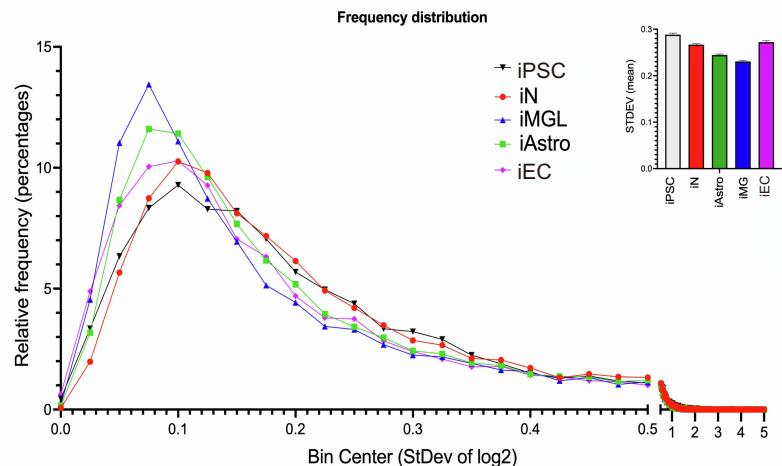
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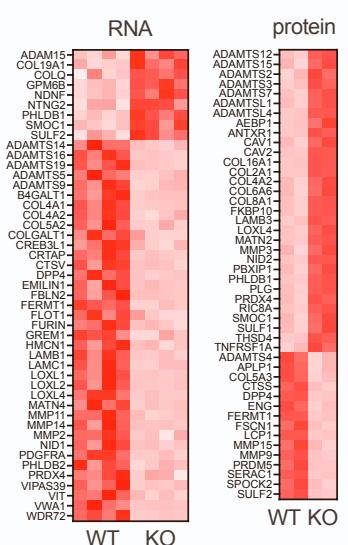


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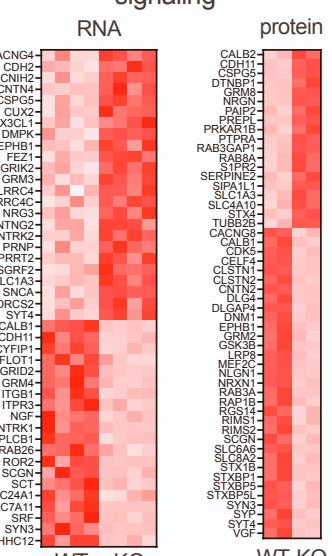


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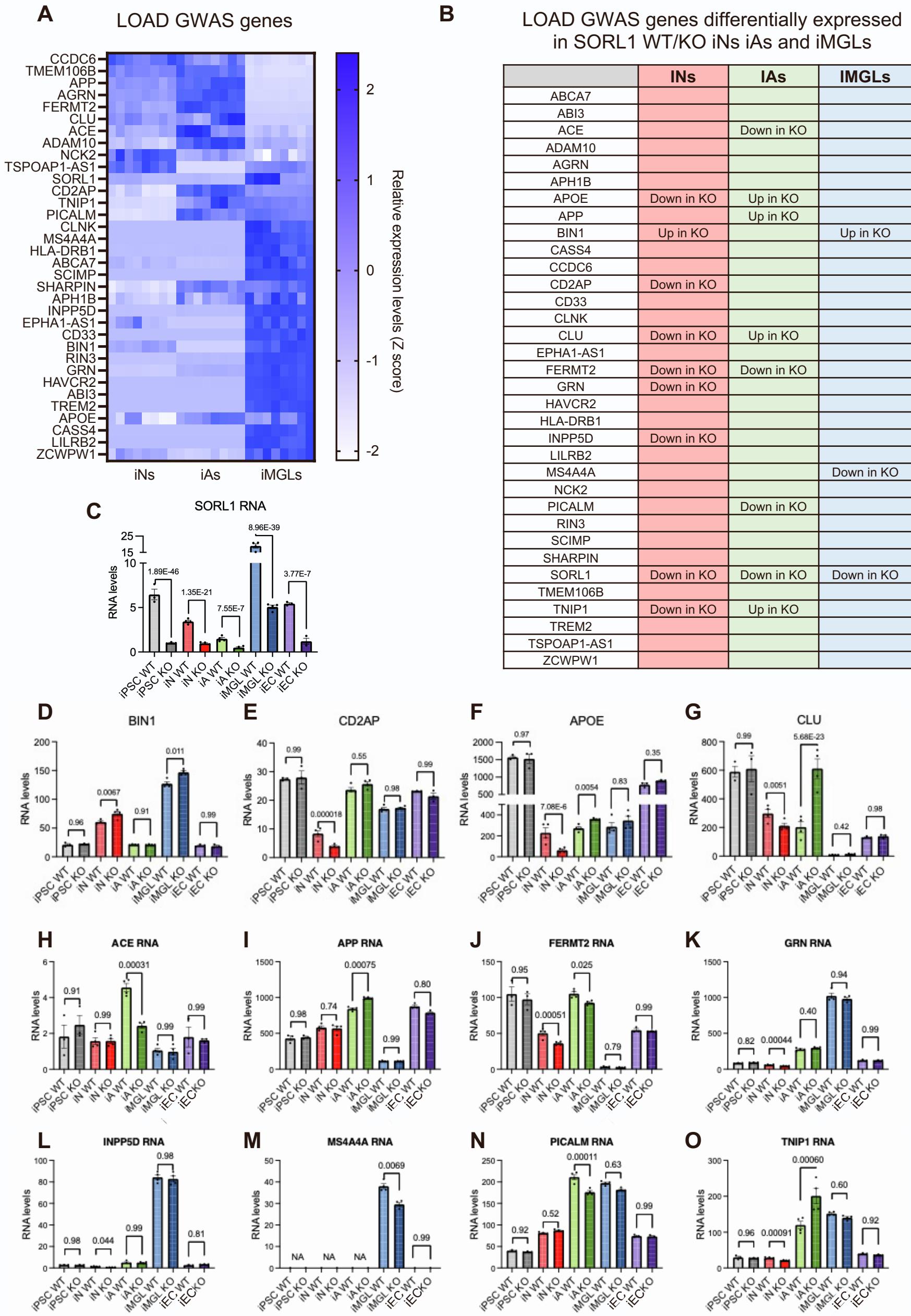
## Neurons



## regulation of trans-synaptic signaling



## Supplemental Figure 2



## Supplemental Figure 3

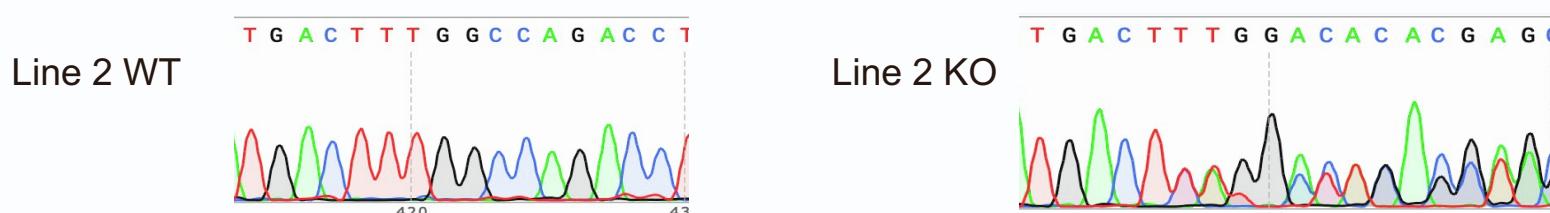
**A**

Line name	iPSC Source	Sex	APOE genotype	Ethnicity	Publication
Line 1	Young lab, University of Washington	Male	ε3, ε4	>90% European ancestry (Levy et al. 2007)	Young et al. 2015 Mishra et al. 2022 Coriell Institute ID: GM25430
Line 2	Young-Pearse lab, Brigham and Women's Hospital	Female	ε3, ε3	>90% European ancestry	Muratore et al. 2014 Muratore et al. 2017

**B**

	SORL1 gRNA	Edited sequence	Genetic Analysis (Stem Cell Technologies Genetic Analysis kit)	Publication
Line 1 WT	ATTGAACGACATGAACCCTC	None	no significantly detected karyotype abnormalities	Knupp et al. 2020
Line 1 KO	ATTGAACGACATGAACCCTC	1 bp deletion	no significantly detected karyotype abnormalities	Knupp et al. 2020
Line 1 G511R	CTCTTGCATTTAGGCTCAG	G511R	no significantly detected karyotype abnormalities	Mishra et al. 2022b
Line 2 WT	ATGTTCCCTGAATCATGATCC	None	no significantly detected karyotype abnormalities*	Generated for this study
Line 2 KO	ATGTTCCCTGAATCATGATCC	16 bp deletion	no significantly detected karyotype abnormalities*	Generated for this study

**C**



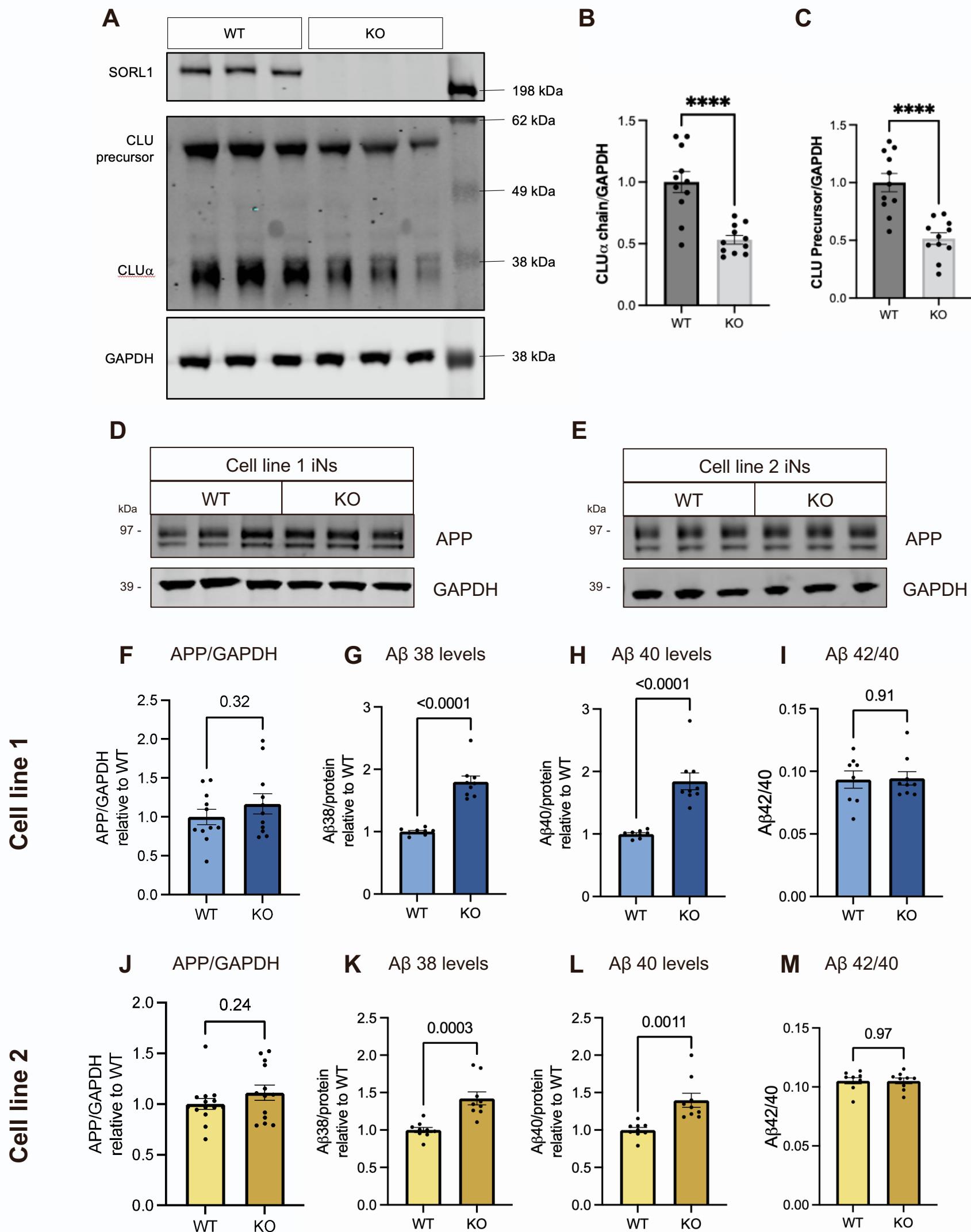
**D**

PCR Primer Sequences	Gene	Potential off-target sequence	PAM	Line1 WT	Line1 KO
CAT CTA ACT TCC AGC CTC TTC TC	SLC22A9	ACTGAAATACATGAACCGTC	GAG	WT	WT
AGA ACC CAA TTC TCC CTT TCC					
CCA TGA TCT CCT CAA GCT GAA A	SECISBP2	CTTGAACAAAATGAAGCCTC	AAG	WT	WT
GAG ACA ACT GGG AAG CCA AA					
AGC ACC TAT GTT GTT CTC ACT C	LOC107984387	TCTGAACGAGATGAACCTTC	CAG	WT	WT
GTC AGC CTG AAC TCT CCT TTC					
GCG ATC TCT GGT CGA GAA TAA A	E2F8	ATTGGACTACTTGAACCATC	GGG	WT	WT
CCA GGT CTT TAC AGA GGC TAA C					
TGG AAG GAA GCA GTT TGG AG	LOC105375202	ATTGAACCAC-TGAACGCTC	CAG	WT	WT
GAA CGA CAC CTC TGG GAT TAA A					

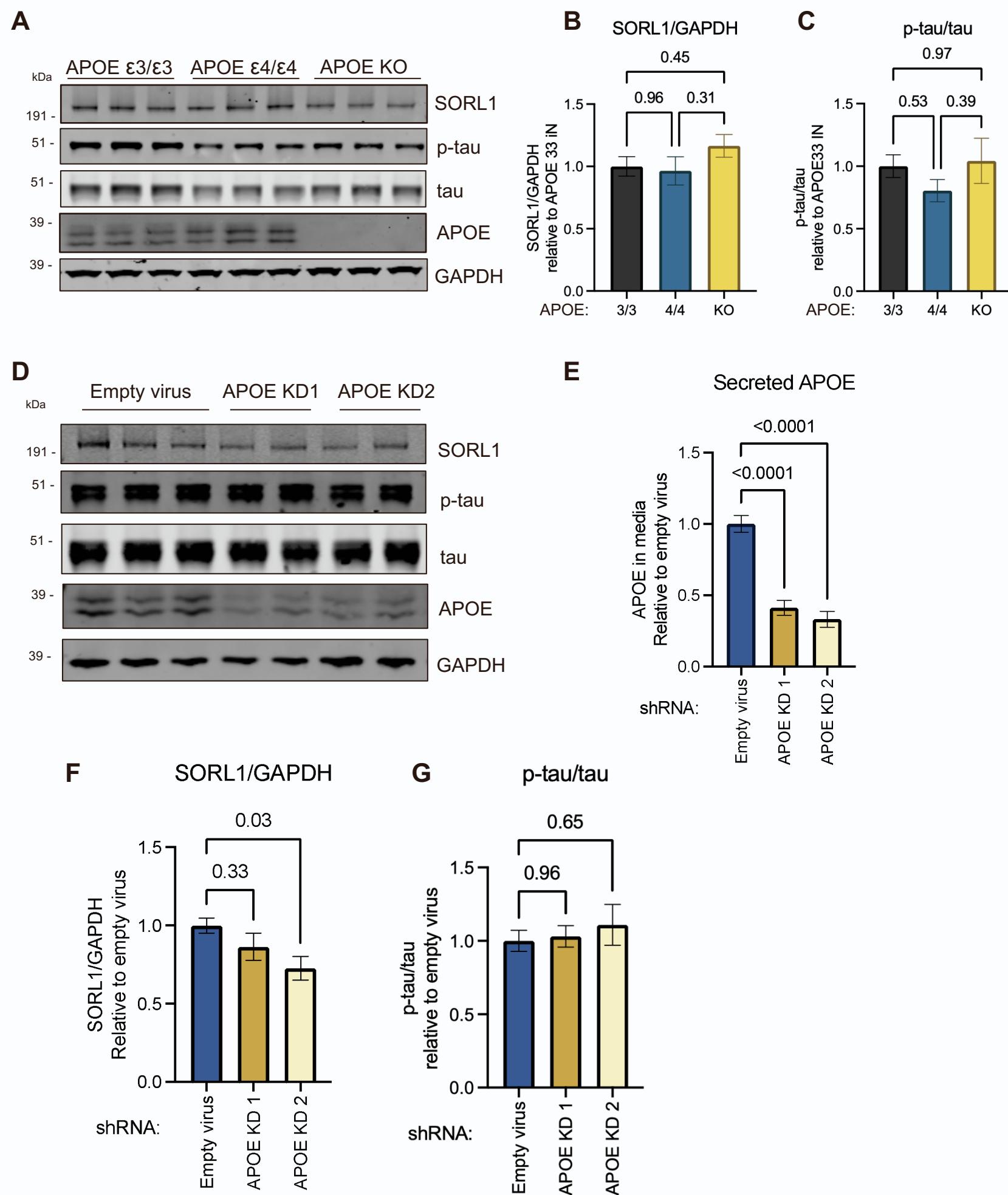
**E**

PCR Primer Sequences	Gene	Potential off-target sequence	PAM	Line2 WT	Line2 KO
CAG GGC CAC ATA GTG AGT TAA G	TNFSF15	GTCTTCCTGAACCATAATCC	TGG	WT	WT
GAG TAT CTC CAG GGA CTG AGA A					
CCT AGC CCT CTT TCC TCT GAT A	ZNF428	ACGATCCTGAAT-ATGATCC	TGG	WT	WT
GTT CCC TCC TCC TCC TCT T					
GAA GCC CTC CAC TTA CAA TCA	GPNMB	ATG-TCCTGATTATGATCC	TAG	WT	WT
TCA CTG CGA AAC CTT ACC TAA A					
CCG TTC TAT CCT TTC CCG TAT C	SLX4IP	CTGTTCCCTGTCCCATGATCC	CAG	WT	WT
GAG AGC TGC TTC TTG GTC TAT C					
TGG CAA ACT GGA TGA GGA TTA C	DICER1	ATGTTCCCTG-ATCATACTCC	AGG	WT	WT
TAG CAC ATT GCA TCC CAA GAG					
GTT CAG GAG GAA GGT GTG ATG	MRM3	AAGTTCCCTGAAGCATCATCC	TGG	WT	WT
TGA TCA GCT GTG AGT GAT GTG					
ACA GAG GAT GTG TGG GAA AG	OSBP	ATGCCCTGAATCATAATCT	TAG	WT	WT
CGC CTG AGG AAG TCC ATA AA					
GGG CTA TCT CTA AGG TGC TAC T	ADAMTS19	ATGTTCATGAA-CATGATCA	AAG	WT	WT
CAG GGA AAT GGG TGG ATC AAT					

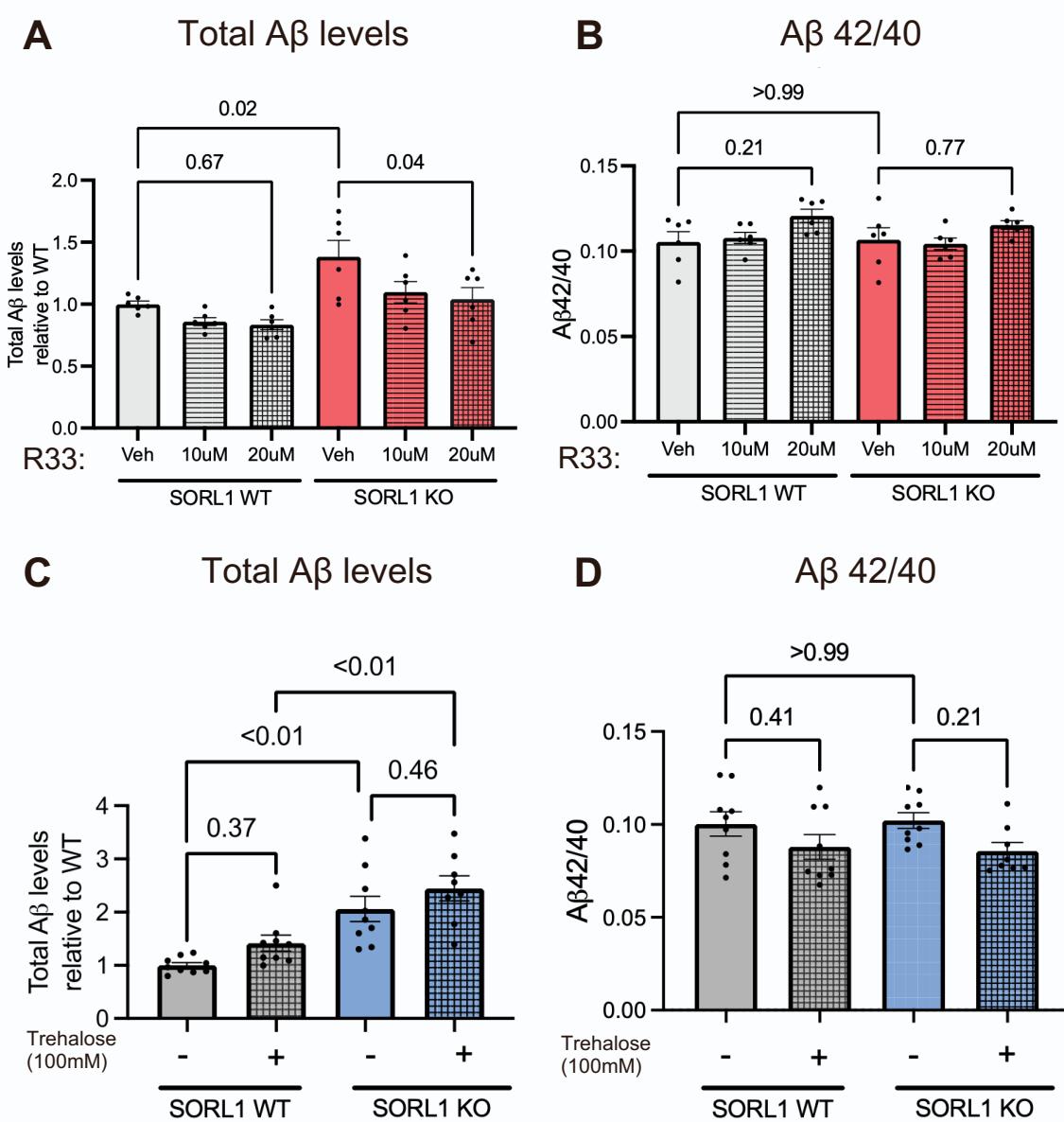
## Supplemental Figure 4



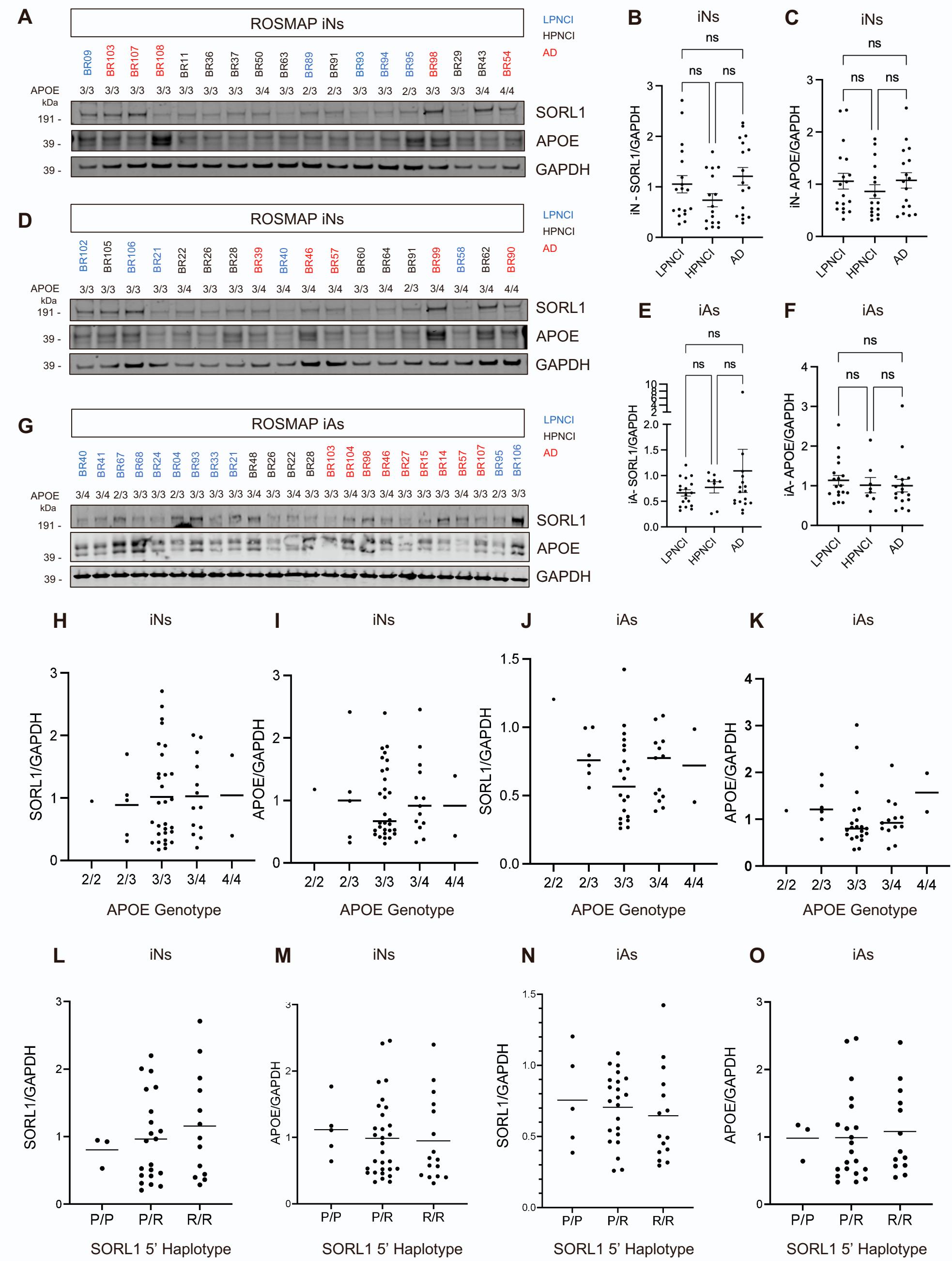
## Supplemental Figure 5.



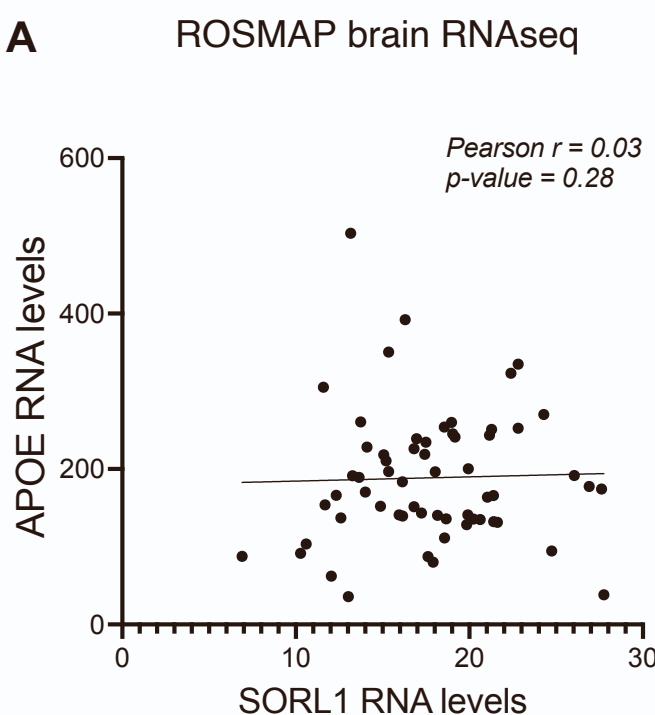
## Supplemental Figure 6



## Supplemental Figure 7



## Supplemental Figure 8

**A****B**