

1 **Fluorine mass balance, including total fluorine, extractable organic**
2 **fluorine, oxidizable precursors and target PFAS, in pooled human**
3 **serum from the Tromsø population in 1986, 2007 and 2015**

4 Lara Cioni^{1,2}, Merle Plassmann³, Jonathan P. Benskin³, Ana Carolina M. F. Coêlho², Therese
5 H. Nøst², Charlotta Rylander², Vladimir Nikiforov¹, Torkjel M. Sandanger^{1,2}, Dorte Herzke¹,
6 4*

7 1. NILU, Fram Centre, Tromsø, Norway, NO-9296

8 2. UiT – The Arctic University of Norway, Department of Community Medicine, Tromsø,
9 Norway, NO-9037

10 3. Stockholm University, Department of Environmental Science, Stockholm, Sweden, SE-106
11 91

12 4. Norwegian Institute for public Health, Oslo, Norway, NO-0213

13 ***Corresponding authors**

14 Dorte Herzke - NILU, Fram Centre, Tromsø, Norway, NO-9296

15 [*dhe@nilu.no](mailto:dhe@nilu.no)

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17 Summary: 21 pages, 6 figures 10 tables

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20 The following information is included:
21 Chemicals and consumables (Page S3); characteristics of Tromsø Study samples and pools
22 (Page S3-S4); quality control measures for TF, EOF, TOP assay and target PFAS (Page S4-
23 S11); data evaluation equations (Page S11-S12); PFAS concentrations used for FMB
24 calculations (Page S13); TF and EOF concentrations in human blood from this study and from
25 the literature (Page S14); multiple linear regression coefficients estimates and 95% confidence
26 intervals for $\ln(\text{TF})$, $\ln(\text{EOF})$, $\ln(\sum_{12} \text{PFAS})$, % UEOF and TOP (Page S14); multiple linear
27 regression (including sex and sampling year interaction terms) coefficients estimates and 95%
28 confidence intervals for $\ln(\sum_{12} \text{PFAS})$ and % UEOF (Page S15); UEOF concentrations in
29 human blood from this study and from the literature (Page S15); TF, EOF, TOP, $\sum_{12} \text{PFAS}$ and
30 UEOF concentrations in serum pools containing the same individuals in 1986, 2007 and 2015
31 (Page S16); UpSet plot showing the intersection of PFAA with increased concentrations after
32 oxidation (Page S17); individual target PFAS in pooled serum from 1986, 2007 and 2015 (Page
33 S18); $\sum_{12} \text{PFAS}$ concentrations in relationship with mean age of the individuals in the pools
34 (Page S19); individual target PFAS concentrations in men and women (Page S20).

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45 **1. Materials and methods**

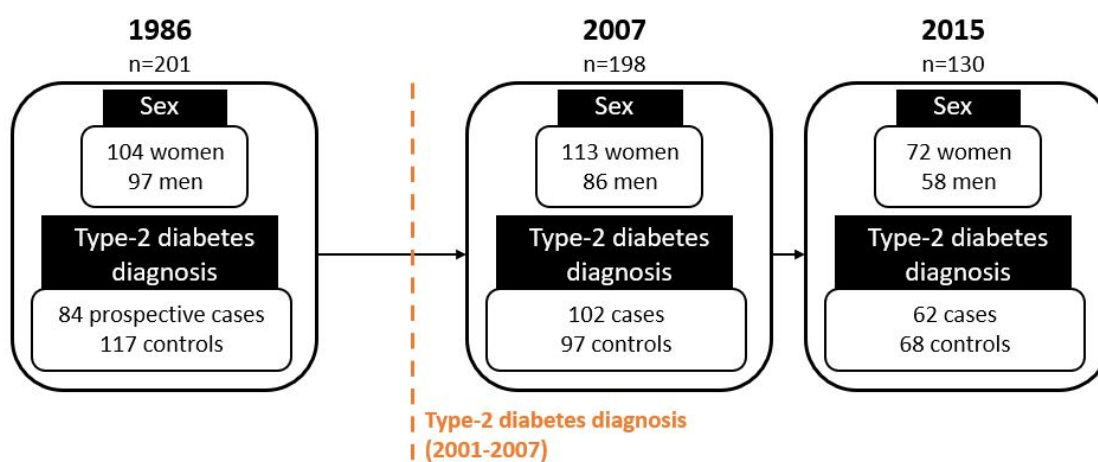
46 **1.1. Chemicals and consumables**

47 Acetonitrile (ACN, LiChrosolv®), tert-butyl methyl ether (MTBE, Suprasolv®), fuming
48 hydrochloric acid (HCl, p.a. 37%) and sodium hydroxide (NaOH, EMSURE®, ≥ 99.0%) were
49 obtained from Merck (Darmstadt, Germany). Sodium persulfate (Na₂S₂O₈, reagent grade, ≥
50 98%, lot #BCCC8760) and ammonium acetate (NH₄OAc, LiChropur™) were obtained from
51 Sigma-Aldrich (Steinheim, Germany). Ammonia (NH₃, solution 25%, AnalaR NORMAPUR)
52 was purchased from VWR (Fontenay-sous-Bois, France). All native and isotopically labelled
53 PFAS standards were obtained from Wellington Laboratories Inc. (Guelph, Ontario, Canada).

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55 **1.2. Serum samples and pooling strategy**

56 **Figure S1**– Tromsø Study serum samples selection.



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64 **Table S1** – Characteristics of the Tromsø Study serum pools.

1986 (n=167)					2007 (n=175)					2015 (n=130)				
<i>Pool ID</i>	<i>n</i>	<i>Sex</i>	<i>Age mean (range)</i>	<i>Diabetes</i>	<i>Pool ID</i>	<i>n</i>	<i>Sex</i>	<i>Age mean (range)</i>	<i>Diabetes</i>	<i>Pool ID</i>	<i>n</i>	<i>Sex</i>	<i>Age mean (range)</i>	<i>Diabetes</i>
1	14	Women	36 (25-45)	Controls	1	14	Women	57 (46-66)	Controls	1	14	Women	65 (54-74)	Controls
2	12	Women	49 (46-57)	Controls	2	12	Women	70 (67-78)	Controls	2	12	Women	78 (75-86)	Controls
3	11	Women	41 (30-45)	Prospective cases	3	11	Women	62 (51-66)	Cases	3	11	Women	70 (59-74)	Cases
4	8	Women	49 (46-53)	Prospective cases	4	8	Women	70 (67-74)	Cases	4	8	Women	78 (75-82)	Cases
5	10	Men	34 (17-47)	Controls	5	10	Men	55 (38-68)	Controls	5	10	Men	63 (46-76)	Controls
6	10	Men	51 (48-55)	Controls	6	10	Men	72 (69-76)	Controls	6	10	Men	80 (77-84)	Controls
7	13	Men	44 (33-58)	Prospective cases	7	13	Men	65 (54-79)	Cases	7	13	Men	73 (62-87)	Cases
8	15	Women	31 (25-43)	Controls	8	15	Women	56 (46-64)	Controls	8	15	Women	63 (54-72)	Controls
9	15	Women	45 (43-47)	Controls	9	15	Women	67 (65-69)	Controls	9	15	Women	78 (74-82)	Controls
10	15	Women	52 (48-60)	Controls	10	15	Women	74 (70-81)	Controls	10	15	Women	70 (59-74)	Cases
11	15	Women	45 (43-48)	Prospective cases	11	15	Women	60 (51-64)	Cases	11	15	Women	78 (75-83)	Cases
12	15	Men	37 (17-49)	Controls	12	15	Women	67 (65-69)	Cases	12	15	Men	61 (46-73)	Controls
13	15	Men	55 (50-61)	Controls	13	15	Women	74 (70-81)	Cases	13	15	Men	79 (74-84)	Controls
14	15	Men	40 (25-48)	Prospective cases	14	15	Men	56 (38-66)	Controls	14	15	Men	81 (75-89)	Cases
15	15	Men	55 (49-60)	Prospective cases	15	15	Men	72 (68-76)	Controls					
					16	15	Men	62 (54-66)	Cases					
					17	15	Men	71 (67-76)	Cases					

Pool ID: cells highlighted in green indicate pools with same individuals across 1986, 2007 and 2015
n = number of individuals

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66 **1.3. TF quality control**

67 A 9-point calibration curve ranging from 2.5 to 2500 ng of NaF in water ($R^2 > 0.999$) was
68 included at the beginning and end of each run. Quality control measures for each run included:
69 (1) three sample boat blanks for limit of detection (LOD) calculation, (2) two sample boats
70 spiked with 100 ng of PFOS standard, and (3) three measurements of a certified reference
71 material (fluorine in clay, CRM 461). Blanks ranged between 18 and 21 ng F/mL (n=9) and
72 LOD (average boat blanks + 3 times the standard deviation of the blanks) ranged between 23
73 and 25 ng F/mL. The recovery of the PFOS standard ($120 \pm 6\%$, n=6) confirmed complete

74 combustion and measurements of the certified reference material showed good accuracy and
75 precision (recovery: $123 \pm 9 \%$, $n=9$).

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77 **1.4. EOF quality control**

78 For each extraction batch (14 serum samples), the quality control measures included: (1) three
79 extraction blanks, (2) three reference serum samples not spiked, (3) one reference serum sample
80 spiked with 239 ng of PFOS, (4) one reference serum sample spiked with 500 ng of NaF. The
81 reference serum was obtained from the Arctic Monitoring and Assessment Programme
82 (AMAP) Ring Test for Persistent Organic Pollutants [1]. Each extraction batch was run
83 separately and included a calibration curve at the beginning and end of the run (2.5-1000 ng of
84 NaF in water, $R^2 > 0.999$) and two sample boats spiked with 100 ng of PFOS standard. The
85 extraction blanks ranged from 5 to 7 ng F/mL ($n=12$) and the EOF LOD (average extraction
86 blanks + 3 times the standard deviation of the blanks) ranged from 6 to 9 ng F/mL. The analysis
87 of the reference serum samples spiked with PFOS confirmed good recovery and reproducibility
88 of the EOF analysis in human serum (recovery: $77 \pm 14 \%$, $n=8$). The analysis of the controls
89 spiked with NaF confirmed the removal of fluoride upon extraction (NaF recoveries ranging
90 from 0 to 2 %, $n=4$).

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92 **1.5. TOP assay quality control**

93 For each TOP assay batch (18 samples), a blank and an AMAP reference serum sample were
94 included and processed as the samples. Blanks before and after oxidation showed low levels of
95 PFAA (Table S2). LODs were calculated as the average concentration in the blanks plus 3
96 times the standard deviation of the blanks and in case of no detection in the blanks, LODs were
97 calculated by multiplying the noise of the blanks by 3. LODs before and after oxidation were
98 comparable for most compounds (Table S2). Measured PFAA concentrations before oxidation

99 in the AMAP serum samples were within $\pm 20\%$ of the reference values. Mean recoveries
100 before TOP assay ranged from 61 to 78 % and mean recoveries after TOP assay ranged from
101 55 to 65 %. Model precursors spiking oxidation experiments were performed as part of the
102 validation described in our method paper and showed complete conversion for all spiked
103 precursors and yields of PFAA ranging from 35-100% [2].

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106 **Table S2** - Average blank concentrations and LODs before and after TOP assay in ng/mL of
 107 serum (n=3).

Compound	Before TOP assay		After TOP assay	
	Blank concentration	LOD	Blank concentration	LOD
PFBA	0.15	0.47	0.12	0.49
PFPeA	0.20	0.32	0.26	0.47
PFHxA	0.03	0.10	0.13	0.39
PFHpA	0.01	0.02	0.02	0.13
PFOA	0.05	0.10	0.08	0.18
PFNA	0.00	0.02	0.01	0.03
PFDA	0.00	0.02	0.03	0.10
PFUnDA	0.00	0.02	0.00	0.04
PFDoDA	0.00	0.02	0.00	0.04
PFTTrDA	0.00	0.07	0.00	0.09
PFTeDA	0.00	0.13	0.00	0.13
PFBS	0.00	0.04	0.02	0.08
PFPeS	0.00	0.07	0.00	0.07
PFHxS	0.10	0.13	0.01	0.04
PFHpS	0.00	0.03	0.00	0.03
PFOS	0.09	0.16	0.06	0.14
PFNS	0.00	0.04	0.00	0.04
PFDS	0.00	0.05	0.00	0.05

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110 **Table S3** – Recoveries in pooled serum samples before and after TOP assay (n=46).

Compound	Before TOP assay	After TOP assay
¹³ C-PFBA	74 ± 7	58 ± 10
¹³ C-PFPeA	78 ± 5	62 ± 5
¹³ C-PFHxA	75 ± 7	63 ± 4
¹³ C-PFHpA	70 ± 5	65 ± 4
¹³ C-PFOA	73 ± 6	62 ± 6
¹³ C-PFNA	71 ± 5	58 ± 5
¹³ C-PFDA	78 ± 5	55 ± 3
¹³ C-PFUnDA	61 ± 8	57 ± 5
¹³ C-PFDoDA	72 ± 6	61 ± 7
¹³ C-PFTeDA	75 ± 6	58 ± 7
¹³ C-PFHxS	74 ± 5	62 ± 6
¹³ C-PFOS	75 ± 3	61 ± 8

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112 **1.6. Target PFAS quality control**

113 Target PFAS analyses on the EOF extracts included also the EOF extraction blanks (n=9). No
 114 PFAA were detected in the blanks and the LODs were calculated using the standard error of
 115 the regression divided the slope of the calibration curve multiplied by 3. LODs ranged from
 116 0.03 to 0.13 ng/mL (Table S4). Measured PFAA concentrations in the AMAP serum samples
 117 use as quality control were within +/- 20% of the reference values.

118 After the TOP assay the extracts were also analysed for C₂ and C₃-PFAA using a Raptor Polar
 119 X column. Trifluoroacetic acid (TFA) was analysed in a 5 minute isocratic run with 80 % 2mM
 120 ammonium acetate in methanol and 20 % 2mM ammonium acetate in 90:10 water:methanol.
 121 Perfluoropropionic acid (PFPrA), trifluoromethane sulfonic acid (TFMS), difluoro
 122 (perfluoromethoxy) acetic acid (1,2-PFECA), difluoroacetic acid (DiFA) and chlorodifluoro
 123 acetic acid (Cl-DiFA) were analysed in a 10 minute isocratic run using 80% 60:40

124 methanol:water with 0.05% formic acid and 20% 10 mM ammonium formate in water with
125 0.05% formic acid, based on an application note from Restek [3]. For these analyses, serum
126 extracts were spiked with ¹³C-TFA before oxidation and recoveries ranged from 56 to 65 %
127 (n=46). Concentrations in the blanks ranged from 0.00 to 0.25 ng/mL. LODs were calculated
128 as the average concentration in the blanks plus 3 times the standard deviation of the blanks and
129 in case of no detection in the blanks, LODs were calculated by multiplying the noise of the
130 blanks by 3 (Table S4).

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Table S4 – Target PFAS analysed on EOF extracts by UHPLC-Orbitrap.

Abbreviation	Name	LOD (ng/mL)
PFCA (Perfluoroalkyl carboxylic acids)		
PFBA	Perfluorobutanoic acid	0.07
PFPeA	Perfluoropentanoic acid	0.06
PFHxA	Perfluorohexanoic acid	0.07
PFHpA	Perfluoroheptanoic acid	0.07
PFOA	Perfluorooctanoic acid	0.06
PFNA	Perfluorononanoic acid	0.07
PFDA	Perfluorodecanoic acid	0.09
PFUnDA	Perfluoroundecanoic acid	0.10
PFDoDA	Perfluorododecanoic acid	0.10
PFTrDA	Perfluorotridecanoic acid	0.10
PFTeDA	Perfluorotetradecanoic acid	0.13
PFPeDA	Perfluoropentadecanoic acid	0.13
PFHxDA	Perfluorohexadecanoic acid	0.14
PFOcDA	Perfluorooctadecanoic acid	0.13
PFSA (Perfluoroalkyl sulfonic acids)		
PFBS	Perfluorobutane sulfonic acid	0.06
PFPeS	Perfluoropentane sulfonic acid	0.06
PFHxS	Perfluorohexane sulfonic acid	0.06
PFHpS	Perfluoroheptane sulfonic acid	0.06
PFOS	Perfluorooctane sulfonic acid	0.03
PFNS	Perfluorononane sulfonic acid	0.04
PFDS	Perfluorodecane sulfonic acid	0.05
PFUnDS	Perfluoroundecane sulfonic acid	0.06
PFECA (Perfluoroalkyl ether sulfonic acids)		
GenX	Ammonium perfluoro-4,8-dioxa-3H-nonanoic acid	0.08
ADONA	Perfluoro-4,8-dioxa-3H-nonanoic acid	0.08
FTCA (Fluorotelomer carboxylic acids)		
3:3 FTCA	3:3 Fluorotelomer carboxylic acid	0.06
5:3 FTCA	5:3 Fluorotelomer carboxylic acid	0.08
7:3 FTCA	7:3 Fluorotelomer carboxylic acid	0.08
FTS (Fluorotelomer sulfonates)		
4:2 FTS	4:2 Fluorotelomer sulfonic acid	0.06
6:2 FTS	6:2 Fluorotelomer sulfonic acid	0.08
8:2 FTS	8:2 Fluorotelomer sulfonic acid	0.08
Perfluorooctane sulfonamido substances		
FOSA	Perfluorooctane sulfonamide	0.07
Me-FOSA	N-Methyl perfluorooctane sulfonamide	0.07
Et-FOSA	N-Ethyl perfluorooctane sulfonamide	0.07
FOSAA	Perfluorooctane sulfonamidoacetic acid	0.06
Me-FOSAA	N-Methyl perfluorooctane sulfonamidoacetic acid	0.06
Et-FOSAA	N-Ethyl perfluorooctane sulfonamidoacetic acid	0.06
Me-FOSE	N-Methyl perfluorooctane sulfonamido ethanol	0.08
Et-FOSE	N-Ethyl perfluorooctane sulfonamido ethanol	0.08
CI-PFAES		
9Cl-PF3ONS	9Cl-Perfluoro-3-oxononane sulfonic acid	0.10
11Cl-PF3OUs	11Cl-Perfluoro-3-oxoundecane sulfonic acid	0.10
PAPs		
4:2 monoPAP	4:2 Fluorotelomer phosphate monoester	0.10
4:2 diPAP	4:2 Fluorotelomer phosphate diester	0.10
6:2 monoPAP	6:2 Fluorotelomer phosphate monoester	0.10
6:2 diPAP	6:2 Fluorotelomer phosphate diester	0.10
6:2/8:2 diPAP	6:2/8:2 Fluorotelomer phosphate diester	0.10
6:2/10:2 diPAP	6:2/10:2 Fluorotelomer phosphate diester	0.12
6:2/12:2 diPAP	6:2/12:2 Fluorotelomer phosphate diester	0.12

6:2/14:2 diPAP	6:2/14:2 Fluorotelomer phosphate diester	0.12
8:2 diPAP	8:2 Fluorotelomer phosphate diester	0.13
8:2/10:2 diPAP	8:2/10:2 Fluorotelomer phosphate diester	0.13
10:2 monoPAP	10:2 Fluorotelomer phosphate monoester	0.13
10:2 diPAP	10:2 Fluorotelomer phosphate diester	0.13

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137 **Table S5** - Average blank concentrations and LODs before and after TOP assay in ng/mL of

138 serum (n=3).

Compound	Blank concentration	LOD
TFA	0.28	0.32
PFPrA	0.10	0.13
TFMS	0.00	0.07
1,2-PFECA	0.00	0.07
DiFA	0.00	0.07
Cl-DiFA	0.00	0.07

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141 **1.7. Data evaluation**

142 For comparison with EOF values, target PFAS concentrations measured in the EOF extracts

143 and Δ PFAA concentrations from the TOP assay were converted to F equivalents using the

144 following equation:

$$Concentration \left(\frac{ng\ F}{mL} \right) = \frac{concentration \left(\frac{ng}{mL} \right) \cdot nF \cdot AW_F}{MW_{PFAS}} \quad (S1)$$

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146 where n_F is the number of fluorine atoms in the PFAS structure, A_F is the atomic weight of
147 fluorine and MW_{PFAS} is the molecular weight of the PFAS which concentration is being
148 converted.

149 Differences in TF, EOF, \sum_{12} PFAS, unidentified EOF and TOP between sampling years were
150 assessed by multiple linear regression to account for the influence of sex and age (as weighted
151 mean of the age of the individuals in the pools expressed in years) using the following equation:

$$y = \beta_0 + \beta_1 dummy\ 1 + \beta_2 dummy\ 2 + \beta_3 sex + \beta_4 age \quad (S2)$$

152 where y is the log transformed concentration for TF, EOF and \sum_{12} PFAS, the $\Delta PFAA$
153 concentration in ng/mL for TOP and the percentage contribution to EOF for UEOF; β_0 is the
154 intercept of the multiple linear regression; β_1 , β_2 , β_3 and β_4 are the regression coefficients for
155 the predictor variables; dummy 1 is a dummy variable equal to 1 if sampling year is 1986, equal
156 to 0 if sampling year is 2007 or 2015; dummy 2 is a dummy variable equal to 1 if sampling
157 year is 2015, equal to 0 if sampling year is 1986 and 2007; sex is categorical variable equal to
158 0 for women and equal to 1 for men; age is the weighted mean age of the individuals making
159 up each pool expressed in years.

160 When sex was a significant predictor, differences in concentrations between men and women
161 at each sampling year were assessed by adding an interaction term between sex and each
162 sampling year dummy variable as described by equation S3.

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$$y = \beta_0 + \beta_1 dummy\ 1 + \beta_2 dummy\ 2 + \beta_3 sex + \beta_4 age + \beta_5 dummy1\ sex + \beta_6 dummy2\ sex \quad (S3)$$

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166 **2. Results**

167 **Table S6 – PFAS concentrations (ng/mL) used for fluorine mass-balance calculations**

168 (concentrations are not recovery corrected).

	1986 (n=15)				2007 (n=17)				2015 (n=14)			
	DF	Median	Mean	Range	DF	Median	Mean	Range	DF	Median	Mean	Range
PFHpA	0/15	-	-	-	0/17	-	-	-	0/14	-	-	-
PFOA	15/15	1.60	1.50	0.88-2.04	17/17	2.32	2.40	2.00-2.96	14/14	1.52	1.56	1.12-2.24
PFNA	15/15	0.24	0.25	0.08-0.64	17/17	1.04	0.99	0.72-1.52	14/14	1.14	1.18	0.68-1.60
PFDA	1/15	<0.09	<0.09	<0.09-0.24	17/17	0.40	0.39	0.20-0.84	14/14	0.50	0.50	0.20-0.84
PFUnDA	15/15	0.32	0.32	0.12-0.56	17/17	0.60	0.60	0.24-1.56	14/14	0.62	0.58	0.24-1.20
PFDoDA	0/15	-	-	-	0/17	-	-	-	0/14	-	-	-
PFHxS	15/15	0.40	0.38	0.16-0.72	17/17	1.44	1.59	1.04-4.68	14/14	1.24	1.36	0.72-3.12
PFHpS	4/15	<0.03	<0.03	<0.03-0.08	17/17	0.20	0.17	0.04-0.36	14/14	0.08	0.09	0.04-0.20
br-PFOS	15/15	3.68	3.58	2.48-5.16	17/17	5.44	5.26	3.68-7.16	14/14	3.14	3.40	2.56-4.88
lin-PFOS	15/15	10.9	10.4	6.72-15.4	17/17	16.2	16.5	11.1-30.2	14/14	10.4	11.3	5.52-18.8
FOSAA	9/15	0.08	0.08	<0.06-0.20	0/17	-	-	-	0/14	-	-	-
Me-FOSAA	14/15	0.16	0.13	<0.06-0.28	10/17	0.08	0.08	<0.06-0.20	0/14	-	-	-
Et-FOSAA	15/15	0.28	0.27	0.12-0.52	0/17	-	-	-	0/14	-	-	-
∑ 12 PFAS	15/15	17.8	17.2	11.0-24.1	17/17	27.9	28.3	21.5-46.1	14/14	19.2	20.3	11.4-30.0

DF = detection frequency: number of pools with PFAS concentration > LOD.

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178 **Table S7** – Descriptive statistics for TF and EOF concentrations (ng F/mL) in the Tromsø
 179 Study pooled serum samples from 1986, 2007 and 2015 and in samples from previous studies
 180 available in the literature (n=number of pools/number of individual samples).

Study	Country	Sampling year	Matrix	n	TF (ng F/mL)			EOF (ng F/mL)		
					Median	Mean	Range	Median	Mean	Range
This study	Norway	1986	Serum (pooled)	15	79.1	112	<25.0-1330	22.2	23.3	13.3-45.3
This study	Norway	2007	Serum (pooled)	17	74.2	74.8	<25.0-1212	20.8	20.5	16.2-30.3
This study	Norway	2015	Serum (pooled)	14	68.3	71.6	<25.0-265	18.5	18.4	12.6-22.6
Miyake et al. (2007)	Japan	2003-2004	Whole blood	3	208	214	181-262	<6	<6	<6-8.89
Miyake et al. (2007)	USA	2001	Plasma	4	149	163	140-189	45.2	38.3	17.8-59.0
Yeung et al. (2008)	China	2004	Whole blood	30	-	-	60.6-166	-	-	<6-43.4
Yeung and Mabury (2016)	China	2004	Whole blood	34	-	-	-	17	18.4	8.22-94.4
Yeung and Mabury (2016)	Germany (Halle)	1995-2009	Plasma	42	-	-	-	-	15.9	5.29-43.9
Yeung et al. (2016)	Germany (Munster)	1982-2009	Plasma	80	-	-	-	-	23.7	9.20-115
Miaz et al. (2020)	Sweden	1996-2017	Serum (pooled)	57	-	-	-	-	-	8.10-32.0
Aro et al. (2021)	Sweden	2015	Whole blood	9	-	-	-	-	24.8	17.6-37.8
Aro et al. (2021)	Sweden (Ronneby)	2014-2016	Whole blood	20	-	-	-	-	234	<107-592
Aro et al. (2021)	Sweden	2018-2019	Whole blood	130	-	-	-	-	-	0.51-48.7
Kaiser et al. (2021)	Austria	2021	Serum (pooled)	6	-	-	-	-	3.83	2.85-7.17

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182 **Table S8** – Multiple linear regression coefficients estimates and 95% confidence intervals for
 183 $\ln(\text{TF})$, $\ln(\text{EOF})$, $\ln(\sum 12 \text{ PFAS})$, % UEOF and TOP in pooled serum samples from the Tromsø
 184 Study.

	$\ln(\text{TF})$	$\ln(\text{EOF})$	$\ln(\sum 12 \text{ PFAS})$	% UEOF	TOP
	Estimate (95% CI)	Estimate (95% CI)	Estimate (95% CI)	Estimate (95% CI)	Estimate (95% CI)
β_0 (intercept)	1.17 (2.68 to 5.03)	2.55 (1.91 to 3.20)	2.76 (2.33 to 3.08)	61.4 (18.8 to 104)	0.49 (-0.48 to 1.45)
β_1 (1986-2007)	1.41 (-0.15 to 2.97)	0.29* (0.03 to 0.55)	-0.48*** (-0.61 to -0.35)	22.8* (5.60 to 40.0)	0.14 (-0.25 to 0.53)
β_2 (2015-2007)	-0.39 (-1.47 to 0.68)	-0.16 (-0.34 to 0.02)	-0.41*** (-0.51 to -0.30)	18.2** (6.35 to 30.0)	0.02 (-0.24 to 0.29)
β_3 (sex)	0.04 (-0.77 to 0.85)	-0.05 (-0.18 to 0.09)	0.18*** (0.10 to 0.26)	-14.3** (-23.3 to -5.32)	-0.16 (-0.36 to 0.05)
β_4 (age mean)	0.05 (-0.01 to 0.11)	0.01 (-0.002 to 0.017)	0.02*** (0.01 to 0.02)	-0.64 (-1.27 to 0.01)	-0.001 (-0.015 to 0.013)
R ²	0.084	0.209	0.796	0.594	0.091
F-test p-value	0.445	0.042	0.000	0.000	0.409
*p < 0.05 **p < 0.01 *** p < 0.001					

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187 **Table S9** – Multiple linear regression (including sex and sampling year interaction terms)
 188 coefficients estimates and 95% confidence intervals for $\ln(\sum 12 \text{ PFAS})$ and % UEOF in pooled
 189 serum samples from the Tromsø Study.

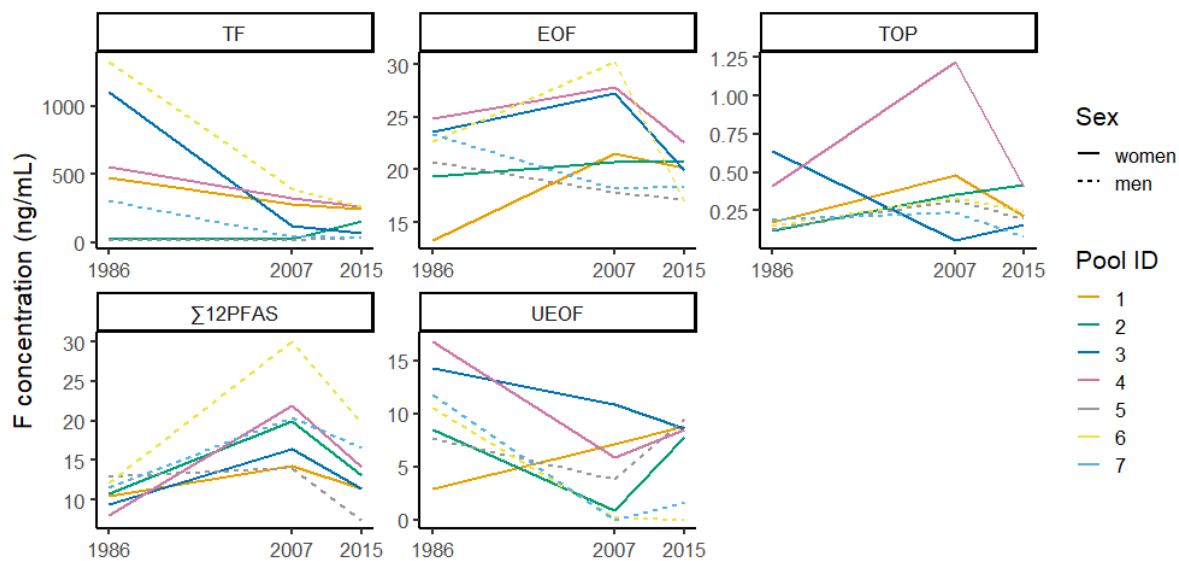
	$\ln(\sum 12 \text{ PFAS})$	% UEOF
	Estimate (95% CI)	Estimate (95% CI)
β_0 (intercept)	2.72 (2.34 to 3.11)	64.4 (20.3 to 109)
β_1 (1986-2007)	-0.17 (-0.35 to 0.01)	18.3 (-2.27 to 38.8)
β_2 (2015-2007)	-0.40*** (-0.53 to -0.27)	17.1* (1.91 to 32.3)
β_3 (2007 sex)	0.16* (0.02 to 0.28)	-18.3* (-33.4 to -3.14)
β_4 (age mean)	0.02*** (0.01 to 0.02)	-0.66* (-1.31 to -0.01)
β_5 (1986 sex)	0.08 (-0.11 to 0.27)	9.24 (-12.7 to 31.2)
β_6 (2015 sex)	-0.01 (-0.02 to 0.18)	3.04 (-19.4 to 25.5)
R ²	0.802	0.602
F-test p-value	0.000	0.000
*p < 0.05 **p < 0.01 *** p < 0.001		

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191 **Table S10** – Descriptive statistics for UEOF concentrations (ng F/mL and/or %) in the Tromsø
 192 Study pooled serum samples from 1986, 2007 and 2015 and in samples from previous studies
 193 available in the literature (n=number of pools/number of individual samples).

Study	Country	Sampling year	Matrix	n	UEOF		
					Median	Mean	Range
This study	Norway	1986	Serum (pooled)	15	10.5 ng F/mL 46%	10.9 ng F/mL 46%	2.93-34.8 ng F/mL 21-77%
This study	Norway	2007	Serum (pooled)	17	2.26 ng F/mL 10%	3.17 ng F/mL 14%	0.00-10.9 ng F/mL 0-40%
This study	Norway	2015	Serum (pooled)	14	7.54 ng F/mL 37%	5.32 ng F/mL 27%	0.00-9.74 ng F/mL 0-56%
Miyake et al. (2007)	Japan	2003-2004	Whole blood	3	-	-	0.00-1.38 ng F/mL 0-15%
Miyake et al. (2007)	USA	2001	Plasma	4	-	-	0.00-4.40 ng F/mL 0-15%
Yeung et al. (2008)	China	2004	Whole blood	30	-	-	15-43%
Yeung and Mabury (2016)	China	2004	Whole blood	34	-	-	14-69%
Yeung and Mabury (2016)	Germany (Halle)	1995-2009	Plasma	42	-	-	0.0-9.5 ng F/mL
Yeung et al. (2016)	Germany (Munster)	1982-2009	Plasma	80	-	-	0.0-9.9 ng F/mL
Miaz et al. (2020)	Sweden	1996-2017	Serum (pooled)	57	-	-	11-75%
Aro et al. (2021)	Sweden	2015	Whole blood	9	-	84%	71-97%
Aro et al. (2021)	Sweden (Ronneby)	2014-2016	Whole blood	20	-	37%	0-76%
Aro et al. (2021)	Sweden	2018-2019	Whole blood	130	-	0-99%	-
Kaiser et al. (2021)	Austria	2021	Serum (pooled)	6	-	1.17 ng F/mL 24%	-

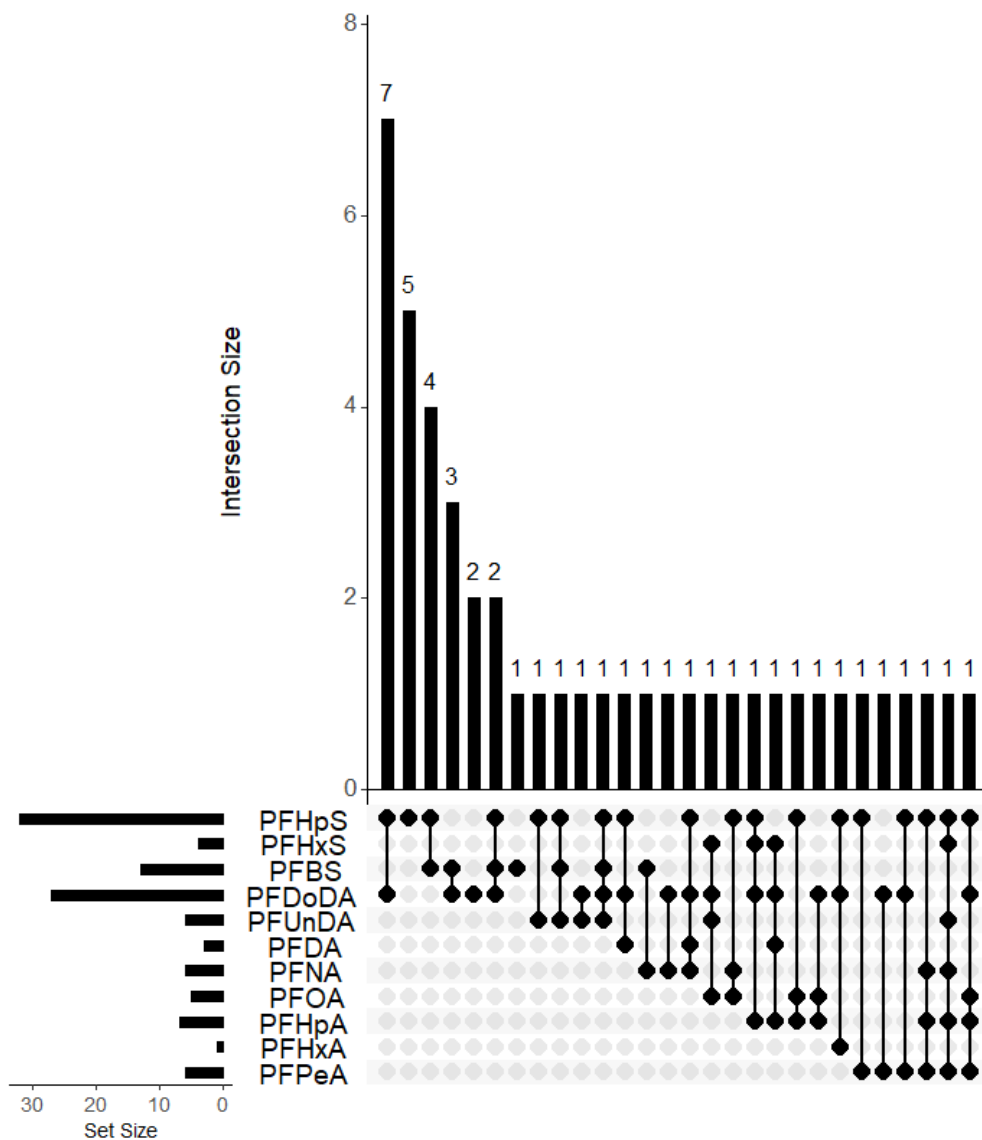
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196 **Figure S2** – TF, EOF, TOP, Σ_{12} PFAS and UEOF concentrations (ng F/mL) in serum pools
 197 from the Tromsø Study containing the same individuals in 1986, 2007 and 2015.

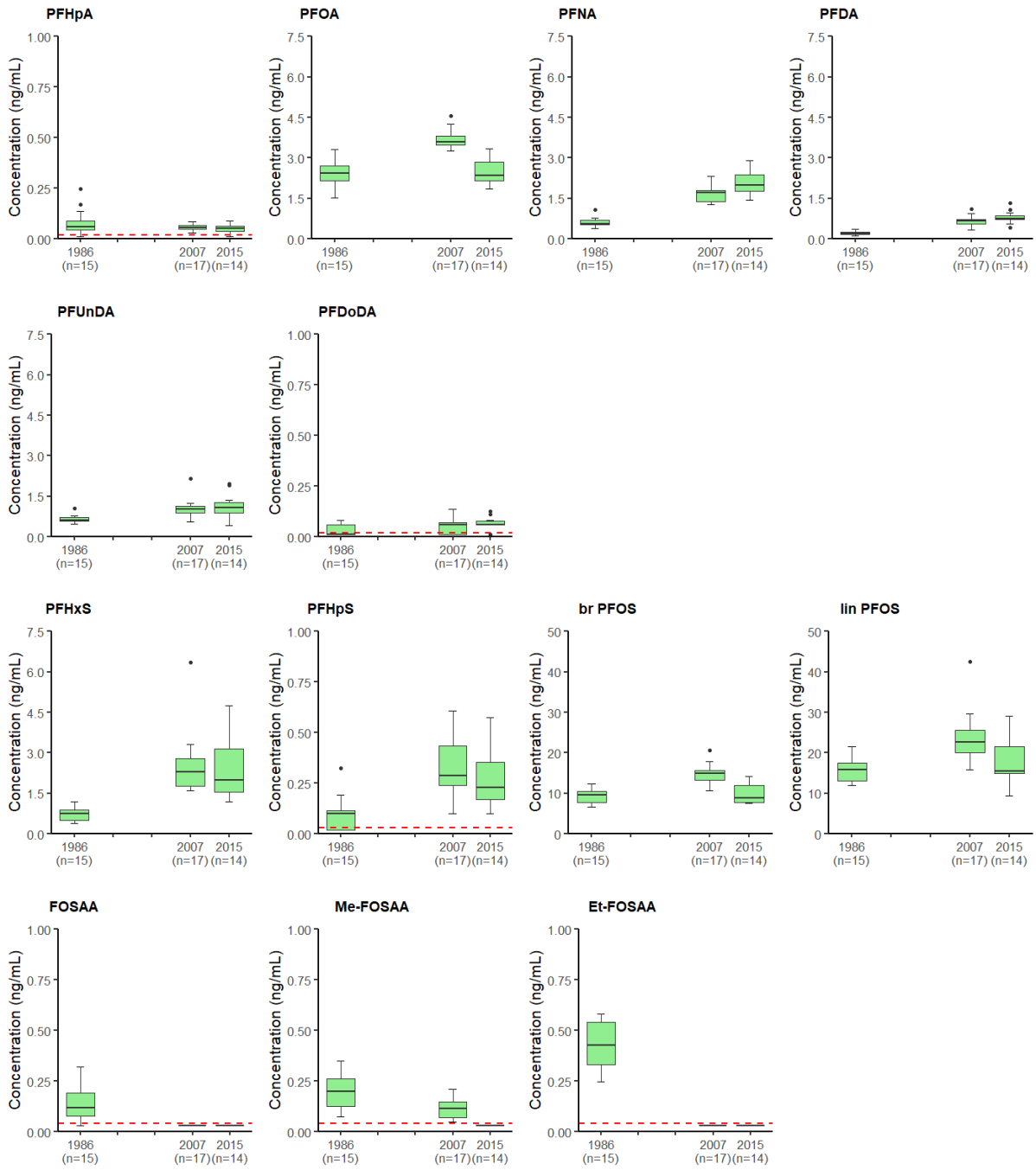
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200 **Figure S3** - UpSet plot showing the intersection of PFAs with increased concentrations after
 201 oxidation. The bar chart shows the number of pools with increases in concentrations of a
 202 combination of PFAs. The graphical table underneath indicates the PFAA combinations (black
 203 dots and lines). The frequency count of each PFA across all subsets is shown as a smaller bar
 204 chart on the left side of the graphical table.

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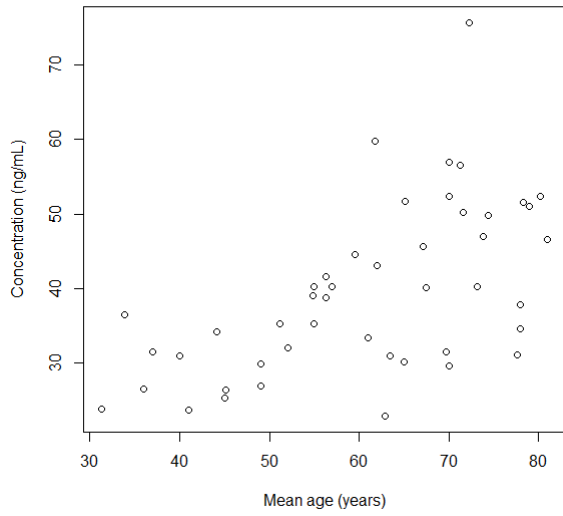
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209 **Figure S4 – Target PFAS (ng/ml) in pooled serum samples from the Tromsø Study collected**

210 **in 1986, 2007 and 2015.**

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215 **Figure S5** – Concentrations of $\Sigma 12$ PFAS in pooled serum samples from the Tromsø Study
216 collected in 1986, 2007 and 2015 in relationship with mean age of the individuals in the pools
217 in years.

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233 **References**

- 234 1. AMAP, *AMAP Assessment 2021: Human Health in the Arctic*. 2022, Arctic
235 Monitoring and Assessment Programme (AMAP): Tromsø, Norway. p. 240.
- 236 2. Cioni, L., et al., *Total Oxidizable Precursors Assay for PFAS in Human Serum*.
237 Environment International, 2022(170).
- 238 3. Restek. *Raptor Polar X: Separate a Wide Variety of Polar Analytes with a Novel*
239 *Hybrid Stationary Phase* [https://www.restek.com/row/technical-literature-](https://www.restek.com/row/technical-literature-library/articles/raptor-polar-x-separate-a-wide-variety-of-polar-analytes-with-a-novel-hybrid-stationary-phase/)
240 [library/articles/raptor-polar-x-separate-a-wide-variety-of-polar-analytes-with-a-](https://www.restek.com/row/technical-literature-library/articles/raptor-polar-x-separate-a-wide-variety-of-polar-analytes-with-a-novel-hybrid-stationary-phase/)
241 [novel-hybrid-stationary-phase/](https://www.restek.com/row/technical-literature-library/articles/raptor-polar-x-separate-a-wide-variety-of-polar-analytes-with-a-novel-hybrid-stationary-phase/). 2020.

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