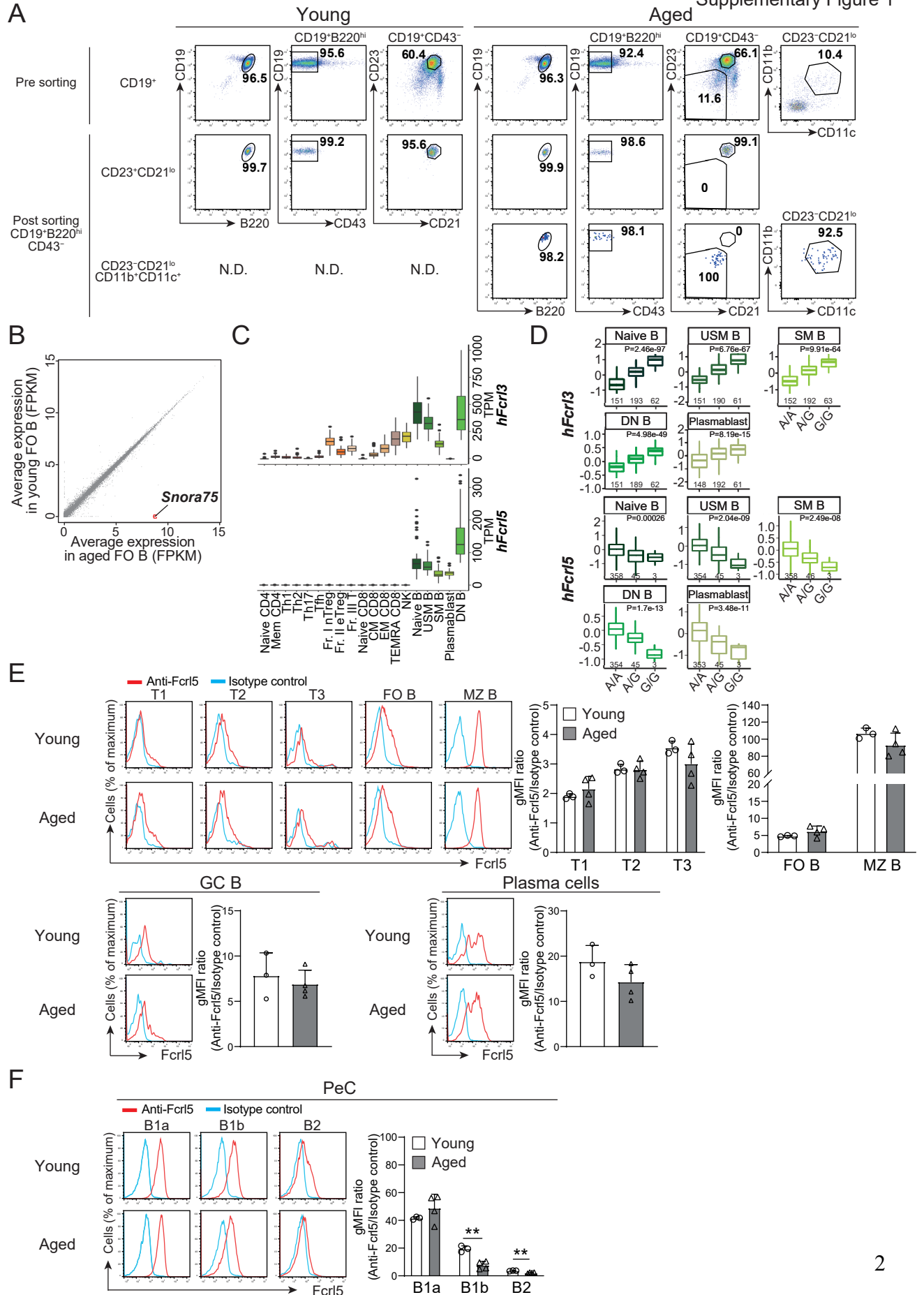


Supplementary Material

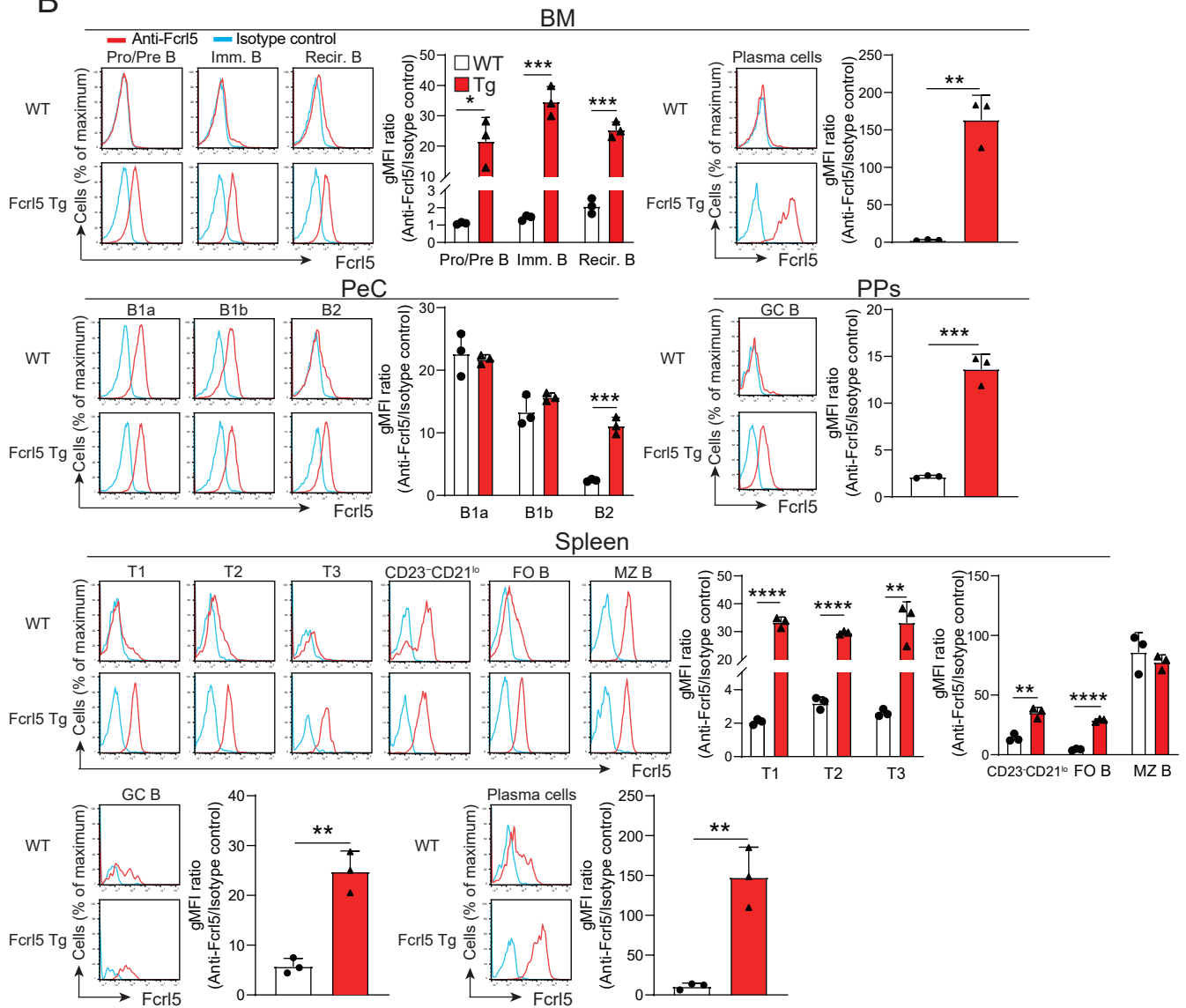
Supplementary Figures



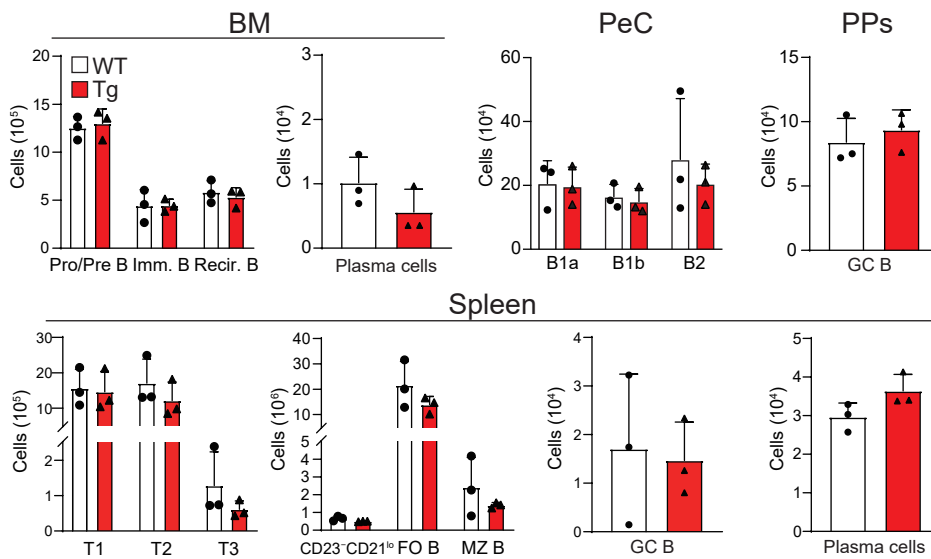
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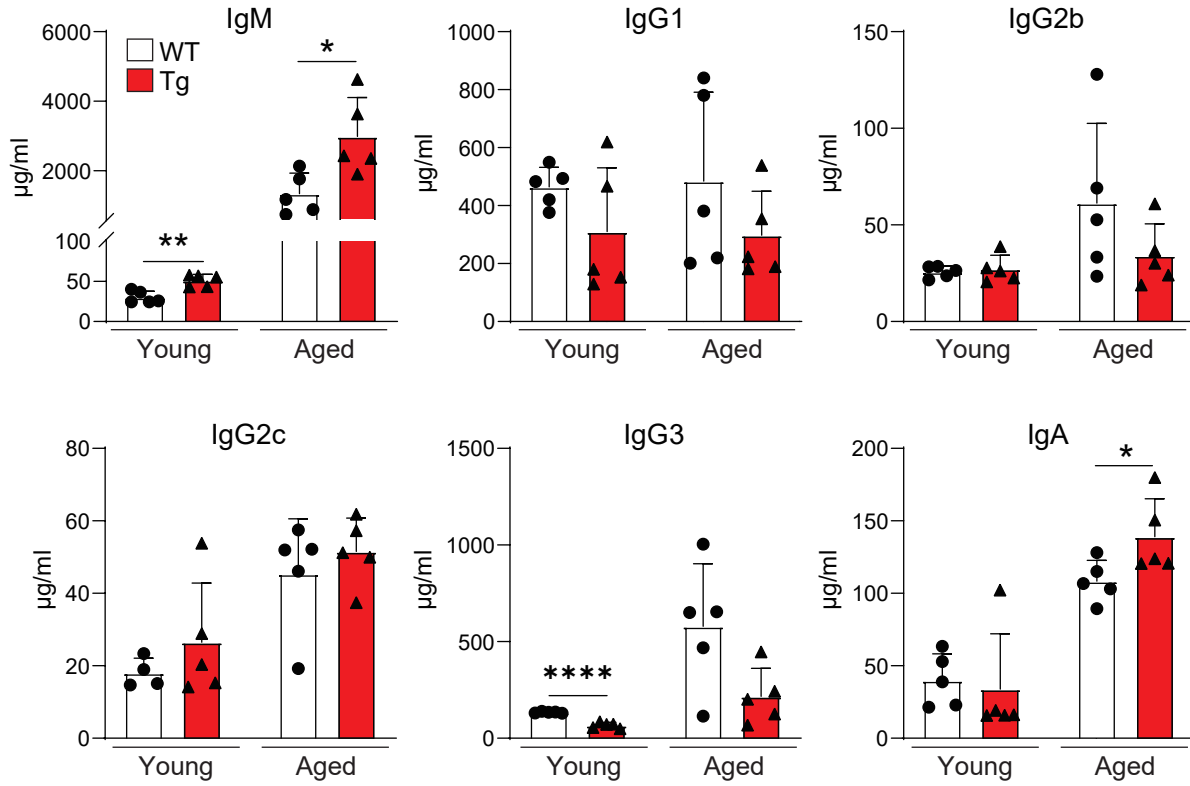
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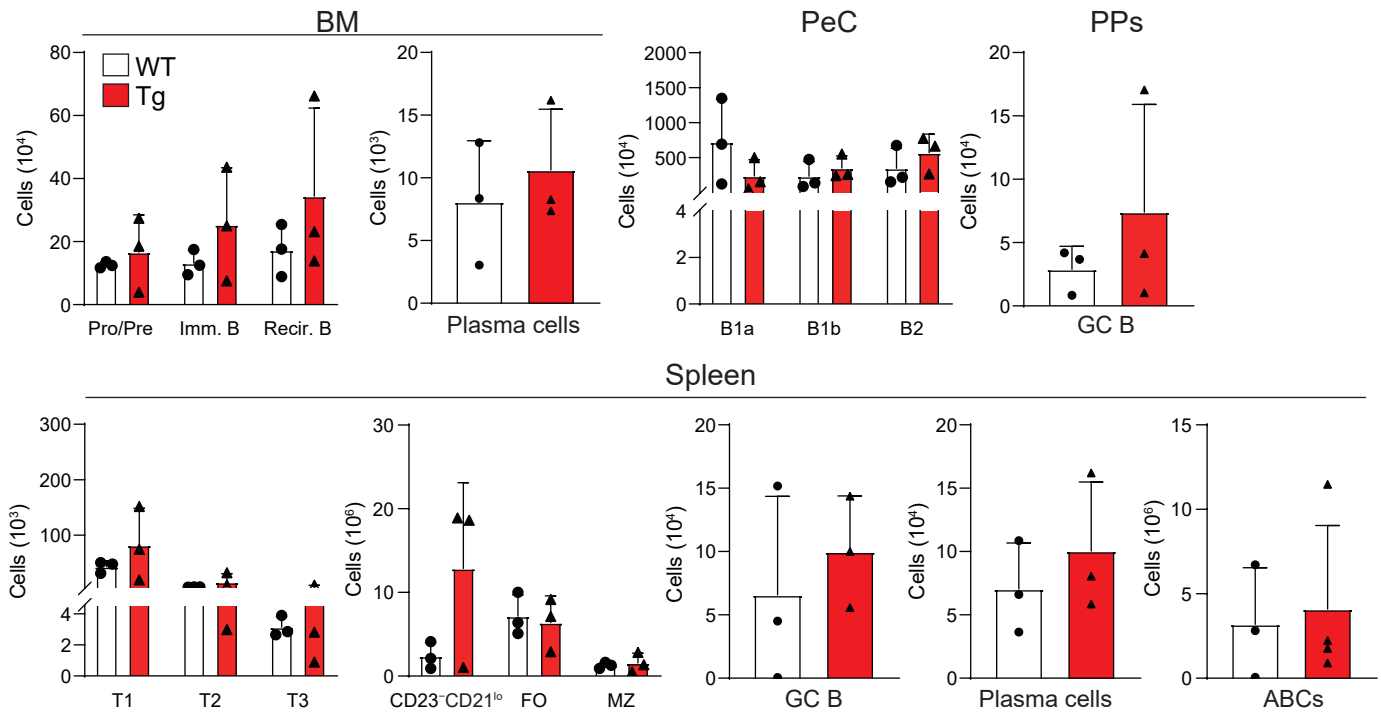
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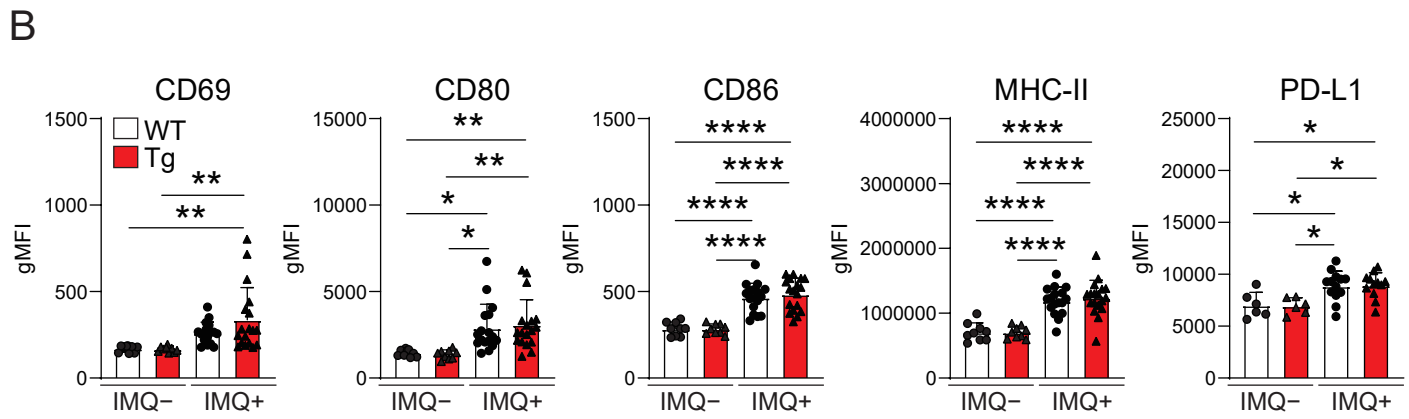
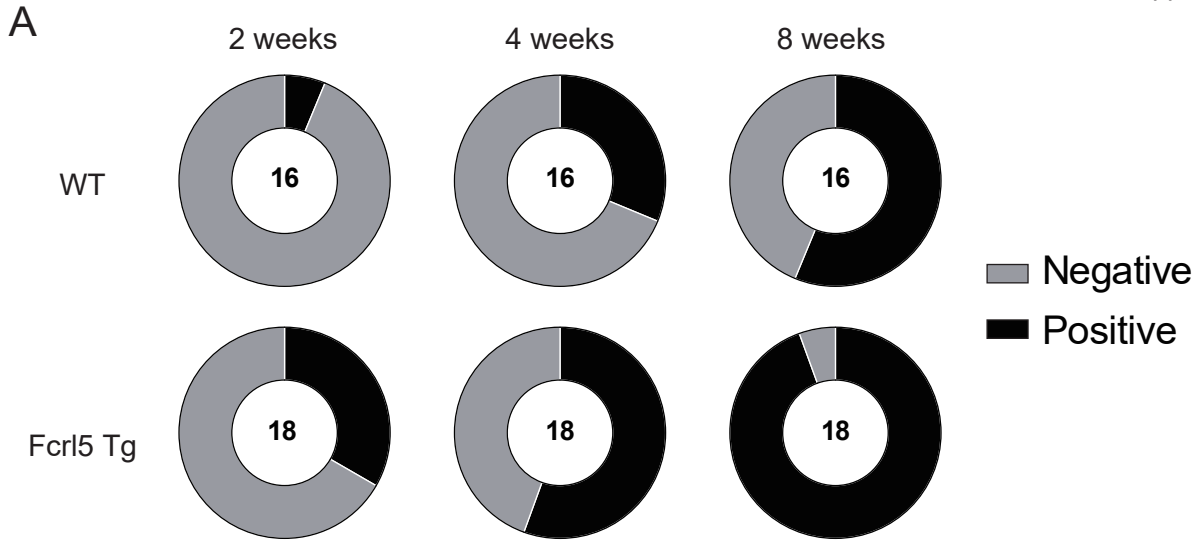


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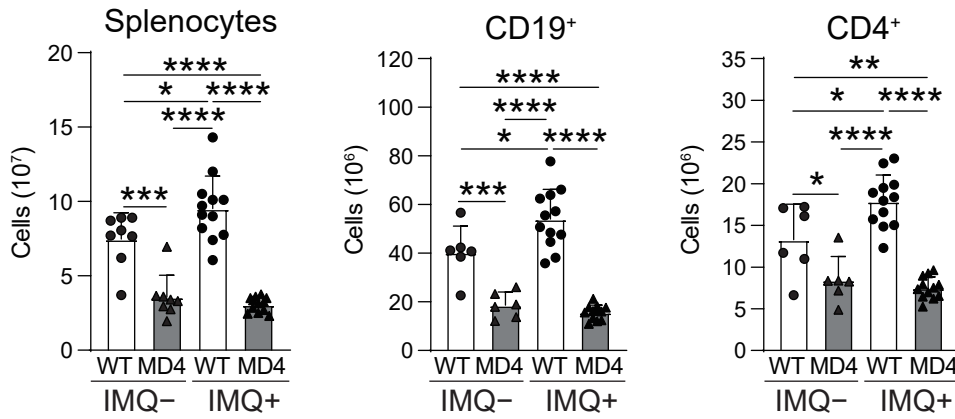


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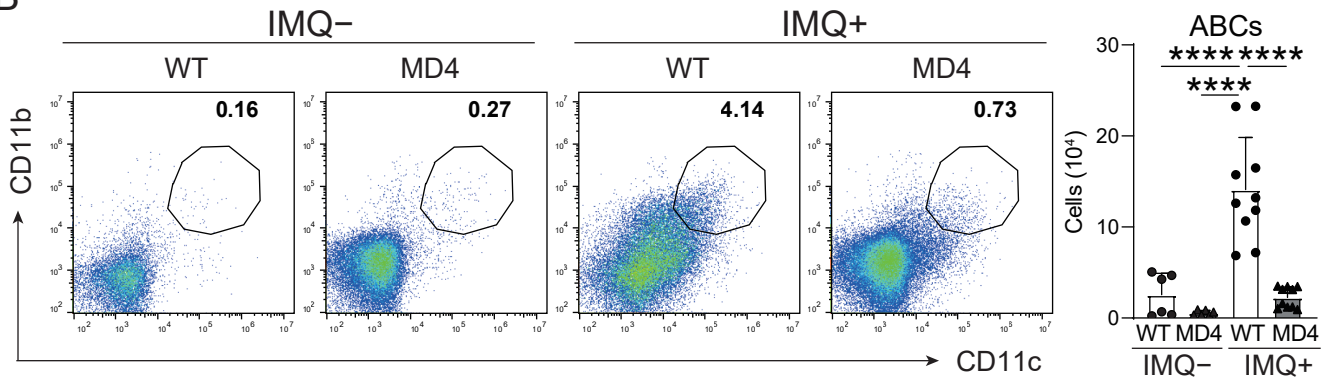




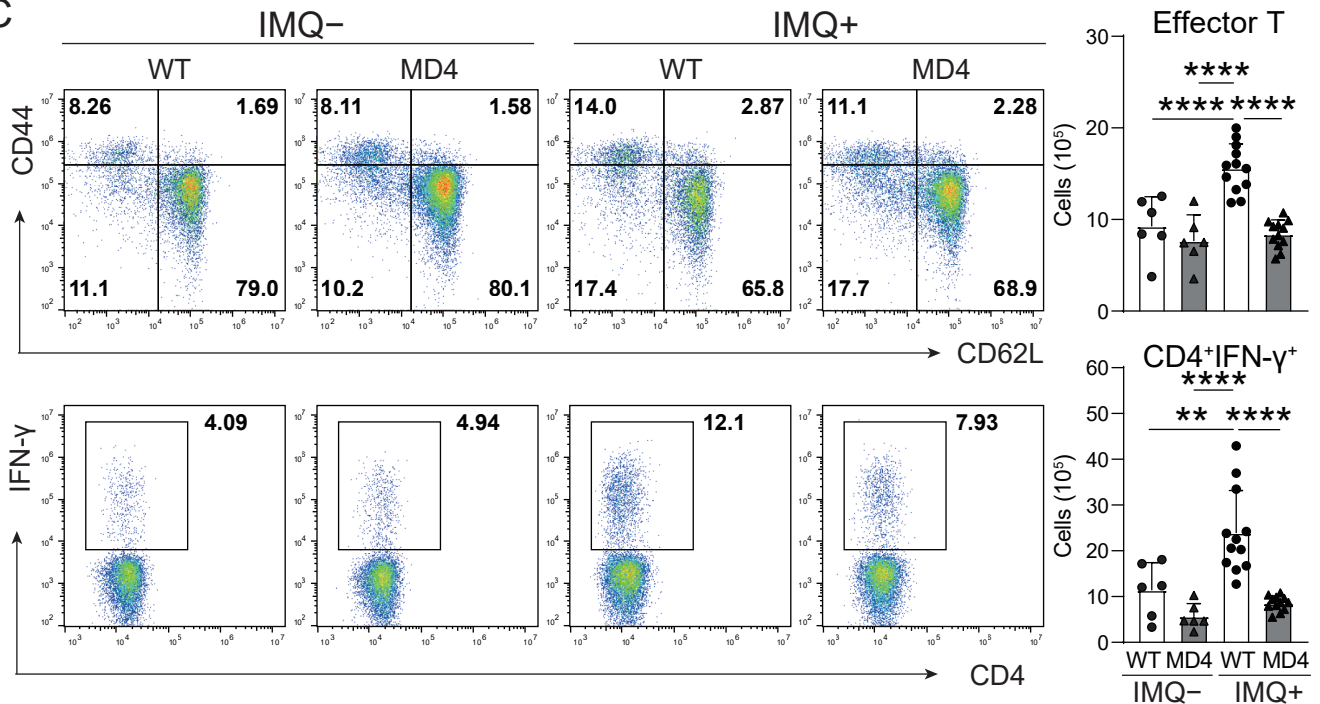
A



B



C



Supplementary Figure legends

Supplementary Figure 1. Gating strategy and distribution of Fcrl5 expression. (A) Gating strategy employed to sort FO B (CD19⁺B220^{hi}CD43⁻CD23⁺CD21^{lo}) cells and ABCs (CD19⁺B220^{hi}CD43⁻CD23⁻CD21^{lo}CD11b⁺CD11c⁺) from young and aged WT mice (n=3 mice each). (B) Differential gene expression of FO B cells from young WT mice and FO B cells from aged WT mice (n=3 mice each) presented as a scatter plot. (C) Expression of *hFcrl3* and *hFcrl5* in immune subsets: unswitched memory B (USM B), switched memory B (SM B), and double negative B (DN B) cells in healthy individuals (n=79). TPM, Transcript per million. (D) eQTL effect of GWAS SNP rs7528684 (*hFcrl3*) and rs11264750 (*hFcrl5*) in the immune subsets from patients with 10 immunological disorders (n=416). P-values were obtained by testing the alternative hypothesis that the slope of a linear regression model between genotype and expression deviates from 0. (E) Representative histograms and gMFIs of Fcrl5 expression on T1 (CD19⁺CD93⁺IgM⁺CD23⁻), T2 (CD19⁺CD93⁺IgM⁺CD23⁺), T3 (CD19⁺CD93⁺IgM⁻CD23⁺), FO B (CD19⁺CD93⁻CD23⁺CD21^{lo}), MZ B (CD19⁺CD93⁻CD23⁻CD21^{hi}), GC B (B220⁺Fas⁺GL7⁺), and plasma (CD138⁺TACI⁺) cells from young WT (n=3) and aged WT mice (n=4). Data are representative of two independent experiments. (F) Representative histograms and gMFIs of Fcrl5 expression of B1a (CD19⁺CD5⁺CD23⁻), B1b (CD19⁺CD5⁻CD23⁻), and B2 (CD19⁺CD5⁻CD23⁺) cells in PeC from young WT (n=3) and aged WT (n=4) mice. Data are representative of two independent experiments. Statistical data are shown as mean values with s.d., and data were analyzed by unpaired Student's t-test. **P<0.01.

Supplementary Figure 2. Analysis of Fcrl5 expression and B cell development in young Fcrl5 Tg mice. (A) A scheme of the constructs used to generate Fcrl5 Tg mice. (B) Representative histograms and gMFIs of Fcrl5 expression on Pro/Pre B (B220⁺IgM⁻), Immature B (Imm. B; B220^{lo}IgM⁺), recirculating B (Recir. B; B220^{hi}IgM⁺) and plasma cells (CD138⁺TACI⁺) in BM, B1a (CD19⁺CD5⁺CD23⁻), B1b (CD19⁺CD5⁻CD23⁻), and B2 (CD19⁺CD5⁻CD23⁺) cells in PeC, GC B (B220⁺Fas⁺GL7⁺) in PPs and T1 (CD19⁺CD93⁺IgM⁺CD23⁻), T2 (CD19⁺CD93⁺IgM⁺CD23⁺), T3 (CD19⁺CD93⁺IgM⁻CD23⁺), CD23⁻CD21^{lo} (CD19⁺CD93⁻CD23⁻CD21^{lo}), FO B (CD19⁺CD93⁻CD23⁺CD21^{lo}), MZ B (CD19⁺CD93⁻CD23⁻CD21^{hi}), GC B (B220⁺Fas⁺GL7⁺), and plasma (CD138⁺TACI⁺) cells in spleen from young WT and young Fcrl5 Tg mice (n=3 mice each). Data are representative of four independent experiments. (C) Absolute numbers of B cell subsets described in (B) from young WT and young Fcrl5 Tg mice (n=3 mice each). Data are representative of five independent experiments. Statistical data are shown as mean values with s.d., and data were analyzed by unpaired Student's t-test. *P<0.05, **P<0.01, ***P<0.001, ****P<0.0001.

Supplementary Figure 3. Analysis of natural antibody titers in young and aged Fcrl5 Tg mice and B cell development in aged Fcrl5 Tg mice. (A) Antibody production in serum isolated from young WT, young Fcrl5 Tg, aged WT, and aged Fcrl5 Tg mice (n=5 mice each), assayed by ELISA. Data are pooled from two independent experiments. (B) Absolute numbers of Pro/Pre B (B220⁺IgM⁻), Immature B (Imm. B; B220^{lo}IgM⁺), recirculating B (Recir. B; B220^{hi}IgM⁺), and plasma cells (CD138⁺TACI⁺) in BM, B1a (CD19⁺CD5⁺CD23⁻), B1b (CD19⁺CD5⁻CD23⁻), and B2 (CD19⁺CD5⁻CD23⁺) cells in PeC, GC B (B220⁺Fas⁺GL7⁺) in PPs and T1 (CD19⁺CD93⁺IgM⁺CD23⁻), T2 (CD19⁺CD93⁺IgM⁺CD23⁺), T3 (CD19⁺CD93⁺IgM⁻CD23⁺), CD23⁻CD21^{lo} (CD19⁺CD93⁻CD23⁻CD21^{lo}), FO B (CD19⁺CD93⁻CD23⁺CD21^{lo}), MZ B (CD19⁺CD93⁻CD23⁻CD21^{hi}), GC B (B220⁺Fas⁺GL7⁺), plasma (CD138⁺TACI⁺) cells, and ABCs (CD19⁺B220^{hi}CD23⁻CD21^{lo}CD11b⁺CD11c⁺) in spleen from aged WT (n=3) and aged Fcrl5 Tg (n=3-4) mice. Data are representative of two or three independent experiments. Statistical data are

shown as mean values with s.d., and data were analyzed by Mann-Whitney test. * $P < 0.05$, ** $P < 0.01$, **** $P < 0.0001$.

Supplementary Figure 4. Autoantibody production and B cell activation in imiquimod-treated Fcrl5 Tg mice. (A) ANAs production in serum from imiquimod-treated (IMQ+) WT (n=16) and Fcrl5 Tg (n=18) for 2, 4, and 8 weeks, detected by immunofluorescence assay using HEp-2 cells. Data are pooled from three independent experiments. (B) Representative histograms and gMFIs of CD69, CD80, CD86, MHC-II, and PD-L1 expression levels on splenic CD19⁺ cells isolated from imiquimod-untreated (IMQ-) WT (n=9) and Fcrl5 Tg (n=9) or -treated (IMQ+) WT (n=17) and Fcrl5 Tg (n=18) mice. Data are pooled from three independent experiments. Statistical data are shown as mean values with s.d., and data were analyzed by one-way ANOVA with Tukey's multiple comparisons test. * $P < 0.05$, ** $P < 0.01$, **** $P < 0.0001$.

Supplementary Figure 5. Requirement of antigen recognition via BCR for imiquimod-induced inflammation. (A) Absolute number of splenocytes, CD19⁺, and CD4⁺ cells in spleen from imiquimod-untreated (IMQ-) WT (n=6-8) and MD4 Tg (n=6-8) or -treated (IMQ+) WT (n=12) and MD4 Tg (n=12) mice. Data are pooled from two independent experiments. (B) Representative flow cytometry plots and absolute numbers of ABCs (CD19⁺B220^{hi}CD23⁻CD21^{lo}CD11b⁺CD11c⁺) from imiquimod-untreated (IMQ-) WT (n=6) and MD4 Tg (n=6) or -treated (IMQ+) WT (n=10) and MD4 Tg (n=10) mice. Data are pooled from two independent experiments. (C) Representative flow cytometry plots (left) and absolute numbers (right) of CD4⁺ T cell subsets: effector T (CD4⁺CD25⁻CD62L⁻CD44^{hi}) and CD4⁺IFN- γ ⁺ cells from imiquimod-untreated (IMQ-) WT (n=6) and MD4 Tg (n=6) or -treated (IMQ+) WT (n=12) and MD4 Tg (n=12) mice. Data are pooled from two independent experiments. Statistical data are shown as mean values with s.d., and data were analyzed by one-way ANOVA with Tukey's multiple comparisons test. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$.