

Supplementary Materials for
**A marmoset brain cell census reveals regional specialization of
cellular identities**

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The PDF file includes:

Figs. S1 to S9
Legends for tables S1 to S7
Legends for data S1 and S2

Other Supplementary Material for this manuscript includes the following:

Tables S1 to S7
Data S1 and S2

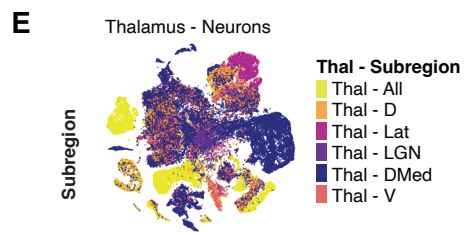
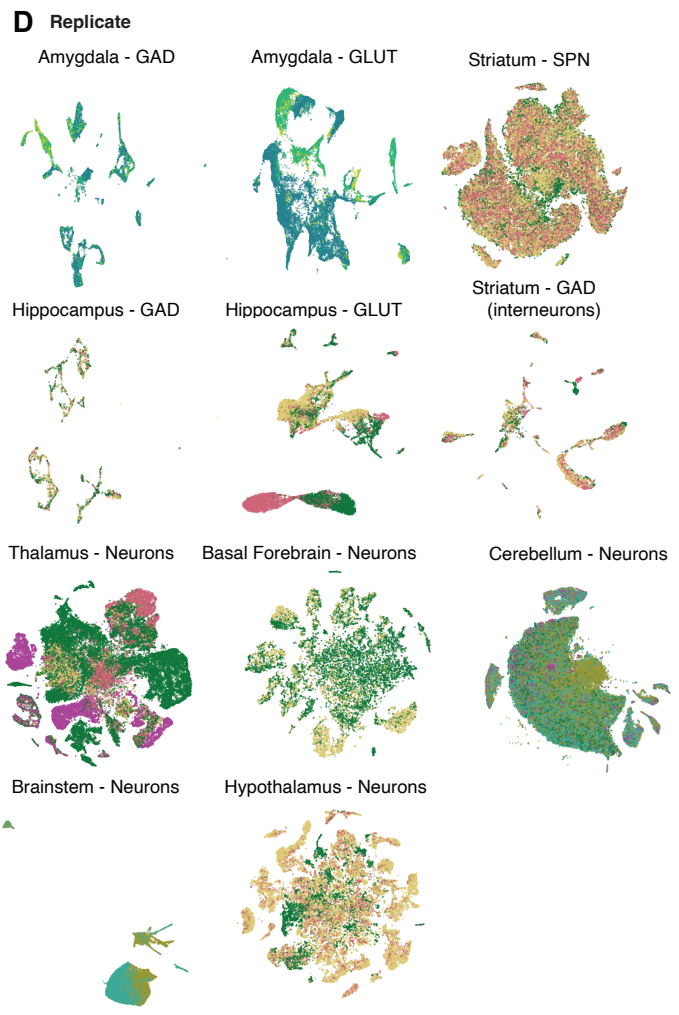
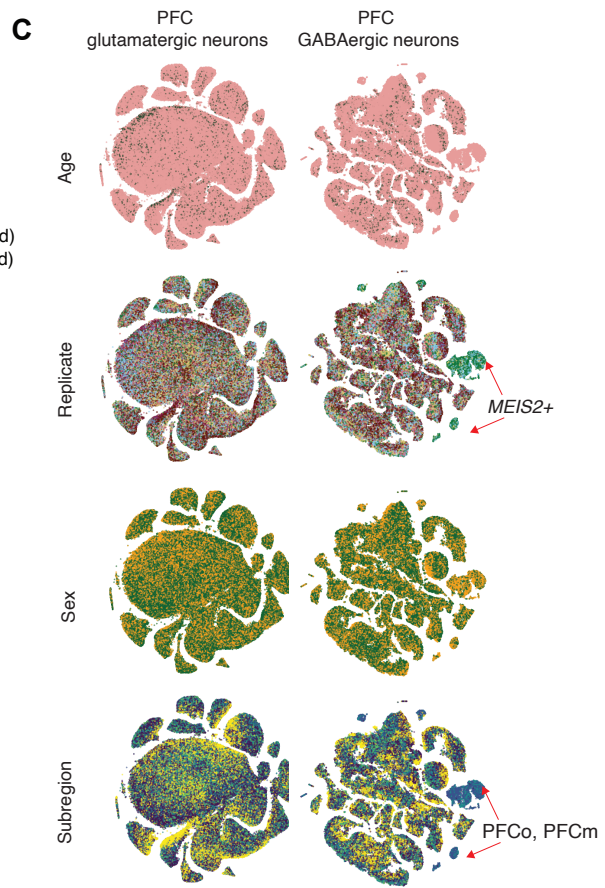
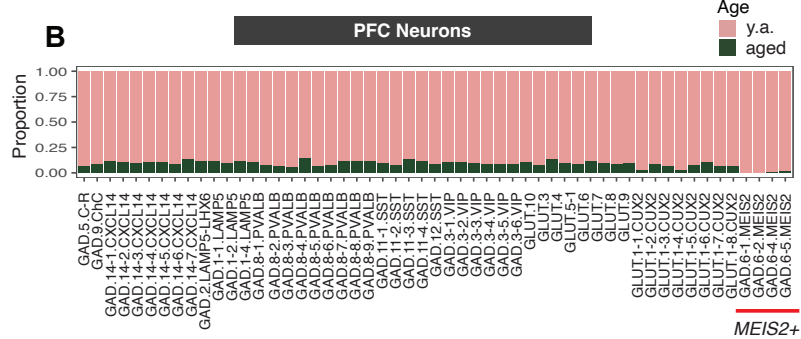
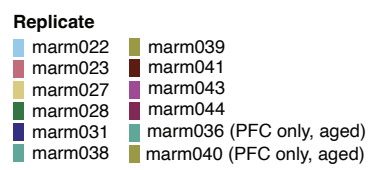
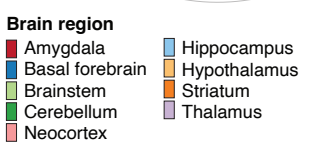
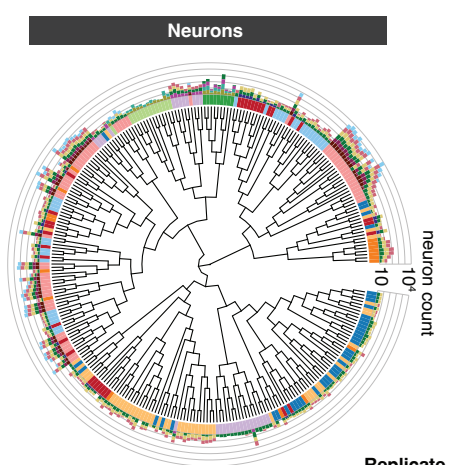


Fig. S1. Neuron counts by donor across brain regions. (A) Neuronal dendrogram as in Fig. 1C, with outer barplots depicting number of nuclei per cell type and replicate. Ring colors are brain regions, colors in barplots correspond to replicates. (B) Proportional per-cluster representation of PFC neurons between young adult donors and aged (n=2) donors. While our snRNA-seq collection focused on post-sexual maturity young adults, we acquired an additional dataset of PFC sampled from 2 aged animals (1 M, 11y5m; 1F, 14y5m, 37,260 cells total; Table S1). Individual replicates contributed similar proportions of neurons to each prefrontal neuron subtype, and clusters generally had proportional representation across young adults and aged animals, as well as across males and females, suggesting that these variables do not dramatically impact neuronal ensembles and identities in prefrontal cortex. (C) t-SNE embeddings of PFC neurons (*top row*, GABAergic; *bottom row*, glutamatergic) with colors representing different metadata: age (young vs aged), replicate, PFC subregion. There was notable enrichment of *MEIS2*+ GABAergic neurons in medial prefrontal and orbital prefrontal dissections. Based on their gene expression profiles, these cells likely correspond to the recently described population of LGE-derived *MEIS2*+ neurons that populate the olfactory bulb in mice, and which are routed to medial prefrontal cortex in macaques and humans(46). (D) t-SNEs of neurons in each brain structure, with cells colored by replicate (colors as in (A)). Telencephalic neurons are plotted separately by class: GABAergic and glutamatergic classes (neocortex, hippocampus, amygdala), or GABAergic interneurons and spiny projection neurons (striatum). Compared with neocortex, greater cross-donor variability was observed in some subcortical structures such as hypothalamus and thalamus, though this was likely driven more by dissection variability than by donor variability, as the donor-specific clusters tended to be from subregions that were only sampled in one individual (see E). (E) t-SNEs of thalamic neurons with cells colored by thalamic subdivision.

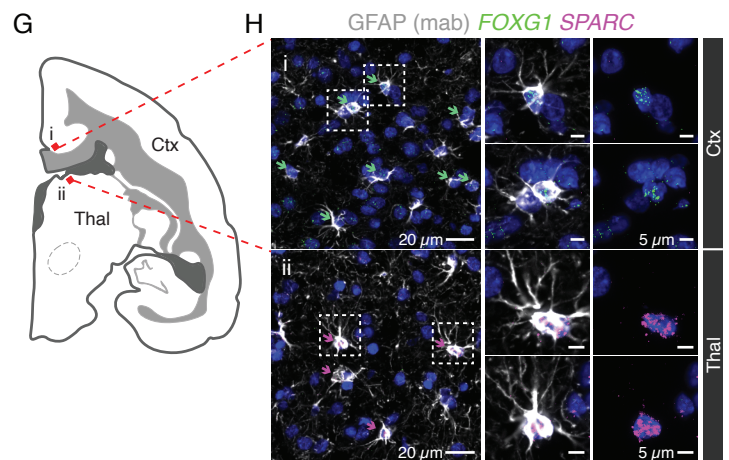
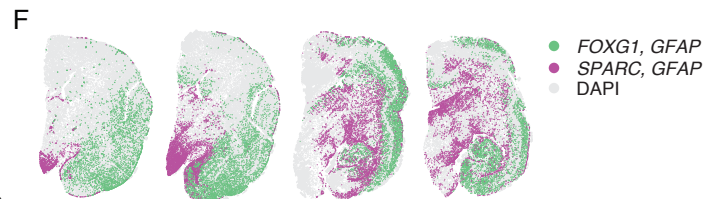
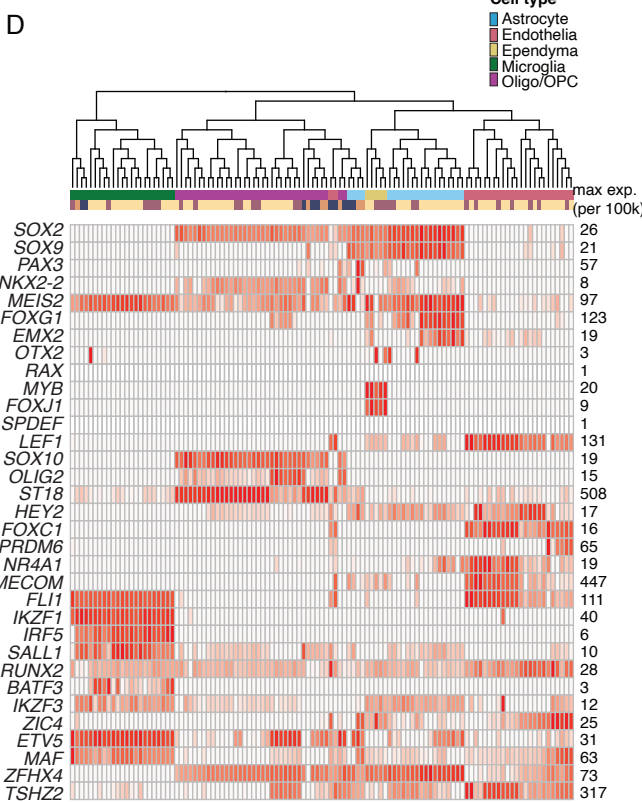
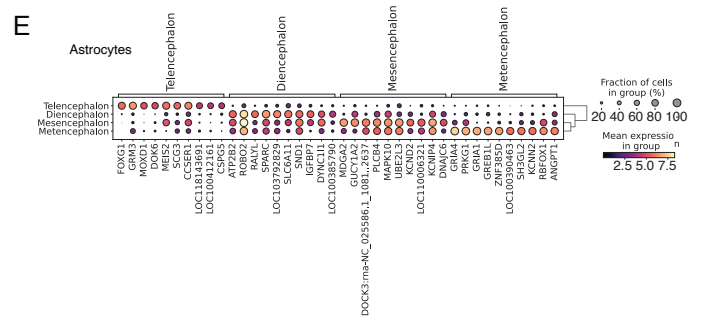
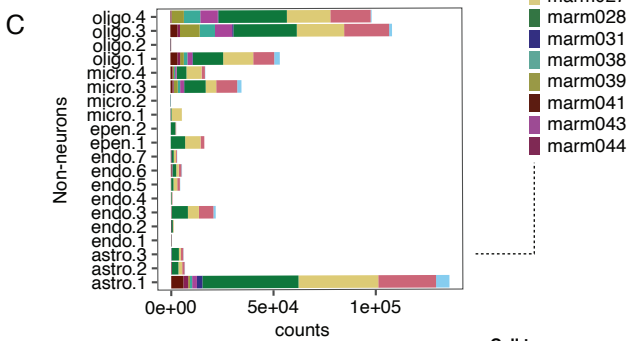
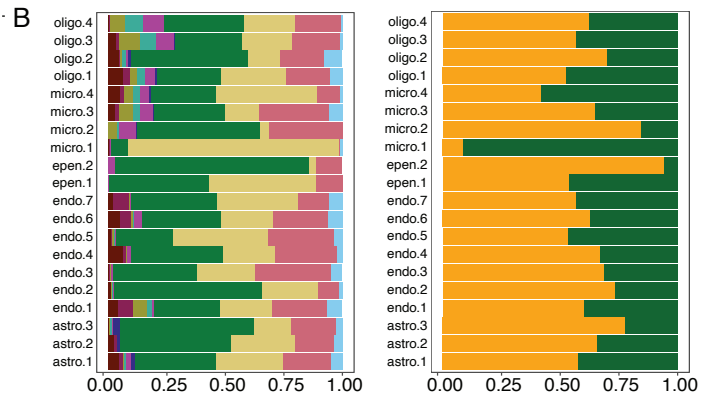
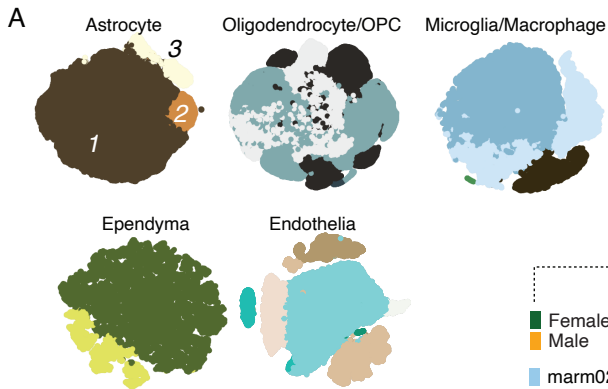
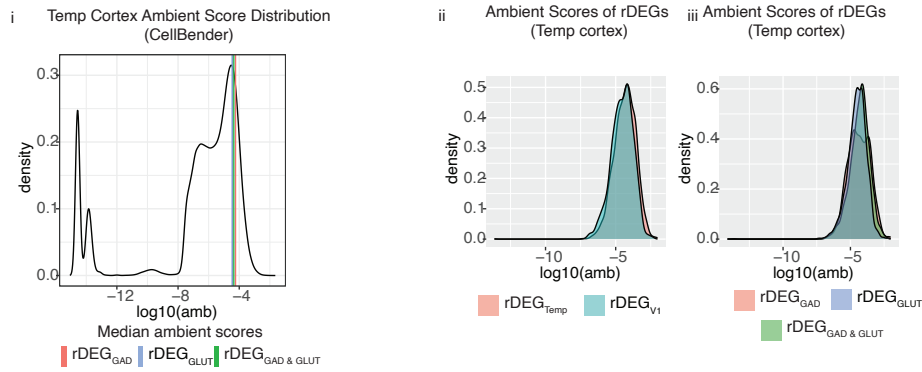


Fig. S2. Glial diversity across brain structures. (A) t-SNE embeddings of major non-neuronal types colored by cluster. (B) Barplots of glial proportions colored by donor and by sex. (C) Non-neuronal nuclei counts. Colors indicate donor, same as (B). (D) Expression of marker genes in non-neurons. Genes as in (1). Heatmap colors are scaled to max normalized expression for each row (gene). Dendrogram and metadata colors as in **Fig. 1D**. (E) Differentially expressed genes in astrocytes across cephalic compartments. (F) Tissue validation for astrocyte differentially expressed genes (*FOXG1*, *SPARC*) in coronal sections of marmoset brain. Green dots indicate locations of cells that stain positive for *GFAP* (IHC, mAb, **Table S4**) and *FOXG1* (smFISH). Magenta dots indicate cell positions for *GFAP* (IHC) and *SPARC* (smFISH). (G) Cartoon of coronal section imaged; Red boxes (*i-ii*) correspond to tissue validation in (H). (H) (*Left*) Fields of view from neocortex and thalamus stained for *GFAP* antibody (gray), *FOXG1* (green), and *SPARC* (magenta). Green arrows highlight *GFAP* cells colocalized with *FOXG1*, magenta arrows highlight *GFAP* cells colocalized with *SPARC*. (*Right*) Magnified examples of double positive cells in neocortex and thalamus. Ctx = cortex, Thal = thalamus.

A



B

Neuron _{GAD}								
	pfc	m1	temp	a1	s1	par	v2	v1
pfc	0	27	2	6	86	34	101	231
m1	5	0	5	1	1	2	2	24
temp	25	33	0	86	102	117	155	352
a1	16	22	13	0	77	70	97	190
s1	57	6	36	25	0	171	108	189
par	19	9	15	10	89	0	33	181
v2	27	1	36	10	18	5	0	54
v1	72	14	109	27	53	62	19	0

Neuron _{GLUT}								
	pfc	m1	temp	a1	s1	par	v2	v1
pfc	0	118	65	136	202	292	401	413
m1	80	0	132	37	9	47	90	108
temp	96	242	0	28	96	173	321	364
a1	152	98	16	0	111	254	314	368
s1	154	35	55	44	0	272	308	358
par	131	36	87	53	105	0	74	280
v2	201	38	175	111	89	31	0	82
v1	211	57	189	120	150	141	58	0

Oligodendrocyte lineage								
	pfc	m1	temp	a1	s1	par	v2	v1
pfc	0	39	13	3	65	17	81	90
m1	15	0	7	9	12	3	5	3
temp	37	57	0	41	103	47	25	41
a1	201	10	2	0	121	62	76	128
s1	186	5	3	14	0	79	88	99
par	153	2	1	16	121	0	10	99
v2	191	13	4	40	96	34	0	83
v1	141	6	9	37	84	45	33	0

Astrocytes								
	pfc	m1	temp	a1	s1	par	v2	v1
pfc	0	71	23	29	149	42	72	151
m1	22	0	30	3	4	1	8	8
temp	44	38	0	12	32	24	51	50
a1	98	24	6	0	95	59	46	86
s1	104	3	12	21	0	86	55	83
par	221	19	11	57	204	0	95	248
v2	81	11	29	26	36	6	0	23
v1	134	14	43	44	68	53	14	0

Fig. S3. Cortical rDEGs do not reflect ambient RNA contamination and vary across region pairs. (A) (i) rDEG ambient score distributions (from CellBender (111)) in temporal cortex samples, which had amongst the highest numbers of rDEGs compared to other neocortical regions. Despite glutamatergic neurons being more numerous and having more expressed genes/transcripts per cell, the median ambient contamination scores for glutamatergic rDEGs were not higher than median contamination scores for GABAergic rDEGs. rDEGs shared between glutamatergic and GABAergic neurons had indistinguishable scores compared with rDEGs private to one neuronal class. (ii) Ambient scores in temporal cortex of temporal cortex rDEGs are indistinguishable from V1 rDEGs. (iii) Distributions of temporal cortex rDEG ambient scores by neuron class, again showing no difference between rDEGs that are shared or private to a neuronal class. (B) Numbers of regionally differentially expressed genes (rDEGs) between pairs of cortical regions for neurons, astrocytes, and oligodendrocyte lineage types.

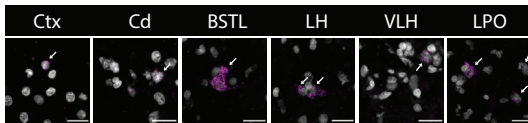
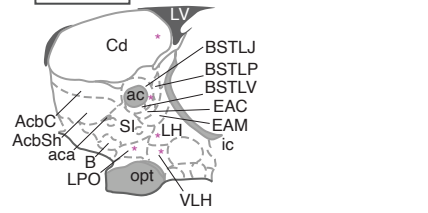
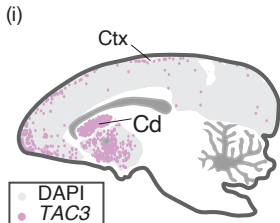
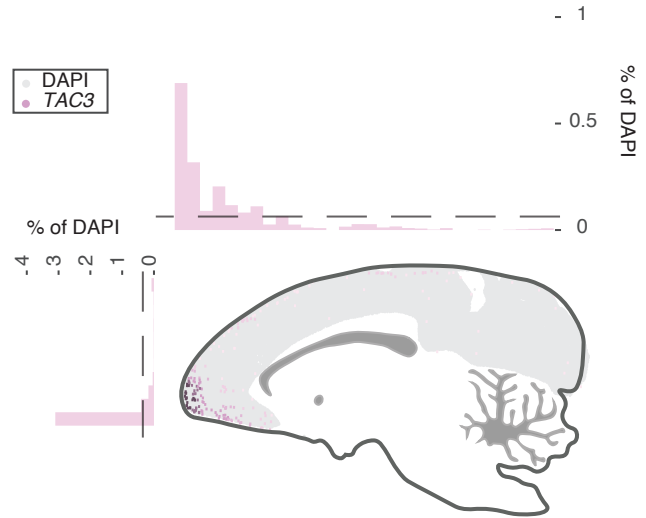
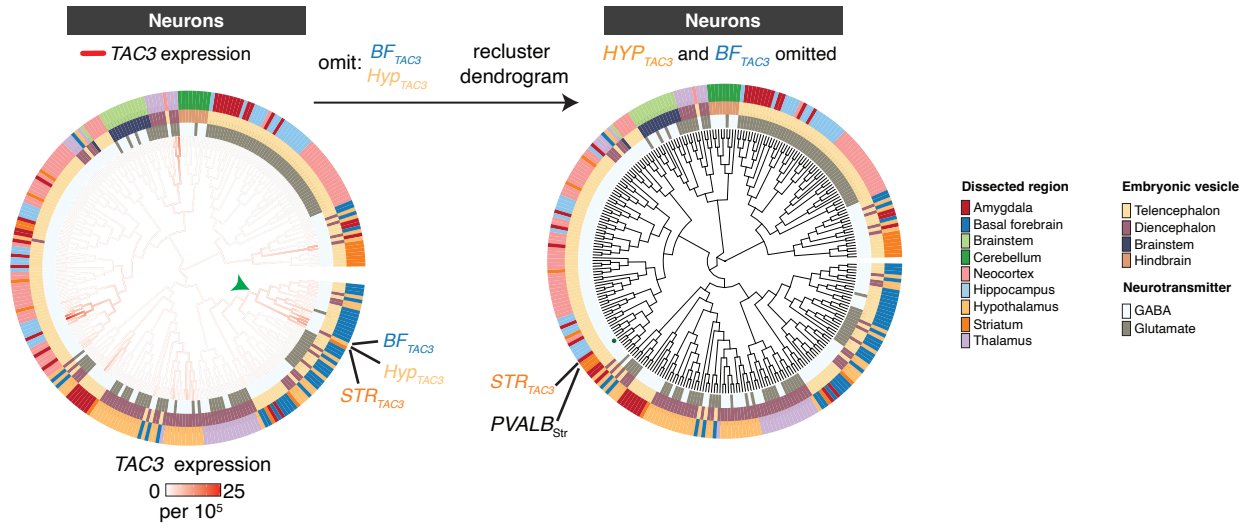
A**TAC3 expression (smFISH)****B****TAC3 distribution in neocortex (smFISH)****C**

Fig. S4. Locations of *TAC3*+ cells in marmoset forebrain. (A) smFISH reveals anatomical locations and expression levels *TAC3*+ types in different brain regions. (i) Schematic of *TAC3*+ cells imaged across cortex, dorsal striatum. Ctx = neocortex, Cd = Caudate. (ii) Cartoon close-up of nuclei in striatum, basal forebrain and hypothalamus. Magenta stars = locations of *TAC3*+ cells in lower image panel. Cd = Caudate, AcbC = nucleus accumbens core, AcbSh = nucleus accumbens shell, SI = Substantia innominata, B = basal nucleus of Meynert, EAM = extended amygdala, medial, EAC = extended amygdala, central, ac = anterior commissure, BSTLP = bed nuc st, lateral posterior, BSTLJ = bed nuc st, juxtacap, BSTLV = bed nuc st, lateral ventral, LH = lateral hypothalamus, VLH = ventrolateral hypothalamus, LPO = lateral preoptic area. (B) Density and location of *TAC3*+ cells as proportion of all DAPI+ cells. Barplots show percentages in bins (approximately 1,290 μm per bin) taken across the anterior-posterior (top) and dorsal-ventral (left side) axes. (C) Effect on placement of the *TAC3*+ striatal type on the neuronal dendrogram when omitting the two *TAC3*+ types in hypothalamus and basal forebrain. When these types are omitted and hierarchical clustering is repeated (using HCA_{PCA}), the *TAC3*+ striatal type is most similar to *PVALB*+ striatal interneurons, consistent with previous reports that only compared telencephalic interneurons(10).

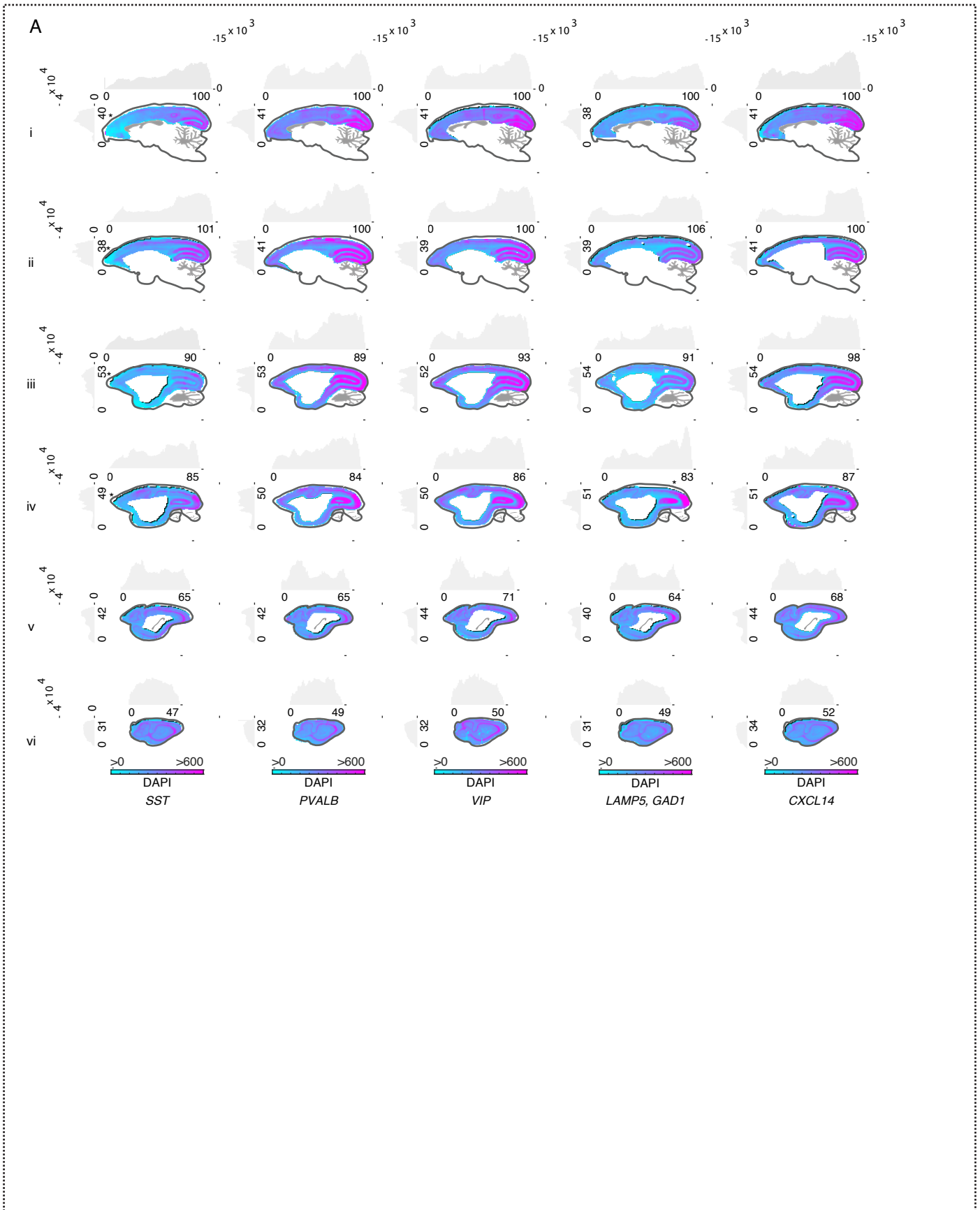


Fig. S5. Estimated cell numbers across marmoset neocortex. (A) Total numbers of DAPI+ cells per unit area (approximately 387 μm per bin) for each of the sections shown in **Fig. 6C**.

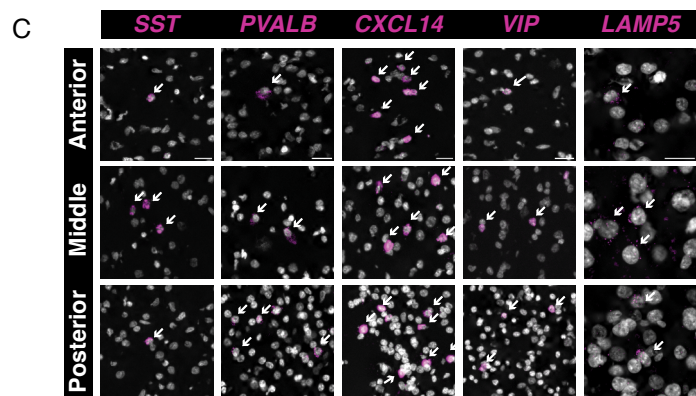
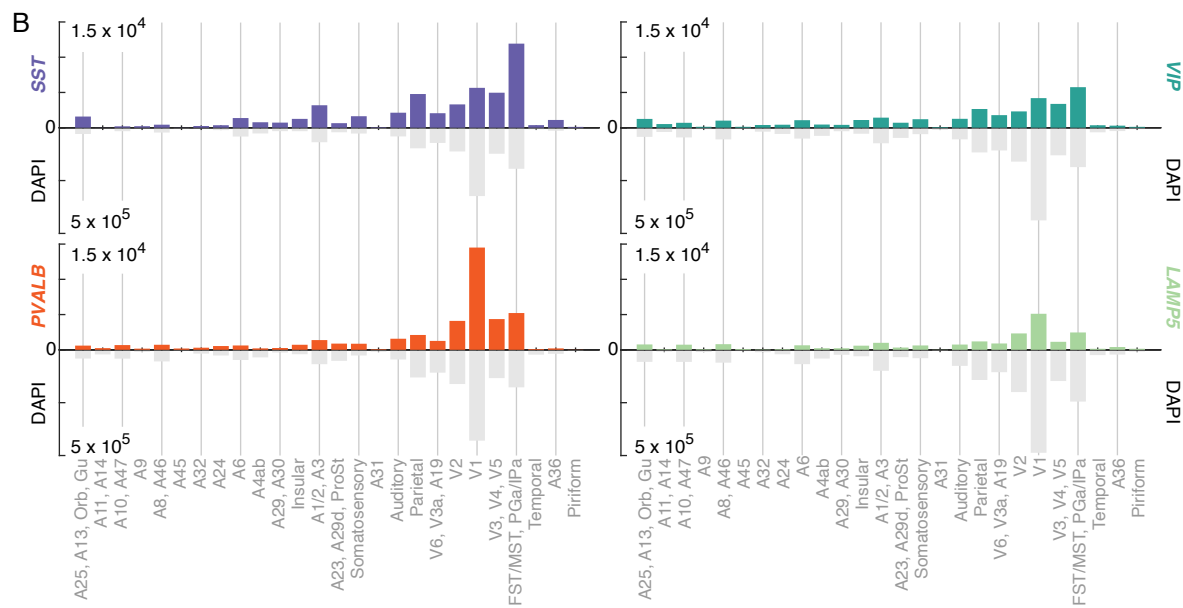
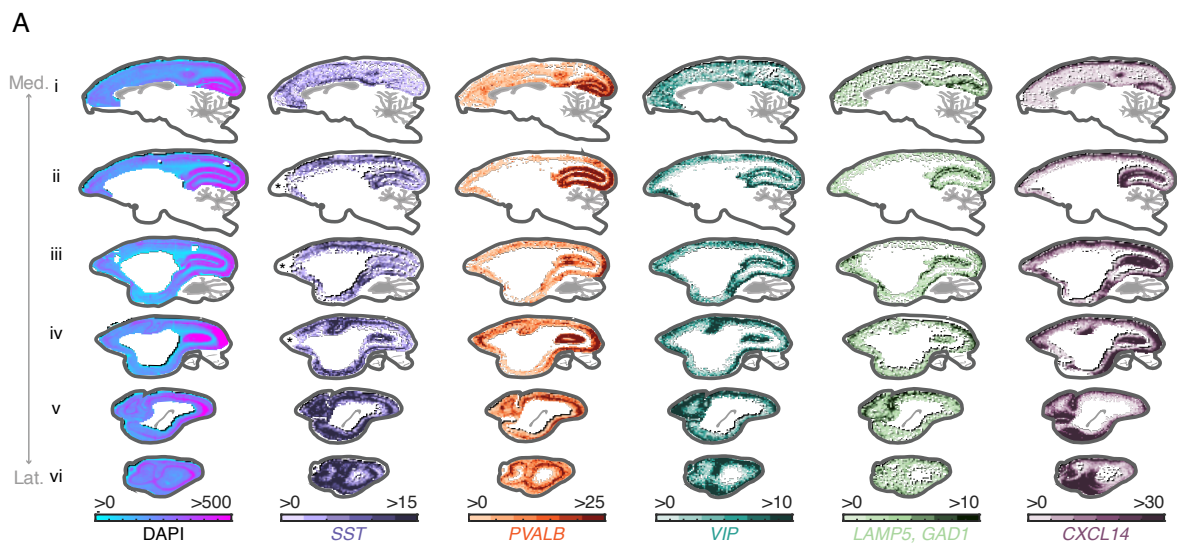


Fig. S6. Estimated interneuron numbers across marmoset neocortex. (A) smFISH for neocortical interneuron subclass markers showing locations of cells positive for each marker across 6 sagittal sections of the marmoset neocortex. Heatmap scale shows absolute density per unit area (approximately 387 μm per bin). First column shows DAPI and area profiled. (B) Quantification of interneurons (*SST*, *PVALB*, *VIP*, and *LAMP5 (GAD1+)*) and DAPI cells by cortical area in marmoset parcellated according to **Fig. 6F** (anterior to posterior). DAPI counts are plotted on the inverse *y*-axis. (C) Examples of smFISH images quantitated in **Fig. 6B-E**. Panels for each marker show example positive cells in anterior, middle, and posterior locations across neocortex. White arrows indicate positive cells. Scale bar = 20 μm .

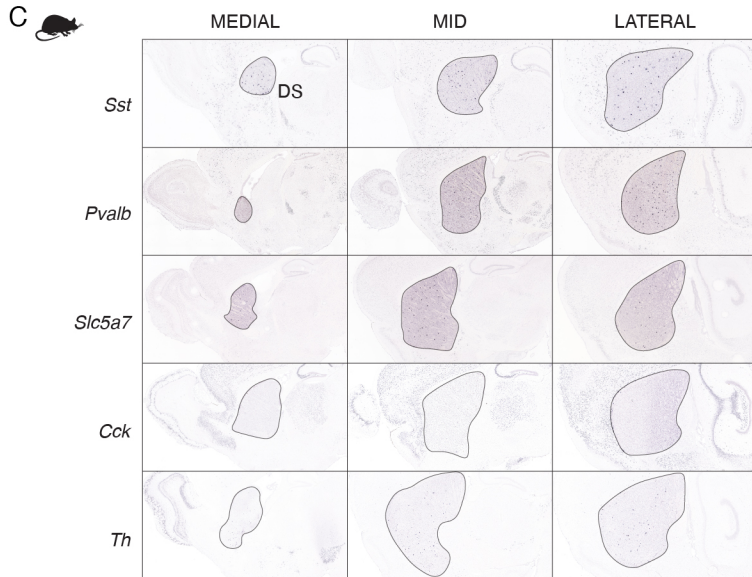
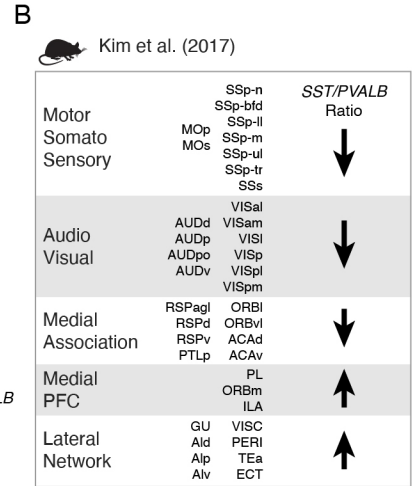
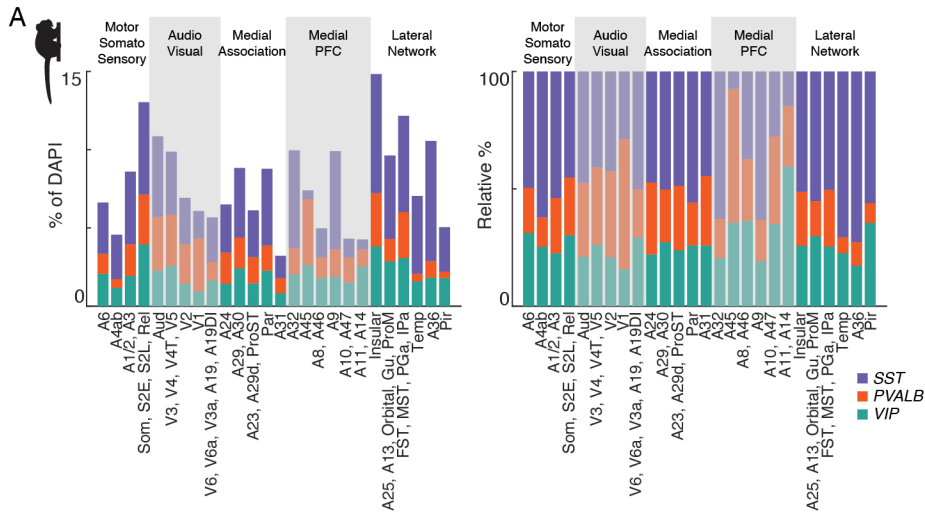


Fig. S7. Interneuron proportions in marmoset and mouse. (A) Quantitation of interneuron proportions by cortical area in marmoset parcellated according to **Fig. 6F**. (B) Schematization of interneuron proportions by cortical area in mouse reproduced from (22). (C) Mouse striatal *in situ* from Allen Brain Atlas for *Sst*, *Pvalb*, *Slc5a7*, *Cck*, *Th*; medial to lateral. DS = dorsal striatum.

Cortex

BIL: 79e50b4a8a5c2a91
Type: CXCL14



BIL: 735ad76f1301ab79
Type: CXCL14



BIL: a08d0191a7f0636a
Type: LAMP5



BIL: a378a612a104daa8
Type: LAMP5



BIL: a08d0191a7f0636a
Type: PVALB



BIL: c57067b0137f63e7
Type: SST



BIL: c57067b0137f63e7
Type: SST



Striatum

BIL: fefc9e1a639ec580
Type: SST



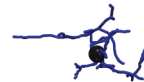
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Type: TH



BIL: c5aeb41a58b8eb45
Type: PVALB



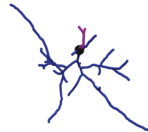
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Type: TH, PVALB



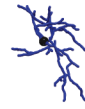
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Type: SLC5A7, CHAT



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Type: CCK



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BIL: 073974be411daf5a
Type: VIP

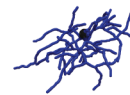


Fig. S8. Morphology examples using NeuTube reconstructions. Example morphological reconstructions of striatal and neocortical interneurons using the NeuTube pipeline. Each cell, along with associated smFISH staining, is available for download at <https://doi.org/10.35077/g.609>.

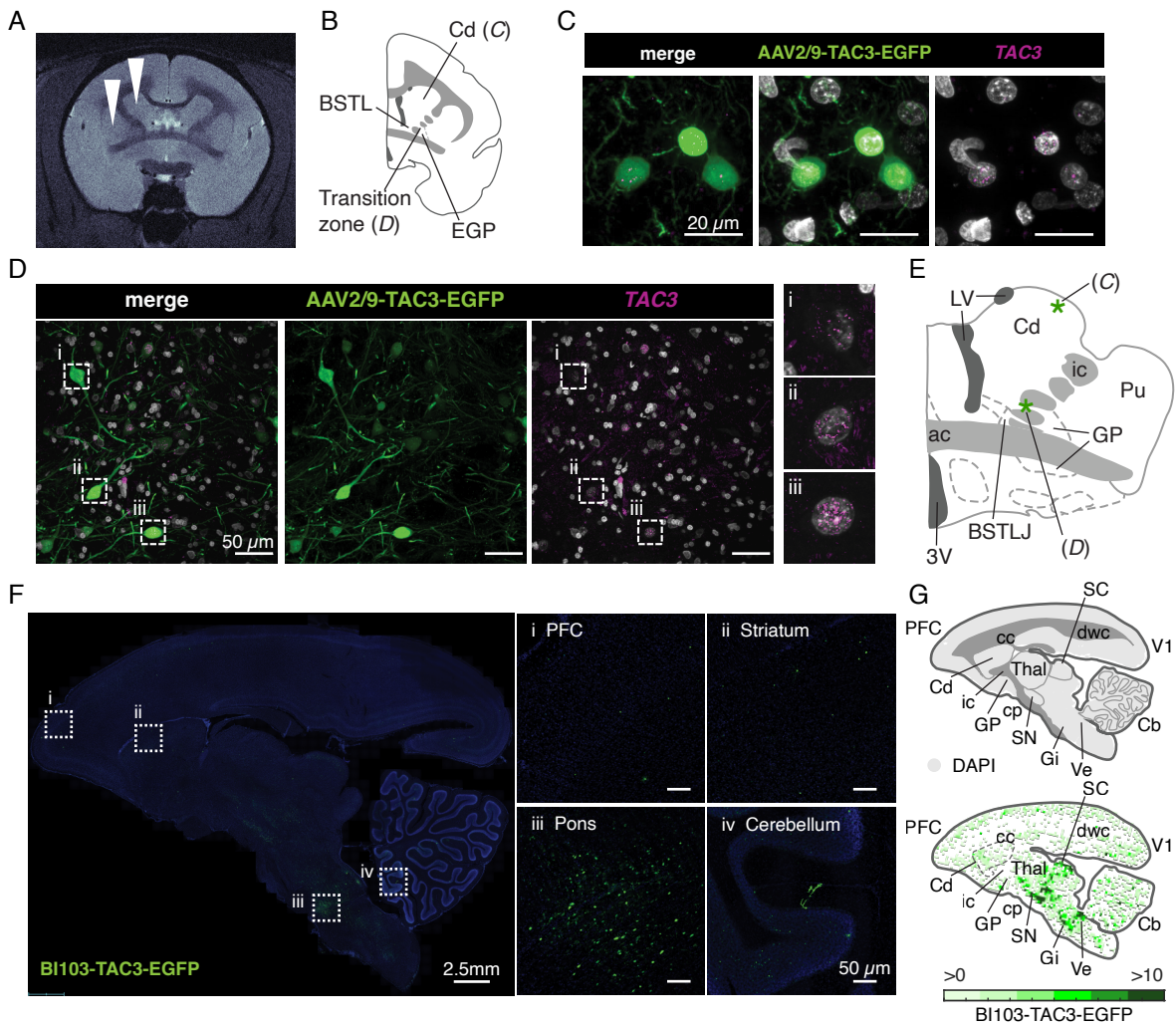


Fig. S9. Examples of striatal and peri-striatal *TAC3*+ neurons labeled by AAV-tandemE-TAC3-EGFP. (A) MRI showing injection location (white arrowheads) of virus into unilateral striatum (caudate and putamen) in one animal (Cj 17-B111, **Table S1**). (B) Cartoon showing location of positive cells in (C) as well as labeled cells in transition zone (D). (C) EGFP antibody-amplified confocal image of a labeled cell (position shown in (B)) with smFISH for *TAC3* showing colocalization. Scale bar = 20 μm . (D) Examples of extra-striatal labeled cells from injections in (A). Position shown in (B). Scale bar = 50 μm . (E) Close up of position of cells in (C) and (D). Cd = caudate, Pu = putamen, ic = internal capsule, ac = anterior commissure, BSTLJ = bed nuc st, juxtacap, EGP = external globus pallidus, 3V = third ventricle, LV = lateral ventricle. (F) Whole sagittal section (20x image) of adult marmoset showing cells transduced by the AAV-BI103-tandemE-TAC3-EGFP virus in marmoset, Cj 20-214 (**Table S1**). EGFP+/*TAC3*+ cells were detected sparsely in neocortex as well as striatum, cerebellum, substantia nigra, superior colliculus, and brainstem. Scale bar = 2.5 mm. (i) Prefrontal cortex, (ii) Striatum, (iii) Pons, (iv) Cerebellum. Scale bar = 50 μm . (G) Heatmap showing the locational density of cells transduced by the AAV-BI103-tandemE-TAC3-EGFP virus across one sagittal section of the marmoset brain after systemic IV injection - (top) reference cartoon superimposed on a scatterplot of DAPI cells profiled (light gray) and (bottom) density plot of EGFP+ cells. Heatmap scale shows absolute density per unit area (approximately 344 μm per bin). PFC = prefrontal cortex, Cd = caudate, cc = corpus callosum, ic = internal capsule, GP = globus pallidus, Thal = thalamus, cp = cerebral peduncle, SN = substantia nigra, Gi = gigantocellular reticular nucleus, SC = superior colliculus, dwc = deep cortical white matter, Ve = vestibular nucleus, V1 = visual cortex V1, Cb = cerebellum.

Supplementary Table Legends (separate files)

Table S1. Marmoset sample information. Table of animals, metadata, and experimental information.

Table S2. snRNA-seq dataset by donor and brain area. Tables of the number of cells per brain structure and samples per donor, dendrogram of metacells, PC scores.

Table S3. Neocortical rDEGs across three donors. Neocortical rDEGs across three donors with pairwise comparisons between neocortical locations for major clusters of cortical excitatory neurons, inhibitory neurons, astrocytes and oligodendrocyte lineage types.

Table S4. Fluorescent *in situ* hybridization probes and antibodies used. A list of all FISH probes (RNA-Scope and Molecular Instruments) and antibodies used for validation of gene and protein expression *in situ*.

Table S5. Morphological reconstructions performed with Neutube. A list of all cells reconstructed with Neutube with their corresponding morphological measurements.

Table S6. Morphological reconstructions performed with Imaris. A list of all cells reconstructed with Imaris with their corresponding morphological measurements.

Table S7. Links and DOIs. A list of all links and DOIs referenced.

Supplementary Data Legends (separate files)

Data S1. *TAC3* medio-lateral quantification. Tabular data for *TAC3* medio-lateral histograms and barplots shown in **Fig. S4**.

Data S2. Striatal and cortical medio-lateral quantification. Tabular data for striatal and cortical medio-lateral histograms/bar plots shown in **Figs. 6-7**.