

A Differential Microbiological Assay for Vitamin B₁₂ and Pseudovitamin B₁₂^{1,2}

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Although vitamin B₁₂ can be assayed biologically with mice, chicks or rats, the microbiological method of assay is preferred since it is more rapid and economical. One serious shortcoming of many microbiological B₁₂ assay procedures is the lack of specificity when crude materials are assayed. Crude materials can contain inhibitory or stimulatory factors which cause serious errors in assay results.

One of the interfering substances in the microbiological assay of vitamin B₁₂ with *Lactobacillus leichmannii* is pseudovitamin B₁₂. Pfiffner *et al.* (1951) isolated an organism from bovine rumen contents which produced a new form of vitamin B₁₂ which was completely inactive for chicks, rats, and humans, but was active for *L. leichmannii* and *Lactobacillus lactis* var. Dorner. These workers crystallized two major red pigments from an anaerobic fermentation broth containing this rumen organism and named the pigments pseudovitamin B₁₂ and pseudovitamin B_{12b}. The chemical differences between pseudovitamin B₁₂ and vitamin B₁₂ have been described (Dion *et al.*, 1952; Pfiffner *et al.*, 1952). Lewis *et al.* (1952) reported that a vitamin B₁₂ analog which they designated vitamin B_{12f} appeared to be identical with pseudovitamin B₁₂. Using paper chromatography in conjunction with a bioautographic technique, these workers found that vitamin B_{12f} was present in the fecal matter of the rat, cow, sheep, pig, horse, chicken, guinea pig, and man. Sheep rumen contents were also a source of vitamin B_{12f}. No vitamin B_{12f} was detected in rat or beef liver or in a sample of injectable liver extract.

The experimental work described in this paper was conducted in an attempt to develop a simple microbiological procedure for detecting the presence of pseudovitamin B₁₂ in crude samples. In addition, the application of the method in the differential quantitative assay of vitamin B₁₂ and pseudovitamin B₁₂ was also investigated.

The observation by O. D. Bird (personal communica-

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tion, 1950) that pseudovitamin B₁₂ was almost as potent as vitamin B₁₂ for *L. leichmannii* 7830, but was much more active than vitamin B₁₂ for *Lactobacillus acidophilus* 832, was the basis for this work.

MATERIALS AND METHODS

The test organisms, *Lactobacillus leichmannii* strain ATCC 7830 and *L. acidophilus* strain ATCC 832 were carried in stock cultures using a tomato juice-yeast extract agar culture medium (A.O.A.C., 1953). The same medium minus agar was used for broth subcultures of the organisms. In preparing the inoculum for assays, 24-hour subcultures of the organisms were centrifuged and the cells washed three times with saline. The turbidity of the suspended cells was adjusted to 70 per cent light transmission (Coleman Model 14 Spectrophotometer) for *L. leichmannii* and 50 per cent transmission for *L. acidophilus*. One loop of the cell suspension was used to inoculate each assay tube.

The assay medium for both organisms was a commercial vitamin B₁₂ assay broth⁴ (Pharmacopeia of the United States, 1951) supplemented with 400 µg of pantethine⁵ per liter of double strength medium. Five ml of the double strength medium were added to each assay tube. The assay tubes were 16 x 20 mm and were graduated at 10 ml. The volume of each tube was adjusted to 10 ml with water after the samples were added. The tubes were covered with glass caps and sterilized at 15 pounds pressure for 5 minutes. After cooling, the tubes were inoculated with a wire loop and then incubated for 20 hours at 37 C. Growth was determined turbidimetrically using a Coleman Spectrophotometer containing a PC-4 filter at a wave-length setting of 650 mµ.

Samples were extracted with 0.1 N acetate buffer containing 1 per cent sodium bisulfite. In most experiments a trace of cyanide was also added to prevent the conversion of vitamin B₁₂ to B_{12a}. The assay results on crude samples were the same in the presence or absence of cyanide. After autoclaving at 15 pounds pressure for 20 min, the sample was adjusted to pH 6, brought to

⁴ Difco Bacto-B₁₂ Assay Medium USP, dehydrated.

⁵ Kindly supplied by Dr. O. D. Bird, Parke, Davis and Co., Detroit, Michigan.

TABLE 1. The growth of *Lactobacillus leichmannii* and *Lactobacillus acidophilus* in Difco medium with and without added pantethine*

Organism	Vitamin B ₁₂ per Tube	No Pantethine	Plus Pantethine*
		Light transmission	Light transmission
	μg	<i>per cent</i>	<i>per cent</i>
<i>L. leichmannii</i>	0.0	100.0	100.0
	0.04	67.0	67.0
	0.08	45.5	42.5
	0.12	35.5	36.5
	0.20	27.5	25.0
<i>L. acidophilus</i>	0.0	100.0	100.0
	0.4	93.5	69.5
	0.8	100.0	54.0
	1.2	100.0	41.5
	1.6	98.0	34.5

* Four hundred μg of pantethine added per liter of double strength medium.

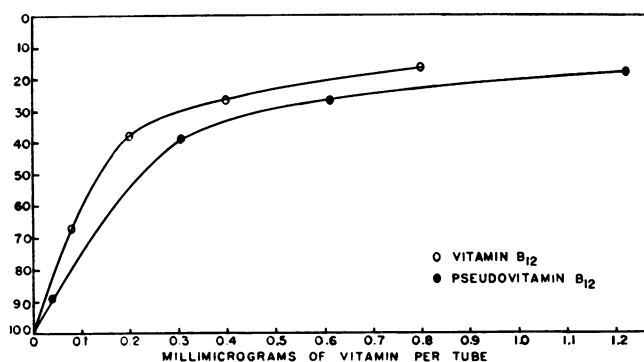


FIG. 1. The growth response of *Lactobacillus leichmannii* ATCC 7830 to vitamin B₁₂ and pseudovitamin B₁₂.

a convenient volume, filtered and the filtrates assayed. Four or more levels of each sample were used in an assay. Duplicate tubes were used for each level of sample. None of the samples assayed contained appreciable alkali-stable activity, thus indicating the absence of desoxyribosides.

Crystalline vitamin B₁₂⁶ was used as a standard in the range of 0.01 to 0.8 millimicrograms per tube for *L. leichmannii*, and from 0.05 to 2.0 millimicrograms for *L. acidophilus*. Crystalline pseudovitamin B₁₂⁷ was used in the range of 0.04 to 1.2 and 0.02 to 0.2 millimicrograms per tube for *L. leichmannii* and *L. acidophilus* respectively.

RESULTS AND DISCUSSION

In order to employ *L. leichmannii* and *L. acidophilus* in a differential assay to detect pseudovitamin B₁₂, it was deemed advisable to first find a convenient assay

⁶ Kindly supplied by Dr. E. L. R. Stokstad, Lederle Labs. Pearl River, New York

⁷ Kindly supplied by Dr. J. J. Piffner, Parke, Davis and Co., Detroit, Michigan

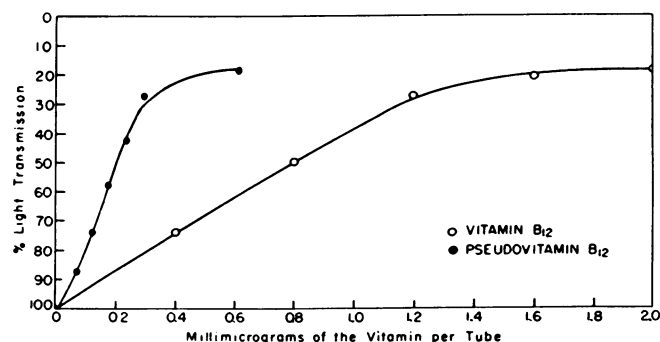


FIG. 2. The growth response of *Lactobacillus acidophilus* ATCC 832 to vitamin B₁₂ and pseudovitamin B₁₂.

medium which would support growth of both organisms.

The Difco medium was found to be a satisfactory assay medium for *L. leichmannii*; however, *L. acidophilus* would not grow in this medium unless pantethine was added. Table 1 shows that the Difco medium supported good growth of *L. acidophilus* when supplemented with pantethine. The addition of pantethine had no effect on the response of *L. leichmannii* to vitamin B₁₂.

Repeated assays were conducted in which the relative potency of vitamin B₁₂ and pseudovitamin B₁₂ was determined for both test organisms. The results of two typical assays are shown in figures 1 and 2. The average of a number of assays showed that pseudovitamin B₁₂ was 0.66 as active as vitamin B₁₂ for *L. leichmannii*. This agrees with the potency of 0.7 observed by Piffner *et al.* (1954). In the *L. acidophilus* assay, pseudovitamin B₁₂ was 3.3 times as active as vitamin B₁₂.

Using crystalline vitamin B₁₂ as a standard, mixtures of crystalline vitamin B₁₂ and crystalline pseudovitamin B₁₂ were assayed with both organisms to learn if there was any interaction between the two analogs. The results of these assays are shown in table 2. The experimental activity of the mixtures was in good agreement with the calculated theoretical activity. The method of calculation using the factors 0.66 and 3.3 for the potency of pseudovitamin B₁₂ is shown at the bottom of table 2.

Based on these results, it is obvious that a sample containing only vitamin B₁₂ would yield identical assay results with either *L. leichmannii* or *L. acidophilus*. If the sample contained pseudovitamin B₁₂ as well as vitamin B₁₂, the *L. acidophilus* assay would give a higher value since pseudovitamin B₁₂ is 3.3 times as active as B₁₂ for *L. acidophilus*. If we let x equal the vitamin B₁₂ content, then $x + 0.66y$ would equal the assay value obtained using *L. leichmannii* and $x + 3.3y$ would equal the assay values obtained using *L. acidophilus*. Thus we have two equations and two unknowns which can be solved simultaneously to determine the vitamin B₁₂ and pseudovitamin B₁₂ content of the sample.

TABLE 2. Microbiological assay of mixtures containing crystalline vitamin B₁₂ and pseudovitamin B₁₂

Assay Organism	Composition of Mixture		Theoretical Activity*	Experimental Activity†	Per Cent of Theoretical
	Vitamin B ₁₂	Pseudovitamin B ₁₂			
	μg	μg	μg	μg	
<i>Lactobacillus leichmannii</i> (Typical assay)...	0.05	0.030	0.070	0.066	95
	Avg of 4 experiments involving 20 mixtures, 90. Range, 85-97.				
<i>Lactobacillus acidophilus</i> (Typical assay)...	0.10	0.029	0.20	0.21	105
	Avg of 4 experiments involving 28 mixtures, 98. Range, 92-104.				

* For *L. leichmannii* theoretical activity was calculated using the factor of 0.66 as the potency of pseudovitamin B₁₂. That is, B₁₂ in mixture + (0.66) × (pseudovitamin B₁₂ in mixture) = theoretical B₁₂ activity. For *L. acidophilus* theoretical activity was calculated using the factor 3.3 as the potency of pseudovitamin B₁₂. That is, B₁₂ in mixture + (3.3) × (pseudovitamin B₁₂ in mixture) = theoretical B₁₂ activity.

† Experimental B₁₂ activity was determined by assaying the mixture using crystalline vitamin B₁₂ as the standard.

To test the accuracy of the above calculations, an unknown mixture of the crystalline vitamins was prepared and assayed with both microorganisms. The unknown mixture contained 3.0 millimicrograms of vitamin B₁₂ and 0.47 millimicrograms of pseudovitamin B₁₂ per ml. The average results of 4 assays calculated by use of simultaneous equations showed that the unknown mixture contained 3.12 millimicrograms of vitamin B₁₂ and 0.423 millimicrograms of pseudovitamin B₁₂ per ml. The experimental data was 104 per cent of theoretical for vitamin B₁₂ and 90 per cent of theoretical for pseudovitamin B₁₂.

The results obtained from the assay of various samples are presented in table 3. Liver and kidney samples and vitamin B₁₂ supplement #2 appear to contain only vitamin B₁₂; fish solubles, rabbit feces and chick droppings contain appreciable amounts of non-B₁₂ activity. Vitamin B₁₂ supplement #1 may contain a small quantity of non-B₁₂ activity. These results are in good agreement with the distribution of B₁₂ analogs reported by other workers (Lewis *et al.*, 1952; Coates *et al.*, 1953).

Assuming that vitamin B₁₂ and pseudovitamin B₁₂ are the only two growth stimulatory factors present in fish solubles, rabbit feces and chick droppings, the concentration of both of these vitamins can be calculated using the simultaneous equations mentioned previously. The results of such theoretical calculations showed the following vitamin content per g: fish solubles #1, 0.26 μg of B₁₂ and 0.17 μg of pseudovitamin B₁₂; fish solubles #2, 0.23 μg of B₁₂ and 0.095 μg of pseudovitamin B₁₂;

TABLE 3. Microbiological assay of various materials with *Lactobacillus leichmannii* and *Lactobacillus acidophilus*

Sample	Avg. B ₁₂ Activity		Pseudovitamin B ₁₂ or Other Analogs Present
	<i>L. leichmannii</i>	<i>L. acidophilus</i>	
	μg/g	μg/g	
Lyophilized beef liver.....	2.14	2.22	No
Lyophilized beef kidney.....	0.85	0.89	No
Fresh chick liver.....	0.13	0.14	No
Fresh chick kidney.....	0.14	0.14	No
Fish solubles #1*.....	0.37	0.83	Yes
Fish solubles #2*.....	0.29	0.54	Yes
Vitamin B ₁₂ feed supplement #1.....	11.50	14.20	?
Vitamin B ₁₂ feed supplement #2.....	2.53	2.40	No
Rabbit feces.....	0.31	1.40	Yes
Fresh chick droppings†.....	0.07	0.30	Yes

* These samples were stored at room temperature for several months before they were assayed.

† Obtained from vitamin B₁₂ deficient chicks which were 5 weeks old.

chick droppings, 0.013 μg of B₁₂ and 0.087 μg of pseudovitamin B₁₂.

The reports of Holdsworth (1953) and Ford and Porter (1953) show that crude products such as feces contain B₁₂ analogs called Factors A, B, and C in addition to pseudovitamin B₁₂. The presence of numerous analogs of B₁₂ in natural products and the difference in microbiological potency of these analogs makes it very unlikely that the differential assay procedure described in this paper is applicable to quantitative assays of crude samples.

DISCUSSION

The detection of non-B₁₂ activity in crude samples appears to be possible by means of assaying the sample with the two test organisms using the method described here. This method involves one medium and that medium can be obtained commercially. The method is therefore more simple than the chromatographic and bioautographic procedures of Lewis *et al.* (1952) or the microbiological procedures used by Coates *et al.* (1953).

The use of the quantitative differential assay procedure is satisfactory when using pure mixtures of vitamin B₁₂ and pseudovitamin B₁₂, but is probably not valid for crude products since they may contain additional B₁₂ analogs.

Ford (1953) developed a microbiological B₁₂ assay which he reported to be specific for vitamin B₁₂. The procedure employs the organism *Ochromonas malhamensis* in a 72-hour assay at 29 C with constant shaking during the incubation period. The procedure outlined in this paper for the detection of non-B₁₂ activity in crude samples might have application in the rapid screening of a large number of samples. Those

samples which show higher values with *L. acidophilus*, thus indicating the presence of non-B₁₂ activity, could then be reassayed using the more tedious method employing *Ochromonas malhamensis*.

The presence of non-B₁₂ activity in samples of fish solubles is of interest since fish solubles are used commercially as a source of vitamin B₁₂ in animal feeds. Fish solubles are stored in large tanks without refrigeration and it is quite possible that the B₁₂ analogs are synthesized by microorganisms which contaminate this product.

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SUMMARY

A simple microbiological procedure is described for the detection of pseudovitamin B₁₂ and other non-B₁₂ activity in crude samples. The method employs a differential assay using *Lactobacillus leichmannii* 7830 and *Lactobacillus acidophilus* 832 as the test organisms.

Using this method it appeared that lyophilized beef liver, lyophilized beef kidney, fresh chick liver, and fresh chick kidney contained only vitamin B₁₂. Fish solubles, rabbit feces and fresh chick droppings contained appreciable amounts of non-B₁₂ activity.

The differential assay procedure was quantitative when mixtures of pure vitamin B₁₂ and pseudovitamin B₁₂ were assayed, but was probably not quantitative for crude samples due to the presence of other vitamin B₁₂ analogs.

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