

## **Supplementary Information**

### **Lipid nanoparticles with PEG-variant surface modifications mediate genome editing in the mouse retina**

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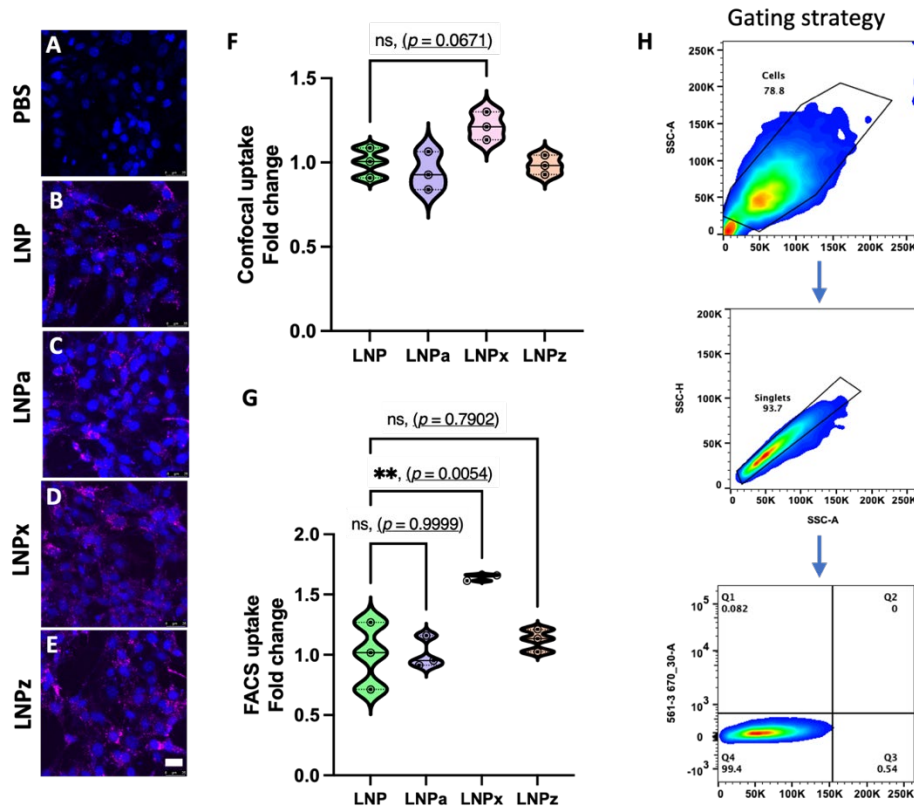
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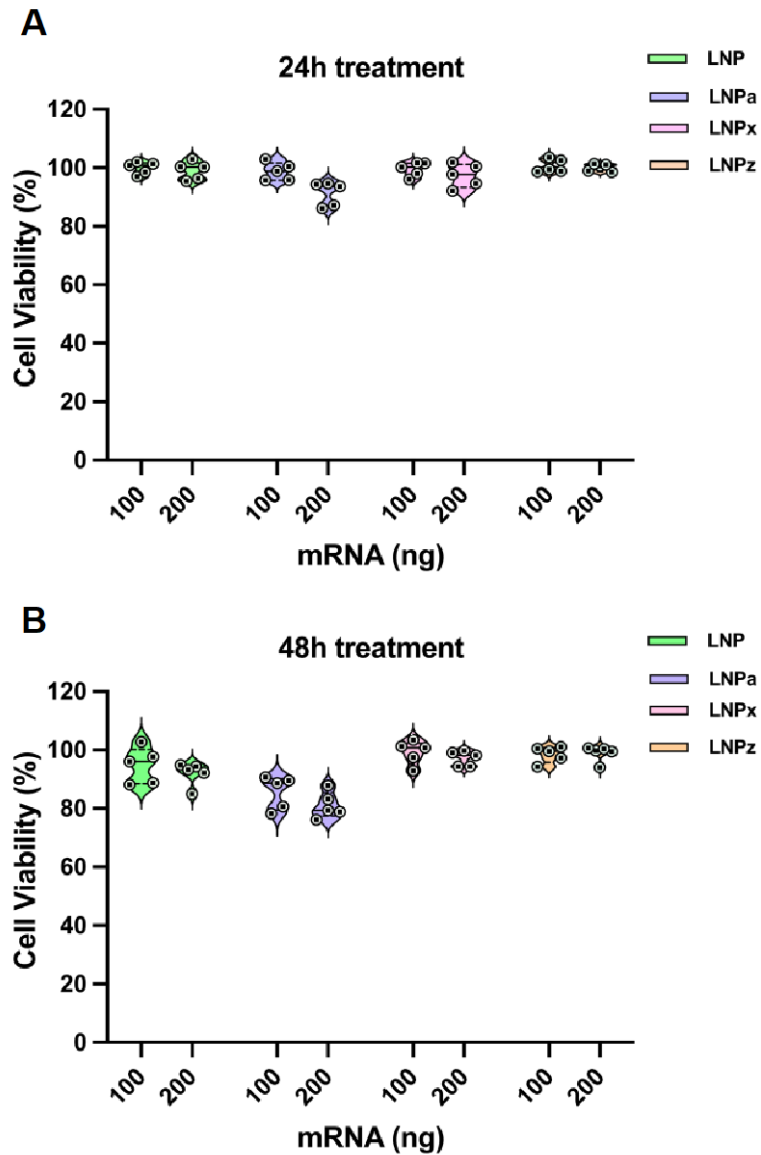
Supplementary Figures 1-7

Supplementary Tables 1-4

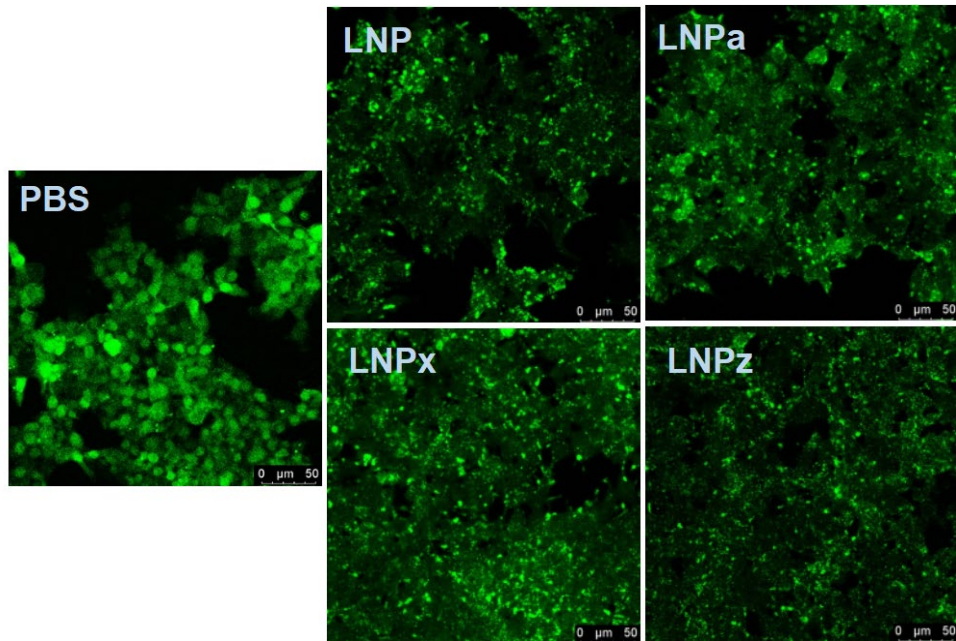


### Supplementary Figure 1. Intracellular uptake of Cy-5 tagged LNP variants. (A-E)

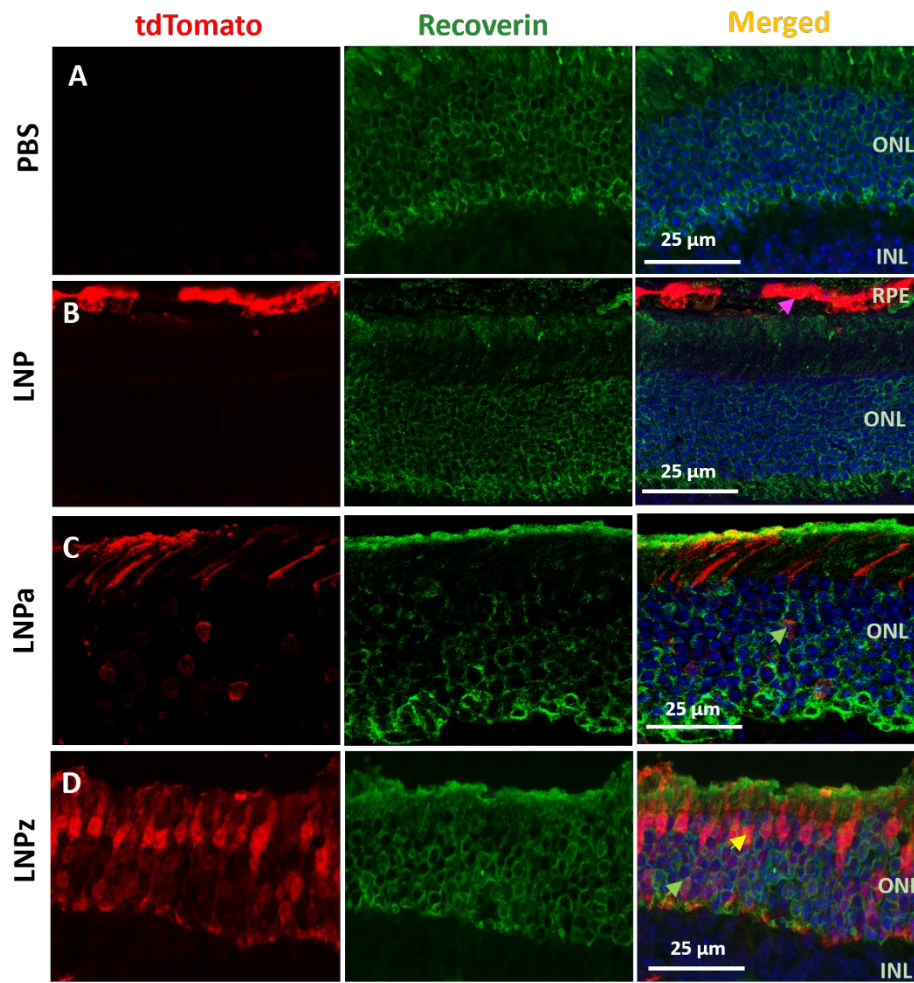
Representative confocal images of 661W cone cells treated with Cy-5 tagged LNP variants at 24 hours treatment. All the cells were treated with Cy5-tagged LNP dose equivalent to 100 ng/well of mRNA cargo (scale bar: 25  $\mu$ m). **(F)** Relative cellular uptake of Cy-5 tagged LNP variants obtained by quantitative FACS estimation of confocal images against unmodified LNP. **(G)** Quantification of cellular uptake using flow cytometry. **(H)** Gating strategy used during cellular uptake study for untreated cells and same strategy was used for other LNP variant treated cells. All data are presented as Mean  $\pm$  SD. An ordinary one-way ANOVA, with Tukey's correction for multiple comparisons test was used for comparisons. Unmodified LNP values were normalized, and fold changes were compared with LNP variants. ns-not significant, \*\* $p < 0.01$ . All representative figures and graphs have  $n = 3$  replicates. Source data are provided as a Source Data file.



**Supplementary Figure 2. *In-vitro* cell viability assay.** 661w cone cells were treated with LNP variants at a 100 and 200 ng fluc mRNA dose. **(A)** 24 hours post-treatment and **(B)** 48 hours post-treatment. All graphs have n=5 replicates. Source data are provided as a Source Data file.

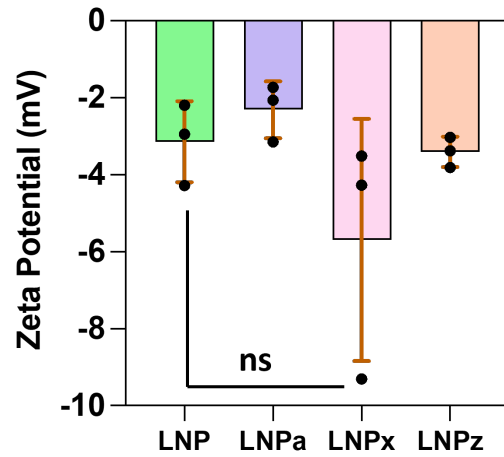


**Supplementary Figure 3. Endosomal escape of LNP variants.** Representative images of the Gal8-GFP reporter cells, 24 hours after *cre* mRNA-loaded LNP variant treatments. All representative figures from n = 3 replicates.



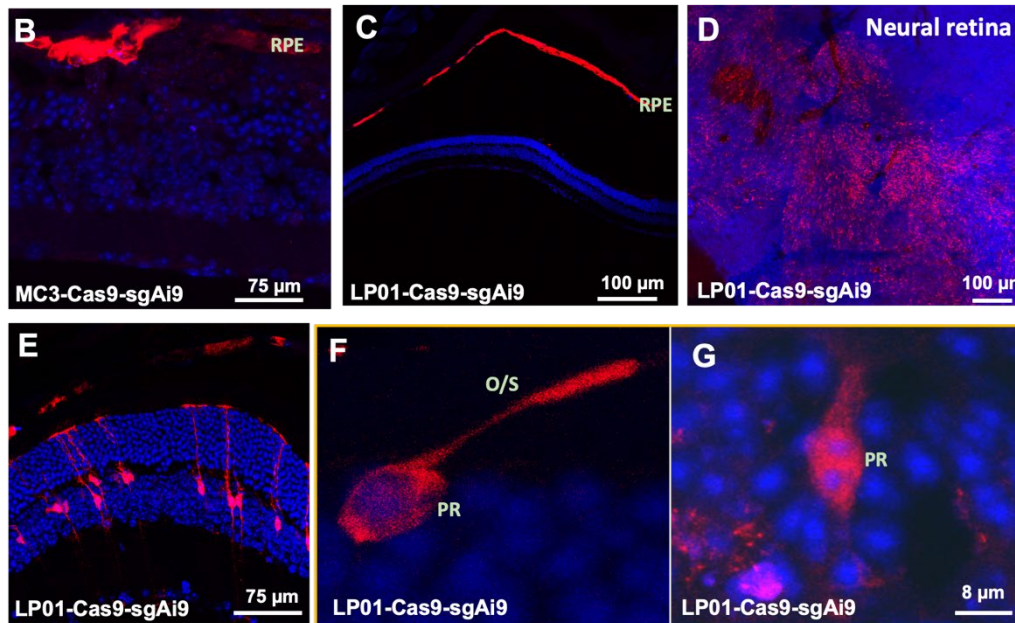
**Supplementary Figure 4. Photoreceptor colocalization study in Ai9 mice. (A-D)**

Representative confocal images showing tdTomato expression in the RPE (LNP) and photoreceptors (LNPa and LNPz), which are colocalized with recoverin. Arrow indicates tdTomato expression (pink: RPE, yellow: cone photoreceptor nuclei, green: rod photoreceptor nuclei). RPE: retinal pigment epithelium, ONL: outer nuclear layer, INL: inner nuclear layer. All representative figures from n = 6 single eyes per group.

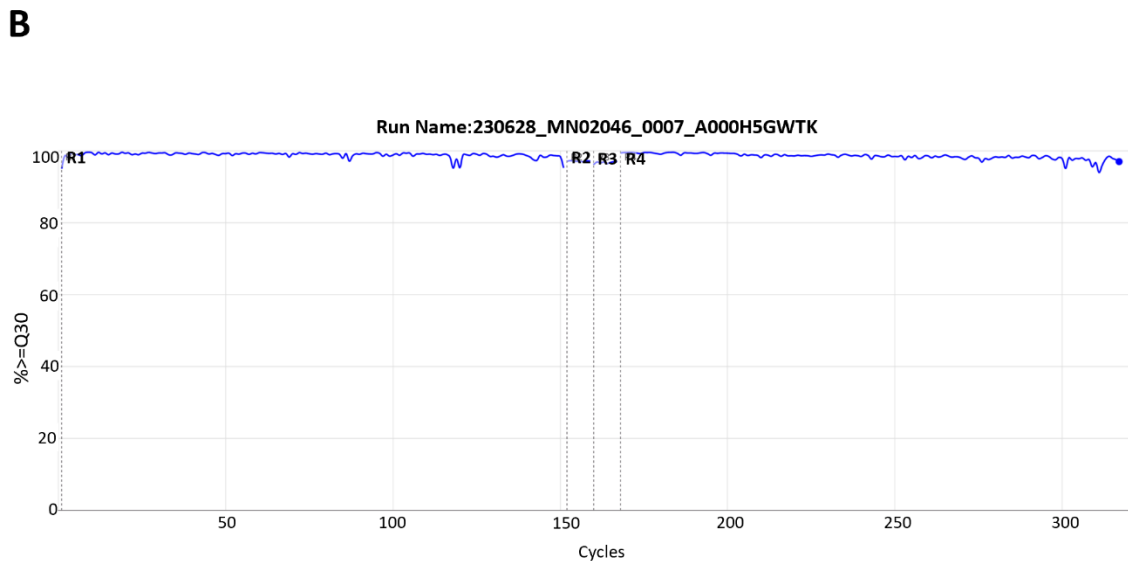
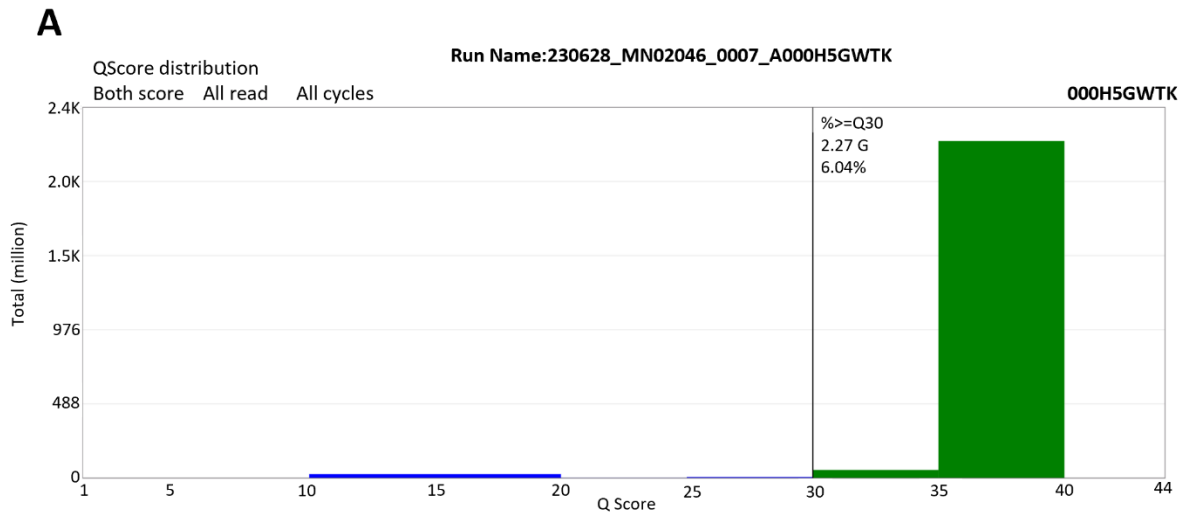


**Supplementary Figure 5.** Zeta potential value from mCherry encapsulated LNP variants measured by Malvern zeta sizer. n=3 LNPs per group. All Data are presented as Mean  $\pm$  SD. An ordinary one-way ANOVA, with Tukey's correction was used for comparison with unmodified LNP. ns-not significant. Source data are provided as a Source Data file.

A) A\*A\*G\*UAAAACCUACUACAAAUGGUUUUAGAGCUAGAAAUAGCAAGUUAA  
 AAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUG  
 CU\*U\*U\*U



**Supplementary Figure 6. CRISPR-Cas9 delivery and sgRNA modification in Ai9 mice.** (A) sgAi9 guide with modifications highlighted. In the sgRNA structure, the asterisks (blue) and red font indicate the phosphorothioate bond and 2'-O-methyl ribonucleotides, respectively. (B) Representative confocal image showing outer nuclear thinning and disruption 7-days post-subretinal injection with MC3-Cas9-sgAi9-LNPx (n=6 single eyes). (C-G) Representative confocal images showing editing events post-LP01-Cas9-sgAi9-LNPx injection. (C) Retinal cross section highlighting RPE editing (n=6 single eyes). (D) Neural retina flatmount showing tdTomato fluorescence signal within the treated area of the retina (n=9 single eyes). (E) Retinal cross sections highlighting editing in the Müller glia and (F-G) photoreceptors (n=6 single eyes). O/S: outer segment, PR: photoreceptor, RPE: retinal pigment epithelium.



**Supplementary Figure 7. Illumina quality control data from NGS. (A)** QScore distribution indicating high quality sequencing data (over 96% Q30 and above). **(B)** Percentage of bases with  $\geq$  Q30 plotted by cycle showing high accuracy of base call across the whole sequencing run (n=9).



**Supplementary Table 1: Antibodies used for staining**

<b>S.N.</b>	<b>Primary Antibody</b>	<b>Host</b>	<b>Dilution</b>
1.	Anti-recoverin (Cat. # AB5585)	Anti-rabbit	1:500
2.	Anti-mCherry (Cat. # NBP2-25157)	Anti-rabbit	1:250
3.	Visual Arrestin (E-3) (SC-166388)	Anti-mouse	1:100

<b>S.N.</b>	<b>Secondary Antibody</b>	<b>Host</b>	<b>Dilution</b>
1.	Alexa Fluor 700-IgG (Cat. # A21038)	Goat anti-rabbit	1:1000
2.	Alexa Fluor 594-IgG (H+L) (Cat. # A-21207)	Donkey anti-rabbit	1:500
3.	Alexa Fluor Plus 488-IgG (H+L) (Cat. # A21038)	Goat anti-mouse	1:200

**Supplementary Table 2: Guide RNA, primers, and cycling conditions used for editing studies**

S.N.	sgRNA Name	Sequence
1.	sgAi9	AAGUAAAACCUACAAAUG
2.	sgGFP	CUCGUGACCACCCUGACCUA

S.N.	Primer Name	Sequence (5' -> 3')
1.	Ai9_NGS_F1_F	ACACTCTTTCCTACACGACGCTCTTCCGATCTATACGAAGTTATTCGCGATG
2.	Ai9_NGS_F2_F	ACACTCTTTCCTACACGACGCTCTTCCGATCTGTTGTGGTTTGTCCAAAC
3.	Ai9_NGS_R	GACTGGAGTTCAGACGTGTGCTCTTCCGATCTTGTTCAGGTTCAGGGGGAG

**Supplementary Table 3: Thermocycling methods for 1<sup>st</sup> PCR for NGS library preparation**

<b>Step</b>	<b>Temperature</b>	<b>Time</b>
Initial Denaturation	95.0 °C	3 min
30 Cycles: Denaturation	98.0 °C	20 sec
Annealing	60.0 °C	20 sec
Extension	72.0 °C	30 sec
Final Extension	72.0 °C	30 sec

**Supplementary Table 4: Thermocycling methods for 2<sup>nd</sup> PCR for NGS library preparation**

<b>Step</b>	<b>Temperature</b>	<b>Time</b>
Initial Denaturation	98.0 °C	2 min
12 Cycles: Denaturation	98.0 °C	10 sec
Annealing	62.0 °C	20 sec
Extension	72.0 °C	30 sec
Final Extension	72.0 °C	2 min