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Appendix Figure S1: DNA content analysis of U2OS cells after drug-treatment

A: Representative histograms of U2OS cells of the indicated genotypes before or after treatment with ZM (2 μ M) or DHCB (4 μ M) for up to 48h. DNA content was analyzed after fixation and propidium iodide staining in a flow cytometer.

B: Quantification of different ploidy stages based on DNA profiles obtained in (A). Bars represent means of two independent experiments.



Appendix Figure S2: Evaluation of centrosome counts in A549-PLK4 and RPE1-PLK4 cells

A: Representative IF images of A549-PLK4 cells treated with DMSO (control) or Doxycycline (1µg/ml) for 72h. Cells were fixed and stained with an antibody specific for γ -Tubulin and DAPI.

B: Evaluation of centriole counts after treatment with either solvent (DMSO), ZM ($2 \mu M$), DHCB (4µM) or Doxycycline (1 µg/ml) for the indicated periods of time. A minimum of 50 cells per time point, condition and experiment were evaluated in a blinded manner. Bars represent means of two experiments.

C: RPE1 cells of the indicated genotypes were treated with ZM (2 μ M) or DHCB (4 μ M) for 72h, or left untreated. A minimum of 50 cells per time point, condition and experiment were evaluated in a blinded manner.Bars represent means of two experiments.



Appendix Figure S3: Assessment of DNA damage and mitotic timing

A: U2OS-PLK4 cells were exposed to DMSO or ZM (2µg/ml) for the indicated times. Etoposide treatment (10µg/ml) for 24h was used as a postive control. Cell extracts were analyzed by western blot analysis using antibodies specific for γ H2AX or GAPDH to control for protein loading.

B: 293T-Rex Flp-In cells were exposed to a single thymidine block and released into fresh medium. Cells were fixed at the indicated times after release and subjected to immunofluorescence analysis using an anti-phosho histone H3 specific antibody to evaluate the percentage of cells in mitosis over time.

C: Westerm blot analysis of A549 RELA/B double mutants and controls confirming lack of RELA and RELB expression.

Appendix Table S1: cloning and CRISR primers

primer name	seqence
PIDD_BamHI_Exon_DD_R	CCCCTCGGACCCTCGGGATCCCGGCAGCTTGATGG
F_hPIDD_G876S	GCAGTCTTGGAGCTCAGCCGCCGCAAGTACC
R_hPIDD_G876S	GGTACTTGCGG GGCTGAGCTCCAGACTGC
F_hPIDD_R815W	CTGGGGGTGTCCTACTGGAGGTGCAGCGCA
R_hPIDD_R815W	TGCGCTGCACCTCCCAGTAGGACACCCCCAG
hPIDD_flag_Q863stop_F	GAGCAGAGTGACCGGGATTACAAGGATGACGATGACAAATAAGTCTTGGAGCTCGG
hPIDD_flag_Q863stop_R	CCGAGCTCCAAGACTTATTTGTCATCGTCATCCTTGTAATCCCGGTCACTCTGCTC
PIDD1_C_fwd1	AACTTAAGCTTGCCACCATGTCCTGGTTCCTTGTGG
PIDD1_CC_fwd1	AACTTAAGCTTGCCACCATGTCCTGGTACTGGCTCTGG
PIDD1_S588A_frw1	CCAGGTCACACACTTCGCCTGGTACTGGC
PIDD1_S588A_rev1	GCCAGTACCAGGCGAAGTGTGTGACCTGG
PIDDC_S446A_F	GGTGCCCCACTTCGCCTGGTTCCTTGTGG
PIDDC_S446A_R	CCACAAGGAACCAGGCGAAGTGGGGCACC
rtTA-pInd fw	CTGGAACTAATCATATGTGG
IRES-pInd rv	GGTGGACCGGTAAGCTTATC
HygR_pInduc21 fw	GCTTACCGGTCCACCATGAAAAAGCCTGAACTCAC
HygR_pInduc21 rv	TCGCCGTCGGGCATGCTATTCCTTTGCCCTCGGAC
HA-PIDD1-N	AACTTAAGCTTGCCACCATGTACCCATACGACGTACCAGATTACGCTGAATTCGGATCCATGGCTGCA

Name	gRNA sequence
PIDD1 1	GCCGATAGCGGATGGTGATG
PIDD1 2	GGCCCGGCGCTGCCGTGAAG
CASP 2	TGGTGAGCAACATATCCTCC
RAIDD	CCAGCTCCAGGCGAAGTGAG
RIPK1	TAGTCTGACGGATAAACACC
NEMO	ACGCCGCCGTCCGAGAGACG
SCLT1	GGGCCTCAGTCATATGTTCC
RELA	ACTACGACCTGAATGCTGTG
RELB	GAAAGCACAGGATCCATC
sg Control 1	TATCTAATCGCGGAGTCGTA
sg Control 2	GCTGGGTGAGTCGATTATCC