nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	A description of all covariates tested
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
	Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection

LI-COR Odyssey Imaging system (model 2802) software, InuCyte 2020C Rev1 software

Data analysis

GraphPad Prism 8.0 & 9.1.1 software, ImageJ FIJI 2.9, Zen 2.6 blue ZEISS software, OriginLab Origin 2021b, XDS, COOT 0.9.8, MAIN, PyMOL 2.5, Chimera 1.14

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio <u>guidelines for submitting code & software</u> for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

PDB: 7Z58, 7QKB, 7Z3U, 7QKA, 7QGW, 8C3D

Research inv	olving hu	man participants, their data, or biological material
Policy information	about studies w	vith

Antibodies

Antibodies used

Plants

Dual use research of concern

TMPRSS2 rabbit antibody (Sigma-Aldrich; ZRB1633), horseradish peroxidase-conjugated anti-rabbit (Cell Signaling; 7074S), HRP-

Antibodies used (κPL - SeraCare; 041806), anti-α-tubulin (Thermo Fisher Scientific; MA1-19401)

Validation

Describe the validation of each primary antibody for the species and application, noting any validation statements on the manufacturer's website, relevant citations, antibody profiles in online databases, or data provided in the manuscript.

Eukaryotic cell lines

Policy information about cell lines and Sex and Gender in Research

Cell line source(s)

VERO-CCL81 cell was initiated from the kidney tissue derived from a normal, adult African green monkey Cercopithecus aethiops, obtained from ATCC (ATCC® CCL-81). LC-HK2 cell line was derived from a tumor induced in nude rat by the inoculation of LC-HK1 cells which was spontaneously established from an explant of a cervical human non-small cell lung cancer metastasis. The cellular and biochemical characterization of LC-HK2 in reported in: Bonaldo, M. de F. et al. Comparative characterization of a human large cell lung carcinoma cell line and the xenograft derived cell line. Cell Biol Int Rep 15, 229–241 (1991); Manelli-Oliveira, R. & Machado-Santelli, G. M. Cytoskeletal and nuclear alterations in human lung tumor cells: a confocal microscope study. Histochem Cell Biol 115, 403–411 (2001); Cortez, B. A., Rezende-Teixeira, P., Redick, S., Doxsey, S. & Machado-Santelli, G. M. Multipolar mitosis and aneuploidy after chrysotile treatment: a consequence of abscission failure and cytokinesis regression. Oncotarget 7, 8979–8992 (2016).

Authentication

VERO-CCL81 was authenticated via ATCC and LC-HK2 was not authenticated

Mycoplasma contamination

VERO-CCL81 and LC-HK2 were regularly analyzed for mycoplasma via PCR reaction and mycoplasma contamination was not detected.

Commonly misidentified lines (See ICLAC register)

. Name any commonly misidentified cell lines used in the study and provide a rationale for their use.

Animals and other research organisms

Policy information about <u>studies involving animals</u>; <u>ARRIVE guidelines</u> recommended for reporting animal research, and <u>Sex and Gender in Research</u>

Laboratory animals Mesocricetus auratus, 6-8 weeks old, male

Wild animals Not applicable

Reporting on sex Only males were used, thus, results are only applicable to male sex.

Field-collected samples Not applicable

Ethics oversight

All procedures were approved by the Committee on Animal Use and Experimentation from the Institute of Biomedical Sciences and the College of Veterinary Medicine, University of São Paulo, Brazil (protocols # 9498230321 and # 8711260321).

Note that full information on the approval of the study protocol must also be provided in the manuscript.