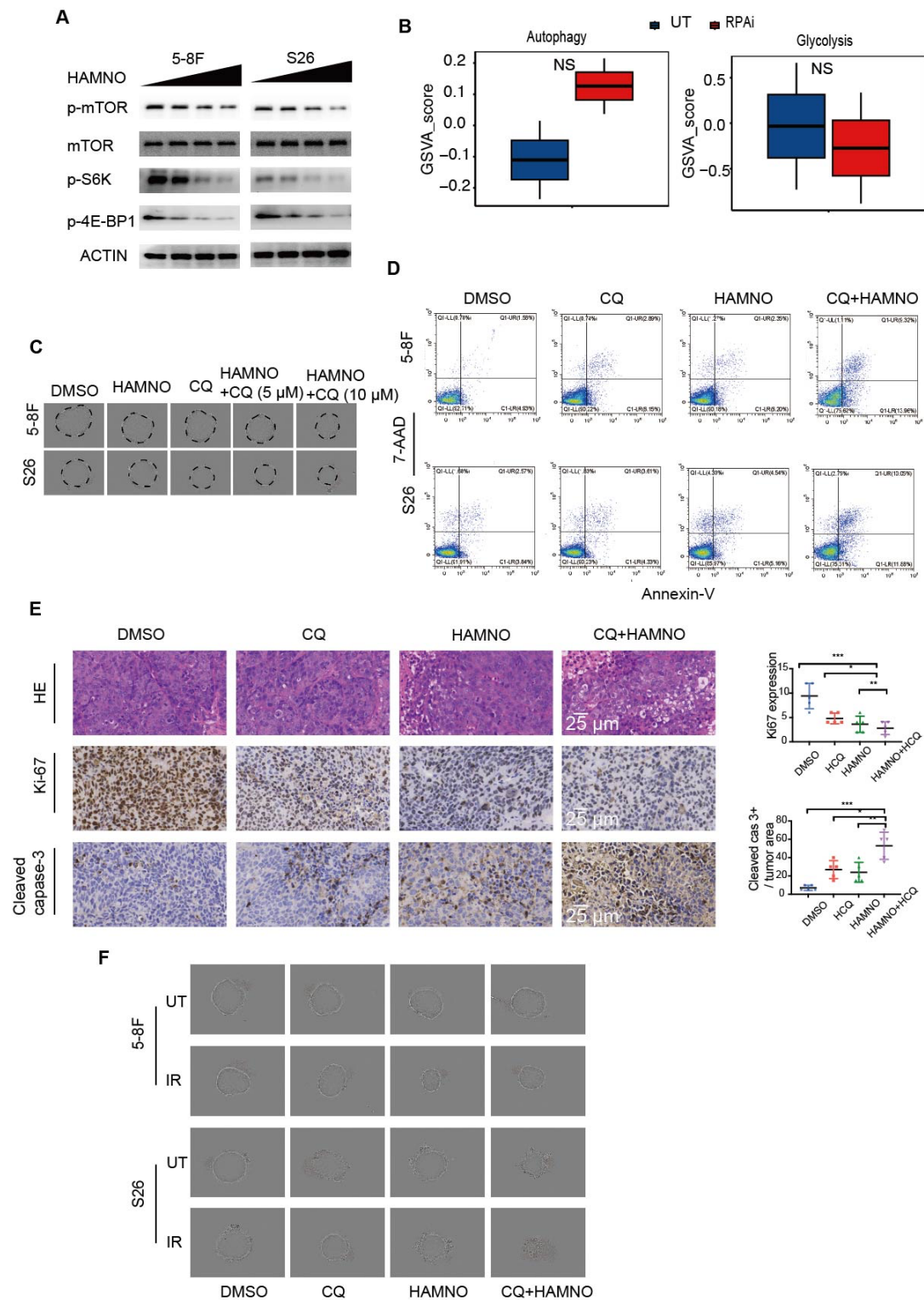


Additional file 1: Fig. S1

a Colony formation analysis of NPC cell lines treated with HAMNO, IR, or their combination. **b** Representative image of the tumor sphere formation assay in NPC cells treated with a single inhibitor or combination with IR after 12 days of cell

culture. Scale bar = 25/50 μm . **c** Percentage of cells in apoptosis induced by RPAi or IR alone or in combination for 48 hours was determined via Annexin V/7-AAD staining and flow cytometry. **d** Representative images of γH2A staining. Scale bar = 50 μm .



Additional file 1: Fig. S2

a 5-8F and S26 cells were treated with DMSO or HAMNO (10 μ M, 20 μ M or 30 μ M) for 48 hours. Immunoblot analyses of cell lysates were performed to determine the levels of pmTOR, total mTOR, pS6K, p-4E-BP1 and ACTIN. **b** 5-8F and S26 were

treated with HAMNO (15 μ M) for 48 h. Cells were collected and subjected to RNA-seq analysis. GSVA was completed to estimate the variation of different metabolic pathway activities in a nonsupervised manner. Each of the two cell lines was treated as a biological replicate. The GSVA scores for a gene set composed of autophagy-related and glycolysis genes were presented. GSVA scores of the two groups were compared using the Mann-Whitney U test. **c** Tumoroids treated with RPAi (5 μ M) or CQ or IR alone or in combination were captured by the IncuCyte system. **d** Percentage of cells in apoptosis induced by RPAi or CQ alone or in combination for 48 hours was determined via flow cytometry and Annexin V/7-AAD staining. **e** Representative images of HE and IHC with indicated antibodies in tumor tissues from S26 models (left). Scale bar = 25 μ m. IHC analysis of Ki67 and cleaved caspase-3 in tumor sections (right). **f** Tumoroids treated with HAMNO (5 μ M), CQ or IR alone or in combination were captured by the IncuCyte system.

Additional file 1: Fig. S3

primers

QP-hGABARAPL1-Forward	TTGTAGAGAAGGCTCCAAAAGCC
QP-hGABARAPL1-Reverse	GGTCTCAGGTGGATTCTCTTCC
QP-hWIP11- Forward	CTTCAAGCTGGAACAGGTCACC
QP-hWIP11- Reverse	CGGAGAAGTTCAAGCGTGCAGT
QP-hSQSTM11- Forward	TGTGTAGCGTCTGCGAGGGAAA
QP-hSQSTM11- Reverse	AGTGTCCGTGTTTCACCTTCCG